Critical Examinations of Neurodegenerative Disorders

1 2 5 EBCCO Publication Condition (BBCCODOL) Inted on 2/10/2023 11:28 PM Condition Condition

Handbook of Research on Critical Examinations of Neurodegenerative Disorders

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A volume in the Advances in Medical Diagnosis, Treatment, and Care (AMDTC) Book Series



Published in the United States of America by IGI Global Medical Information Science Reference (an imprint of IGI Global) 701 E. Chocolate Avenue Hershey PA, USA 17033 Tel: 717-533-8845 Fax: 717-533-8661 E-mail: cust@igi-global.com Web site: http://www.igi-global.com

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Library of Congress Cataloging-in-Publication Data

Names: Sahab Uddin, Md., 1992- editor. | Shah Amran, Md., 1969- editor.
Title: Handbook of research on critical examinations of neurodegenerative disorders / Md. Sahab Uddin and Md. Shah Amran, editors.
Description: Hershey PA : Medical Information Science Reference, [2018] | Includes bibliographical references.
Identifiers: LCCN 2017038751| ISBN 9781522552826 (hardcover) | ISBN 9781522552833 (ebook)
Subjects: | MESH: Neurodegenerative Diseases

Classification: LCC RC522 | NLM WL 358.5 | DDC 616.8/3--dc23 LC record available at https://lccn.loc.gov/2017038751

This book is published in the IGI Global book series Advances in Medical Diagnosis, Treatment, and Care (AMDTC) (ISSN: 2475-6628; eISSN: 2475-6636)

British Cataloguing in Publication Data

A Cataloguing in Publication record for this book is available from the British Library.

All work contributed to this book is new, previously-unpublished material. The views expressed in this book are those of the authors, but not necessarily of the publisher.

For electronic access to this publication, please contact: eresources@igi-global.com.



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ISSN:2475-6628 EISSN:2475-6636

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Section 1 Triggers of Neurodegeneration

Chapter 1

Neurodegeneration is the progressive and gradual dysfunction and loss of axons in the central nervous system. It is the main pathological characteristic of chronic and acute neurodegenerative conditions like Alzheimer's disease (AD), Parkinson's disease (PD), and multiple sclerosis (MS). The usual aspects of pathogenesis of disease can be abridged with regards to the downstream implications of uncontrollable protein oligomerization and aggregation from postmitotic cells. The brain structure constantly changes in normal aging without any dysfunction accompanying the structural changes in brain. The decline in cognitive capabilities, for example, processing speed, memory, and functions related to decision making are the sign of healthy aging. The reduction in brain volume in healthy aging is possibly related to neuronal loss at some marginal extent. The following chapter discusses the structural and functional alterations in the brain in ageing and neurodegeneration.

Chapter 2

Neurons are the building units of the nervous system and are therefore critical units for the health of the brain and the spinal cord. This is necessitated by their inability to be either replaced or reproduced once lost. Their losses are implicated in a number of conditions which have been elaborated in this chapter. Oxidative stress has been strongly implicated in neurodegeneration through blockade of neuroprotection by a number of mechanisms including inhibitory effect on insulin-like growth factor I (IGF-1) via stimulation of the transcription factor, Forkhead box O3 (FOXO3). This chapter elaborates on these two phenomena which cannot be decoupled.

Chapter 3

Free radicals are intricately woven into the fabric of oxidative stress and are significant in the development of neurodegenerative disorders (NDs). This chapter examines free radicals in the context of neurodegeneration and provides overview of the multiple roles they play in the pathophysiology and clinical progression of varying NDs including Pick's disease (PiD), Parkinson's disease (PD), Alzheimer's disease (AD), prion diseases (PrD), traumatic brain injury, and aging. The molecular mechanisms of degeneration in Huntington's disease (HD) are also examined with respect to free radicals. Different antioxidant systems and their mechanisms of action are briefly reviewed in addition to the role of diet in aging. The effectiveness of selected synthetic drugs and natural products used in oxidative stress is also reviewed. Lastly, the chapter examines challenges associated with the use of antioxidants and how promising future directions like the endocannabinoid system is being pursued in the race to effectively manage NDs.

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Mitochondrial Dysfunction in Aging and Neurodegeneration	76
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Mitochondria are a dynamic organelle of the cell involved in the various biological processes. Mitochondria are the site of the adenosine triphosphate (ATP) production, electron transport chain (ETC), oxidation of fatty acids, tricarboxylic acid (TCA), and cellular apoptosis. Besides these, mitochondria are the site of production of reactive oxygen species (ROS), which further disrupts the normal functioning of this organelle also making mitochondria itself as an important target of oxidative stress. Thus, mitochondria serve as an important target in the process of neurodegeneration. In the present chapter, the authors describe mitochondria and its functioning, dynamics, and the mitochondrial dysfunction in aging and neurodegenerative disorders (NDs).

Chapter 5

Neurodegenerative disorders (NDs) are characterized by dysfunction and loss of neurons associated with altered proteins that accumulate in the human brain and peripheral organs. Mitochondrial and Golgi apparatus (GA) dysfunctions are supposed to be responsible for various NDs. Damaged mitochondria do not produce sufficient adenosine triphosphate (ATP) and produce reactive oxygen species (ROS) and pro-apoptotic factors. Mitochondrial dysfunctions may be caused by various factors such as environmental causes, mutations in both nuclear or mitochondrial deoxyribonucleic acid (DNA), that code many mitochondrial components. Three factors that are mainly responsible for the morphological changes in GA are certain pathological conditions, drugs, and over expression of Golgi associated proteins. In this chapter, common aspects of mitochondrial and GA dysfunction concerned about NDs are summarized and described for Alzheimer's disease (AD), Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), and Huntington's disease (HD).

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Neurodegenerative Disorders Progression: From Synaptic Dysfunction to Transmission Failure..... 129 Ramneek Kaur, Jaypee Institute of Information Technology, India Harleen Kaur, Amity Institute of Biotechnology, India Rashi Rajput, Jaypee Institute of Information Technology, India Sachin Kumar, Jaypee Institute of Information Technology, India Manisha Singh, Jaypee Institute of Information Technology, India

Neurodegenerative disorders (NDs) are a diverse group of disorders characterized by selective and progressive loss of neural systems that cause dysfunction of the central nervous system (CNS). The examples of NDs include Alzheimer's disease (AD), amyotrophic lateral sclerosis (ALS), Parkinson's disease (PD), and Huntington's disease (HD). The aggregated proteins block or disrupt the normal proteosomal turnover, autophagy, and become abnormally modified with time, generating toxicity via pathways thereby resulting in neurodegeneration and neuron death. The chapter highlights the understanding in the areas of AD, PD, HD as illustrative of major research so as to define the key factors and events in the improvement of NDs. It defines the physiological functioning of neural transmission (presynaptic, postsynaptic activity) at neural signaling pathway, then the dynamics of neuronal dysfunctioning and its molecular mechanism. Further, it also discusses the progression from synaptic dysfunction to transmission failure followed by NDs.

Chapter 7

The brain relies on a specialized endothelial system, the blood brain barrier (BBB), which is capable of regulating the transfer of substances from the blood to the neurons. Stroke is the most frequent cause of disability in adulthood. Lesions of vascular origin also include asymptomatic small infarcts, microbleeds, dilated perivascular spaces, and atrophy. Vascular cognitive impairment (VCI) is the second most

prevalent cause of dementia. Several mechanisms are implied, including strategic infarct dementia, poststroke dementia, cerebral amyloid angiopathy, and subcortical vascular dementia. As there is no disease modifying therapies currently available, treatment of comorbidities and adequate control of the vascular risk factors remain the standard strategies to reduce the vascular contributions to neurodegeneration. This chapter represents the basic concepts of pathophysiology of cerebrovascular diseases, and describes the subtypes of VCI, as well as treatment and primary prevention strategies.

Chapter 8

Zinc and Neurodegenerative Disorders	
Olakunle Bamikole Afolabi, Afe Babalola University, Nigeria	
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Zinc (Zn) is an essential trace element that is abundantly present in humans. Despite its importance in normal brain functions, alterations in zinc homeostasis cause various neurological pathologies such as dementia, Parkinson's disease, Prion's disease, etc. A growing body of evidence has shown that zinc might play a dual role: in which both zinc depletion and excess zinc cause severe damage and hence neurotoxicity develops. Homeostatic controls are put in place to avoid the accumulation of excess zinc or its deficiency. This cellular zinc homeostasis results from the actions of a coordinated regulation effected by different proteins involved in the uptake, excretion, and intracellular storage or trafficking of zinc. Further investigation has also shown the role of endogenous carnosine (beta-alanyl-L-histidine) in binding excess zinc. Hence, it has the ability to prevent neurotoxicity. Also, the role of a zinc-rich diet cannot be overemphasized. The authors of the chapter, however, provide an insight into the link between zinc homeostasis and neurodegenerative disorders (NDs).

Section 2 Imperious Neurodegenerative Disorders

Chapter 9

Neurodegenerative diseases (NDs) are characterized by specific dysfunction and damage of neurons related to pathologically changed proteins that deposit in the patient brain but also in peripheral organs. These proteins can be used for therapy or used as biomarkers. Except for a plethora of alterations revealed for dissimilar neurodegeneration-related proteins, amyloid- β , prion protein, TAR DNA-binding protein 43 (TDP-43, transactive response DNA binding protein 43 kDa), tau and α -synuclein, or fused in sarcoma protein (FUS), molecular classification of NDs depend on the full morphological assessment of protein deposits, their spreading in the brain, and their correspondence to clinical signs with specific genetic modifications. The current chapter represents the etiology of neurodegeneration, classification of NDs, concentrating on the maximum applicable biochemical and anatomical characteristics and most imperative NDs.

Chapter 10

Tau Pathology: A Step Towards Understanding Neurodegenerative Disorders Network	
Complexity	
Ankush Bansal, Jaypee University of Information Technology, India	

Mehul Salaria, Jaypee University of Information Technology, India Tiratha Raj Singh, Jaypee University of Information Technology, India

A number of neurodegenerative disorders (NDs) are usually referred as tauopathies and characterized by the disappearance or disintegration of tau protein from microtubules. Alzheimer's disease (AD), Pick's disease (PiD), Parkinson's disease (PD) are directly or indirectly associated with tauopathy. Tau is a protein which is usually associated with microtubule. Microtubules are the backbone of neurons, and tau provides a support to microtubule stability. Hyperphosphorylation of tau leads to its separation from microtubule, consequently forming neurofibrillary tangles and resulting in a condition of dementia. Therapeutic implication on tauopathy is symptomatic as there is no exact regulation mechanism known till date. This chapter helps in the comprehensive study of biomarkers and pathways involved in tauopathy to decipher the complexity of the system, resulting in candidate drug target for the management of NDs.

Chapter 11

Amyloid Beta: The Foremost Protagonist in Alzheimer's Disease	
Abhinav Anand, Lovely Professional University, India	
Neha Sharma, Lovely Professional University, India	
Monica Gulati, Lovely Professional University, India	
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Alzheimer's disease (AD), exhibiting accumulation of amyloid beta (A β) peptide as a foremost protagonist, is one of the top five causes of deaths. It is a neurodegenerative disorder (ND) that causes a progressive decline in memory and cognitive abilities. It is characterized by deposition of A β plaques and neurofibrillary tangles (NFTs) in the neurons, which in turn causes a decline in the brain acetylcholine levels. A β hypothesis is the most accepted hypothesis pertaining to the pathogenesis of AD. Amyloid Precursor Protein (APP) is constitutively present in brain and it is cleaved by three proteolytic enzymes (i.e., alpha, beta, and gamma secretases). Beta and gamma secretases cleave APP to form A β . Ubiquitin Proteasome System (UPS) is involved in the clearing of A β plaques. AD also involves impairment in UPS. The novel disease-modifying approaches involve inhibition of beta and gamma secretases. A number of clinical trials are going on worldwide with moieties targeting beta and gamma secretases. This chapter deals with an overview of APP and its enzymatic cleavage leading to AD.

Chapter 12

Parkinson's Disease: A Progressive Disorder of the Nervous System That Affects Movement....... 252 Vaibhav Walia, Maharshi Dayanand University, India Ashish Gakkhar, Maharshi Dayanand University, India Munish Garg, Maharshi Dayanand University, India

Parkinson's disease (PD) is a neurodegenerative disorder in which a progressive loss of the dopaminergic neurons occurs. The loss of the neurons is most prominent in the substantia nigra region of the brain. The prevalence of PD is much greater among the older patients suggesting the risk of PD increases with the increase of age. The exact cause of the neurodegeneration in PD is not known. In this chapter, the authors introduce PD, demonstrate its history, pathogenesis, neurobiology, sign and symptoms, diagnosis, and pharmacotherapy.

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Alpha-synuclein is a protein that forms a major component of abnormal neuronal aggregates known as Lewy bodies. A particular group of neurodegenerative disorders (NDs) is characterized by the abnormal accumulation of α -synuclein; termed the α -synucleinopathies, this group includes Parkinson's disease (PD), dementia with Lewy bodies (DLB), and multiple system atrophy (MSA). Lysosomal storage diseases have also been linked to α -synuclein toxicity. Several therapeutic targets have been chosen among steps of metabolism of α -synuclein. Reducing α -synuclein synthesis or expression and increasing the clearance can be achieved in many ways. The development of immunotherapeutic approaches targeting α -synuclein has received considerable attention in recent years. The aim of this chapter is to present the α -synucleinopathies, as well as to present the most recent researches about treatment of synucleinopathies based on knowledge of the pathophysiology of α -synuclein pathways.

Chapter 14

Fritz Heinrich Lewy described the intracytoplasmic inclusions found in the neurons for the very first time. In 1919 these inclusions were termed as "LBs" by Tretiakoff. LBs were found in the brain of the patients suffering from Lewy body disease (LBD). LBD is characterized by the presence of Parkinsonian symptoms in the earlier stages and dementia in the later stages of the disease. LBs were classified on the basis of the region of the brain in which they are distributed and so is the case of the LBD means the type of the LBD depends on the anatomical areas of the brain involved. LBD is not a single disorder. It is a spectrum of disorders. This chapter addresses the entire profile of LBs, types, composition, formation, and various LB pathologies as well as diagnostic criteria and pharmacotherapy.

Chapter 15

Amyotrophic Lateral Sclerosis: A Predominant Form of Degenerative Disease of the Motor
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 Oduro Kofi Yeboah, Kwame Nkrumah University of Science and Technology, Ghana
 Prince Amankwah Baffour Minkah, Kwame Nkrumah University of Science and Technology,
 Ghana

Amyotrophic lateral sclerosis (ALS) is a progressive neurodegenerative disorder (ND) that primarily comprises the neurons responsible for controlling voluntary muscle movement. The unique neuropathologic findings include anterior horn cell degeneration producing muscle atrophy or amyotrophy, degeneration, and sclerosis of the corticospinal tracts. It is a common neuromuscular disease worldwide and has been

identified in people of all races. There seems to be neither identified risk factors nor family history associated with most of the documented ALS cases. There exists no treatment for ALS that can prevent neither its progression nor reverse its development. However, there are treatments available that can help control symptoms, prevent unnecessary complications, and make living with the disease easier. This chapter extensively discusses this neurodegenerative disorder based on the currently available knowledge on the condition.

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Kirti Rani, Amity University Noida, India	

Autism spectrum disorder (ASD) is neurodevelopmental disorder which is characterized by lack of social behaviors and impaired non-verbal interactions that start early in childhood. It can also lead to progressive neurodegeneration like schizophrenia disorder, Alzheimer's disease, Parkinson's disease, and dementia. Genetic studies of ASD have confirmed the mutations that interfere with neurodevelopment in mother's womb through childhood and these mutations are further involved in synaptogenesis and axon motility. Crucial role of amygdala is found to be deficit in ASD individuals whose association cognition with nucleus accumbens lead to impaired social behaviors and cognitive stimulus. Educational and behavioral treatments are considered the key steps used for its management along with pharmacological and interventional therapies. In this chapter, the author presents the etiology of ASD, proof of neurodegeneration in ASD, as well as the clinical feature and the management of ASD.

Chapter 17

Creutzfeldt-Jakob disease (CJD) is a rare disease associated with neurodegeneration mostly characterized by damage to the neurons. CJD is caused by aggregation of misfolded proteins known as prions; thus, CJD is said to be a prion-related illness. CJD and other prion-related illnesses such as Kuru and Gerstmann-Sträussler-Scheinker disease (GSS) have been reported to have complex mechanisms due to their association with the brain and the nervous system in general. A lot of questions have been raised about the mechanism, diagnosis, and pathogenesis of this disease. The complexity of prion proteins themselves have contributed to more questions about the complications of CJD, whether misfolding of the prions are responsible for neurodegeneration or the misfolding are mere symptoms of the disease. This chapter attempts to explore some details about CJD and answers most related questions about the disease's mechanism. The author finally attempts to explore recent development in pathogenesis, diagnosis, and treatment of CJD.

Chapter 18

Neurodegenerative disorders (NDs) are a group of disorders with the deterioration of attained skills often with no solutions, usually ending with death or crippling disabilities. This chapter contains a classification for childhood NDs as well as an algorithmic approach for easy management of these disorders. Genetic defects and pathophysiology of disorders like Canavan, Krabbe, subacute sclerosing panencephalitis (SSPE), etc. are written in detail. Suggestions regarding management of some of these conditions are

described as a lifecycle approach, from birth to death, to enable those who are taking care of such kids. Allogeneic hematopoietic stem cell transplantation (HSCT), gene therapy, combination therapy, and other experimental therapies have enlarged the scope of diagnosis and treatment options for these disorders. The author aims to brush up the existing and latest possibilities in NDs, including those at the experimental stage, for an easy understanding and for further research, especially as treatment options.

Section 3 Therapeutic Interventions for Neurodegenerative Disorders

Chapter 19

Functional neurosurgery consists of procedures that either promotes judicious destruction or chronic stimulation of the nervous system in order to treat disordered behavior or aberrant function, as it is expected in neurodegenerative disorders ([NDs], e.g., movement disorders [Parkinson's disease, Tourette's syndrome, essential tremor, ballism, and dystonia]). Over the past 20 years, approximately 100,000 deep brain stimulation implant procedures have been performed worldwide. Neurosurgery is also a well-established therapeutic option for people with epilepsy whose seizures are not controlled by anti-epilepsy drugs. The most common pathological finding in patients with drug-resistant mesial temporal lobe epilepsy is hippocampal sclerosis. The aim of this chapter is to present the main NDs that can be treated through surgical procedures, and to describe the surgeries with a focus on the pathophysiology of diseases.

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Preface

Neurodegenerative disorders have become one of the most exciting and glowing researched areas of neuroscience. The Handbook of Research on Critical Examinations of Neurodegenerative Disorders accentuate on the diverse genres of diseases, comprising Alzheimer's disease, Parkinson's disease, Huntington's disease, Lewy body disease, amyotrophic lateral sclerosis, autism spectrum disorder, Creutzfeldt-Jakob disease, and multiple system atrophy. This book introduces various factors such as aging, oxidative stress, pathogenic protein, dysfunction of mitochondria and Golgi apparatus, alpha-synuclein, stroke, and zinc that serve as protagonist to neurodegeneration. Furthermore, this book provides a diaphanous explanation of various neurodegenerative disorders based on existing studies to clarify etiology, pathological mechanisms, diagnosis, therapeutic interventions, as well as current status and future opportunities and challenges. This book also focuses on numerous childhood neurodegenerative disorders and neurosurgical treatments.

This book represents the copious set of specific research updates. All over the world numerous erudite, experienced and eminent academicians, researchers and scientists had participated to write the texts of this book to give a concise thorough understanding of neurodegenerative disorders at a more advanced level with excellent presentation.

This book is suitable for professionals, academicians, students, researchers, scientists and industrialists around the world. Biomedical, health, and life science departments can use this book as a crucial textbook. Researchers and scientist from research institutes can use this book as efficient research info. Pharmacists, physicians, and other healthcare professionals can use this book as a complete reference book. Furthermore, for interested readers, this book is a storehouse of knowledge to comprehend neurodegenerative disorders complexity. The organizations of this book provided a profound knowledge and also maintain the reader's interest.

This book contains 19 chapters divided into 3 sections. The contents of the book cover the peril factors that are likely to the development and propagation of neurodegenerative disorders, and concise thorough understanding of these disorders as well as existing and forthcoming prevention and treatment strategy of neurodegenerative disorders.

Section 1 represents the spectrum of ample factors that can initiate and propagate chronic neuronal dysfunction that leads to neurodegenerative disorders. This section consists of eight chapters. A description of each chapter follows.

Chapter 1 is "The Aging Brain: From Physiology to Neurodegeneration". This chapter explores the physiological functioning of the brain. Further, it discusses the process, mechanism and various factors that cause aging. The pathway and mechanism of neurodegeneration are also introduced. Lastly, it highlights the functional and structural changes in the brain in aging and neurodegeneration.

Preface

Chapter 2 is "Oxidative Stress and Neurodegeneration". This chapter clarifies the precarious link between neurodegeneration and oxidative stress and how this could inform future research on the neurodegenerative disorders.

Chapter 3 is "Free Radicals in Oxidative Stress, Aging and Neurodegenerative Disorders". This chapter reviews the causes of susceptibility of the central nervous system to oxidative damage and the contribution of the aging in neurodegenerative disorders. Additionally, the role of free radicals in some neurodegenerative disorders as well as novel targets which are being explored to effectively manage these neurodegenerative disorders is presented.

Chapter 4 is "Mitochondrial Dysfunction in Aging and Neurodegeneration". This chapter explains the alteration in the mitochondrial dysfunction by aging and how this alteration is linked with the neuronal degeneration.

Chapter 5 is "Alternation of Mitochondrial and Golgi Apparatus in Neurodegenerative Disorders". This chapter focuses on that how mitochondria and Golgi apparatus get affected in neurodegenerative disorders. The target is to explain the factors that are mainly responsible for the morphological changes in these organelles.

Chapter 6 is "Neurodegenerative Disorders Progression: From Synaptic Dysfunction to Transmission Failure". This chapter describes the dynamics of neuronal dysfunction, the models that are used to study neurodegenerative disorders and the factors that cause the dysfunctioning. Further, it also highlights the progression of the pathological basis from synaptic deregulation to synaptic transmission deficits.

Chapter 7 is "Stroke: A Potential Risk Factor of Neurodegenerative Disorders". This chapter represents the relationship between stroke and neurodegenerative disorders, predominantly those related to cognitive impairment. Moreover, describes the pathophysiological mechanisms of vascular action on neurodegeneration reaching prevention and treatment suggestions.

Chapter 8 is "Zinc and Neurodegenerative Disorders". This chapter relates the significant role of zinc in relation to several biochemical and pathophysiologic activities in humans and its relation to neurological disarranges. The basic impact of zinc on homeostasis, oxidative stress, aging, and neuro-degeneration is also stated.

Section 2 focuses on the clear explanation of the utmost imperative neurodegenerative disorders with a momentous impact on etiology, pathological mechanisms, disease management, current status, future opportunities, and challenges. This section consists of ten chapters. A description of each chapter follows.

Chapter 9 is "Neurodegenerative Disorders: An Introduction". This chapter portrays the causes and concepts of the classification of neurodegenerative disorders. Moreover, this chapter deals a concise depiction of the neuropathological features of numerous neurodegenerative disorders.

Chapter 10 is "Tau Pathology: A Step Towards Understanding Neurodegenerative Disorders Network Complexity". This chapter discusses various types of neurodegenerative disorders to understand the underlying mechanism of tauopathy. Additionally, presented a comprehensive study of biomarkers and pathways involved in pathological aggregation of tau protein in the human brain.

Chapter 11 is "Amyloid Beta: The Foremost Protagonist in Alzheimer's Disease". This chapter focuses on the amyloid beta hypothesis and the biomolecular targets, including the proteins and enzymes that contribute towards the development of Alzheimer's disease. A brief insight into the currently existing pipeline of potential drug candidates involving these targets has also been provided.

Chapter 12 is "Parkinson's Disease: A Progressive Disorder of the Nervous System that Affects Movement". This chapter provides a detailed overview of the Parkinson's disease from pathogenesis to therapeutics with existing challenges and the future directions.

Chapter 13 is "Alpha-Synucleinopathies: Parkinson's Disease, Dementia with Lewy Bodies, and Multiple System Atrophy". This chapter presents the alpha-synucleinopathies and the most recent researches about the treatment of alpha-synucleinopathies based on knowledge of the pathophysiology of alpha-synuclein pathways.

Chapter 14 is "Lewy Body Disease: Point Towards Progressive Dementia". This chapter represents the different pathologies, characterized by the presence of Lewy bodies in the different regions of the brain, which further form the basis of the progression of the Lewy pathology.

Chapter 15 is "Amyotrophic Lateral Sclerosis: A Predominant Form of Degenerative Disease of the Motor Neuron System". This chapter delves the mechanisms for motor neuron degeneration in amyotrophic lateral sclerosis as informed by up to date knowledge on this neurodegenerative disorder. In addition, diagnosis, pharmacotherapy, and forthcoming research tips are offered.

Chapter 16 is "Spectrum of Neurodegeneration in Autism Spectrum Disorder". This chapter expresses the effect of multi-factors such as oxidative stress, neuronal loss, microglia activation and proinflammatory cytokines that further lead to progressive neurodegeneration in autism spectrum disorder patients despite of showing its regression in neurodevelopmental practices.

Chapter 17 is "Creutzfeldt-Jakob Disease: A Prion Related Neurodegenerative Disorder". This chapter attempts to answer based on recent studies on the mechanisms and pathogenesis of Creutzfeldt-Jakob disease as well other details including epidemiology, diagnosis, and treatment of the disease. The chapter also highlights current and future perspectives in various studies aimed at finding the cure for the disease.

Chapter 18 is "Childhood Neurodegenerative Disorders". This chapter focuses on almost all details of neurodegenerative disorders of the children including epidemiology, pathophysiology, and management of these disorders. Moreover, the chapter gives an insight into the recent advances, experimental trials, and possible treatment and investigational options which may come up in the near future.

Section 3 offers the neurosurgical treatments strategy for the neurodegenerative disorders consistent with current studies, recent futuristic technologies, and techniques. This section consists of only one chapter. A description of this chapter follows.

Chapter 19 is "Neurosurgical Treatments of Neurodegenerative Disorders". This chapter presents the main neurodegenerative disorders that can be treated through surgical procedures, and to describe the surgeries with a focus on the pathophysiology of the disorders.

It is expected that readers shall find this book very informative and enormously useful. Since science is constantly changing readers are strongly recommended to check the recent update. The editors are ebulliently ready to accept any comment, suggestion, advice or critique.

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Acknowledgment

Editing is a complex process, the editors would like to thank the people involved in this project for their quality time, expertise, and countless efforts. The editors are extremely indebted to the following:

The authors for the countless time and expertise that they have put into their texts. Those authors who submitted works to contribute to this project but unfortunately rejected. The reviewers for their constructive reviews in improving the quality and presentation of the contents. Furthermore, the editorial advisory board for their prodigious supports and suggestions. The teachers, colleagues, friends, and supporters for their endless inspiration and assists. Especially, IGI Global for giving us a great opportunity to edit this book and their aid in the preparation of this edition.

Last but not least, the editors would like to express the heartiest gratitude to almighty Allah and their parents.

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Section 1 Triggers of Neurodegeneration

Chapter 1 **The Aging Brain:** From Physiology to Neurodegeneration

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ABSTRACT

Neurodegeneration is the progressive and gradual dysfunction and loss of axons in the central nervous system. It is the main pathological characteristic of chronic and acute neurodegenerative conditions like Alzheimer's disease (AD), Parkinson's disease (PD), and multiple sclerosis (MS). The usual aspects of pathogenesis of disease can be abridged with regards to the downstream implications of uncontrollable protein oligomerization and aggregation from postmitotic cells. The brain structure constantly changes in normal aging without any dysfunction accompanying the structural changes in brain. The decline in cognitive capabilities, for example, processing speed, memory, and functions related to decision making are the sign of healthy aging. The reduction in brain volume in healthy aging is possibly related to neuronal loss at some marginal extent. The following chapter discusses the structural and functional alterations in the brain in ageing and neurodegeneration.

DOI: 10.4018/978-1-5225-5282-6.ch001

INTRODUCTION

Aging of the brain is described through various anatomical, molecular and functional changes directing to enhance susceptibility to numerous diseases (Glorioso, Oh, Douillard, & Sibille, 2011). During aging, structural brain changes include volumetric shrinkage of brain and change in specific brain morphology in some region (Peters, 2006). Apart from the damage in structural integrity and neural plasticity, progressive declining in reserves of cellular homeostatic and variation in the mechanism of calcium dependent signaling has been recommended as the significant events associated in brain aging (Cai & Tammineni, 2017). Brain aging is also associated with some chemical and molecular alteration such as changes in hormones and neurotransmitter levels, accelerated formation of reactive oxygen species (ROS), dysfunctioning of mitochondria, accretion of nuclear and mitochondrial DNA damage convoyed by age related failure in DNA repairing and aggregation of intracellular and extracellular proteins (Wallace, 2010). Such severe changes in cellular and molecular levels cause difficulties in regular activities like attention, sleep, language, speech, decision making and cognitive abilities like work memory and long term memory (Alhola & Polo-Kantola, 2007). Higher-order brain system breakdown in aging is associated partially to myelinated fibres disruption which connects neurons to various cortical regions. Though the minimal neuronal loss in most of the cortical region is a part of normal brain aging, variation in synaptic physiology of neurons aging may participate in alteration of connectivity and higher order integration (Bishop, Lu, & Yankner, 2010). Significantly, these changes reduced down the coordination of brain activity which leads to weak performance in numerous cognitive domains.

Neurodegeneration is continuous loss of structural and functional properties of neurons corresponds to some pathological conditions which including neurons death (Gorman, 2008). In manner, neurodegenerative disorders (NDs) signify a huge assembly of neurological disorders with varied pathological and clinical expressions disturbing particular neuronal sets in detailed functional anatomic systems that progress in a persistent manner (Lin & Beal, 2006). Different NDs like PD, AD, Huntington disease (HD) and amyotrophic lateral sclerosis (ALS) are common in therapeutic research studies. NDs are mostly characterized through the factors like genetic risk factors, certain age ranges, courses of progression, clinical symptoms, dysfunctioning and neuronal death particular biochemical abnormalities, and presence of extracellular and intracellular protein (Kovacs, 2016). In the advance study, the beginning of NDs are instigated by the protein aggregation which are called proteinopathies (Shelkovnikova, Kulikova, Tsvetkov, Peters, Bachurin, Bukhman, et al., 2012). In this case, the aggregation of soluble monomers or oligomers cause toxic conditions subjected to consideration like some slight variations in the synthesis of α -synuclein and tau protein cause the occurrence of certain stressful condition which lead to PD and frontotemporal dementia respectively (Wang & Roberts, 2010). For these proteins, it is common that the holoprotein comparatively or entirely benign and with a series of cleavage of these protein lead to the production of toxic species or fragments. Likely, the rate of full length protein production is not a significant issue with respect to the rate of cleavage level of that protein. The example of ongoing studies on this fact is the cleavage of full-length amyloid precursor protein (APP) resulted to the production of toxic product called amyloid β (A β) peptide which get aggregated onto the neurons and cause AD (O'Brien & Wong, 2011). Similarly, this toxic-fragment model appears to be appropriate to some polyglutamine diseases like spinocerebellar ataxia type 3 and HD (Shao & Diamond, 2007). This notion also proves to be helpful in therapeutically tracing of NDs. The chapter highlights the physiological functioning of brain, structural and functional brain changes in ageing and neurodegeneration. The chapter also focusses on the factors which are responsible for aging and neurodegeneration.

BACKGROUND

Relatively, neurons shrinkage, synaptic spines reduction and decrease in amount of synapses expected to justify the reductions in gray issue (Zhou, Homma, & Poo, 2004). Additionally, the length of axons covered with myelin sheath is significantly reduced, nearly 50%. In neurodegeneration, major signs of alteration are structural, however far not functional and consistently found to be associated with clinical phenotype and disease severity which further inspected for its connectivity correlation of molecular pathology (Alexander, 2004) (Figure 1). Gradual change in neural cells cannot trigger these variations instead differences in the action of neural networks and, possibly, chronic intoxication by irregular proteins that can be overcome by the brain and help in the concept of therapeutic implications. ND, for example HD and PD, are linked with the intracellular aggregation of toxic proteins and some are produced by aggregate prone proteins (Gorman, 2008). These diseases are called proteinopathies that depends on the conditions wherein the proteins are predominantly cytosolic in HD and PD, mostly intranuclear (like spinocerebellar ataxia type 1), aggregation of endoplasmic reticulum (like mutations in neuroserpin causes familial encephalopathy with neuroserpin inclusion bodies) and extracellular secretion (for example amyloid- β aggregation cause AD) (Shelkovnikova, Kulikova, Tsvetkov, Peters, Bachurin, Buchman, et al., 2012). Study of transgenic and genetic data indicate that numerous mutations accountable for proteinopathies cause illness by conferring a toxicity amplification of function on the relevant proteins (Sweeney et al., 2017). For example, some minor mutations in α -synuclein and tau protein can be the major cause for types of PD and frontotemporal dementia respectively that promptly aggregate as compared to wild type proteins (Recasens & Dehay, 2014; Schulz-Schaeffer, 2010). Similarly, in case of HD, mutation in polyglutamine expansion cause increased tendency of their aggregation with polyglutamine tract length (Daldin et al., 2017). Consequently, there is a need to understand their levels of regulation in cellular processes, rates of synthesis and degradation.

PHYSIOLOGICAL FUNCTIONING OF BRAIN

Brain is the most integral and perhaps highly intricate organ of the human body. It is the powerhouse which controls all physiological and psychological functioning of the body. There are the three major parts of the brain called forebrain, midbrain and hindbrain which are further classified into smaller division with their specific functionality. Forebrain is accountable for processing of sensory information gathered from five sensory organs i.e. skin, nose, eyes, tongue and ears (Posner & Rothbart, 2007).

Forebrain is further divided into diencephalon and telencephalon. Diencephalon included thalamus and hypothalamus region which coordinately manage the sensory and autonomic processes whereas telencephalon comprise of the biggest part of the brain known as cerebrum. The outermost surface of the neuronal tissues in cerebrum is called cerebral cortex. The medial longitudinal fissure of the cortex divides it into two main sections called right cerebral hemisphere and left cerebral hemisphere. The basic functioning of cerebral cortex comprises of motor areas, sensory areas, and association areas. The motor areas positioned in both the hemispheres have its major concern with the regulation of voluntary movement. With the significance of the name, sensory areas are connected with the processing of the data collected from the senses. The association areas are primarily assist the abstract thinking and language and also support in producing expressive perceptual experience of the surrounding world (Bailey, 2017).

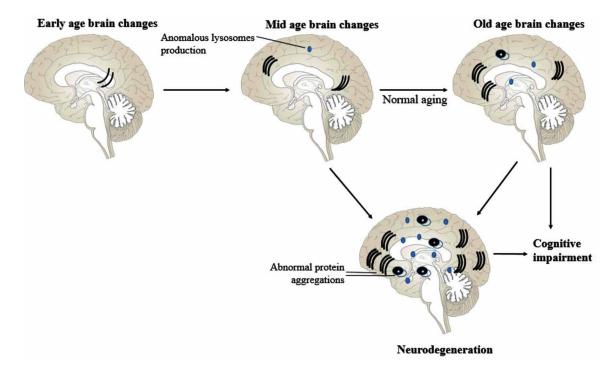


Figure 1. Schematic diagram representing structural brain changes in neurodegeneration

Midbrain is situated between the forebrain and hindbrain like a bridge which transmit the signals form hindbrain to forebrain coming from touch and hearing senses, gathered by sense organs i.e. skin and ears respectively. Optic tectum is the upper region of the midbrain that helps in combining visionary and auditory records (Bailey, 2017).

The main function of hindbrain is to regulate specific visceral functions in human body like heartbeat, blood pressure and breathing. Hindbrain is also divided into three parts namely medulla oblongata, pons and cerebellum. Pons are assigned with task of monitoring the waking and sleep functions in harmonization to the other parts of nervous system whereas the cerebellum synchronizes the association of arms and legs and also manage the processing of information of sensory which is received from visual and auditory systems (Tyler, Danilov, Bach-y-Rita, & Bach-y-Rita, 2007).

NDs in central nervous systems are grouped in diseases on the basis of the region of occurrence like basal ganglia, cerebral cortex, cerebellum and brainstem and further classification is done based on their major characteristics and properties. For example, the disorders that mainly concern the cerebral cortex are segregated to nondementing and dementing conditions. Notably, while AD is probably the most often mentioned source of dementing cortex pathology, dementia can seemingly be detected in no less than 50 various disorders. Furthermore, dementia is witnessed in NDs and various toxic, metabolic, ischemic, traumatic and infectious insults of the brain. However, on the basis of abnormal movement phenomena, the basal ganglia diseases are categorized as hyperkinetic or hypokinetic. Hypokinetic disorders of the basal ganglia are characterized by PD, where the speed and amplitude of actions are reduced and in severe situations, even nonexistent, subject suffocates like a prisoner in its own body (Purves et al., 2001). On the other end, hyperkinetic basal ganglion disorders that are exemplified by essential tremor and HD (Magrinelli et al., 2016). In mentioned circumstances, extreme abnormal movements like tremor or

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chorea are overlaid onto and restrict the normal voluntary movements. Whereas, division of NDs of the cerebellum and its associated networks is mostly demanding due to the noticeable connection amongst the several pathological conditions. Certainly, diseases related to cerebellum are classified into three foremost neuropathological categories: pontocerebellar atrophy (lesion altering some brain structures and cerebellar) (Namavar, Barth, & Baas, 2011), friedreich ataxia (lesion influencing the spinal cord at the posterior column, heart and peripheral nerves) (Bidichandani & Delatycki, 2017) and cerebellar cortical atrophy (injuries constricted to the inferior olives and purkinje cells) (MacKenzie-Graham et al., 2006). There are some diseases of cerebellum which can't be grouped in mentioned classifications like dentatorubral degeneration, where the most pronounced injuries are located in the red nuclei and dentate. Machado Joseph disease, where degeneration includes the upper and lower motor neurons, dentate system and substantia nigra (Paulson et al., 1997).

PROCESS AND MECHANISM OF AGING

With the time passes, brain displays several anatomical, molecular, and functional variations and leads to escalated frequency of neurological disorders and traumatic brain lesions (McKee & Daneshvar, 2015). Aging of brain comprises of depletion of brain structural integrity, changes in the levels of hormones, enzymes, deregulated metabolism, elevated oxidative stress, genetic and epigenetic modulation, varied protein processing and synaptic function and together these alterations directed to disability in physiological and cognitive functions (Chakrabarti et al., 2011). Initially in brain aging, prevalent volumetric brain damage is observed in frontal cortex, temporal cortex, putamen, thalamus, and nucleus accumbens. Also in MRI analysis, it was observed that there is a reduction in grey matter volume whereas the white matter volume increases in frontal, parietal and temporal cortices, insula and superior parietal gyri in both males and females (Terribilli et al., 2011). The reductions in the brain volume cause because of neuronal shrinkage, deduction in synaptic spines, dropped level of synapses and length shortening of myelinated axons. With the aging progression cerebral ventricles enlarge, this phenomenon is called ventriculomegaly and further was studied biomechanically in depth with significant applications to hydrocephalus by Wilkie et al. 2012 (Wilkie et al., 2012). Also, permeability through blood brain barrier (BBB) increases with aging (Rosenberg, 2014). Incorporating neuroanatomical changes during aging with molecular variation comprising alteration in neurochemicals, genetic and epigenetic factors.

Chemical Changes

Aging of brain includes noted variation in the levels of neurotransmitters, hormones, enzymes and other metabolites. In the study conducted by Ota et al. 2006, the level of dopamine reduce down by 10% per decade from initial adulthood as a result of dopaminergic neurons damaging present in between frontal cortex and striatum and due to the reduction in binding affinity and amount of dopamine receptors (Ota et al., 2006). Loss in dopamine level is related with the decline in motor and cognitive performance due to progression of aging. The level of serotonin (Meltzer et al., 1998) and glutamate (McEntee & Crook, 1993) also goes down with aging which is responsible for synaptic plasticity loss in old brain. Activities of neurotransmitters regulated by enzymes such as monoamine oxidase enhance with aging and cause the release of free radicals from reactions which reserve the inherent antioxidants (Kumar et al., 2012).

Other important elements that affect the brain aging and cognitive performance are hormones such as growth hormones, melatonin, thyroxine and sex hormones involving testosterone, estrogen, progesterone and DHEA (dehydroepiandrosterone). During brain aging, these hormones declines and the stress hormones cortisol increases significantly (Sorwell & Urbanski, 2013) which is believed to be a severe risk factor for cardiovascular disorders, instant brain aging and obesity. Functional and structural changes during aging are credited largely to modulate the level of estrogen that primarily acts through its estrogen receptor (ER α and ER β) and intracellular receptors. The expression of those receptors is controlled by various factors comprising of their very own ligand estrogen, and others like thyroid hormone and growth bodily hormone and. The amounts of these factors reduce with aging that signaling resulting in alterations in brain functions (Näntö-Salonen, Muller, Hoffman, Vu, & Rosenfeld, 1993). Recent report which ER 13 interacts with casein kinase2. Phosphokinase N and C - myristoylation, present in mitochondrial and nuclear proteins may be useful to regulate gene regulation in mind for therapeutics (Giorgi, De Stefani, Bononi, Rizzuto, & Pinton, 2009).

Insulin and Insulin-like growth factors (IGFs) have been progressively recorded as molecules that are significant in regulating the speed of aging in humans (Hammerman, 1987). Increased insulin levels and immunity with advancing age influences integrity, brain blood vessel function and energy metabolic process and imposes the chance of developing dementia in late life. The brain aging may suffer from impaired glucose metabolic process or an input of sugar due to collapse of cerebrovascular efficiency. Recent studies show that lactate level in the brain produced by a change from the lactate dehydrogenase (Bergersen, 2015).

Genetic Changes

Expansion in gene expression studies like microarray analysis of genes has helped to comprehend the molecular mechanisms underlying the changes which occur during brain aging. Most of gene expression studies in human brain across the lifetime show changes in molecules associated with anxiety, inflammation, immune response, mitochondrial function, expansion factors, neuronal survival, synaptic plasticity and calcium homeostasis. Evaluation of phylogenetically distant organisms has shown some widely conserved functional classes genes with expression changes which are age-dependent (Gout, Kahn, Duret, & Paramecium Post-Genomics, 2010). While enzymes associated with stress response, inflammation and DNA repair are up regulated. Mitochondrial genes are repressed in process of aging in all the individuals studied. As purposed by Bishop et al. (2010), in analysis of transcriptional profiling of the aging process of cortex in rhesus macaques, humans and mice, the biggest conserved shift was age-dependent increased regulation of apolipoprotein D which functions as a lipid anti-oxidant conferring immunity and is induced in the brain of organisms with AD (Bishop et al., 2010). Different areas of the forebrain exhibit gene profile vary with age. For example, superior frontal gyrus shows a remarkable alteration in expression of gene although changes in the entorhinal cortex are mild. Changes happening in 60-70 years old individuals across the whole cortical region indicate it to be a transition point in aging of brain.

In hippocampus, CA1 (cornu ammonis 1) region is more vulnerable to aging and exhibits a higher number of changed genes relative to dentate gyms (DO) and the CA3 and an increase in expression of genes linked to the apoptosis and immune response (Shah & Zeier, 2013). Male brain is illustrated by global decrease in anabolic and catabolic capacity with aging, and down-regulating genes which are involved in protein transport/synthesis and energy production. Improved immune activity is a leading feature of aging in both genders, with correspondingly greater female brain activation. Besides regional

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and gender differences, gene expression has also been shown to be specific for neurons and glia. It is suggested that glial-enrich genes is largely correlated with immune system and complement activation is up regulated, whereas neuronal enriched genes related to synaptic structure and function, calcium regulation, signal transduction and transmembrane receptors, are largely down-regulated with age. Even among glial genes, astrocyte markers are up -regulated while oligodendrocytic genes are repressed during aging (Loerch et al., 2008).

Epigenetic Changes

Besides genetic alterations, the emerging field of epigenetics indicates that building-up of abnormal epigenetic marks comprising histone and DNA modifications can be a driver of aging-related physiological and cellular alterations. Study of Zeng et al. (2011) reveal that age-reliant shortfalls in long-term loss of hippocampal dendritic spines and synaptic plasticity in the aged Fischer 344 rats are closely related with upregulation of histone deacetylase (HDAC2), reduction in histone acetylation, and decrease in expression of histone acetyltransferase (Zheng, Seabold, Horak, & Petralia, 2011). Further examination discloses brain-derived neurotrophic factor (BDNE) as one of the crucial genes affected by such changes. Age-dependent reductions in 113 and 114 acetylation are noticed in multiple promoter regions of the BUM gene indicating the impairment of downstream signaling in the aged hippocampus and a substantial reduction in BDNE expression. These discrepancies could be salvaged by enhancing the expression of trk13 and 13DNI by inhibition of FIDAC or via directly activating trkB receptors with 7,8-dihydroxyflavone which is a newly recognized, selective agonist for trkB. Epigenetic changes in genes like Arc and zif268 in vulnerable brain regions of hippocampus and prefrontal cortex are also related with age-associated cognitive deficits (Penner, Blair, Albrecht, & Dovidio, 2014). Dynamic alterations in the structure of chromatin across the lifetime could potentially either contribute to enhanced aging and age-related dysfunctions in other cases, or counter aging and age -linked disorders in some cases. Despite the identification of several genes influencing brain aging and overall lifespan, an important question still remains unanswered if age-associated cognitive alterations are facilitated by any of the chief regulators of aging and lifespan identified in model organisms.

NEUROBIOLOGICAL MECHANISMS ASSOCIATED WITH AGE-RELATED COGNITIVE DECLINE IN HUMANS

Memory processes are strongly affected by experiences of life. Among life experiences, stress is proved to modulate memory function (Sandi, 2004). Stress is one of the major factors involved in cognitive decline in humans. Stress is a condition that varies as per the situation. There are circumstances that require moderately acquired adaptations from the individual and there are situations that may be tremendously persistent and adverse. But it is an inevitable entity since life constantly deals with challenges and changes. Since the concept of stress varies from person to person, the impacts of stressful experiences on cognitive functions vary. Stress within limits can facilitate information storage, which is helpful while learning and retaining things but experiencing excessive stress can be highly harmful to the memory function (de Kloet, Oitzl, & Joëls, 1999; Roozendaal, 2002; Sandi, 1998).

Stress

Stress is a biologically substantial factor and has a profound effect on the functioning of the brain. It can amend the properties of brain cell and is also able to interrupt cognitive processes, subsequently limiting human life quality. Increasing stress increases the glucocorticoid hormone in the body (Rajeswaran & Bennett, 2013). The stimuli that exert harmful impacts on individual fall under the category of stressors. The way in which an individual interprets a situation and responds to it plays a crucial role in determining whether a stimulus is a stressor or not. Usually, these responses work in accordance with the previous experiences of individuals (Council, 2001; Hibberd, Yau, & Seckl, 2000; McEwen, 2002). Lifetime exposure to anxiety and the consistent upsurges in glucocorticoid hormones are suggested to be crucial factors leading to the acceleration of cognitive decline in the aging individuals. Therefore, in addition to increasing the extent of cognitive disturbances detected in aged individuals, stress can also increase their appearance. Aging is related with decreased feedback sensitivity of the HPA (hypothalamic-pituitary-adrenal) axis to pharmacological challenges and higher basal cortisol levels and (Wilkinson, Peskind, & Raskind, 1997; Wolf, Convit, de Leon, Caraos, & Qadri, 2002). Studies have shown that increasing stress with increasing age leads to inferior learning abilities. Middle age is more susceptible to younger age to stress-induced cognitive disturbances.

FUNCTIONAL CHANGES IN AGING

Aging impacts a variety of brain functions including attention, speech, sleep, decision making, working and long term memory (Hedden & Gabrieli, 2004). Experiments with rodent models show that old age exhibits decline in cognition and motivation, decreased interest in novel tasks, motor disabilities and increased anxiety. These changes show corresponding pattern in primates and humans.

Sleep

8

Sleep dysregulation is a usual ailment amongst the elder people. Sleep alteration which are related to age include sleep fragmentation, insomnia, circadian advance, and loss of slow, deep wave sleep whereas daytime indicator consists drowsiness, increase in napping and breakthrough sleep (Blalock, Buechel, Popovic, Geddes, & Landfield, 2011). Also, healthy younger adults subjected to experimentally induced selective deficiency of night time (stationary phase) deep sleep demonstrates some aging- for example phenotypes, including daytime sleepiness, biochemical and metabolic dysfunctions and cognitive deficits. Recent studies propose that slow, deep wave sleep during the inactive period promotes memory, possibly through localized synaptic protein synthesis effects (Buechel et al., 2011). Therefore, the impaired slow wave sleep dysregulation and age on cognition, and high incidence of sleep variations with age, relatively few studies have examined probable mechanistic relations between sleep architecture changes and cognitive decline which is related to age.

Working Memory

Older adults exhibit significant deficits in working memory (Schulze et al., 2017). Whilst, the mechanisms of these ages-associated deficits are still obscure, decline in the activity of prefrontal cortex has been attributed to age dependent deficits in working memory.

Episodic memory is principally affected by ageing. It stands for the memory for specific experiences or events that occurred in the lifespan (Fjell & Walhovd, 2010). A great problem for elderly is the recollection of context or remembering of source information; when or where something was heard or read, or even whether something actually occurred or was just thought about, what has been known as "reality monitoring". Although semantic memory is principally conserved during aging, older people provide general information rather than specific details.

Attention

Older adults show significant impairment in attention. Frontal lobe that requires switching or dividing of concentration among various tasks or inputs, plays an integral role in conduct of cognitive functioning of attention (Fjell & Walhovd, 2010). Prominently, these types of tasks appear to be responsive to training and show benefits of cardiovascular fitness. Attention deficits can have a noteworthy effect on old person's ability to function properly and independently in day to day life. One vital aspect of the day to day functioning affected by attention problem is driving. Driving is an activity that is essential to independence for many old people. Driving necessitates a constant switching of attention in response to surrounding eventualities. Attention must be distributed between observing the surroundings, driving, and selecting relevant from irrelevant stimuli in a jumbled visual array. Research has shown that increased automobile accidents in older adults result from divided attention impairments (Owsley & McGwin, 2010).

Decision Making

Comparatively little research has been done on the effect of aging on decision-making. It has been noticed that while making decision, old people tend to rely more on previous information about the problem and less on new information however young people probably have less knowledge about the prior information and incline to sample and assess more recent information and ponder upon more alternatives before making their decisions (Miller, 2010).

FACTORS AFFECTING BRAIN AGING

Multiple factors affect the rate of brain aging, while some of them accelerate, others slow down agerelated worsening and slow down the completion of pathological levels. Recognizing such factors is crucial to design therapeutic strategies. Some of the factors that modify brain aging are discussed below:

Hypertension

Hypertension is a chronic age-linked illness which is related to numerous changes in the vascular system (Cabeza, Nyberg, & Park, 2016). Chronic raise of blood pressure affects structure of brain and people with medically controlled hypertension are at lower threat for cognitive decline than those who are undiagnosed or untreated. Hypertension accelerates age-related shrinkage of the hippocampus (Raz, Rodrigue, & Haacke, 2007). Individuals with vascular disease factors like hypertension display longitudinal decline in the regions that are generally stable in normal aging, for instance the primary visual cortex. Indeed, in contrary to the hale and hearty adults, individuals with vascular risk factors and vascular disease show rapid expansion in parietal areas and this increase in the pace of expansion is connected with higher systolic blood pressure. Unfortunately, the knowledge of neuroanatomical correlates of hypertension in standard animal models is scarce. Though, selective susceptibility of the prefrontal regions to hypertension is observed in impulsively hypertensive rats, and treatment with antihypertensive agents displays remarkable neuroprotective effect in the prefrontal cortex (Raz & Rodrigue, 2006).

In addition to hypertension, metabolic markers of cardiovascular risk may be linked with structural differences in the brain usually credited to aging. Homocysteine (Hey), an amino acid produced with involvement of vitamins of the 13 group as co-factors, is one such marker. In healthy adults, Hey escalation is connected with deterioration of the hippocampus, reduced total Grey matter volume and ventriculo-megaly. Increased plasma Hey levels predict cognitive decline in nondemented elder individuals, and is related to poor performance on a wide variety of neuropsychological tests, exclusively those measuring delayed executive control and recall (Raz & Rodrigue, 2006).

Stress and Depression

Another cause backing to variable changes in brain function associated with age is individual differences in the stress system arising either due to natural genetic variation or through exposure to a variety of stress causing elements over the lifespan of the individual. Stress has an important influence on the brain throughout the lifespan. Early life stressful experiences initially impair learning and memory processes but enhances emotional memory formation later in life (Shors, 2006). Recent research has reported that people who are exposed to chronic stress or who have shown recurring depression exhibit accelerated brain aging. With growing age, telomeres become short, and studies have shown that this shortening is accelerated by oxidative stress and inflammation (Kordinas, Ioannidis, & Chatzipanagiotou, 2016). On this basis, length of telomere has been linked to age -related diseases and, unhealthy lifestyle and longevity. Also, it has been proposed that telomere length is a determinant of biological aging. Recent research demonstrates that tinier telomere length is linked with both cortisol levels revealing the exposure to prolonged stress and repeated depression (Wikgren et al., 2012).

Vascular Factors and Dementia

White matter lesions (WML), small vessel disease, strokes and dementia increase with growing age and all of them are the damages linked to vascular factors and blood pressure and. Increased cardiovascular risk, decrease in cerebral blood flow, vascular density and cerebral reactivity are associated with WML or hyperintensities (Artero et al., 2004; Kuo & Lipsitz, 2004) (Marstrand et al., 2002). It is yet vague if the

WML causes the vessel loss or vice versa. (Moody et al., 2004). WML are more prominent in frontal than posterior regions of the brain and is related to reduced cognition (Artero et al., 2004; Petkov et al., 2004).

Along with contributing to the cognitive complications in ageing, vascular factors also give rise to the two most common dementias. The incidence of dementia increases exponentially with the increase in age. It is around 20% of those aged 80 and rising to 40% of those aged 90 (Lobo et al., 2000). The two types of dementia seen most frequently in the elder people are Alzheimer's disease (AD) and vascular dementia (VaD) which account for around 40%–70% and 15%–30%. of dementias respectively (Fratiglioni et al., 2000; Lobo et al., 2000). There is an intersection between these two dementias and studies have exposed that AD falls under the category of vascular disorders. A postmortem study found that 77% of VaD cases showed AD pathology (Barker et al., 2002) and high blood pressure has been associated with increased neurofibrilliary tangles characteristic of AD (Sparks et al., 1995). Multiple types of vascular pathology have been associated with AD including microvascular degeneration, disorders of the blood-brain barrier, WML, microinfarctions, and cerebral haemorrhages (K. Jellinger, 2002). It has been suggested that large vessel factors, for example, atherosclerosis, increase the risk of AD and may play a part in cerebral vessel amyloid deposition (Ellis et al., 1996). AD patients do show significantly higher levels of cerebrovascular pathology when compared with controls at postmortem examination (K. A. Jellinger & Mitter-Ferstl, 2003) although this did not correlate with severity of cognitive decline. The characteristic neurofibrillary tangles and plaques found in AD are also evident to some degree in most elderly brains at postmortem examination even those without symptoms, as are white matter lesions (Wen & Sachdev, 2004).

Ageing and development of dementia has shown to be related to occurrence of diabetes, hyperhomocysteinaemia, hypertension, and a high cholesterol even though the proof for all but hypertension is far from clear (Areosa & Grimley, 2002; Budge et al., 2000; Fontbonne, Berr, Ducimetière, & Alpérovitch, 2001; Joosten, 2001; Shepherd et al., 2002). This could be avoided by incorporating changes in diet, reduction/avoidance of alcohol, and by doing regular exercise.

NEURODEGENERATION: PATHWAY AND MECHANISM

The specific conformational changes in proteins happen during neurodegeneration and these transformed proteins intrude the homeostasis of major ions and mitochondrial energetics of the cell (Ward, Zucca, Duyn, Crichton, & Zecca, 2014). These protein alterations are intermingled with multiple domains of cell dysfunction, including: genetic composition; epigenetic modification of genes; post-translational alterations in RNA; endoplasmic reticulum-related protein modifications, including phosphorylation and ubiquitination; cofactors of proteins like metal ions, chaperones; endosomal and lysosomal clearance. These altered proteins lead to the death of neurons by eliciting downstream microglial responses and neuroinflammation. The cell to cell transfer of these altered proteins leads to progression of neurodegeneration across the brain (Prusiner, 2013). The culprit proteins are specific for each ND. In AD, aggregation of amyloid, tau and TDP-43 proteins happen. In the case of PD, neuronal α -synuclein gets deposited. Generally, in all the NDs, monomers are generated and then they aggregate to give rise to oligomers and eventually they turn into insoluble form of the protein. The major abnormalities present in these disorders are unregulated autophagy and poor protein processing. These abnormalities hinder the protein disposal and proteostasis biogenesis, folding, trafficking and degradation of proteins. As stated above, these abnormal proteins lead to mitochondrial dysfunction and disrupted cellular energet-

ics (Guo, Sun, Chen, & Zhang, 2013). These abnormalities result in disturbed oxidative mechanism and increased free radical production.

STRUCTURAL BRAIN CHANGES IN NEURODEGENERATION

Progressive cerebral atrophy is a peculiar attribute of neurodegeneration. The dendritic and neuronal losses are the major causatives of atrophy (Johnson, Fox, Sperling, & Klunk, 2012). In neurodegeneration, volume of the grey matter (GM) and microstructural wholeness of the white matter (WM) tracts gets changed. The white matter consists of inferior longitudinal fasciculus (ILF) and inferior frontooccipital fasciculus (IFOF). IFOF touches the superior frontal cortex rostrally and dorsal parietal and occipital cortex caudally. ILF connects the anterior temporal and the occipital cortices. IFOF is involved in visuospatial function, while ILF plays a role in visual memory. The pattern of loss differs between diseases reflecting regional disease expression and/or selective neuronal vulnerability. Studies have revealed that in individuals with Parkinson's disease with dementia have dispersed prefrontal cortex and parietal-temporal atrophy (Burton, McKeith, Burn, Williams, & O'Brien, 2004; Camicioli et al., 2003; Junqué et al., 2005; Kenny, Burton, & O'Brien, 2008; Tam, Burton, McKeith, Burn, & O'brien, 2005). The individuals distressed with Parkinson's disease with non-demented or have minor cognitive impairment have displayed varying degrees of atrophy for the amygdala, hippocampus, parietal-temporal cortex and prefrontal cortex (Beyer, Janvin, Larsen, & Aarsland, 2007). In Primary open angle glaucoma (POAG) patients, GM atrophy was observed in the lateral region, medial region, most anterior, and most posterior parts of the visual cortex (Boucard et al., 2009). Some studies have revealed that the thickness in cortical region in primary and secondary visual cortex gets reduced in POAG patients (Yu et al., 2013). Additionally, individuals suffering from POAG exhibited GM atrophy in regions involved in processing of cognitive functions such as the fronto-orbital cortex (decision-making), hippocampus (memory) and superior parietal lobule (spatial orientation). Remarkably, several of these structures are significantly involved in the tau pathology of AD (Frisoni, Fox, Jack, Scheltens, & Thompson, 2010). Studies reported that AD is identified by a sneaky start and unstoppable progression of atrophy that is manifested firstly in the medial temporal lobe (Scahill, Schott, Stevens, Rossor, & Fox, 2002). Usually, the earliest site of atrophy is the entorhinal cortex, shadowed by the hippocampus, amygdala, and parahippocampus (Chan et al., 2001; Dickerson et al., 2001; Killiany et al., 2002; Lehericy et al., 1994). Other structures like posterior cingulate are also affected earlier. These neural damages then binge on spreading to include the temporal neocortex and then all neocortical association zones.

TECHNIQUES AND ADVANCE IMAGING TO IDENTIFY THE PROGRESSION OF NEURODEGNERATION

Usually, NDs have a long latency period, during which pathological changes happen, followed by the inevitable neuronal damage. This period is known as pre-symptomatic period. There are certain bio-markers and advance imaging techniques which helps in identifying pre-symptomatic individuals with indication of neurodegenerative change. They are also used assess the stage of their disease.

Biomarkers

A biomarker could be a naturally occurring molecule or gene or a characteristic that plays a key role as an indicator of usual biological developments, pharmacological reactions, or pathogenic processes to a therapeutic intervention. Brain imaging or patient-derived fluids (CSF, blood, urine, saliva) may provide the relevant information about the biomarker (Gonzalez-Cuyar, Sonnen, Montine, Keene, & Montine, 2011). Biomarkers for NDs are less well developed and explored, but the discovery of useful biomarkers across NDs is anticipated. But in the AD, amyloid protein is measured by CSF biomarker and provides total insight into disease pathophysiology and progression. Fluid biomarkers include both trait (e.g., genetic) and state (e.g., CSF markers) measures. Genetic biomarkers are used to detect individuals who are at risk b identifying mutation carriers, risk polymorphisms etc. State biomarkers are used in the identification of pre-symptomatic individuals and to support the accuracy of disease diagnosis. Genetic biomarkers are used to detect individuals who are at risk. State biomarkers are used in identification of pre-symptomatic individuals and to support diagnostic accuracy of disease diagnosis.

Brain Imaging Techniques

The most extensively used neuroimaging technique for examination of neurodegeneration in vivo is structural MRI. It has the ability to do both local and global assessment of atrophic brain changes. It can also be used for assessing function of brain. Magnetic resonance spectroscopy [MRS], diffusion tensor and diffusion weighted imaging [DTI/DWI], and perfusion imaging are more advanced structural MRI techniques and are used in investigation of dementia (Risacher & Saykin, 2013). DTI/DWI techniques quantify the wholeness of tissue using mean diffusivity (MD) or apparent diffusion coefficient (ADC) and fractional anisotropy (FA) (Huisman, 2010). Decreased FA and elevated MD/ADC are considered to be indicators of reduced gray matter and white matter integrity and neuronal fiber loss. Biological metabolites in target tissue can be quantified by MRS. Two major metabolites that leads to alterations are myoinositol (mIns) and N-acetylaspartate (NAA). Former is a measure of glial cell production and neuronal destruction while later is a marker of neuronal integrity.

Along with other imaging techniques, cerebral perfusion is also usually measured in studies of neurodegeneration (Risacher & Saykin, 2013). It is done by using techniques like PET and SPECT, which incorporate the use radiolabeled ligands to measure perfusion, neurochemical, and metabolic processes in vivo. In studies of neurodegeneration, SPECT is used to estimate brain perfusion while PET (positron emission tomography) is used to evaluate the functional changes in the neurotransmitter, metabolism of brain, and the level of other proteins. This assessment can deliver significant information about degenerative alterations occurring in the brains of diseased individuals.

Functional MRI (fMRI) is used for measuring the activity of brain during a sensory, cognitive, or motor task or at rest by quantifying blood oxygen levels and blood flow. The primary outcome measured in most fMRI studies is blood oxygenation level dependent (BOLD) contrast signal in which regional brain activity is measured via changes in local blood flow and oxygenation (Ogawa et al., 1992). The BOLD signal is a useful measure for brain activation (Logothetis, Pauls, Augath, Trinath, & Oeltermann, 2001). The altered neuronal metabolism and blood flow due to brain atrophy may cause alterations in the BOLD signal. fMRI studies often evaluate brain activity during cognitive or functional motor tests. In addition to estimates of regional task-related brain activity, quantification of brain networks can provide a unique measure of brain activity (Greicius, Krasnow, Reiss, & Menon, 2003).

FUTURE RESEARCH DIRECTIONS

One of the major hurdles of forthcoming research will be the identification of significant factors that govern individual differences in susceptibility to stress, hypertension, and aging.

The biomarkers for NDs are less well developed and explored, so future studies will focus on the discovery of useful biomarkers across NDs. It will help in detection of disease prior to its onset and thus could help in treating it effectively. Future studies will be focusing on utilization of novel techniques for understanding the pathology of NDs, unveiling of disease at initial stage, and the development of targeted therapeutics. The advances in neuroimaging will help in improving the clinical diagnosis by detecting the disease even prior to its commencement, and its early intervention by embracing the potential therapeutics (Risacher & Saykin, 2013). It is anticipated that advanced neuroimaging techniques for diagnostics will significantly enhance the specificity, sensitivity and accuracy of diagnosis. The framework for development of preventive therapies is also expected. Future studies may also include developing a probable and multicentre design of novel tools meant for the purpose of diagnosis; assessment of various potential biomarkers and conduct of a clinical trial (till death as end-point) that can assess both clinical features and determine the biological diagnostics and ultimately neuropathological confirmation by inspecting the brains of patients at death (Agrawal & Biswas, 2015). However, pre-symptomatic detection and preventive treatment management of neurodegeneration and age-related cognitive decline will be the greatest future impacts of existing techniques and those under development (Small et al., 2008).

CONCLUSION

Stress and hypertension are the most common factors associated with nearly all NDs, resulting in neurofibrillary tangles, A β plaques and brain atrophy which lead to modulation of structure and functions of the brain. The increasing incidence of hypertension due to the aging population worldwide and rising occurrence of stress can also escalate the count of people suffering from ND, since midlife hypertension nearly doubles the possibility of developing diseases like AD in later life. These factors lead to the decline in cognition and memory function. Unnecessary and enormous stress experienced at difficult time windows of life can intensely affect cognitive function at later stages, with a particular effect on cognitive aging. There are certain biomarkers both genetic and state which are helpful in pre-determination of disease. They can be observed by neuroimaging or by analyzing patient-derived fluids. Imaging studies of NDs are extremely enlightening regarding structural, functional, and molecular changes in brain. Neuroimaging techniques cannot be taken into account for differential diagnosis of various diseases, but they have proved to be helpful. Further studies with advanced MRI techniques and future PET tracers for proteinopathies beyond amyloid (i.e., tau, α -synuclein, and TDP-43) will probably provide even more data about pathology associated with the various degenerative and dementing syndromes. Additionally, neuroimaging techniques may prove to be useful in conduct of clinical trials of novel therapeutics intended to treat these disorders for both monitoring changes related to disease or as end-points to complement present clinical outcome measures.

ACKNOWLEDGMENT

All authors have contributed equally and there are no conflicts of interest.

REFERENCES

Agrawal, M., & Biswas, A. (2015). Molecular diagnostics of neurodegenerative disorders. *Frontiers in Molecular Biosciences*, 2. PMID:26442283

Alexander, G. E. (2004). Biology of Parkinson's disease: Pathogenesis and pathophysiology of a multisystem neurodegenerative disorder. *Dialogues in Clinical Neuroscience*, 6(3), 259. PMID:22033559

Alhola, P., & Polo-Kantola, P. (2007). Sleep deprivation: Impact on cognitive performance. *Neuropsychiatric Disease and Treatment*, *3*(5), 553. PMID:19300585

Areosa, S., & Grimley, E. (2002). Effect of the treatment of Type II diabetes mellitus on the development of cognitive impairment and dementia. *Cochrane Database of Systematic Reviews*, (4): CD003804–CD003804. PMID:12519608

Artero, S., Tiemeier, H., Prins, N., Sabatier, R., Breteler, M., & Ritchie, K. (2004). Neuroanatomical localisation and clinical correlates of white matter lesions in the elderly. *Journal of Neurology, Neurosurgery, and Psychiatry*, 75(9), 1304–1308. doi:10.1136/jnnp.2003.023713 PMID:15314121

Bailey, R. (2017). *Divisions of the Brain. Retrieved from Divisions of the Brain - Forebrain, Midbrain, Hindbrain.* Retrieved from https://www.thoughtco.com/divisions-of-the-brain-4032899

Barker, W. W., Luis, C. A., Kashuba, A., Luis, M., Harwood, D. G., Loewenstein, D., ... Sevush, S. (2002). Relative frequencies of Alzheimer disease, Lewy body, vascular and frontotemporal dementia, and hippocampal sclerosis in the State of Florida Brain Bank. *Alzheimer Disease and Associated Disorders*, *16*(4), 203–212. doi:10.1097/00002093-200210000-00001 PMID:12468894

Bergersen, L. H. (2015). Lactate Transport and Signaling in the Brain: Potential Therapeutic Targets and Roles in Body—Brain Interaction. *Journal of Cerebral Blood Flow and Metabolism*, *35*(2), 176–185. doi:10.1038/jcbfm.2014.206 PMID:25425080

Beyer, M. K., Janvin, C. C., Larsen, J. P., & Aarsland, D. (2007). A magnetic resonance imaging study of patients with Parkinson's disease with mild cognitive impairment and dementia using voxel-based morphometry. *Journal of Neurology, Neurosurgery, and Psychiatry*, 78(3), 254–259. doi:10.1136/jnnp.2006.093849 PMID:17028119

Bidichandani, S. I., & Delatycki, M. B. (2017). Friedreich ataxia. Academic Press.

Bishop, N. A., Lu, T., & Yankner, B. A. (2010). Neural mechanisms of ageing and cognitive decline. *Nature*, 464(7288), 529–535. doi:10.1038/nature08983 PMID:20336135

Blalock, E. M., Buechel, H. M., Popovic, J., Geddes, J. W., & Landfield, P. W. (2011). Microarray analyses of laser-captured hippocampus reveal distinct gray and white matter signatures associated with incipient Alzheimer's disease. *Journal of Chemical Neuroanatomy*, *42*(2), 118–126. doi:10.1016/j. jchemneu.2011.06.007 PMID:21756998

Boucard, C. C., Hernowo, A. T., Maguire, R. P., Jansonius, N. M., Roerdink, J. B., Hooymans, J. M., & Cornelissen, F. W. (2009). Changes in cortical grey matter density associated with long-standing retinal visual field defects. *Brain*, *132*(7), 1898–1906. doi:10.1093/brain/awp119 PMID:19467992

Budge, M., Johnston, C., Hogervorst, E., De Jager, C., Milwain, E., Iversen, S., ... Smith, A. (2000). Plasma total homocysteine and cognitive performance in a volunteer elderly population. *Annals of the New York Academy of Sciences*, 903(1), 407–410. doi:10.1111/j.1749-6632.2000.tb06392.x PMID:10818531

Buechel, H. M., Popovic, J., Searcy, J. L., Porter, N. M., Thibault, O., & Blalock, E. M. (2011). Deep Sleep and Parietal Cortex Gene Expression Changes Are Related to Cognitive Deficits with Age. *PLoS One*, *6*(4), e18387. doi:10.1371/journal.pone.0018387 PMID:21483696

Burton, E. J., McKeith, I. G., Burn, D. J., Williams, E. D., & O'Brien, J. T. (2004). Cerebral atrophy in Parkinson's disease with and without dementia: A comparison with Alzheimer's disease, dementia with Lewy bodies and controls. *Brain*, *127*(4), 791–800. doi:10.1093/brain/awh088 PMID:14749292

Cabeza, R., Nyberg, L., & Park, D. C. (2016). *Cognitive neuroscience of aging: Linking cognitive and cerebral aging*. Oxford University Press. doi:10.1093/acprof:oso/9780199372935.001.0001

Cai, Q., & Tammineni, P. (2017). Mitochondrial aspects of synaptic dysfunction in Alzheimer's disease. *Journal of Alzheimer's Disease*, 57(4), 1087–1103. doi:10.3233/JAD-160726 PMID:27767992

Camicioli, R., Moore, M. M., Kinney, A., Corbridge, E., Glassberg, K., & Kaye, J. A. (2003). Parkinson's disease is associated with hippocampal atrophy. *Movement Disorders*, *18*(7), 784–790. doi:10.1002/mds.10444 PMID:12815657

Chakrabarti, S., Munshi, S., Banerjee, K., Thakurta, I. G., Sinha, M., & Bagh, M. B. (2011). Mitochondrial Dysfunction during Brain Aging: Role of Oxidative Stress and Modulation by Antioxidant Supplementation. *Aging and Disease*, 2(3), 242–256. PMID:22396876

Chan, D., Fox, N. C., Scahill, R. I., Crum, W. R., Whitwell, J. L., Leschziner, G., ... Rossor, M. N. (2001). Patterns of temporal lobe atrophy in semantic dementia and Alzheimer's disease. *Annals of Neurology*, *49*(4), 433–442. doi:10.1002/ana.92 PMID:11310620

Council, N. R. (2001). *New horizons in health: An integrative approach* (Vol. 277). National Academies Press.

Daldin, M., Fodale, V., Cariulo, C., Azzollini, L., Verani, M., Martufi, P., ... Macdonald, D. (2017). Polyglutamine expansion affects huntingtin conformation in multiple Huntington's disease models. *Scientific Reports*, *7*(1), 5070. doi:10.103841598-017-05336-7 PMID:28698602

de Kloet, E. R., Oitzl, M. S., & Joëls, M. (1999). Stress and cognition: Are corticosteroids good or bad guys? *Trends in Neurosciences*, 22(10), 422–426. doi:10.1016/S0166-2236(99)01438-1 PMID:10481183

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Dickerson, B. C., Goncharova, I., Sullivan, M., Forchetti, C., Wilson, R., Bennett, D., ... deToledo-Morrell, L. (2001). MRI-derived entorhinal and hippocampal atrophy in incipient and very mild Alzheimer's disease. *Neurobiology of Aging*, 22(5), 747–754. doi:10.1016/S0197-4580(01)00271-8 PMID:11705634

Ellis, R., Olichney, J., Thal, L., Mirra, S., Morris, J., Beekly, D., & Heyman, A. (1996). Cerebral amyloid angiopathy in the brains of patients with Alzheimer's disease The CERAD experience, part XV. *Neurology*, *46*(6), 1592–1596. doi:10.1212/WNL.46.6.1592 PMID:8649554

Fjell, A. M., & Walhovd, K. B. (2010). Structural brain changes in aging: Courses, causes and cognitive consequences. *Reviews in the Neurosciences*, 21(3), 187–221. doi:10.1515/REVNEURO.2010.21.3.187 PMID:20879692

Fontbonne, A., Berr, C., Ducimetière, P., & Alpérovitch, A. (2001). Changes in cognitive abilities over a 4-year period are unfavorably affected in elderly diabetic subjects. *Diabetes Care*, 24(2), 366–370. doi:10.2337/diacare.24.2.366 PMID:11213894

Fratiglioni, L., Launer, L., Andersen, K., Breteler, M., Copeland, J., Dartigues, J., ... Hofman, A. (2000). Incidence of dementia and major subtypes in Europe: A collaborative study of population-based cohorts. Neurologic Diseases in the Elderly Research Group. *Neurology*, *54*(11Suppl 5), S10–S15. PMID:10854355

Frisoni, G. B., Fox, N. C., Jack, C. R., Scheltens, P., & Thompson, P. M. (2010). The clinical use of structural MRI in Alzheimer disease. *Nature Reviews. Neurology*, *6*(2), 67–77. doi:10.1038/nrneurol.2009.215 PMID:20139996

Giorgi, C., De Stefani, D., Bononi, A., Rizzuto, R., & Pinton, P. (2009). Structural and functional link between the mitochondrial network and the endoplasmic reticulum. *The International Journal of Biochemistry & Cell Biology*, *41*(10), 1817–1827. doi:10.1016/j.biocel.2009.04.010 PMID:19389485

Glorioso, C., Oh, S., Douillard, G. G., & Sibille, E. (2011). Brain molecular aging, promotion of neurological disease and modulation by Sirtuin5 longevity gene polymorphism. *Neurobiology of Disease*, *41*(2), 279–290. doi:10.1016/j.nbd.2010.09.016 PMID:20887790

Gonzalez-Cuyar, L. F., Sonnen, J. A., Montine, K. S., Keene, C. D., & Montine, T. J. (2011). Role of Cerebrospinal Fluid and Plasma Biomarkers in the Diagnosis of Neurodegenerative Disorders and Mild Cognitive Impairment. *Current Neurology and Neuroscience Reports*, *11*(5), 455–463. doi:10.100711910-011-0212-0 PMID:21725901

Gorman, A. M. (2008). Neuronal cell death in neurodegenerative diseases: Recurring themes around protein handling. *Journal of Cellular and Molecular Medicine*, *12*(6a), 2263–2280. doi:10.1111/j.1582-4934.2008.00402.x PMID:18624755

Gout, J.-F., Kahn, D., Duret, L., & Paramecium Post-Genomics, C. (2010). The Relationship among Gene Expression, the Evolution of Gene Dosage, and the Rate of Protein Evolution. *PLOS Genetics*, *6*(5), e1000944. doi:10.1371/journal.pgen.1000944 PMID:20485561

Greicius, M. D., Krasnow, B., Reiss, A. L., & Menon, V. (2003). Functional connectivity in the resting brain: A network analysis of the default mode hypothesis. *Proceedings of the National Academy of Sciences of the United States of America*, 100(1), 253–258. doi:10.1073/pnas.0135058100 PMID:12506194

Guo, C., Sun, L., Chen, X., & Zhang, D. (2013). Oxidative stress, mitochondrial damage and neurodegenerative diseases. *Neural Regeneration Research*, 8(21), 2003. PMID:25206509

Hammerman, M. (1987). Insulin-like growth factors and aging. *Endocrinology and Metabolism Clinics* of North America, 16(4), 995–1011. PMID:3322823

Hedden, T., & Gabrieli, J. D. (2004). Insights into the ageing mind: A view from cognitive neuroscience. *Nature Reviews. Neuroscience*, *5*(2), 87–96. doi:10.1038/nrn1323 PMID:14735112

Hibberd, C., Yau, J. L., & Seckl, J. R. (2000). Glucocorticoids and the ageing hippocampus. *Journal of Anatomy*, *197*(4), 553–562. doi:10.1046/j.1469-7580.2000.19740553.x PMID:11197528

Huisman, T. (2010). Diffusion-weighted and diffusion tensor imaging of the brain, made easy. *Cancer Imaging; the Official Publication of the International Cancer Imaging Society*, *10*(1A), S163–S171. doi:10.1102/1470-7330.2010.9023 PMID:20880787

Jellinger, K. (2002). Alzheimer disease and cerebrovascular pathology: An update. *Journal of Neural Transmission (Vienna, Austria)*, *109*(5), 813–836. doi:10.1007007020200068 PMID:12111471

Jellinger, K. A., & Mitter-Ferstl, E. (2003). The impact of cerebrovascular lesions in Alzheimer disease. *Journal of Neurology*, 250(9), 1050–1055. doi:10.100700415-003-0142-0 PMID:14504965

Johnson, K. A., Fox, N. C., Sperling, R. A., & Klunk, W. E. (2012). Brain Imaging in Alzheimer Disease. *Cold Spring Harbor Perspectives in Medicine*, 2(4), a006213. doi:10.1101/cshperspect.a006213 PMID:22474610

Joosten, E. (2001). Homocysteine, vascular dementia and Alzheimer's disease. *Clinical Chemistry and Laboratory Medicine*, *39*(8), 717–720. doi:10.1515/CCLM.2001.119 PMID:11592440

Junqué, C., Ramírez-Ruiz, B., Tolosa, E., Summerfield, C., Martí, M. J., Pastor, P., ... Mercader, J. M. (2005). Amygdalar and hippocampal MRI volumetric reductions in Parkinson's disease with dementia. *Movement Disorders*, 20(5), 540–544. doi:10.1002/mds.20371 PMID:15645532

Kenny, E. R., Burton, E. J., & O'Brien, J. T. (2008). A volumetric magnetic resonance imaging study of entorhinal cortex volume in dementia with Lewy bodies. *Dementia and Geriatric Cognitive Disorders*, 26(3), 218–225. doi:10.1159/000153432 PMID:18781072

Killiany, R., Hyman, B., Gomez-Isla, T., Moss, M., Kikinis, R., Jolesz, F., ... Albert, M. (2002). MRI measures of entorhinal cortex vs hippocampus in preclinical AD. *Neurology*, *58*(8), 1188–1196. doi:10.1212/ WNL.58.8.1188 PMID:11971085

Kordinas, V., Ioannidis, A., & Chatzipanagiotou, S. (2016). The Telomere/Telomerase System in Chronic Inflammatory Diseases. Cause or Effect? *Genes*, 7(9), 60. doi:10.3390/genes7090060 PMID:27598205

Kovacs, G. G. (2016). Molecular pathological classification of neurodegenerative diseases: Turning towards precision medicine. *International Journal of Molecular Sciences*, *17*(2), 189. doi:10.3390/ ijms17020189 PMID:26848654

The Aging Brain

Kumar, H., Lim, H.-W., More, S. V., Kim, B.-W., Koppula, S., Kim, I. S., & Choi, D.-K. (2012). The Role of Free Radicals in the Aging Brain and Parkinson's Disease: Convergence and Parallelism. *International Journal of Molecular Sciences*, *13*(8), 10478–10504. doi:10.3390/ijms130810478 PMID:22949875

Kuo, H.-K., & Lipsitz, L. A. (2004). Cerebral white matter changes and geriatric syndromes: Is there a link? *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, 59(8), M818–M826. doi:10.1093/gerona/59.8.M818 PMID:15345732

Lehericy, S., Baulac, M., Chiras, J., Pierot, L., Martin, N., Pillon, B., ... Marsault, C. (1994). Amygdalohippocampal MR volume measurements in the early stages of Alzheimer disease. *AJNR. American Journal of Neuroradiology*, *15*(5), 929–937. PMID:8059663

Lin, M. T., & Beal, M. F. (2006). Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature*, 443(7113), 787–795. doi:10.1038/nature05292 PMID:17051205

Lobo, A., Launer, L., Fratiglioni, L., Andersen, K., Di Carlo, A., Breteler, M., ... Martinez-Lage, J. (2000). Prevalence of dementia and major subtypes in Europe: A collaborative study of population-based cohorts. *Neurology*, *54*(5), S4. PMID:10854354

Loerch, P. M., Lu, T., Dakin, K. A., Vann, J. M., Isaacs, A., Geula, C., ... Li, C. (2008). Evolution of the aging brain transcriptome and synaptic regulation. *PLoS One*, *3*(10), e3329. doi:10.1371/journal. pone.0003329 PMID:18830410

Logothetis, N. K., Pauls, J., Augath, M., Trinath, T., & Oeltermann, A. (2001). Neurophysiological investigation of the basis of the fMRI signal. *Nature*, *412*(6843), 150–157. doi:10.1038/35084005 PMID:11449264

MacKenzie-Graham, A., Tinsley, M. R., Shah, K. P., Aguilar, C., Strickland, L. V., Boline, J., ... Jacobs, R. E. (2006). Cerebellar cortical atrophy in experimental autoimmune encephalomyelitis. *NeuroImage*, *32*(3), 1016–1023. doi:10.1016/j.neuroimage.2006.05.006 PMID:16806982

Magrinelli, F., Picelli, A., Tocco, P., Federico, A., Roncari, L., Smania, N., ... Tamburin, S. (2016). Pathophysiology of motor dysfunction in Parkinson's disease as the rationale for drug treatment and rehabilitation. *Parkinson's Disease*. PMID:27366343

Marstrand, J., Garde, E., Rostrup, E., Ring, P., Rosenbaum, S., Mortensen, E. L., & Larsson, H. (2002). Cerebral perfusion and cerebrovascular reactivity are reduced in white matter hyperintensities. *Stroke*, *33*(4), 972–976. doi:10.1161/01.STR.0000012808.81667.4B PMID:11935046

McEntee, W. J., & Crook, T. H. (1993). Glutamate: Its role in learning, memory, and the aging brain. *Psychopharmacology*, *111*(4), 391–401. doi:10.1007/BF02253527 PMID:7870979

McEwen, B. S. (2002). Sex, stress and the hippocampus: Allostasis, allostatic load and the aging process. *Neurobiology of Aging*, 23(5), 921–939. doi:10.1016/S0197-4580(02)00027-1 PMID:12392796

McKee, A. C., & Daneshvar, D. H. (2015). The neuropathology of traumatic brain injury. *Handbook of Clinical Neurology*, *127*, 45–66. doi:10.1016/B978-0-444-52892-6.00004-0 PMID:25702209

Meltzer, C. C., Smith, G., DeKosky, S. T., Pollock, B. G., Mathis, C. A., Moore, R. Y., ... Reynolds, C. F. (1998). Serotonin in aging, late-life depression, and Alzheimer's disease: The emerging role of functional imaging. *Neuropsychopharmacology*, *18*(6), 407–430. doi:10.1016/S0893-133X(97)00194-2 PMID:9571651

Miller, S. (2010). Aging and decision making. Academic Press.

Moody, D. M., Thore, C. R., Anstrom, J. A., Challa, V. R., Langefeld, C. D., & Brown, W. R. (2004). Quantification of afferent vessels shows reduced brain vascular density in subjects with leukoaraiosis. *Radiology*, *233*(3), 883–890. doi:10.1148/radiol.2333020981 PMID:15564412

Namavar, Y., Barth, P. G., & Baas, F. (2011). Classification, diagnosis and potential mechanisms in pontocerebellar hypoplasia. *Orphanet Journal of Rare Diseases*, *6*(1), 50. doi:10.1186/1750-1172-6-50 PMID:21749694

Näntö-Salonen, K., Muller, H. L., Hoffman, A. R., Vu, T. H., & Rosenfeld, R. G. (1993). Mechanisms of thyroid hormone action on the insulin-like growth factor system: All thyroid hormone effects are not growth hormone mediated. *Endocrinology*, *132*(2), 781–788. doi:10.1210/endo.132.2.7678799 PMID:7678799

O'Brien, R. J., & Wong, P. C. (2011). Amyloid precursor protein processing and Alzheimer's disease. *Annual Review of Neuroscience*, 34(1), 185–204. doi:10.1146/annurev-neuro-061010-113613 PMID:21456963

Ogawa, S., Tank, D. W., Menon, R., Ellermann, J. M., Kim, S. G., Merkle, H., & Ugurbil, K. (1992). Intrinsic signal changes accompanying sensory stimulation: Functional brain mapping with magnetic resonance imaging. *Proceedings of the National Academy of Sciences of the United States of America*, 89(13), 5951–5955. doi:10.1073/pnas.89.13.5951 PMID:1631079

Ota, M., Yasuno, F., Ito, H., Seki, C., Nozaki, S., Asada, T., & Suhara, T. (2006). Age-related decline of dopamine synthesis in the living human brain measured by positron emission tomography with L-[β-11 C] DOPA. *Life Sciences*, *79*(8), 730–736. doi:10.1016/j.lfs.2006.02.017 PMID:16580023

Owsley, C., & McGwin, G. Jr. (2010). Vision and Driving. *Vision Research*, 50(23), 2348–2361. doi:10.1016/j.visres.2010.05.021 PMID:20580907

Paulson, H. L., Das, S. S., Crino, P. B., Perez, M. K., Patel, S. C., Gotsdiner, D., ... Pittman, R. N. (1997). Machado-Joseph disease gene product is a cytoplasmic protein widely expressed in brain. *Annals of Neurology*, *41*(4), 453–462. doi:10.1002/ana.410410408 PMID:9124802

Penner, L. A., Blair, I. V., Albrecht, T. L., & Dovidio, J. F. (2014). Reducing racial health care disparities: A social psychological analysis. *Policy Insights from the Behavioral and Brain Sciences*, *1*(1), 204–212. doi:10.1177/2372732214548430 PMID:25705721

Peters, R. (2006). Ageing and the brain. *Postgraduate Medical Journal*, 82(964), 84–88. doi:10.1136/ pgmj.2005.036665 PMID:16461469

The Aging Brain

Petkov, C. I., Wu, C. C., Eberling, J. L., Mungas, D., Zrelak, P. A., Yonelinas, A. P., ... Jagust, W. J. (2004). Correlates of memory function in community-dwelling elderly: The importance of white matter hyperintensities. *Journal of the International Neuropsychological Society*, *10*(3), 371–381. doi:10.1017/S1355617704103056 PMID:15147595

Posner, M. I., & Rothbart, M. K. (2007). *Educating the human brain*. American Psychological Association. doi:10.1037/11519-000

Prusiner, S. B. (2013). Biology and genetics of prions causing neurodegeneration. *Annual Review of Genetics*, 47(1), 601–623. doi:10.1146/annurev-genet-110711-155524 PMID:24274755

Purves, D., Augustine, G. J., Fitzpatrick, D., Katz, L. C., LaMantia, A.-S., McNamara, J. O., & Williams, S. M. (2001). *Circuits within the basal ganglia system*. Academic Press.

Rajeswaran, J., & Bennett, C. N. (2013). The Neuropsychology of Stress. *Stress and Work: Perspectives on Understanding and Managing Stress*, 13.

Raz, N., & Rodrigue, K. M. (2006). Differential aging of the brain: Patterns, cognitive correlates and modifiers. *Neuroscience and Biobehavioral Reviews*, *30*(6), 730–748. doi:10.1016/j.neubiorev.2006.07.001 PMID:16919333

Raz, N., Rodrigue, K. M., & Haacke, E. M. (2007). Brain Aging and Its Modifiers: Insights from in Vivo Neuromorphometry and Susceptibility Weighted Imaging. *Annals of the New York Academy of Sciences*, *1097*(1), 84–93. doi:10.1196/annals.1379.018 PMID:17413014

Recasens, A., & Dehay, B. (2014). Alpha-synuclein spreading in Parkinson's disease. *Frontiers in Neuroanatomy*, 8. PMID:25565982

Risacher, S. L., & Saykin, A. J. (2013). Neuroimaging Biomarkers of Neurodegenerative Diseases and Dementia. *Seminars in Neurology*, *33*(4), 386–416. doi:10.1055-0033-1359312 PMID:24234359

Roozendaal, B. (2002). Stress and memory: Opposing effects of glucocorticoids on memory consolidation and memory retrieval. *Neurobiology of Learning and Memory*, 78(3), 578–595. doi:10.1006/ nlme.2002.4080 PMID:12559837

Rosenberg, G. A. (2014). Blood-Brain Barrier Permeability in Aging and Alzheimer's Disease. *The Journal of Prevention of Alzheimer's Disease*, 1(3), 138–139. doi:10.14283/jpad.2014.25 PMID:26301207

Sandi, C. (1998). The role and mechanisms of action of glucocorticoid involvement in memory storage. *Neural Plasticity*, *6*(3), 41–52. doi:10.1155/NP.1998.41 PMID:9920681

Sandi, C. (2004). Stress, cognitive impairment and cell adhesion molecules. *Nature Reviews. Neuroscience*, *5*(12), 917. doi:10.1038/nrn1555 PMID:15550947

Scahill, R. I., Schott, J. M., Stevens, J. M., Rossor, M. N., & Fox, N. C. (2002). Mapping the evolution of regional atrophy in Alzheimer's disease: Unbiased analysis of fluid-registered serial MRI. *Proceedings of the National Academy of Sciences of the United States of America*, 99(7), 4703–4707. doi:10.1073/pnas.052587399 PMID:11930016

Schulz-Schaeffer, W. J. (2010). The synaptic pathology of α -synuclein aggregation in dementia with Lewy bodies, Parkinson's disease and Parkinson's disease dementia. *Acta Neuropathologica*, *120*(2), 131–143. doi:10.100700401-010-0711-0 PMID:20563819

Schulze, T., Ahel, M., Ahlheim, J., Aït-Aïssa, S., Brion, F., Di Paolo, C., ... Hollert, H. (2017). Assessment of a novel device for onsite integrative large-volume solid phase extraction of water samples to enable a comprehensive chemical and effect-based analysis. *The Science of the Total Environment*, *581*, 350–358. doi:10.1016/j.scitotenv.2016.12.140 PMID:28062104

Shah, J., & Zeier, J. (2013). Long-distance communication and signal amplification in systemic acquired resistance. *Frontiers in Plant Science*, *4*, 30. doi:10.3389/fpls.2013.00030 PMID:23440336

Shao, J., & Diamond, M. I. (2007). Polyglutamine diseases: Emerging concepts in pathogenesis and therapy. *Human Molecular Genetics*, *16*(R2), R115–R123. doi:10.1093/hmg/ddm213 PMID:17911155

Shelkovnikova, T., Kulikova, A., Tsvetkov, F., Peters, O., Bachurin, S., Bukhman, V., & Ninkina, N. (2012). Proteinopathies--forms of neurodegenerative disorders with protein aggregation-based pathology. *Molekuliarnaia biologiia*, *46*(3), 402–415. PMID:22888630

Shelkovnikova, T., Kulikova, A., Tsvetkov, P. O., Peters, O., Bachurin, S., Buchman, V. L., & Ninkina, N. (2012). Proteinopathies, neurodegenerative disorders with protein aggregation-based pathology. *Molecular Biology*, *46*(3), 362–374. doi:10.1134/S0026893312020161 PMID:22888630

Shepherd, J., Blauw, G. J., Murphy, M. B., Bollen, E. L., Buckley, B. M., Cobbe, S. M., ... Jukema, J. W. (2002). Pravastatin in elderly individuals at risk of vascular disease (PROSPER): A randomised controlled trial. *Lancet*, *360*(9346), 1623–1630. doi:10.1016/S0140-6736(02)11600-X PMID:12457784

Shors, T. J. (2006). Stressful Experience and Learning Across the Lifespan. *Annual Review of Psychology*, *57*(1), 55–85. doi:10.1146/annurev.psych.57.102904.190205 PMID:16318589

Small, G. W., Bookheimer, S. Y., Thompson, P. M., Cole, G. M., Huang, S., Kepe, V., & Barrio, J. R. (2008). Current and future uses of neuroimaging for cognitively impaired patients. *Lancet Neurology*, 7(2), 161–172. doi:10.1016/S1474-4422(08)70019-X PMID:18207114

Sorwell, K. G., & Urbanski, H. F. (2013). Causes and consequences of age-related steroid hormone changes: Insights gained from nonhuman primates. *Journal of Neuroendocrinology*, *25*(11), 1062–1069. doi:10.1111/jne.12064 PMID:23796387

Sparks, D. L., Scheff, S. W., Liu, H., Landers, T. M., Coyne, C. M., & Hunsaker, J. C. III. (1995). Increased incidence of neurofibrillary tangles (NFT) in non-demented individuals with hypertension. *Journal of the Neurological Sciences*, *131*(2), 162–169. doi:10.1016/0022-510X(95)00105-B PMID:7595642

Sweeney, P., Park, H., Baumann, M., Dunlop, J., Frydman, J., Kopito, R., ... Hodgson, R. (2017). Protein misfolding in neurodegenerative diseases: Implications and strategies. *Translational Neurodegeneration*, *6*(1), 6. doi:10.118640035-017-0077-5 PMID:28293421

Tam, C., Burton, E., McKeith, I., Burn, D., & O'brien, J. (2005). Temporal lobe atrophy on MRI in Parkinson disease with dementia A comparison with Alzheimer disease and dementia with Lewy bodies. *Neurology*, *64*(5), 861–865. doi:10.1212/01.WNL.0000153070.82309.D4 PMID:15753423

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The Aging Brain

Terribilli, D., Schaufelberger, M. S., Duran, F. L. S., Zanetti, M. V., Curiati, P. K., Menezes, P. R., ... Busatto, G. F. (2011). Age-related gray matter volume changes in the brain during non-elderly adulthood. *Neurobiology of Aging*, *32*(2-6), 354–368. doi:10.1016/j.neurobiolaging.2009.02.008 PMID:19282066

Tyler, M., Danilov, Y., Bach-y-Rita, P., & Bach-y-Rita, J. (2007). Systems and methods for altering brain and body functions and for treating conditions and diseases of the same. Google Patents.

Wallace, D. C. (2010). Mitochondrial DNA mutations in disease and aging. *Environmental and Molecular Mutagenesis*, *51*(5), 440–450. PMID:20544884

Wang, W., & Roberts, C. J. (2010). Aggregation of therapeutic proteins. John Wiley & Sons. doi:10.1002/9780470769829

Ward, R. J., Zucca, F. A., Duyn, J. H., Crichton, R. R., & Zecca, L. (2014). The role of iron in brain ageing and neurodegenerative disorders. *Lancet Neurology*, *13*(10), 1045–1060. doi:10.1016/S1474-4422(14)70117-6 PMID:25231526

Wen, W., & Sachdev, P. (2004). The topography of white matter hyperintensities on brain MRI in healthy 60-to 64-year-old individuals. *NeuroImage*, 22(1), 144–154. doi:10.1016/j.neuroimage.2003.12.027 PMID:15110004

Wikgren, M., Maripuu, M., Karlsson, T., Nordfjäll, K., Bergdahl, J., Hultdin, J., ... Adolfsson, R. (2012). Short telomeres in depression and the general population are associated with a hypocortisolemic state. *Biological Psychiatry*, *71*(4), 294–300. doi:10.1016/j.biopsych.2011.09.015 PMID:22055018

Wilkie, S., van Schalkwyk, M. C., Hobbs, S., Davies, D. M., van der Stegen, S. J., Pereira, A. C., ... Maher, J. (2012). Dual targeting of ErbB2 and MUC1 in breast cancer using chimeric antigen receptors engineered to provide complementary signaling. *Journal of Clinical Immunology*, *32*(5), 1059–1070. doi:10.100710875-012-9689-9 PMID:22526592

Wilkinson, C. W., Peskind, E. R., & Raskind, M. A. (1997). Decreased hypothalamic-pituitary adrenal axis sensitivity to cortisol feedback inhibition in human aging. *Neuroendocrinology*, 65(1), 79–90. doi:10.1159/000127167 PMID:9032777

Wolf, O. T., Convit, A., de Leon, M. J., Caraos, C., & Qadri, S. F. (2002). Basal hypothalamo-pituitaryadrenal axis activity and corticotropin feedback in young and older men: Relationships to magnetic resonance imaging-derived hippocampus and cingulate gyrus volumes. *Neuroendocrinology*, 75(4), 241–249. doi:10.1159/000054715 PMID:11979054

Yu, L., Xie, B., Yin, X., Liang, M., Evans, A. C., Wang, J., & Dai, C. (2013). Reduced cortical thickness in primary open-angle glaucoma and its relationship to the retinal nerve fiber layer thickness. *PLoS One*, *8*(9), e73208. doi:10.1371/journal.pone.0073208 PMID:24019910

Zheng, C. Y., Seabold, G. K., Horak, M., & Petralia, R. S. (2011). MAGUKs, synaptic development, and synaptic plasticity. *The Neuroscientist*, *17*(5), 493–512. doi:10.1177/1073858410386384 PMID:21498811

Zhou, Q., Homma, K. J., & Poo, M. (2004). Shrinkage of dendritic spines associated with long-term depression of hippocampal synapses. *Neuron*, 44(5), 749–757. doi:10.1016/j.neuron.2004.11.011 PMID:15572107

Chapter 2 Oxidative Stress and Neurodegeneration

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ABSTRACT

Neurons are the building units of the nervous system and are therefore critical units for the health of the brain and the spinal cord. This is necessitated by their inability to be either replaced or reproduced once lost. Their losses are implicated in a number of conditions which have been elaborated in this chapter. Oxidative stress has been strongly implicated in neurodegeneration through blockade of neuroprotection by a number of mechanisms including inhibitory effect on insulin-like growth factor I (IGF-1) via stimulation of the transcription factor, Forkhead box O3 (FOXO3). This chapter elaborates on these two phenomena which cannot be decoupled.

DOI: 10.4018/978-1-5225-5282-6.ch002

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INTRODUCTION

Neurodegenerative disorders (NDs) are a group of pathological disorders that primarily affect neurons and are associated with progressive loss of neuronal structure and function, and death (Khan et al, 2016). Neurodegeneration is manifested in a number of disorders including Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), amyotrophic lateral sclerosis (ALS), prion disease, ophthalmological diseases, etc (Khan et al, 2016; Tezel, 2006).

Neuronal loss is triggered by a number of mechanisms which include inflammation and oxidative stress (Harris, 2014; Barreto et al, 2011; Cabezas et al, 2012). Protein misfolding, compromised chronic oxidative stress response, excitotoxicity, altered calcium homeostasis, reduced cerebral blood flow and supply, impaired phosphorylation, loss of gene expression regulating ability, changes in proteases/inhibitors and environmental factors also play a role in neuronal loss (Khan et al, 2016; Sayre et al, 2001; Leszek et al, 2016).

Oxidative stress has also been implicated aetiologically in the development of neurodegeneration in other conditions such as HD, a trinucleotide CAG repeat expansion polyglutamate toxicity disorder (Hensley et al, 2006), and ALS (Polidori et al, 1999). Despite the huge hereditary and biochemical confirmation of increased oxidative stress in NDs, and also the established therapeutic benefits of antioxidants in animal models, there is no success with clinical use of antioxidants in human subjects. Accordingly, the role of oxidative stress in the pathogenesis of NDs stays disputable.

A number of causalities have been attributed to the development of neurodegeneration. Such factors include environmental and genetic causes; however, oxidative stress continues to be ascribed to play a focal pathogenic role in their development (Mark, 2004). Reactive oxygen species (ROS) and reactive nitrogen species (RNS) are generated in vivo as a result of oxidative stress and free radical generation. These two groups of reactive metabolites are believed to play a significant role in neurodegeneration (Emerit & Edeas, 2004). ROS are known to have significant impact on the neuronal biochemical composition. This is due to the high levels of unsaturated lipids in these neuronal biochemical molecules and their susceptibility to peroxidation and oxidative alterations (Uttara et al, 2009). Butterfield et al (2002) has also reported on the susceptibility of the double bonds in unsaturated fatty acids to free radical attack. This triggers a cascade of events that ultimately lead to the damage of neighbouring unsaturated fatty acids. Also, the brain tends not to have high levels of antioxidants to mop up free radicals when compared to other internal organs such as the liver (Uttara et al, 2009). Marklund et al (1982) has reported on the brain's reduced levels of catalase. This, coupled with the fact that the substantia nigra (SN) is enriched with iron and dopamine, both pro-oxidants, tend to increase the levels of reactive species in the brain (Zaleska et al, 1989). This chapter therefore discusses the relationship between oxidative stress and the development of NDs.

BACKGROUND

The pathological signature of NDs is the gradual, progressive and selective loss of anatomically or physiologically related neural systems (Leszek et al, 2016). With the loss of neuronal architecture in these multiple circuits comes the altered integrity of neuronal tissues and irreversible loss of brain function. This is mostly seen as cognitive deficits, dementia, dyskinesia, behaviour deviations and psychological disorders (Garcia et al, 2017). Research has suggested the significant relation of these disorders at the subcellular level. This is encouraging finding which would help with the discovery of novel diagnostic and therapeutic potentials for NDs. Some established parallel pathways between various NDs include atypical protein assemblage and induced apoptosis; and these span from systemic to molecular levels. This points to the multi-level nature of neurodegeneration in neuronal circuits (Bredesen et al, 2006).

AD and PD are neuropathologically characterized by abundant insoluble protein deposits e.g., amyoid β peptides (A β [1-40/42]) and hyperphosphorylated tau in AD (Jimenez-Del-Rio & Velez-Pardo, 2012) α - Synuclein in PD (Forno, 1996), specific neuronal and synaptic loss of the hippocampal pyramidal neurons (AD), and substantia nigra dopaminergic neurons (PD), probably via oxidative stress (Wang & Michaelis, 2010). There are over 200 known pathogenic mutations among the three genes that encode for the proteins: A β amyloid precursor protein (APP), presenilin-1 (PSEN1), presenilin-2 (PSEN2), and the six genes that encode for the proteins: α -synuclein (*SNCA*), Leucine-rich repeat kinase 2 (*LRRK2*), *Parkin*, PTEN-induced putative kinase 1 (*PINK1*), *DJ-1*, and P-type ATPase 13A2 (*ATP13A2*). Both Bekris et al (2010a) and Bekris et al (2010b) have reported on the fact that these mutations have been established to play a significant role in the development of familial Alzheimer and Parkinsonism. Interestingly, these mutations are directly related to oxidative stress and mitochondrial dysfuntion (McCoy & Cookson, 2012). In AD, the conformational changes in A β leads to the accumulation of toxic fibrillary A β aggregates. Oxidative stress also causes tau hyperphosphorylation and aggregation (Van Raamsdonk et al, 2017).

There is an increased oxidative stress via mitochondrial dysfunction and this is accounted for by multiple genetic risk factors for AD. These genetic factors include apolipoprotein E allele APOE E4, CD2-associated protein (*CD2AP*), translocase of outer mitochondrial membrane 40 (*TOMM40*) and translocator protein (*TSPO*). Again, Van Raamsdonk et al (2017) identified other risk factors for AD which were the variant genes that should play protective roles against oxidative stress such as clusterin (*CLU*), glutathione S-transferase omega 2 (*GSTO2*), mitochondrial fission regulator 1 (*MTFR1*) and methionine sulfoxide reductase B3 (*MSRB3*). Neuropathological findings identify the involvement of SN depletion of dopaminergic neurons, with an associated decrease in striatal dopamine and formation of intraneuronal alpha-synuclein aggregates in most parts of the brain, to be a major contributing factor to the late-stage PD (Sanders et al, 2014). Although several studies point to the possible therapeutic benefits of antioxidant therapies in improving the pathogenic mechanism of PD, these therapies are yet to be translated into slowing of neurodegeneration in PD clinically (Van Raamsdonk et al, 2017).

OXIDATIVE STRESS AND INFLAMMATION IN THE BRAIN

Molecular damages arise from increased biochemical imbalances that result in enhanced oxidative stress. These molecular alterations can lead to significant failure in biological responses of cells and culminate in cell death (Burxer, 1974; Golikov, 1985). ROS are generated from imbalances in biochemical processes in almost all cellular components which include the cytoplasm, endoplasmic reticulum, lysosomes, nucleus, plasma membrane, peroxisomes, and mitochondria by the actions of enzymes like oxidases, peroxidases, dehydrogenases, lipoxygenases, and cyclooxygenases (Drose et al, 2014). Through the electron carriers of the respiratory chain, the mitochondria tend to play a significant role in the production of ROS hence its importance in neurodegeneration (Lenaz, 1998).

Mitochondrial oxidative damage has been described as crucial in the pathogenesis of AD as evidenced by reduced brain metabolism preceding the development of abnormalities in neuropsychological testing, thus suggesting a casual role of impaired brain metabolism in Alzheimer's pathogenesis (Meraz-Rios et al, 2014; de la Monte et al, 2014). Aluminium neurotoxicity has been attributed to its interaction with peroxisome proliferator activated receptor gamma co-activator- 1α (PGC- 1α) hence a potential target in the prevention of mitochondrial-dependent neurodegeneration observed in NDs (Sharma et al, 2013). Some neurotoxic agents also cause damage to mitochondria via their ability to damage dopamine-producing cells found in the SN (Schuler & Casida, 2001).

Mitochondria is a target of most of the disease specific pathogenic mutant proteins and therefore account for the promotion of oxidative stress and the mitochondrial apoptotic pathway (Ferreiro et al, 2012). The prevalence of NDs increases with aging and aging is now considered as one of the strongest risk factors for PD (Collier et al, 2011). Although de Oliveira et al (2012) and Rikans & Hornbrook (1997) have identified the neuroprotective effect of low levels of ROS, oxidative stress has an important relation with NDs. Nitric oxide, in the presence of superoxide, forms a reactive peroxynitrite which causes cellular and tissue damage which implicates it in several diseases, including atherosclerosis, stroke, and AD (Mao & Reddy, 2010; Shin et al, 2005).

In the pathology of neurodegenerative disorders, the generation of free radicals are quite harmful as they affect nucleic acids, lipids, and proteins (Valco et al, 2006). Studies increasingly point to the involvement of oxidative stress in the development of prion diseases; these are fatal NDs. Henceforth, oxidative stress-induced mitochondrial and neuronal cell damages are suggestive of mitochondrial damage and dysfunction associated with prion disease progression. Acquatella-Tran et al (2013) attribute this to the initial prion disease development.

Although inflammation and oxidative stress are distinct biochemical processes, there is a close relation between them and they function in parallel in the brain due to its high susceptibility to oxidative stress (Leszek et al, 2016). Despite the fact that studies on the linkage between inflammation and oxidative stress are not conclusive, their interaction which explains their tendency to occur together and enhance each other is known. Inflammation activates and triggers oxidative stress and also activated microglia produce ROS as defense mechanism against pathogens or their markers (Barreto et al, 2014) which when exceeds levels that can be contained by the cell's antioxidant capacity may lead to cellular and tissue damage (Lau et al, 2007). Oxidative stress can also enhance inflammation via the activation of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), a transcription factor which controls the expression of many genes including those involved in both acute and chronic inflammatory response (Avila et al, 2014).

GLUTAMATERGIC AND MITOCHONDRIAL INHIBITION IN OXIDATIVE STRESS-INDUCED NEURODEGENERATION

There is a linkage between the glutamatergic mechanisms and the mitochondrial function. There is a close association between glutamatergic alterations and mitochondrial impairment with each feeding the other in a vicious cycle where they tend to strengthen each other in their induction of the onset and progression of NDs as reported by Cassano et al (2016).

Mitochondria

Mitochondria, which are the power house of cells, play integral roles in apoptotic mechanism, lipolysis, regulation of cells redox potential, free radical scavenging and cellular calcium and iron homeostasis under the tight regulation of nuclear and mitochondrial DNA (mtDNA), and associated proteins (Correia-Melo & Passos, 2015). The multiple cellular and metabolic activities they undertake make the mitochondria very susceptible. There is continual generation of ROS within the mitochondria as a result of oxidative phosphorylation occurring within them and can in turn damage their structure and functions as well as those of other organelles within the cell (Camara et al, 2010).

Brain hypoperfusion has been attributed to oxidative stress and mitochondrial failure while coenzyme Q and creatine have also been reported by Aliev et al (2011) to account for the protective effect on metabolism of energy in cerebral hypoperfusion. The mitochondrial dysfunction is particularly problematic in excitable cells, which have exceptional energy requirements, because there is an increase in ROS production, and oxidative damage is responsible for NDs (Chauhan et al, 2012). Mitochondrial dysfunction can be attributed to strenuous exercise, stress, environment toxicants, injury, aging and cancer (Hassan et al, 2013). Intrinsic mitochondrial apoptotic process is the most common form of cell death observed in neurodegeneration (Elmore, 2007). The functionality of the mitochondria is tightly linked with cell biology and this is because the mtDNA of mitochondria is replicated and stabilized by nuclear encoded proteins, as reported by de Moura et al (2010). Chatterjee et al (2011) also observed that ROS can cause direct mutation in mtDNA or can indirectly affect nuclear DNA upon its leakage from organelles.

A number of factors account for the effectiveness of the mitochondria and cell although they have high levels of antioxidant enzymes. These factors include age, stress, nutritional status and the level of immunity the individual possesses (Rahman, 2007). This makes mitochondrial dysfunction a contributing factor to the development of NDs. Sporadic AD has been ascribed to mitochondrial dysfunction (Garc et al, 2013) with amyloid accumulation associated with inherited AD (Tang & Gershon, 2003). Mutation in nuclear gene coding and dysfunction in electron transport chain (ETC) complex I are factors implicated in PD development (Winklhofer & Haass, 2010).

There is an indirect correlation between the increased numbers in CAG trinucleotide repeats (CAG repeats) and the dysfunction in the complex II which arises as a result of mtDNA damage by ROS (Damiano et al, 2010). Reddy & Reddy (2011) also reported that accumulation of mtDNa, perturbation in oxidative phosphorylation and a host of other mitochondrial abnormalities are hallmark of early and late onset NDs. Also histopathological findings establishing mitochondrial damage have also been realized in NDs (Kolesnikova, 2013). It is however worth noting that there has been new body of knowledge on the mechanistic capabilities of mitochondria which has opened alternative avenues of study in this regard (Solis-Herrera et al, 2015; Herrera et al, 2015)

Disordered functioning of the mitochondrial chaperones such as the heat shock protein (Hsp) 60/ Hsp10 complex accounts for the changes in the mitochondrial complexes seen in nervous system diseases such as AD and PD (Macario et al, 2010; Walls et al, 2012). There is a functional relationship between glutamate and the mitochondria (see Figure 1). Mitochondria play a significant role in regulating Ca^{2+} concentration and when there is impaired mitochondrial function, there is an increased cell death (Green & Kroemer, 2004). The uptake of Ca^{2+} by the mitochondria is of importance in the regulation of adenosine triphosphate (ATP) synthesis, the amplitude and spatiotemporal patterns of intracellular Ca^{2+} transients, the mitochondrial fission-fusion, dynamics the opening of mitochondrial permeability transition pores (mPTPs), and the generation of ROS (Gunter & Sheu, 2009; Drago et al, 2011). The knowledge of mitochondrial Ca^{2+} uptake has long been known but the nature of the transporters involved in the uptake were known relatively recently (Baughman et al, 2011). The mitochondrial Ca^{2+} uniporter (MCU) is believed to shuttle Ca^{2+} into the inner membranes of the mitochondria (Cassano et al, 2016)

During resting conditions, the mitochondrial Ca^{2+} content is low, but when a cell is stimulated with a cytosolic Ca^{2+} -increasing agonist (e.g., glutamate, histamine, ATP), the mitochondria accumulate high amounts of Ca^{2+} via the MCU complex (De Stefani et al, 2011; Somlyo et al, 1985). Other findings point to the fact that MCU controls excitotoxicity and is transcriptionally repressed by neuroprotective nuclear Ca^{2+} signals (Qui et al, 2013). In particular, MCU increases mitochondrial Ca^{2+} levels following N-methyl-D-aspartate (NMDA) receptor activation, with a resultant increase in membrane depolarization within the mitochondria which culminates and cell death. There is a reduction in the NMDA-induced increases in mitochondrial Ca^{2+} when the MCU expression is reduced endogenously with resultant decrease and mitochondrial depolarization and resistance to excitotoxicity.

Qiu et al (2013) has reported on the role of MCU on activity-dependent adaptive mechanism that limits mitochondrial Ca^{2+} overload when cytoplasmic Ca^{2+} levels are high. The mechanism entails the transcriptionally repressing MCU via synaptic activity involving Ca^{2+} and Ca^{2+} /calmodulin-dependent protein (CaM) kinase-mediated induction of neuronal PAS domain protein 4 (Npas4). This leads to inhibition of NMDA receptor-induced mitochondrial Ca^{2+} uptake and eventually cell death due to excitotoxicity (Qui et al, 2013). Npas4 can promote negative gene regulation in neurons, and it is an immediate-early gene (Lin et al, 2008).

Several key pro-death pathways are activated in parallel in response to an excitotoxic insult, including NO and ROS production, oxidative stress, *c*-Jun N-terminal kinase (JNK) and poly (ADP-ribose) polymerase- 1 (PARP-1) activation, and cAMP response element-binding protein (CREB) shut-off (Lau & Tymianski, 2010; Borsello et al, 2003). How significant these are hinders on the cell type, stage of development and severity of cell damage. Hopefully, further findings on the MCU-driven Ca²⁺ uptake will elaborate on the number of excitotoxic pathways. There have been reported role of other mitochondrial Ca²⁺ uptake mechanisms although their molecular identities and specific roles in the Ca²⁺ transport cannot be ascertained. Among these, two other Ca²⁺-transporting proteins, mitochondrial ryanodine receptor type 1 and leucine-zipper-EF-hand–containing transmembrane protein 1, have been investigated although further studies on them need to be done (O-Uchi et al, 2012).

Glutamate

As the most common excitatory neurotransmitter, glutamate plays important roles both in physiological and pathological brain function including the regulation of neurogenesis, neurite outgrowth, synaptogenesis, and programmed cell death (apoptosis) (Mattson, 2008; Shepherd & Huganir, 2007; Martin et al, 2000). There are both ionotropic and metabotropic glutamate receptors (Stayte & Vissel, 2014). The ligand-gated ionotropic glutamate receptors (iGluRs) permit the flow of ions through its Na⁺ and/ or Ca²⁺ membranes once they are activated. NMDA, alpha-amino-3-hydroxyl-4-isoxazolepropionic acid (AMPA) and Kainate subtypes belong to this family of iGluRs (Hollmann & Heinemann, 1994; Wright & Vissel, 2012; Wiltgen et al, 2010).

However, metabotropic glutamate receptors (mGluRs) are G-protein coupled receptors. Upon ligand binding to these receptors, there is an initiation of a series of second messenger mediated intracellular reactions, which are involved in the signal transduction (Conn et al, 2005; Kuwajima et al, 2007). Brain levels of glutamate are very high, about 10 mM, but in the extracellular spaces of the brain the concen-

trations of glutamate are almost ten thousand times lower (1-5 μ M), due to the role of the excitatory amino acid carrier 1 (EAAC1), glutamate transporter 1 (GLT1) and glutamate/aspartate transporter (GLAST). EAAC1 is present in the brain and works as a neuronal glutamate reuptake. GLT1, which is mainly expressed in astrocytes, is the most important carrier when it comes to keeping glutamate levels low in the extracellular spaces. GLAST is mainly activated when there is high glutamate levels and they are mainly expressed in glial cells (Niciu et al, 2012).

The metabolic relationship between astrocytes and neurons is critical for energy metabolism as well as for the synthesis of neurotransmitters. During the glutamatergic transmission, the arrival of an action potential causes a depolarization of the membrane, with consequent opening of Ca^{2+} channels and glutamate release into the synaptic cleft. Thereafter, the glutamate binds to its receptors and it is transported into the astrocytes through the GLT1, where it is amidated to glutamine in the glutamine synthetase pathway. Glutamine is metabolized to glutamate, indirectly generating gamma-aminobutyric acid (GABA) as well as the tricarboxylic acid (TCA) cycle intermediate, alpha-ketoglutarate (α -KG) in the neurons (Dienel, 2013; Cooper, 2012). Elevated extracellular glutamate levels can cause neuronal death in a phenomenon called "excitotoxicity" and it is involved in many neurological diseases in the CNS (Atlante et al, 2001; Yang et al, 2011).

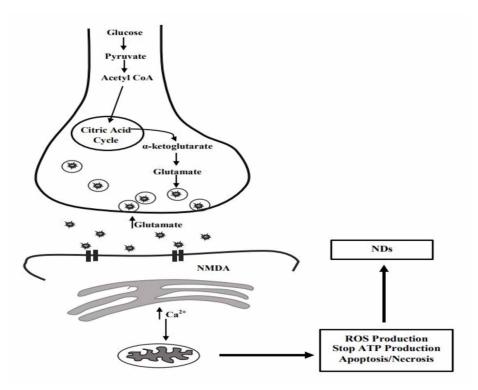
Calcium ion overload accounts for the glutamate-induced excitoxicity. This is attributed to glutamate binding to NMDA receptors (NMDARs), and to a minor extent binding to other receptor subtypes (Yang et al, 2011; Nicholls & Budd, 2000). Other studies also link glutamate overstimulation to the excitotoxicity. Changes in the activation pattern of NMDARs at different subcellular locations have been proposed as critical in activating pathways leading to neuroprotection versus neurodestruction (Hardingham & Bading, 2010). NMDARs are also found at extrasynaptic sites (Clark et al, 1997), raising the possibility that the synaptic and extrasynaptic subsets of NMDARs play different physiological and pathological roles in the cell (Figure 1). In line with this, there is available evidence the role of differential activation of extrasynaptic relative to synaptic NMDARs in the development of glutamatergic neurotoxicity (Leveille et al, 2008; Xu et al, 2009).

There is activation of glutamate receptors (mainly NMDA) when there is increased concentration of glutamate. This leads to elevated Ca^{2+} levels within the mitochondria. Increased Ca^{2+} concentration contributes to impaired mitochondrial function, which triggers excitotoxic cell death via a number of connected pathways. Changes in the mitochondria such as increase in ROS, ATP synthesis failure contribute significantly to cellular disruption (Adapted from Cassano et al, 2016)

Extreme actuation of extrasynaptic pathways particularly initiates apoptotic signal transduction cascade advancing neuronal cell death, while activation of synaptic NMDARs has a neuroprotective part by means of Ca²⁺-mediated signal transduction pathways advancing neuronal survival. The dichotomous neuroprotective and neurotoxic effects induced by initiation of the synaptic and extrasynaptic NMDARs, respectively, are mediated by complex regulatory activities on protective (anti-apoptotic, pro-survival and anti-oxidant) and pro-apoptotic genes (Papadia et al, 2008). Strangely, the noncompetitive NMDAR antagonist, memantine, shows differential consequences for synaptic and extrasynaptic NMDARs (Chen and Lipton, 2006). At low concentrations, memantine does not concentrate in the synaptic cleft to antagonize synaptic NMDRs; rather, it antagonizes extrasynaptic NMDARs, which spare their exposure to high levels of extracellular glutamate in pathological states like neurodegenerative processes (Xia et al, 2010).

Oxidative Stress and Neurodegeneration

Figure 1. Role of Ca²⁺ in the glutamatergic transmission



OXIDATIVE STRESS AND ALZHEIMER'S DISEASE

Oxidative Stress and Amyloid Beta Accumulation in Alzheimer's Disease

There is a convincing proof of oxidative damage in AD brain after death, with critical collection of markers of lipid, protein and DNA oxidation, and transition metals which include copper and zinc within the sight of impaired antioxidant defense (Butterfield et al, 2013). Redox proteomics investigation of after death AD brain shows oxidative harm to key proteins associated with energy metabolism, neurotransmitter-related proteins, mitochondrial proteins and proteasomal components (Butterfield et al, 2014). The cross talk between oxidative stress and A β proteinopathy may happen by means of numerous ways influencing transcription of the *APP* gene or translation of APP messenger RNA (mRNA), processing and degradation of APP and A β peptides and also associations of APP with transition metals. The promoter locales of the *APP* gene have been mapped in various species, and a few translation factors, for example, heat shock factor 1 (HSF-1) and NF- κ B, which are receptive to ROS, can bind and play a role in *APP* gene induction (Theuns and Van Broeckhoven, 2014). APP mRNA contains a 5'-untranslated district (UTR) stem–loop order of the iron-responsive element (IRE), where the IRE-binding protein (IREBP) downregulates translation. There is a reported elevation of iron levels in AD brains which points to a link between oxidative stress and APP production (Huang et al, 2004).

The major processing enzymes of APP in the amyloidogenic pathway are β -secretase (BACE1) and γ -secretase (Dong et al, 2012). The effect on gene expression of BACE1 by the transcription factors

specificity protein 1 (Sp1), NF- κ B, and hypoxia-inducible factor 1-alpha (HIF-1 α), is often very complex, and may result in both up- and downregulation of *BACE1* gene depending on the cell types and the physiological or experimental conditions of the cells (Chen et al, 2013). A number of factors account for the varied response post transcription factors binding. Example, NF- κ B's interaction with ROS is complex and could result in either activation or enhanced nuclear translocation under specific experimental conditions or decreased nuclear binding in others. In the same way, the nature of the subunits binding to the NF- κ B binding sites may determine whether up- or downregulation of *BACE1* gene occurs (Kaur et al, 2015). Then again, there is evidence of an increase in the enzyme activity through stress-activated protein kinase from 4-hydroxynonenal (4-HNE) or oxidants (such as H₂O₂ and iron–ascorbate mixture, which are products of oxidative damage (Ganguly et al, 2017).

Also Ganguly et al (2017) has reported on the ability of oxidative stress to upregulate BACE1 activity at the translational level involving double-stranded RNA-dependent protein kinase (PKR) and eukaryotic initiation factor-2 (eIF2) phosphorylation. These findings give credence to the role of oxidative stree in increasing the activity of BACE1 in AD (Ganguly et al, 2017). From a study which elaborated on this, it was realized that oxidative stress-induced increase in BACE1 activity both in cultured cells and in experimental animals requires the involvement of γ -secretase and activation of the JNK/c-jun pathway. The γ -secretase enzyme complex, which is made up of presenilin1 (PS1), nicastrin, PEN2 and APH1, is also essential for the release of A β 42 from APP, and its activity is heightened by oxidative stress through upregulation of PS1 (Tamagno et al, 2008).

The elevated levels of $A\beta42$ have been linked with oxidative stress in some studies and this happening is attributed to the expression and processing of APP (Misonou et al, 2000). Also, low-density lipoproteins receptor-related protein 1 (LRP1) and the receptor for advanced glycation end products (RAGE) are believed to inhibit the clearance of $A\beta$ in AD due to the effect of oxidative stress (Ramanathan et al, 2015). There are two forms of LRP1; the membrane-bound form is responsible for the removal of cerebral amyloid beta to the circulation across the blood–brain barrier and is expressed in neurons, astrocytes, vascular endothelial cells and smooth muscle cells, while the soluble form binds the $A\beta$ in the peripheral circulation while RAGE is found in the blood-brain barrier and functions to permit reentry of $A\beta$ from the peripheral circulation (Deane et al, 2009). Ramanathan et al (2015) reports that there are changes in the membrane-bound LRP1 when there is oxidative stress and this goes a long way to impair the clearance of $A\beta$ from the brain. Again, failure to properly bind to circulating $A\beta$, will lead to reentry into the brain.

A β is known to also increase the intracellular production of ROS and also contribute to neuronal death via its involvement with ASK 1 (Kadowaki et al, 2005). This A β -induced ROS production is believed to be due to its activation of reduced nicotinamide adenine dinucleotide phosphate (NADPH) oxidase. However, with mitochondrial production of oxygen radicals, mitochondria-targeted antioxidants have been shown to curtail their production (Hu & Li, 2016). An alternate mechanism of A β -induced ROS production is via activation and priming of microglial by soluble and fibrillary forms of the peptide. This is mainly seen in primary and co-culture of microglia and neurones (Qin et al, 2006).

With A β acting via a number of receptors, there is activation of microglial cells which leads to the production of ROS and proinflammatory cytokines such as interleukin (IL)-6, IL-1 β , tumor necrosis factor alpha (TNF- α) which triggers an inflammatory response in AD brain. This means that the microglia contain a B-type of scavenger receptor, called CD36 with studies showing that it acts as a receptor for fibrillar A β . The peptide-mediated activation of this receptor results in an increased microglial ROS production, cytokine expression and phagocytosis (Doens & Fernández, 2014). MAC-1 receptor and

phosphoinositide 3-kinase (PI3K) also regulate $A\beta$ -induced microglial activation and ROS production. Ganguly et al (2017) has reported on their ability to activate NADPH oxidase which functions mainly as a regulator of microglial-mediated ROS production.

Oxidative Stress and Tau Phosphorylation in Alzheimer's Disease

The development of neurofibrillary tangles from intracellular amassing of hyperphosphorylated tau protein is another trademark highlight of AD pathogenesis, which accounts for the axonal degeneration and synaptic dysfunction related with neurodegenerative disorder (Alavi Naini and Soussi-yanicostas, 2015). The microtubule-related tau protein has various phosphorylation sites in the proline-rich and in the C-terminal locales of the protein, and various kinases – particularly glycogen synthase kinase 3 beta (GSK3 β) and cyclin-subordinate kinase 5 (CDK5) – can phosphorylate the previous. Tau can be dephosphorylated by a few phosphatases, dominatingly protein phosphate 2A (PP2A), protein phosphate 1 (PP1) and protein phosphate 2B (PP2B) (Alavi Naini and Soussi-yanicostas, 2015). In AD, the expanded phosphorylation of tau protein is the aftereffect of expanded activities of GSK3 β and CDK5, and a related diminishing in PP2A function (Ganguly et al, 2017). Then again, explanations for such changes in the kinase functions and that of phosphatases in these disorders are not clear.

Chronic oxidative stress as glutathione depletion has appeared to build tau phosphorylation in cultured M17 neuroblastoma cells through JNK and p38 activation and in addition decreased phosphatase function (i.e. PP2A) (Su et al, 2010). Notwithstanding, there have been reports of dephosphorylation of tau under oxidative stress in various models of study through kinases modulation, (for example, GSK3 or CDK5) or phosphatases, (for example, PP1), making the relationship between oxidative stress and tau phosphorylation difficult to ascertain (Zambrano et al, 2004). Figure 2 outlines the different conceivable mechanisms through which oxidative stress prompts a gathering of amyloid β 42, which thus impedes mitochondrial work and interfaces with metal ions in microglia in an AD brain.

A β 42 accumulates as a result of increased expression of APP and BACE1 when there is increased oxidative stress. The clearance of A β 42 from the brain is also retarded by oxidative stress. On the other hand, the multiple interactions of A β 42 with mitochondria, microglia and metal ions lead to further oxidative stress. Arrows suggest interactions; a line with an end bar indicates retardation (Adapted from Ganguly et al, 2017).

Abbreviations: A β 42: Amyloid beta peptide 1–42; APP: Amyloid precursor protein; BACE1: β -Secretase; eIF2: Eukaryotic initiation factor-2; IRE: Iron-responsive element; LRP1: Low-density lipoprotein receptor-related protein 1; PKR: Double-stranded RNA-dependent protein kinase.

OXIDATIVE STRESS AND PARKINSON'S DISEASE

α-Synuclein and Oxidative Stress in Parkinson's Disease Pathogenesis

 α -Synuclein is an unfolded protein that can connect with vesicular and membranous structures and assumes a part in synaptic vesicle reusing, stockpiling and compartmentalization of neurotransmitters (Yavich et al, 2006). Fibrils of α -synuclein, residing in Lewy bodies, are related with expanded oxidative or nitrosative stress, hence keeping up physiologic levels of α -synuclein in neurons is important for neuronal survival (Paxinou et al, 2001). Chaperone-mediated autophagy (CMA) rids the cells off

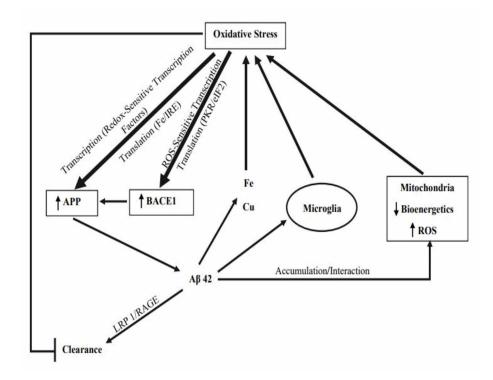


Figure 2. Oxidative stress-mediated amyloid beta proteinopathy in Alzheimer's brain

 α -synuclein and prevents its build up. Be that as it may, its debasement by CMA is impeded by posttranslational alterations (Cuervo and Wong, 2013). Over accumulation and post-translational alteration of α -synuclein result in death of DAergic neurons (Zhou et al, 2013). Oxidative stress effectively controls protein accumulation in PD as seen in the analyses of the brain after death, demonstrating expanded levels of post-translational alterations of the α -synuclein by oxidative stress, including those by 4-hydroxy-2nonenal (HNE- α -synuclein), nitration (n- α -synuclein), and oxidation (o- α -synuclein), which have been ensnared in advancement of oligomerization of α -synuclein (Alam et al, 1997). Among them, HNE- α -synuclein are more inclined to framing oligomers than unmodified α -synuclein.

The cellular toxicity of HNE- α -synuclein is significantly higher than other post-translationally modified species (Xiang et al, 2013). Mitochondrial dysfunction caused by α -synuclein and overexpression of wild or mutant α -synuclein in SH-SY5Y has been shown to increase the intracellular level of ROS (Di Mario et al, 2016). On the other hand, iron which facilitates ROS production and catalyzes peroxidative damage to biomolecules, accumulates in the SN of PD brains in excess amounts and causes translational upregulation of α -synuclein (Febbraro et al, 2012). Furthermore, the binding of transition metals such as iron and copper to α -synuclein has been studied with a variety of biophysical techniques in different studies, and apparently, such binding may lead to enhanced cytotoxicity of α -synuclein through multiple mechanisms. One of such mechanisms is iron-mediated formation of large sodium dodecyl sulfate (SDS)-resistant oligomers of α -synuclein on intracellular organelles (Carboni & Lingor, 2015). Since DAergic neurons are particularly affected in PD, it has been suggested that DA oxidation products such as ROS and toxic quinones could contribute to PD pathology (Ganguly et al, 2017). In this context, it is interesting to note that the interaction of α -synuclein with DA has been studied by several groups, indicating a modulatory role of DA oxidation products on α -synuclein oligomerization and cytotoxicity (Leong et al, 2009). In a very elaborate study, it has been shown that in human DAergic neurons and rat DAergic cell lines exposed to paraquat, increased accumulation and aggregation of α -synuclein occurs, which is crucially dependent upon the activity of NADPH oxidase, implicating the role of ROS in the process. This study has further shown that the systemic injection of paraquat in rats causes increased protein expression of α -synuclein and NADPH oxidase, along with the accumulation of oxidative damage markers in the SN, which can be abolished by knocking down NADPH oxidase (*NOX1*) gene by adeno-associated virus-mediated overexpression of a specific short hairpin RNA (shRNA) (Cristóvão et al, 2010).

The knockdown of *NOX1* likewise keeps the nigral DAergic neuronal misfortune after paraquat treatment of rodent. Another intriguing cross talk between oxidative stress and α -synuclein is likely when oxidatively changed protein turns out to be incompletely impervious to metabolism by the ubiquitinproteasome framework (UPS) or chaperone-intervened autophagy, prompting collection of the misfolded protein (Ganguly et al, 2017). A few sort of oxidative changes of α -synuclein have been illustrated, for example, nitration of tyrosine deposits, oxidation of methionine buildups and covalent adduct synthesis with 4-HNE. Such oxidatively adjusted α -synuclein proteins, when all is said and done, repress oligomerization and fibril formation by the native monomer, and rather may offer ascent to "off-pathway" oligomers, however the unwanted outcome of this has not been certain (Schildknecht et al, 2013). The different mechanistic approaches prompting the aggregation of α -synuclein in the PD mind and the harmfulness of this protein in monomeric or oligomeric structure in the mitochondria are featured in Figure 3.

Increase in *SNCA* and iron/IRE-regulated posttranscriptional mechanism accounts fot the increased expression of α -synuclein in the brains of people with PD. α -Synuclein (monomers and oligomers) have multiple interactions with mitochondria, causing dysfunction of the organelle and increasing ROS production while iron and DA-oxidation products also account for increased oxidative stress in PD brain. The arrows indicate interactions, but arrows with end bars suggest inhibition (Adapted from Ganguly et al, 2017).

Iron-Mediated Oxidative Stress in Parkinson's Disease

In neurodegenerative disorders such as PD and multiple system atrophy, there are aberrantly high levels of iron and oxidative stress (Repetto et al, 2012). Oxidative stress is normally accompanied by increased free iron. Experimentally identifying high intracellular iron does not essentially signal an increased oxidative stress if there are associated increases in proteins that store iron in redox inert forms such as the ferrihydrate core of ferritin (Perry et al, 2002). The entry and release of iron from ferritin occurs via its more coordinately labile ferrous state, active in Fenton generation of hydroxyl radical. Microglia, a major site of ferritin-bound iron, is one source of iron and might contribute to oxidative damage in PD and other neurodegenerative disorders through superoxide-dependent reductive release of ferritin iron, leading to lipid peroxidation, which might be due to Fenton chemistry (Sayre et al, 2001). Besides superoxide, ferritin iron can be released by 6-hydroxydopamine, a neurotoxin implicated in PD, and by other easily oxidized catechols (Sayre et al, 2001).

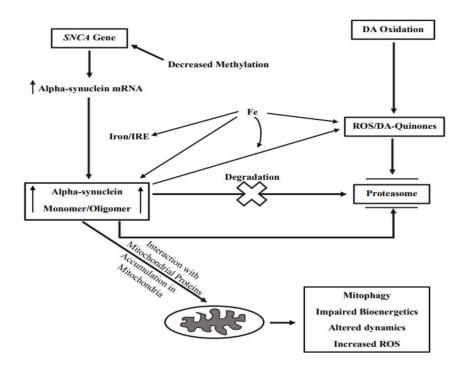
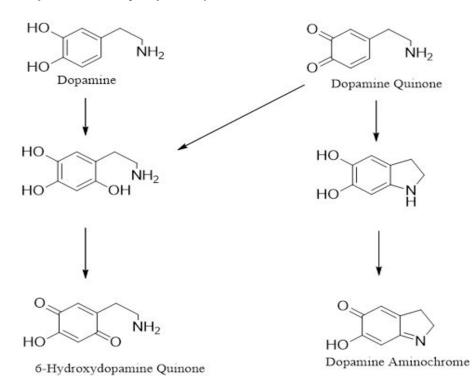


Figure 3. Accumulation of α -synuclein and its toxic role in the development of Parkinson's disease

Figure 4. Oxidation chemistry of dopamine showing the competition between aminochrome and 6-hydroxydopamine formation (Adapted from Sayre et al, 2001)



Sayre et al (2001) reports that sequestration of redox-active iron in the mitochondria of aging nigral astroglia may account for the iron buildup in astrocytes in SN old rats. This may predispose the senescent nervous system to PD (Sayre et al, 2001). Also, anomalous oxidation of dopamine (Figure 4) to 6-hydroxydopamine accounts for redox imbalances, and this will eventually result in autoxidation to its corresponding quinone with the generation of superoxide. In the presence of Fe²⁺ and either H₂O₂ or alkyl peroxide, dopamine is converted to 6-hydroxydopamine and 6,7-dihydroxy-1,2,3,4-tetrahydroisoquinolone, a neurotoxic alkaloid. These reaction pathways may become operative under conditions relevant to neurodegeneration (Sayre et al, 2001).

CONCLUSION

Neuronal proteins and structural components are modified due to oxidative stress in the different neurodegenerative disorders which culminates in neuroinflammation and loss of cognitive function as seen in the above discussed neurodegenerative disorders. As has been enshrined in this chapter, oxidative stress plays a key role in the pathological cause of neurodegeneration. Henceforth, antioxidants may play a significant role in therapeutic management of neurodegeneration upstream. This chapter has clearly discussed the link between neurodegeneration and oxidative stress and should inform future studies on the pathology as well as management of NDs.

REFERENCES

Acquatella-Tran, V. B. I., Imberdis, T., & Perrier, V. (2013). From prion diseases to prion-like propagation mechanisms of neurodegenerative diseases. *International Journal of Cell Biology*, 975832. PMID:24222767

Alam, Z. I., Jenner, A., Daniel, S. E., Lees, A. J., Cairns, N., Marsden, C. D., ... Halliwell, B. (1997). Oxidative DNA damage in the parkinsonian brain: An apparent selective increase in 8-hydroxyguanine levels in substantia nigra. *Journal of Neurochemistry*, 69(3), 1196–1203. doi:10.1046/j.1471-4159.1997.69031196.x PMID:9282943

Alavi Naini, S. M., & Soussi-yanicostas, N. (2015). Tau hyperphosphorylation and oxidative stress, a critical vicious circle in neurodegenerative tauopathies? *Oxidative Medicine and Cellular Longevity*, 151979. PMID:26576216

Aliev, G., Ashraf, G. M., Horecký, J., Vancova, O., Gvozdjakova, A., & Kucharská, J. ... Bachurin, S. (2011). Potential Preventive Effects of Coenzyme Q and Creatine Supplementation on Brain Energy Metabolism in Rats Exposed to Chronic Cerebral Hypoperfusion. In Systems Biology of Free Radicals and Antioxidants. Berlin: Springer.

Atlante, A., Calissano, P., Bobba, A., Giannattasio, S., Marra, E., & Passarella, S. (2001). Glutamate neurotoxicity, oxidative stress and mitochondria. *Federation of European Biochemical Societies (FEBS)*. *Letters*, 497(1), 1–5.

Avila, M. F., Torrente, D., Cabezas, R., Morales, L., García-Segura, L. M., Ganzalez, J., & George, E. B. (2014). Structural insights from GRP78-NF-kappaB binding interactions: A computational approach to understand a possible neuroprotective pathway in brain injuries. *Journal of Theoretical Biology*, *345*, 43–51. doi:10.1016/j.jtbi.2013.12.010 PMID:24361327

Barreto, G. E., Ganzalez, J., Torres, Y., & Morales, L. (2011). Astrocytic-neuronal crosstalk: Implications for neuroprotection from brain injury. *Neuroscience Research*, *71*(2), 107–113. doi:10.1016/j. neures.2011.06.004 PMID:21693140

Barreto, G. E., Santos-Galindo, M., & Garcia-Segura, L. M. (2014). Selective estrogen receptor modulators regulate reactive microglia after penetrating brain injury. *Frontiers in Aging Neuroscience*, *6*, 132. doi:10.3389/fnagi.2014.00132 PMID:24999330

Baughman, J. M., Perocchi, F., Girgis, H. S., Plovanich, M., Belcher-Timme, C. A., Sancak, Y., ... Mootha, V. K. (2011). Integrative genomics identifies MCU as an essential component of the mitochondrial calcium uniporter. *Nature*, 476(7360), 341–345. doi:10.1038/nature10234 PMID:21685886

Bekris, L. M., Mata, I. F., & Zabetian, C. P. (2010). The genetics of Parkinson disease. *Journal of Geriatric Psychiatry and Neurology*, 23(4), 228–242. doi:10.1177/0891988710383572 PMID:20938043

Bekris, L. M., Yu, C. E., Bird, T. D., & Tsuang, D. W. (2010). Review article: Genetics of Alzheimer disease. *Journal of Geriatric Psychiatry and Neurology*, 23(4), 213–227. doi:10.1177/0891988710383571 PMID:21045163

Borsello, T., Clarke, P. G., Hirt, L., Vercelli, A., Repici, M., Schorderet, D. F., ... Bonny, C. (2003). A peptide inhibitor of c-Jun N-terminal kinase protects against excitotoxicity and cerebral ischemia. *Nature Medicine*, *9*(9), 1180–1186. doi:10.1038/nm911 PMID:12937412

Bredesen, D. E., Rao, R. V., & Mehlen, P. (2006). Cell death in the nervous system. *Nature*, 443(7113), 796–802. doi:10.1038/nature05293 PMID:17051206

Burxer, G. V. (1974). Stress in farm animals. Veterinaria, 8, 92-94.

Butterfield, D. A., Castegna, A., Lauderback, C. M., & Drake, J. (2002). Evidence that amyloid β -peptide induced lipid peroxidation and its sequelae in Alzheimer's disease brain contribute to neuronal death. *Neurobiology of Aging*, 23(5), 655–664. doi:10.1016/S0197-4580(01)00340-2 PMID:12392766

Butterfield, D. A., Di Domenico, F., Swomley, A. M., Head, E., & Perluigi, M. (2014). Redox proteomics analysis to decipher the neurobiology of Alzheimer-like neurodegeneration: Overlaps in Down's syndrome and Alzheimer's disease brain. *The Biochemical Journal*, *463*(2), 177–189. doi:10.1042/BJ20140772 PMID:25242166

Butterfield, D. A., Swomley, A. M., & Sultana, S. (2013). Amyloid β -peptide (1–42)-induced oxidative stress in Alzheimer disease: Importance in disease pathogenesis and progression. *Antioxidants & Redox Signalling*, *19*(8), 823–835. doi:10.1089/ars.2012.5027 PMID:23249141

Cabezas, R., El-Bachá, R. S., González, J., & Barreto, G. E. (2012). Mitochondrial functions in astrocytes: Neuroprotective implications from oxidative damage by rotenone. *Neuroscience Research*, 74(2), 80–90. doi:10.1016/j.neures.2012.07.008 PMID:22902554 Camara, A. K. S., Lesnefsky, E. J., & Stowe, D. F. (2010). Potential therapeutic benefits of strategies directed to mitochondria. *Antioxidants & Redox Signalling*, *13*(3), 279–347. doi:10.1089/ars.2009.2788 PMID:20001744

Carboni, E., & Lingor, P. (2015). Insights on the interaction of alpha-synuclein and metals in the pathophysiology of Parkinson's disease. *Metallomics*, 7(3), 395–404. doi:10.1039/C4MT00339J PMID:25648629

Cassano, T., Pace, L., Bedse, G., Lavecchia, A. M., De Marco, F., Gaetani, S., & Gaetano, S. (2016). Glutamate and mitochondria: Two prominent players in the oxidative stress-induced neurodegeneration. *Current Alzheimer Research*, *13*(2), 185–197. doi:10.2174/1567205013666151218132725 PMID:26679860

Chatterjee, A., Dasgupta, S., & Sidransky, D. (2011). Mitochondrial Subversion in Cancer. *Cancer Prevention Research (Philadelphia, Pa.)*, 4(5), 638–654. doi:10.1158/1940-6207.CAPR-10-0326 PMID:21543342

Chauhan, A., Audhya, T., & Chauhan, V. (2012). Brain region-specific glutathione redox imbalance in autism. *Neurochemical Research*, *37*(8), 1681–1689. doi:10.100711064-012-0775-4 PMID:22528835

Chen, H. S., & Lipton, S. A. (2006, June). The chemical biology of clinically tolerated NMDA receptor antagonists. *Journal of Neurochemistry*, 97(6), 1611–1626. doi:10.1111/j.1471-4159.2006.03991.x PMID:16805772

Chen, X. F., Zhang, Y. W., Xu, H., & Bu, G. (2013). Transcriptional regulation and its misregulation in Alzheimer's disease. *Molecular Brain*, *6*(1), 44. doi:10.1186/1756-6606-6-44 PMID:24144318

Clark, B. A., Farrant, M., & Cull-Candy, S. G. (1997). A direct comparison of the single-channel properties of synaptic and extrasynaptic NMDA receptors. *The Journal of Neuroscience*, *17*(1), 107–116. PMID:8987740

Collier, T. J., Kanaan, N. M., & Kordower, J. H. (2011). Ageing as a primary risk factor for Parkinson's disease: Evidence from studies of non-human primates. *Nature Reviews. Neuroscience*, *12*(6), 359–366. doi:10.1038/nrn3039 PMID:21587290

Conn, P. J., Battaglia, G., Marino, M. J., & Nicoletti, F. (2005). Metabotropic glutamate receptors in the basal ganglia motorcircuit. *Nature Reviews. Neuroscience*, *6*(10), 787–798. doi:10.1038/nrn1763 PMID:16276355

Cooper, A. J. (2012). The role of glutamine synthetase and glutamate dehydrogenase in cerebral ammonia homeostasis. *Neurochemical Research*, *37*(11), 2439–2455. doi:10.100711064-012-0803-4 PMID:22618691

Correia-Melo, C., & Passos, J. F. (2015). Mitochondria: Are they causal players in cellular senescence? *Biochimica et Biophysica Acta*, *1847*(11), 1373–1379. doi:10.1016/j.bbabio.2015.05.017 PMID:26028303

Cristóvão, A. C., Guhathakurta, S., Bok, E., Je, G., Yoo, S. D., Choi, D.-H., & Kim, Y.-S. (2012). NADPH oxidase 1 mediates α -synucleinopathy in Parkinson's disease. *The Journal of Neuroscience*, *32*(42), 14465–14477. doi:10.1523/JNEUROSCI.2246-12.2012 PMID:23077033

Cuervo, A. M., & Wong, E. (2013). Chaperone-mediated autophagy: Roles in disease and aging. *Cell Research*, 24(1), 92–104. doi:10.1038/cr.2013.153 PMID:24281265

Damiano, M., Galvan, L., Déglon, N., & Brouillet, E. (2010). Mitochondria in Huntington's disease. *Biochimica et Biophysica Acta*, *1802*(1), 52–61. doi:10.1016/j.bbadis.2009.07.012 PMID:19682570

de la Monte, S. M., & Tong, M. (2014). Brain metabolic dysfunction at the core of Alzheimer's disease. *Biochemical Pharmacology*, 88(4), 548–559. doi:10.1016/j.bcp.2013.12.012 PMID:24380887

de Moura, M. B., dos Santos, L. S., & Van Houten, B. (2010). Mitochondrial dysfunction in neurodegenerative diseases and cancer. *Environmental and Molecular Mutagenesis*, *51*, 391–405. PMID:20544881

de Oliveira, D. M., Ferreira Lima, R. M., & El-Bachá, R. S. (2012). Brain rust: Recent discoveries on the role of oxidative stress in neurodegenerative diseases. *Nutritional Neuroscience*, *15*(3), 94–102. do i:10.1179/1476830511Y.0000000029 PMID:22583954

De Stefani, D., Raffaello, A., Teardo, E., Szabo, I., & Rizzuto, R. (2011). A forty kilodalton protein of the inner membrane is the mitochondrial calcium uniporter. *Nature*, *476*(7360), 336–340. doi:10.1038/ nature10230 PMID:21685888

Deane, R., Bell, R. D., Sagare, A., & Zlokovic, B. V. (2009). Clearance of amyloid-beta peptide across the blood-brain barrier: Implication for therapies in Alzheimer's disease. *CNS & Neurological Disorders* - *Drug Targets*, 8(1), 16–30. doi:10.2174/187152709787601867 PMID:19275634

Di Maio, R., Barrett, P. J., Hoffman, E. K., Barrett, C. W., Zharikov, A., Borah, A., ... Greenamyre, J. T. (2016). α-Synuclein binds to TOM20 and inhibits mitochondrial protein import in Parkinson's disease. *Science Translational Medicine*, 8(342), 342–378. doi:10.1126citranslmed.aaf3634 PMID:27280685

Dienel, G. A. (2013). Astrocytic energetics during excitatory neurotransmission: What are contributions of glutamate oxidation and glycolysis? *Neurochemistry International*, *63*(4), 244–258. doi:10.1016/j. neuint.2013.06.015 PMID:23838211

Doens, D., & Fernández, P. L. (2014). Microglia receptors and their implications in the response to amyloid β for Alzheimer's disease pathogenesis. *Journal of Neuroinflammation*, 11(1), 48. doi:10.1186/1742-2094-11-48 PMID:24625061

Dong, S., Duan, Y., Hu, Y., & Zhao, Z. (2012). Advances in the pathogenesis of Alzheimer's disease: A re-evaluation of amyloid cascade hypothesis. *Translational Neurodegeneration*, *1*(1), 18. doi:10.1186/2047-9158-1-18 PMID:23210692

Drago, I., Pizzo, P., & Pozzan, T. (2011). After half a century mitochondrial calcium in- and efflux machineries reveal themselves. *European Molecular Biology Organization Journal*, *30*(20), 4119–4125. doi:10.1038/emboj.2011.337 PMID:21934651

Drose, S., Brandt, U., & Wittig, I. (2014). Mitochondrial respiratory chain complexes as sources and targets of thiol-based redox-regulation. *Biochimica et Biophysica Acta*, *1844*(8), 1344–1354. doi:10.1016/j. bbapap.2014.02.006 PMID:24561273

Elmore, S. (2007). Apoptosis: A Review of Programmed Cell Death. *Toxicologic Pathology*, *35*(4), 495–516. doi:10.1080/01926230701320337 PMID:17562483

Emerit, J. M., Edeas, F. B., & Bricaire, F. (2004). Neurodegenerative diseases and oxidative stress. *Biomedicine and Pharmacotherapy*, 58(1), 39–46. doi:10.1016/j.biopha.2003.11.004 PMID:14739060 Febbraro, F., Giorgi, M., Caldarola, S., Loreni, F., & Romero-ramos, M. (2012). α -Synuclein expression is modulated at the translational level by iron. *Neuroreport*, 23(9), 576–580. doi:10.1097/WNR.0b013e328354a1f0 PMID:22581044

Fernández-Checa, J. C., Fernández, A., Morales, A., Mari, M., Garcia-Ruiz, C., & Colell, A. (2010). Oxidative stress and altered mitochondrial function in neurodegenerative diseases: Lessons from mouse models. *CNS & Neurological Disorders - Drug Targets*, *9*(4), 439–454. doi:10.2174/187152710791556113 PMID:20522012

Forno, L. S. (1996). Neuropathology of Parkinson's disease. *Journal of Neuropathology and Experimental Neurology*, 55(30), 259–272.

Ganguly, G., Chakrabarti, S., Chatterjee, U., & Saso, L. (2017). Proteinopathy, oxidative stress and mitochondrial dysfunction: Cross talk in Alzheimer's disease and Parkinson's disease. *Drug Design, Development and Therapy*, *11*, 797–810. doi:10.2147/DDDT.S130514 PMID:28352155

Garcia, G., Nanni, S., Figueira, I., Ivanov, I., McDougall, G. J., Stewart, D., ... Santos, C. N. (2017). Bioaccessible (poly)phenol metabolites from raspberry protect neural cells from oxidative stress and attenuate microglia activation. *Food Chemistry*, *215*, 274–283. doi:10.1016/j.foodchem.2016.07.128 PMID:27542476

García-Escudero, V., Martín-Maestro, P., Perry, G., & Avila, J. (2013). Deconstructing mitochondrial dysfunction in Alzheimer disease. *Oxidative Medicine and Cellular Longevity*. PMID:23840916

Golikov, N. (1985). Adaptation in Farm Animals. Sofia, Bulgaria: Agropromizdat.

Green, D. R., & Kroemer, G. (2004). The pathophysiology of mitochondrial cell death. *Science*, *305*(5684), 626–629. doi:10.1126cience.1099320 PMID:15286356

Gunter, T. E., & Sheu, S. S. (2009). Characteristics and possible functions of mitochondr ial Ca²⁺ transport mechanisms. *Biochimica et Biophysica Acta*, *1787*(11), 1291–1308. doi:10.1016/j.bbabio.2008.12.011 PMID:19161975

Hardingham, G. E., & Bading, H. (2010). Synaptic Versus Extrasynaptic NMDA Receptor Signalling: Implications for Neurodegenerative Disorders. *Nature Reviews. Neuroscience*, *11*(10), 682–696. doi:10.1038/nrn2911 PMID:20842175

Harris, R. A. (2014). Spatial, temporal, and functional aspects of macrophages during "the good, the bad, and the ugly" phases of inflammation. *Frontiers in Immunology*, *5*, 612. doi:10.3389/fimmu.2014.00612 PMID:25520719

Hassan, I., Chibber, S., Khan, A. A., & Naseem, I. (2013). Cisplatin-induced neurotoxicity *in vivo* can be alleviated by riboflavin under photoillumination. *Cancer Biotherapy & Radiopharmaceuticals*, 28(2), 160–168. doi:10.1089/cbr.2012.1312 PMID:23215961

Hensley, K., Mhatre, M., Mou, S., Pye, Q. N., Stewart, C., West, M., & Williamson, K. S. (2006). On the relation of oxidative stress to neuroinfl ammation: Lessons learned from the G93A-SOD1 mouse model of amyotrophic lateral sclerosis. *Antioxidants & Redox Signalling*, 8(11-12), 2075–2087. doi:10.1089/ars.2006.8.2075 PMID:17034351

Herrera, A. S., Del, C. A., Esparza, M., Ashraf, M. G., Zamyatnin, A. A., & Aliev, G. (2015). Beyond mitochondria, what would be the energy source of the cell? *Central Nervous System Agents in Medicinal Chemistry*, *15*(1), 32–41. doi:10.2174/1871524915666150203093656 PMID:25645910

Hollmann, M., & Heinemann, S. (1994). Cloned glutamate receptors. *Annual Review of Neuroscience*, *17*(1), 31–108. doi:10.1146/annurev.ne.17.030194.000335 PMID:8210177

Hu, H., & Li, M. (2016). Mitochondria-targeted antioxidant mitotempo protects mitochondrial function against amyloid beta toxicity in primary cultured mouse neurons. *Biochemical and Biophysical Research Communications*, 478(1), 174–180. doi:10.1016/j.bbrc.2016.07.071 PMID:27444386

Huang, X., Moir, R. D., Tanzi, R. E., Bush, A. I., & Rogers, J. T. (2004). Redox-active metals, oxidative stress, and Alzheimer's disease pathology. *Annals of the New York Academy of Sciences*, *1012*(1), 153–163. doi:10.1196/annals.1306.012 PMID:15105262

Jimenez-Del-Rio, M., & Velez-Pardo, C. (2012). The bad, the good, and the ugly about oxidative stress. *Oxidative Medicine and Cellular Longevity*, 2012, 1–13. doi:10.1155/2012/163913 PMID:22619696

Kadowaki, H., Nishitoh, H., Urano, F., Sadamitsu, C., Matsuzawa, A., Takeda, K., ... Ichijo, H. (2005). Amyloid β induces neuronal cell death through ROS-mediated ASK1 activation. *Cell Death and Differentiation*, *12*(1), 19–24. doi:10.1038j.cdd.4401528 PMID:15592360

Kaur, U., Banerjee, P., Bir, A., Sinha, M., Biswas, A., & Chakrabarti, S. (2015). Reactive oxygen species, redox signaling and neuroinflammation in Alzheimer's disease: The NF-κB connection. *Current Topics in Medicinal Chemistry*, *15*(5), 446–457. doi:10.2174/1568026615666150114160543 PMID:25620241

Keller, J. N., Schmitt, F. A., Scheff, S. W., Ding, Q., Chen, Q., Butterfield, D. A., & Markesbery, W. R. (2005). Evidence of increased oxidative damage in subjects with mild cognitive impairment. *Neurology*, *64*(7), 1152–1156. doi:10.1212/01.WNL.0000156156.13641.BA PMID:15824339

Khan, T. A., Hassan, I., Ahmad, A., Perveeen, A., Aman, S., Quddusi, S., ... Aliev, G. (2016). Recent updates on the dynamic association between oxidative stress and neurodegenerative disorders. *CNS & Neurological Disorders - Drug Targets*, *15*, 310–320. doi:10.2174/1871527315666160202124518 PMID:26831262

Kolesnikova, E. É. (2013). Mitochondrial dysfunction and molecular bases of neurodegenerative diseases. *Neurophysiology*, *45*(1), 89–102. doi:10.100711062-013-9341-1

Kuwajima, M., Dehoff, M. H., Furuichi, T., Worley, P. F., Hall, R. A., & Smith, Y. (2007). Localization and expression of group I metabotropic glutamate receptors in the mouse striatum, globus pallidus, and subthalamic nucleus: Regulatory effects of MPTP treatment and constitutive Homer deletion. *The Journal of Neuroscience*, *27*(23), 6249–6260. doi:10.1523/JNEUROSCI.3819-06.2007 PMID:17553998

Lau, A., & Tymianski, M. (2010). Glutamate receptors, neurotoxicity and neurodegeneration. *Pflügers* Archiv, 460(2), 525–542. doi:10.100700424-010-0809-1 PMID:20229265

Lau, F. C., Shukitt-Hale, B., & Joseph, J. A. (2007). Nutritional intervention in brain aging: Reducing the effects of inflammation and oxidative stress. *Sub-Cellular Biochemistry*, *42*, 299–318. doi:10.1007/1-4020-5688-5_14 PMID:17612057

Lenaz, G. (1998). Role of mitochondria in oxidative stress and ageing. *Biochimica et Biophysica Acta*, *1366*(1-2), 53–67. doi:10.1016/S0005-2728(98)00120-0 PMID:9714734

Leong, S. L., Cappai, R., Barnham, K. J., & Pham, C. L. (2009). Modulation of alpha-synuclein aggregation by dopamine: A review. *Neurochemical Research*, *34*(10), 1838–1846. doi:10.100711064-009-9986-8 PMID:19444607

Leszek, J., Barreto, G. E., Gasiorowski, K., Koutsouraki, E., Avila-Rodrigues, M., & Aliev, G. (2016). Inflammatory mechanisms and oxidative stress as key factors responsible for progression of neurodegeneration: Role of brain innate immune system. *CNS & Neurological Disorders - Drug Targets*, *15*(3), 329–336. doi:10.2174/1871527315666160202125914 PMID:26831258

Leveille, F., El Gaamouch, F., Gouix, E., Lecocq, M., Lobner, D., Nicole, O., & Buisson, A. (2008). Neuronal viability is controlled by a functional relation between synaptic and extrasynaptic NMDA receptors. *Federation of American Society for Experimental Biology Journal*, 22(12), 4258–4271. doi:10.1096/fj.08-107268 PMID:18711223

Lin, Y., Bloodgood, B. L., Hauser, J. L., Lapan, A. D., Koon, A. C., Kim, T. K., ... Greenberg, M. E. (2008). Activity dependent regulation of inhibitory synapse development by Npas4. *Nature*, *455*(7217), 1198–1204. doi:10.1038/nature07319 PMID:18815592

Macario, A. J., Cappello, F., Zummo, G., & Conway de Macario, E. (2010). Chaperonopathies of senescence and the scrambling of interactions between the chaperoning and the immune systems. *Annals of the New York Academy of Sciences*, *1197*(1), 85–93. doi:10.1111/j.1749-6632.2010.05187.x PMID:20536837

Mao, P., & Reddy, P. H. (2010). Is multiple sclerosis a mitochondrial disease? *Biochimica et Biophysica Acta*, *1802*(1), 66–79. doi:10.1016/j.bbadis.2009.07.002 PMID:19607913

Mark, P. M. (2004). Metal-catalyzed disruption of membrane protein and lipid signaling in the pathogenesis of neurodegenerative disorders. *Annals of the New York Academy of Sciences*, *1012*(1), 37–50. doi:10.1196/annals.1306.004 PMID:15105254

Marklund, S. L., Westman, N. G., Lundgren, E., & Ross, G. (1982). Copper- and zinc-containing superoxide dismutase, manganese-containing superoxide dismutase, catalase, and glutathione peroxidase in nrmal and neoplastic human cell lines and normal human tissues. *Cancer Research*, *42*, 1955–1961. PMID:7066906

Martin, S. J., Grimwood, P. D., & Morris, R. G. M. (2000). Synaptic plasticity and memory: An evaluation of the hypothesis. *Annual Review of Neuroscience*, *23*(1), 649–711. doi:10.1146/annurev.neuro.23.1.649 PMID:10845078

Mattson, M. P. (2008). Glutamate and neurotrophic factors in neuronal plasticity and disease. *Annals of the New York Academy of Sciences*, *1144*(1), 97–112. doi:10.1196/annals.1418.005 PMID:19076369

McCoy, M. K., & Cookson, M. R. (2012). Mitochondrial quality control and dynamics in parkinson's disease. *Antioxidants & Redox Signalling*, *16*(9), 869–882. doi:10.1089/ars.2011.4019 PMID:21568830

Meraz-Ríos, M. A., Franco-Bocanegra, D., & Rios, D. T. & Campos-Peña, V. (2014). Early onset Alzheimer's disease and oxidative stress. *Oxidative Medicine and Cellular Longevity*. PMID:24669286 Misonou, H., Morishima-kawashima, M., & Ihara, Y. (2000). Oxidative stress induces intracellular accumulation of amyloid beta-protein (Abeta) in human neuroblastoma cells. *Biochemistry*, *39*(23), 6951–6959. doi:10.1021/bi000169p PMID:10841777

Napolitano, A., Pezzella, A., & Prota, G. (1999). New reaction pathways of dopamine under oxidative stress conditions: Nonenzymatic iron-assisted conversion to norepinephrine and the neurotoxins 6-hydroxydopamine and 6,7-dihydroxytetrahydroisoquinoline. *Chemical Research in Toxicology*, *12*(11), 1090–1097. doi:10.1021/tx990079p PMID:10563835

Nicholls, D. G., & Budd, S. L. (2000). Mitochondria and neuronal survival. *Physiological Reviews*, 80(1), 315–360. doi:10.1152/physrev.2000.80.1.315 PMID:10617771

Niciu, M. J., Kelmendi, B., & Sanacora, G. (2012). Overview of glutamatergic neurotransmission in the nervous system. *Pharmacology, Biochemistry, and Behavior, 100*(4), 656–664. doi:10.1016/j. pbb.2011.08.008 PMID:21889952

O-Uchi, J., Pan, S., & Sheu, S.-S. (2012). Perspectives on: SGP symposium on mitochondrial physiology and medicine: molecular identities of mitochondrial Ca²⁺ influx mechanism: updated passwords for accessing mitochondrial Ca²⁺-linked health and disease. *The Journal of General Physiology*, *139*(6), 435–443. doi:10.1085/jgp.201210795 PMID:22641638

Papadia, S., Soriano, F. X., Léveillé, F., Martel, M. A., Dakin, K. A., Hansen, H. H., ... Hardingham, G. E. (2008). Synaptic NMDA receptor activity boosts intrinsic antioxidant defenses. *Nature Neuroscience*, *11*(4), 476–487. doi:10.1038/nn2071 PMID:18344994

Paxinou, E., Chen, Q., Weisse, M., Giasson, B. I., Norris, E. H., Rueter, S. M., ... Ischiropoulos, H. (2001). Induction of alpha-synuclein aggregation by intracellular nitrative insult. *Journal of Neurology*, *15*, 8053–8061. PMID:11588178

Perry, G., Sayre, L. M., Atwood, C. S., Castellani, R. J., Cash, A. D., & Rottkamp, C. A. (2002). The role of iron and copper in the aetiology of neurodegenerative disorders. *Central Nervous System Drugs*, *16*(5), 339–352. PMID:11994023

Polidori, M. C., Meccoci, P., Browne, S. E., Senin, U., & Beal, M. F. (1999). Oxidative damage to mitochondrial DNA in Huntington's disease parietal cortex. *Neuroscience Letters*, 272(1), 53–56. doi:10.1016/ S0304-3940(99)00578-9 PMID:10507541

Qin, B., Cartier, L., Dubois-dauphin, M., Li, B., Serrander, L., & Krause, K. H. (2006). A key role for the microglial NADPH oxidase in APP-dependent killing of neurons. *Neurobiology of Aging*, 27(11), 1577–1587. doi:10.1016/j.neurobiolaging.2005.09.036 PMID:16260066

Qui, J., Tan, Y. W., Hagenston, A. M., Martel, M. A., Kneisel, N., Skehel, P. A., ... Hardingham, G. E. (2013). Mitochondrial calcium uniporter Mcu controls excitotoxicity and is transcriptionally repressed by neuroprotective nuclear calcium signals. *Nature Communications*, *4*, 2034. PMID:23774321

Quintanilla, R. A., Jin, Y. N., Bernhardi, R. V., Gail, V., & Johnson, W. (2013). Mitochondrial permeability transition pore induces mitochondria injury in Huntington disease. *Molecular Neurodegeneration*, *8*(1), 45. doi:10.1186/1750-1326-8-45 PMID:24330821 Rahman, K. (2007). Studies on free radicals, antioxidants, and co-factors. *Clinical Interventions in Aging*, *2*, 219–236. PMID:18044138

Ramanathan, A., Nelson, A. R., Sagare, A. P., & Zlokovic, B. V. (2015). Impaired vascular-mediated clearance of brain amyloid beta in Alzheimer's disease: The role, regulation and restoration of LRP1. *Frontiers in Aging Neuroscience*, *7*, 136. doi:10.3389/fnagi.2015.00136 PMID:26236233

Reddy, P. H., & Reddy, T. P. (2011). Mitochondria as a therapeutic target for aging and neurodegenerative diseases. *Current Alzheimer Research*, *8*, 393–409. doi:10.2174/156720511795745401 PMID:21470101

Repetto, M. G., Domínguez, R. O., Marschoff, E. R., & Serra, J. A. (2012). Free radicals, oxidative stress and oxidative damage in Parkinson's disease. In J. Dushanova (Ed.), *Mechanisms in Parkinson's Disease – Models and Treatments*. InTech Open.

Rikans, L. E., & Hornbrook, K. R. (1997). Lipid peroxidation, antioxidant protection and aging. *Biochimica et Biophysica Acta*, *1362*(2-3), 116–127. doi:10.1016/S0925-4439(97)00067-7 PMID:9540842

Sanders, L. H., McCoy, J., Hu, X., Mastroberardino, P. G., Dickinson, B. C., Chang, C. J., ... Greenamyre, J. T. (2014). Mitochondrial DNA damage: Molecular marker of vulnerable nigral neurons in Parkinson's disease. *Neurobiology of Disease*, *70*, 214–223. doi:10.1016/j.nbd.2014.06.014 PMID:24981012

Sayre, L. M., Smith, M. A., & Perry, G. (2001). Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Current Medicinal Chemistry*, 8(7), 721–738. doi:10.2174/0929867013372922 PMID:11375746

Schildknecht, S., Gerding, H. R., Karreman, C., Drescher, M., Lashuel, H. A., Outeiro, T. F., ... Leist, M. (2013). Oxidative and nitrative alpha-synuclein modifications and proteostatic stress: Implications for disease mechanisms and interventions in synucleinopathies. *Journal of Neurochemistry*, *125*(4), 491–511. doi:10.1111/jnc.12226 PMID:23452040

Schuler, F., & Casida, J. E. (2001). Functional coupling of PSST and ND1subunits in NADH:ubiquinone oxidoreductase established by photoaffinity labeling. *Biochimica et Biophysica Acta*, *1506*(1), 79–87. doi:10.1016/S0005-2728(01)00183-9 PMID:11418099

Selkoe, D. J. (2001). Alzheimer's disease: Genes, proteins, and therapy. *Physiological Reviews*, 81(2), 741–766. doi:10.1152/physrev.2001.81.2.741 PMID:11274343

Sharma, D. R., Sunkaria, A., Wani, W. Y., Sharma, R. K., Kandimalla, R. J., Bal, A., & Gill, K. D. (2013). Aluminium induced oxidative stress results in decreased mitochondrial biogenesis via modulation of PGC-1α expression. *Toxicology and Applied Pharmacology*, 273(2), 365–380. doi:10.1016/j. taap.2013.09.012 PMID:24084166

Shepherd, J. D., & Huganir, R. L. (2007). The cell biology of synaptic plasticity: AMPA receptor trafficking. *Annual Review of Cell and Developmental Biology*, 23(1), 613–643. doi:10.1146/annurev. cellbio.23.090506.123516 PMID:17506699

Shin, S.-G., Kim, J. Y., Chung, H. Y., & Jeong, J. C. (2005). Zingerone as an antioxidant against peroxynitrite. *Journal of Agricultural and Food Chemistry*, 53(19), 7617–7622. doi:10.1021/jf051014x PMID:16159194 Solís-Herrera, A., & Ashraf, G. M., Esparza, M., Arias, R. I., Bachurin, S. O., Barreto, G. E., & Aliev, G. (2015). Biological activities of QIAPI 1 as a melanin precursor and its therapeutic effects in wistar rats exposed to arsenic poisoning. *Central Nervous System Agents in Medicinal Chemistry*, *15*(2), 99–108. doi:10.2174/1871524915666150424113831 PMID:25909193

Somlyo, A. P., Bond, M., & Somlyo, A. V. (1985). Calcium content of mitochondria and endoplasmic reticulum in liver frozen rapidly *in vivo*. *Nature*, *314*(6012), 622–625. doi:10.1038/314622a0 PMID:3990795

Stayte, S., & Vissel, B. (2014). Advances in non-dopaminergic treatments for Parkinson's disease. *Frontiers in Neuroscience*, *8*, 113. PMID:24904259

Su, B., Wang, X., Lee, H. G., Tabaton, M., Perry, G., Smith, M. A., & Zhu, X. (2010). Chronic oxidative stress causes increased tau phosphorylation in M17 neuroblastoma cells. *Neuroscience Letters*, *468*(3), 267–271. doi:10.1016/j.neulet.2009.11.010 PMID:19914335

Tamagno, E., Guglielmotto, M., Aragno, M., Borghi, R., Autelli, R., Giliberto, L., ... Tabaton, M. (2008). Oxidative stress activates a positive feedback between the γ - and β -secretase cleavages of the β -amyloid precursor protein. *Journal of Neurochemistry*, *104*(3), 683–695. PMID:18005001

Tang, Y.-P., & Gershon, E. S. (2003). Genetic studies in Alzheimer's disease. *Journal of Clinical Neuroscience*, *5*, 17–26. PMID:22033785

Tezel, G. (2006). Oxidative stress in glaucomatous neurodegeneration: Mechanisms and consequences. *Progress in Retinal and Eye Research*, 25(5), 490–513. doi:10.1016/j.preteyeres.2006.07.003 PMID:16962364

Theuns, J., & Van Broeckhoven, C. (2000). Transcriptional regulation of Alzheimer's disease genes: Implications for susceptibility. *Human Molecular Genetics*, *9*(16), 2383–2394. doi:10.1093/hmg/9.16.2383 PMID:11005793

Uttara, B., Singh, A. V., Zamboni, P., & Mahajan, R. T. (2009). Oxidative stress and neurodegenerative diseases: A review of upstream and downstream antioxidant therapeutic options. *Current Neuropharmacology*, *7*(1), 65–74. doi:10.2174/157015909787602823 PMID:19721819

Van Raamsdonk, J. M., Vega, I. E., & Brundin, P. (2017). Oxidative stress in neurodegenerative disease: Causation or association? *Oncotarget*, 8(7), 10777–10778. doi:10.18632/oncotarget.14650 PMID:28099897

Walls, K. C., Coskun, P., Gallegos-Perez, J. L., Zadourian, N., Freude, K., Rasool, S., ... LaFerla, F. M. (2012). Swedish Alzheimer mutation induces mitochondrial dysfunction mediated by HSP60 mislocalization of amyloid precursor protein (APP) and beta-amyloid. *The Journal of Biological Chemistry*, 287(36), 30317–30327. doi:10.1074/jbc.M112.365890 PMID:22753410

Wang, X., & Michaelis, E. K. (2010). Selective neuronal vulnerability to oxidative stress in the brain. *Frontiers in Aging Neuroscience*, *2*, 12. PMID:20552050

Wiltgen, B. J., Royle, G. A., Gray, E. E., Abdipranoto, A., Thangthaeng, N., Jacobs, N., ... Vissel, B. (2010). A role for calcium-permeable AMPA receptors in synaptic plasticity and learning. *PLoS One*, *5*(9), 12818. doi:10.1371/journal.pone.0012818 PMID:20927382

Winklhofer, K. F., & Haass, C. (2010). Mitochondrial dysfunction in Parkinson's disease. *Biochimica et Biophysica Acta*, *1802*(1), 29–44. doi:10.1016/j.bbadis.2009.08.013 PMID:19733240

Wright, A., & Vissel, B. (2012). The essential role of AMPA receptor GluR2sub-unit RNA editing in the normal and diseased brain. *Frontiers in Molecular Neuroscience*, *5*, 34. doi:10.3389/fnmol.2012.00034 PMID:22514516

Xia, P., Chen, H. S., Zhang, D., & Lipton, S. A. (2010). Memantine preferentially blocks extrasynaptic over synaptic NMDA receptor currents in hippocampal autapses. *The Journal of Neuroscience*, *30*(33), 11246–11250. doi:10.1523/JNEUROSCI.2488-10.2010 PMID:20720132

Xiang, W., Schlachetzki, J. C., Helling, S., Bussmann, J. C., Berlinghof, M., Schäffer, T. E., ... Becker, C. M. (2013). Oxidative stress-induced posttranslational modifications of alpha-synuclein: Specific modification of alphasynuclein by 4-hydroxy-2-nonenal increases dopaminergic toxicity. *Molecular and Cellular Neurosciences*, *54*, 71–83. doi:10.1016/j.mcn.2013.01.004 PMID:23369945

Xu, J., Kurup, P., Zhang, Y., Goebel-Goody, S. M., Wu, P. H., Hawasli, A. H., ... Lombroso, P. J. (2009). Extrasynaptic NMDA receptors couple preferentially to excitotoxicity via calpain-mediated cleavage of STEP. *The Journal of Neuroscience*, *29*(29), 9330–9343. doi:10.1523/JNEUROSCI.2212-09.2009 PMID:19625523

Yang, J. L., Sykora, P., Wilson, D. M. III, Mattson, M. P., & Bohr, V. A. (2011). The excitatory neurotransmitter glutamate stimulates DNA repair to increase neuronal resiliency. *Mechanisms of Ageing and Development*, *132*(8-9), 405–411. doi:10.1016/j.mad.2011.06.005 PMID:21729715

Yavich, L., Jakala, P., & Tanila, H. (2006). Abnormal compart-mentalization of norepinephrine in mousedentate gyrus in alpha-synuclein knockout and A30P transgenic mice. *Journal of Neurochemistry*, *99*(3), 724–732. doi:10.1111/j.1471-4159.2006.04098.x PMID:16824047

Zaleska, M. M., Nagy, K., & Floyd, R. A. (1989). Iron-induced lipid peroxidation and inhibition of dopamine synthesis in striatum synaptosomes. *Neurochemistry*, *14*(7), 597–605. doi:10.1007/BF00964867 PMID:2550829

Zambrano, C. A., Egaña, J. T., Núñez, M. T., Maccioni, R. B., & González-billault, C. (2004). Oxidative stress promotes tau dephosphorylation in neuronal cells: The roles of cdk5 and PP1. *Free Radical Biology & Medicine*, *36*(11), 1393–1402. doi:10.1016/j.freeradbiomed.2004.03.007 PMID:15135175

Zhou, M., Xu, S., Mi, J., Uèda, K., & Chan, P. (2013). Nuclear translocation of alpha-synuclein increases susceptibility of MES23.5 cells to oxidative stress. *Brain Research*, *1500*, 19–27. doi:10.1016/j. brainres.2013.01.024 PMID:23337620

^{Chapter 3} Free Radicals in Oxidative Stress, Aging, and Neurodegenerative Disorders

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ABSTRACT

Free radicals are intricately woven into the fabric of oxidative stress and are significant in the development of neurodegenerative disorders (NDs). This chapter examines free radicals in the context of neurodegeneration and provides overview of the multiple roles they play in the pathophysiology and clinical progression of varying NDs including Pick's disease (PiD), Parkinson's disease (PD), Alzheimer's disease (AD), prion diseases (PrD), traumatic brain injury, and aging. The molecular mechanisms of degeneration in Huntington's disease (HD) are also examined with respect to free radicals. Different antioxidant systems and their mechanisms of action are briefly reviewed in addition to the role of diet in aging. The effectiveness of selected synthetic drugs and natural products used in oxidative stress is also reviewed. Lastly, the chapter examines challenges associated with the use of antioxidants and how promising future directions like the endocannabinoid system is being pursued in the race to effectively manage NDs.

DOI: 10.4018/978-1-5225-5282-6.ch003

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INTRODUCTION

Since Moses Gomberg's first description of triphenylmethyl radical in literature in 1900, free radicals have remained an important subject of discussion owing to their numerous physiological effects (Gomberg, 1900). Initially they were not believed to be present in biological systems due its excessive reactivity. However, by the end of the second half of the 20th century, the scientific world had come to terms with the fact that free radicals are found in biological systems and contribute to several pathologies as well as aging (Lushchak, 2014). Today it is recognized that free radicals instigate oxidative stress and propagate neuronal injury thus playing a major role in NDs like AD, PD, amyotrophic disorders, PrD and several others (Khan et al., 2016). It is important to note that free radicals are not always deleterious such as the generation of nitric oxide in neurotransmission and the production of superoxide anion (O_2^{\bullet}) by activated microglia. There is a focus on free radicals in the central nervous system (CNS) and mechanisms involving free radicals in selected NDs. Free radicals are reviewed as potential targets of drug action in the management of NDs. Thus this chapter will evaluate what free radicals are and review their contribution to oxidative stress induced neurodegenerative disorders as well as aging. It is expected that at the end of this chapter further knowledge on the mechanisms of free-radical induced neurodegeneration in some NDs would have been gained in addition to an understanding of the prospects of novel therapeutic approaches. Overall, this chapter examines the interplay of free radicals and oxidative stress in the development and progression of several NDs.

BACKGROUND

Free radicals are exceptionally reactive atoms or molecules with one or more unpaired electrons and capable of independent existence (Halliwell, 1992). In some instances, free radicals have been used interchangeably with reactive oxygen species (ROS). While this may be correct sometimes, not all ROS are free radicals. Although both generate oxidative stress, ROS are chemically reactive species that contain oxygen and may or may not necessarily be a radical. For example, whereas hydrogen peroxide (H_2O_2) is a reactive oxygen species, it is not a free radical. Other examples include lipid, protein and nucleic acid peroxides. There are also reactive species that are not oxygen species such as reactive species of nitrogen (peroxynitrite (ONOO[•]) and nitric oxide ([•]NO)), carbon and sulfur.

However, the common denominator in all these terminologies is oxidative stress. Oxidative stress is a chemical process resulting from excessive free radical production due to an insufficiency of the counteracting antioxidant response system (Birben, Sahiner, Sackesen, Erzurum, & Kalayci, 2012). Free radicals and reactive species of oxygen or otherwise participate in chain reactions that culminate in oxidative stress. Normally in aerobic organisms, molecular oxygen is reduced to water via intermediate steps of oxygen reduction that forms O_2^{\bullet} , H_2O_2 and the hydroxyl radical ($^{\bullet}OH$) (Halliwell & Gutteridge, 1990). Free radicals and other reactive species' production in the body is approximately balanced by antioxidant mechanisms needed to mop up these reactive species. However NDs as well as aging, the production of free radical is higher than antioxidant defense. Antioxidants are molecules which at minimal concentrations compared with that of an oxidizable substrate, appreciably slows or stops oxidation of that substrate (Halliwell & Gutteridge, 1990). These enzymatic or non-enzymatic antioxidants, reduce the potential damage of the reactive species thus only minor reactive species induced-damage occurs. Therefore, oxidative stress arises in the event of a significant disparity between the production of free radicals and other ROS and antioxidants production (Halliwell, 2007). In other words, it is a disturbance in the oxidant–antioxidant balance in favor of oxidant mechanisms with a resulting potential tissue damage regarded as oxidative damage (Sies, 1997).

CENTRAL NERVOUS SYSTEM, FREE RADICALS AND NEURODEGENERATIVE DISORDERS

During normal physiological processes, the brain utilizes about 20% of the total oxygen in the body which leads to the generation of more free radicals than in any other tissue (Joshi & Praticò, 2014; Smith, Cappai, & Barnham, 2007). Reactive oxygen species (ROS) are important in numerous cellular and signaling pathways at physiological concentrations such as cell cycle regulation and enzyme activation (Dröge, 2002; Zorov, Juhaszova, & Sollott, 2014). When in excess, they are however scavenged by defense mechanisms which may be enzymatic and non-enzymatic antioxidants. Overproduction of ROS leads to generation of oxidative stress, which has been shown to cause several harmful effects including Deoxyribonucleic acid (DNA), lipid and protein damage (Ohta, Yashiro, Ohashi, & Imai, 2012). Oxidative stress remains a major feature contributing to cerebral biochemical impairment seen in some NDs (Huang, Zhang, & Chen, 2016).

The brain, in comparison to other body organs, has been shown to be predominantly exposed to oxidative damage. This is due to its high metabolic rate, due to a highly active mitochondria metabolism. Additionally, the brain vulnerability to oxidative damage is because a huge proportion of it is made of lipids (Birben et al., 2012. Also, another important factor is the fact that the brain contains low antioxidant defense mechanisms (Birben et al., 2012). Furthermore, neurons have a longer life span than other cells and so are exposed to accumulated oxidative damage over time (Magrassi, Leto, & Rossi, 2013). These among others make the neurodegeneration due to oxidative damage more significant in the brain and spinal cord.

Neurodegeneration is characterized by the progressive loss of specific neuronal cells with an accompanying protein aggregation leading to disorders such as AD, PD and amyotrophic lateral sclerosis (ALS). The genesis of these disorders is attributed to a myriad of factors which are yet not completely understood (Khan et al., 2016). Among these factors include: a genetic basis and familial inheritance, exposure to environmental risk factors, altered immune and inflammatory responses, idiopathic protein aggregation, ageing and of course, the ubiquitous oxidative stress, and its attendant damage (Khan et al., 2016).

Another conceivable mechanism of oxidative stress-induced pathology implicates a dysfunction of ROS-mediated cell-cell communication. Excessive O_2^{\bullet} , OH[•] production by NADPH oxidases (NOXs) and the free radical peroxynitrite (ONOO⁻) by nitric oxide synthase cause massive cell damage through endoplasmic reticulum (ER) stress, impaired calcium handling, and nitration of tyrosine residue of several proteins. NOX is a multi-subunit enzyme which is composed of membrane and cytosolic components including NOX (NOX1-5) and phox subunits. NADPH produces O_2^{\bullet} by transporting an electron from NADPH to oxygen (Ma et al., 2017). The enzyme has been implicated in diverse roles including host defense and cellular signaling (Panday, Sahoo, Osorio, & Batra, 2015). This cell damage perturbs the structure of proteins and alters the catalytic action of enzymes thus, impairing cell signaling pathways which depends on such proteins and enzymes. Additionally, several cell signaling cascades require ROS

as messengers therefore a constant rise in concentrations of ROS can cause an over-activation of such pathways, with several damaging consequences (Freixes, Rodríguez, Dalfó, & Ferrer, 2006).

The high oxygen demands of the brain yet with limited defense strategies against oxidative stress increases its susceptibility to oxidative damage and makes oxidative damage a significant causative effect of neurodegeneration. Evidence to this is led by the several observations of markers of oxidative damage in the brain, spinal fluid and plasma of AD (Arlt, Beisiegel, & Kontush, 2002; Butterfield, Castegna, Lauderback, & Drake, 2002; Selley, Close, & Stern, 2002), PD (Dexter et al., 1989) and ALS (Pedersen et al., 1998; Sayre, Smith, & Perry, 2001) patients. Additionally, DNA damage due to oxidative stress is observed as DNA nitration and hydroxylation in AD brains and increased 8-hydroxy-2-deoxyguanosine and 8-hydroxyguanine levels in PD brains (Alam et al., 1997; Gabbita, Lovell, & Markesbery, 1998). Furthermore, the brain is rich in oxidizable catecholamines such as noradrenaline and dopamine that can undergo auto-oxidation to generate free radicals in the presence of metal ions and propagate neuronal damage (Gerlach, Ben-Shachar, Riederer, & Youdim, 1994; Halliwell & Gutteridge, 1990).

Antioxidant Response System and Biomarkers of Oxidative Stress

The antioxidant defense system is an endogenous enzymatic and non-enzymatic system in place to protect the body from oxidative damage (Ohta et al., 2012). It comprises of enzymatic (superoxide dismutase, glutathione peroxidase and catalase) and non-enzymatic low molecular weight reductants (glutathione, α -tocopherol, and ascorbic acid). The antioxidant defense system prevents free radical mediated damage of cells by converting the overproduced free radicals to non-destructive cellular molecules. Astrocytes also important in maintaining high intracellular concentrations of some antioxidants, making them resistant to oxidative damage by elevating expression of some antioxidant enzymes; increasing glucose metabolism and transport; increasing manufacturing of glutathione; and salvaging ascorbic acid (Cabezas et al., 2014). Ascorbic acid is very important for sustaining oxidative balance as it acts a cofactor for antioxidant enzymes. Studies have shown that ascorbic acid deficiency plays acute role in oxidative damage seen in AD and normal aging (Dixit et al., 2015). Glutathione (GSH) is the main brain antioxidant molecule (Johnson et al., 2012). A progressive diminution of GSH concentration during aging as well as age-associated diseases like has been suggested is some studies (Carvalho, Lim, Nijland, Witte, & Van Horssen, 2014; Johnson, Wilson-Delfosse, & Mieyal, 2012). GSH protects neurons from oxidative damage, primarily by serving as a redox regulator and hence crucial for the detoxification of ROS in neurons (Johnson et al., 2012).

Several markers of oxidative stress have been measured in peripheral blood; these include lipid peroxides, GSH, ascorbic acid and α -tocopherol (Sinclair et al., 1998). Lipid peroxidation is a central feature of oxidative stress (Lushchak, 2014). Oxidative stress-mediated membrane lipids peroxidation is detrimental as it results in modifications in the biological properties of the membrane i.e.the magnitude of membrane fluidity (Lushchak, 2014). This leads to inactivation of membrane-bound receptors and that may ruin cellular functioning and disrupt tissue permeability. Products of lipid peroxidation such as malondialdehyde (MDA), have been used as possible markers of oxidative stress (Sultana et al., 2013).

Measurement of the plasma antioxidants provides an indication of the level of oxidative stress and is routinely used as biomarker of oxidative stress in NDs. This was shown by previous studies that demonstrated a decrease in peripheral levels of retinol, ascorbic acid and α - tocopherol along as well as a diminished activity of SOD and glutathione peroxidase in AD patients (Sinclair et al., 1998; Ferreira et al., 2015). Other works also correlated the increase in oxidative stress with a decreased GSH activity (Sinclair et al., 1998).

Role of Diet in Oxidative Stress and Neurodegeneration

Diet may play a huge role in providing supplementary antioxidants. A typical meal possibly contains several naturally occurring antioxidant molecules (Bayram et al., 2012). These antioxidant compounds are found in different concentrations in vegetables, legumes, fruits, cereals, olive oil, wine, cocoa, and tea (Bayram et al., 2012; Coelho, Hermsdorff, & Bressan, 2013). Studies have shown that an antioxidant-rich diet potentially averts oxidative damage and cognitive decline due to its free radical or active oxygen scavenging properties (Coelho, Hermsdorff, & Bressan, 2013). This has led to a great need to search for nontoxic active ingredients of natural resources that could reverse the biochemical imbalances that occur in NDs.

Oxidative Stress-Induced Mitochondrial Dysfunction in Neurodegeneration

In the brain, the primary sites of ROS generation include mitochondria in the neurons and glia. The production of free radicals in these areas is exacerbated in disorders like PD due to inflammation, dopamine degradation, mitochondrial dysfunction, aging, GSH depletion, and high levels of iron (Dias, Junn, & Mouradian, 2013). Mitochondrial dysfunction, increased apoptosis together with low antioxidant levels are also evident in the development of AD (Birben et al., 2012). In addition, neurodegeneration-induced ROS causes damage in key cellular proteins and disrupt lipid membranes thus promoting oxidative stress (Dias et al., 2013; Birben et al., 2012). Mitochondrial dysfunction leads to increased free radical production in the respiratory chain (Dias et al., 2013; Huang et al., 2016). Particularly, mitochondrial complex I deficiency has been identified to be strongly associated with PD. Indeed, a large amount of the unfavorable neural apoptosis observed in PD can be attributed to the complex I defect (Huang et al., 2016).

Aging and Oxidative Stress

Aging has been shown to be a major risk factor for ND (Moll, El-Ami & Cohen, 2014; Bickford, Flowers & Grimmig, 2017). Aging is a complex phenomenon that results in reduced organ functioning, decline in cognition and overall decline in the organism's homeostasis. As stated already, the brain is prone to oxidative stress however an aging brain is highly susceptible to a greater extent as a result of a loss of resilience associated with aging (Bickford et al., 2017). This leads to a decreased resistance to various forms of stress as well as an increased susceptibility to oxidative stress and thus several diseases. Even though, aging is poorly understood, one of the most widely accepted theories is the 'Free Radical Theory' which attributes aging to the deleterious effects of free radicals on cell components (Harman, 1956).

The environment of the aged brain has been associated with two main biological processes; oxidative stress and inflammation with microglia being one of the major cell types implicated in both processes (Bickford et al., 2017). In the healthy, young brain, microglia (known as the resident tissue macro-phages in the CNS) perform housekeeping duties and constantly assess the microenvironment thereby effectively protecting the CNS from invading pathogens amid other attacks (Koellhoffer, McCullough & Ritzel, 2017). Thus, microglia in the CNS induce an innate immune response and once activated can perform several functions including phagocytosis (Fenn, Henry, Huang, Dugan & Godbout, 2012). It

has been shown that there is an increase in reactive microglia in aged brains with an associated increase in inflammatory markers (Fenn et al., 2012). Aged microglia is also identified in the pathophysiology of Traumatic Brain Injury, AD, and worsening outcomes in stroke (Koellhoffer et al., 2017).

Some agents with anti-oxidant activity such as polyphenols have been shown to be potent modulators of neuroinflammation. This effect may be due to the stimulation of the transcription factor, nuclear factor erythroid related factor 2 (Nrf2)-antioxidant response element (ARE) pathway thereby reducing the production of pro-inflammatory cytokines and modulating glial function (Bickford et al., 2017). The Nrf2-ARE pathway is very vital for the proper functioning of microglia. Polyphenols may therefore be very important in neuroprotection as shown by a study by Bickford et al., 2017 where NT-020, a polyphenolic rich mixture, was able to improve the aging environment as well as cognition with minimal side effects. In fact, when NT-020 was administered to aged human participants (65-85 years) for two months, it was noted that there was an increase in processing speed (according to memory tests) suggesting the important role that antioxidants play in neurodegeneration (Bickford et al., 2017).

MOLECULAR MECHANISMS OF FREE RADICAL-INDUCED DEGENERATION IN NEURODEGENERATIVE DISORDERS

Alzheimer's Disease and Other Dementias

Dementia is a syndrome caused by several disorders which negatively influence cerebral structures and functions, to cause deterioration of memory functions and behavior (Sosa-Ortiz, Acosta-Castillo, & Prince, 2012). Most common causes of dementia include AD, frontotemporal lobar degeneration (FTLD), vascular dementia, Creutzfeldt-Jakob disease (CJD), PD, and Lewy Body dementia (LBD). There is a very strong association age with the prevalence of dementia, so there is an expected increase in the number of people living with dementia as the aging population increases (Sosa-Ortiz et al., 2012).

AD is a very common ND which affect around 16 million of the elderly population worldwide and is the leading cause of dementia (Butterfield & Boyd-Kimball, 2004). It accounts for 60-80% of dementia in the elderly population (Sosa-Ortiz et al., 2012; Huang et al., 2016). AD is characterized by neurofibrillary tangles composed of hyperphosphorylated tau protein and amyloid-beta (A β)-containing plaques in addition to cognitive impairment and progressive neurodegeneration. The neocortex and hippocampus are the main affected brain areas. AD symptoms usually start with minor amnesic episodes and progress toward a vivid change in personality (Sosa-Ortiz et al., 2012). The etiology of AD is multifaceted including major environmental and genetic risk factors (Huang et al., 2016). Increased lipid peroxidation and inadequate antioxidants were shown in the peripheral tissues of AD patients. Similar to most NDs, the specific biochemical mechanism of the pathogenesis of AD remains unknown although evidence suggests a massive loss of acetylcholine and a possible role of oxidative stress (Butterfield et al., 2001). Oxidative damage plays an important part in A β deposition in AD. Lushchak (2014) reviews complex relationships between excitotoxicity and A β deposition, as well as production of ROS in AD.

FTLD, example PiD, are diseases which degeneration primarily affect the frontal (front) and temporal (side) regions of the brain (Diehl et al., 2004). PiD is characterized by the presence of abnormal bodies also known as Pick's bodies in affected neurons (Rademakers, Neumann, & Mackenzie, 2012). Though the symptoms appear similar to those seen in AD, PiD is unique in that it progresses very rapidly. Symptoms include inability to recognize people as well as deterioration of skilled movement and language abilities, (Mendez & Shapira, 2011).

Lewy Body dementia (LBD) is characterized by the deposition of abnormal proteins called Lewy bodies in affected brain regions (Diehl et al., 2004). Dementia with Lewy bodies and Parkinson's disease dementia (Lewy body dementia), are ranked second among the commonest types of degenerative dementia in patients older than 65 years (Gomperts et al., 2012). LBD patients present with symptoms similar to AD. During early stages, symptoms of LBD may be mild but ultimately result in significant impairment of cognitive function (Postuma, Gagnon, Pelletier, & Montplaisir, 2013).

Motor Neuron Disorders

Motor neuron disorders (MND) are group of progressive conditions in which lower and upper motor neurons degenerate and result in decreasing strength in limb, abdominal, thoracic and bulbar muscles without affecting oculomotor and sphincter muscles (Wijesekera & Leigh, 2009). The term generically applies to a complete spectrum of the diseases including progressive bulbar palsy, amyotrophic lateral sclerosis (ALS) and progressive muscular atrophy and of which ALS is the most common and devastating. It is characterized by progressive injury and cell death of motor neurons in the motor cortex, brain stem and spinal cord. This results in the progressive muscle weakness, wasting upper and lower motor neurons with pathologically brisk reflexes of the limb and bulbar muscles (Bäumer, Talbot, & Turner, 2014; Leigh & Ray-Chaudhuri, 1994). It is now recognized that extra motor parts of the CNS are also involved in MND and thus MND can be regarded as a multisystem disease whereby motor neurons are affected quickest and harshest (Wijesekera & Leigh, 2009).

The disorders occur in about one to two individuals per 100 000 and predominantly in middle aged and elderly patients (Shaw, 1999). Men have a relatively higher prevalence of the disease than women (1.5:1) although this seems to be turning towards a balanced prevalence in recent years (Logroscino et al., 2008). In most instances, onset is sporadic (90%) while heredity has also been reported (10%). MND patients have dire prognoses with median survival of approximately 3.5 years after onset of symptoms as a result of death due to paralysis of respiratory muscles (Leigh & Ray-Chaudhuri, 1994; Shaw, 1999).

Several hypotheses have been postulated as to the pathophysiology of MND including genetic predisposition, increased glutamatergic excitotoxicity, failure of axonal transport, mitochondrial dysfunction and oxidative stress (Turner et al., 2013).

Free Radicals and Oxidative Stress in Motor Neuron Disorders

Free radical-instigated oxidative stress is a significant portion of the already understood aspects of the pathophysiology of MND. Although familial MND is not fully understood, several point mutations in chromosome 21 gene product coding for the enzyme, copper-zinc superoxide dismutase (*SOD 1*), is responsible for 20% in different pedigrees of familial cases and 2% of all cases of MND (Rosen et al., 1993; Shaw, 1999). This ubiquitous antioxidant enzyme converts the superoxide free radical into less harmful hydrogen peroxide (H_2O_2) before it is taken care of by other antioxidant enzymes. Evidence suggest that the mutant enzyme exerts its damaging by an abnormal handling of hydrogen peroxide and peroxynitrite which cases elevated production of hydroxyl radicals and nitrotyrosine residues (Carrì, Valle, Bozzo, & Cozzolino, 2015; Cookson & Shaw, 1999; Shaw, 1999; Shibata, Hirano, Yamamoto, Kato, & Kobayashi, 2000). The mutated Cu/Zn SOD may also form neurotoxic intracellular aggregates

or release cytosolic Cu or Zn which then becomes neurotoxic (Shaw, 1999). The increased expression of Cu/Zn SOD in both axons and cell bodies of motor neurons in comparison to all other cells of the nervous system increases their susceptibility to damage by such genetic alterations. In addition, alterations in apurinic/apyramidinic (AP) endonuclease, an important enzyme for repairing oxidative damage to DNA and forming abasic sites, is also associated with ALS (Olkowski, 1998). Other genes have also been identified that increase the susceptibility to MND to various degrees aside SOD and AP endonuclease.

Neuronal injury instigated by free radicals is a key cause of several age-related neurodegenerative diseases because the effects of oxidative stress in neurons, which naturally are non-replicating, is cumulative. Given the effects of mutations in antioxidant enzyme SOD on the development of certain familial ALS, interest in free radicals and for that matter oxidative stress in MND has been high for a long time. Cultured fibroblasts of skin samples of both familial and sporadic motor neuron patients indicate an amplified sensitivity to oxidative insults (Aguirre et al., 1998; Wijesekera & Leigh, 2009). Also, post-mortem results points towards a change in expression of parts of the intracellular antioxidant defense mechanism. This has been recorded in ALS patients and signifies an attempted compensatory response to the increased oxidative stress (Shaw, Chinnery, Thagesen, Borthwick, & Ince, 1997).

The evidence to support an involvement of an aberrant oxidative damage management is also due to the observation that antioxidant molecule vitamin E exhibits beneficial effects in Cu/Zn *SOD1* mutated transgenics by delaying the onset of MND. Ghadge and colleagues (Ghadge et al., 1997) report that free radicals especially O_2^- are implicated in the mutant SOD-mediated cell death, owing to the fact that SOD and glutathione mimetics reversed this mortality. Their data also show that expression of mutant SOD escalates the vulnerability to oxidative stress and increases rate of accumulation of O_2^- . Cell death regulatory protein B-cell lymphoma 2 (BCl-2), which affects free radical production and for that matter cell survival, also protects the cells against mutant SOD-induced apoptosis (Hockenbery, Oltvai, Yin, Milliman, & Korsmeyer, 1993). These seem to explain an important involvement of free radicals and oxidative stress in ALS and MND in general.

Nonetheless, clinical trials involving potent antioxidant n-acetylcysteine showed non-significant correlates suggesting that the situation is not completely clear.

Huntington's Disease

HD is a devastating autosomal dominant ND with abnormal trinucleotide (CAG) expansion in the *hun-tingtin (HTT)* gene (Velusamy et al., 2017) which is characterized by impaired cognition, movement and psychiatric disorders (Ma et al., 2017). Usually, healthy individuals have 5-6 CAG repeats in exon 1 of the *HTT* gene but affected individuals have more than 36 repeats (Manoharan et al., 2016). The associated neuronal death has been attributed to the accumulation of mutant Huntingtin proteins as there is abnormal folding and protein function (Velusamy et al., 2017).

Oxidative stress is hypothesized as significant mechanism in the progression of this disorder with several studies confirming this hypothesis. For instance, the activity of the mitochondrial complexes II, III and IV have been found to be reduced in the striatum of HD patients' post-mortem (Lee, Gold, & Linker, 2012). It has been shown that in HD patients, there is elevation of biomarkers for oxidative stress such as malondialdehyde in the striatum, cortex and serum. The expansion of the CAG triplets has been shown to induce oxidative stress which causes damage to the cell membrane, DNA and other enzymes involved in adenosine triphosphate (ATP) production (Velusamy et al., 2017). This mitochondrial damage therefore provides a strong mechanism for initiating apoptosis in HD brains (Velusamy et al., 2017).

This is because under oxidative stress, there is the activation of glutamate receptors which leads to an increase in the influx of calcium ions. This increase leads to the translocation of proapoptotic Bcl-2 proteins to the mitochondrial membrane as well as the activation of caspase 8 thus initiating apoptosis (Bansal & Singh, 2017). Manoharan et al. (2016) hypothesized that, the causes of oxidative stress in HD include imbalance in oxidant-antioxidant status, higher lipid concentration and high energy requirement as well as poor antioxidant status. This has been supported by a reduced superoxide dismutase activity in the cortex and cerebellum in HD patients (Ma et al., 2017).

NADPH oxidase (NOX2) enzyme has been proposed to be involved in the increase in ROS in the progression of HD. For instance, elevated ROS levels in PC12 cells expressing huntingtin proteins are lowered by treatment with NOX inhibitors (Valencia et al., 2012). Again, higher levels of NOX activity was found in the brain of HD patients as compared to controls (Valencia et al., 2012).

Elevated 4-hydroxy-2-nonenal (4-HNE) adducts levels have been found in both clinical and preclinical models of HD (striatum of both human and mice HD brains) (Lee et al., 2011). Thus, 4-HNE is proposed to be a relevant marker for determining oxidative damage in HD. Data from this study revealed that 4-HNE immunoreactivity was co-localized with mutant huntingtin (mtHtt) inclusions in the striatal neurons of R6/2 HD mice suggesting the importance of 4-HNE as a possible target for therapeutic intervention (Lee et al., 2011).

Parkinson's Disease

PD is a ND which affects over 10 million people above the age of 65 worldwide (Schneider & Obeso, 2014). In PD there is degeneration of dopaminergic neurons in the substantia nigra pas compacta of the basal ganglia which control motor functions. PD exhibits both motor and non-motor symptoms including resting rigidity and postural instability, tremor, bradykinesia, and akinesia (Schneider & Obeso, 2014). Non-motor symptoms observed in PD include cognitive, mood, autonomic and sleep disturbances (Schneider & Obeso, 2014). James Parkinson and Charcot among others had reported several non-motor symptoms in their classic literature that include delusion, pain, fatigue, dysfunction of the bladder and cognitive deterioration (Garcia-Ruiz, Chaudhuri, & Martinez-Martin, 2014).

The etiology of PD remains unclear, but oxidative stress has been considered as one of the major pathophysiological mechanisms involved (Dias et al, 2013). The main mechanism identified leading to oxidative stress in PD include the dopamine oxidation which produces toxic semiquinones. Secondly, the enhanced catabolism of dopamine by monoamine oxidase B (MAO-B) may instigate increased production of hydrogen peroxide, superoxide anions, and-hydroxyl radicals and the accumulation of alpha-synuclein aggregates (Hwang, 2013). These free radicals reduce the activity of mitochondrial complex I, this in turn contributes to the production of more reactive oxygen species which leads to apoptotic cell death (Dias et al, 2013; Blesa et al., 2015).

Prion's Disease

PrD also referred to collectively as transmissible spongiform encephalopathies (TSEs) are uncommon fatal NDs that are acquired either by direct transmission, inheritance of dominant prion protein gene mutations and in about 85% of cases through idiopathic sporadic causes (Brown, 2005; Prusiner, 1991). A prion is a novel pathogenic particle made up of a self-propagating misfolded protein with an abnormal conformation (Sarnataro, Pepe, & Zurzolo, 2017). It is a cell surface glycosylphosphatidylinositol (GPI)-

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anchored glycoprotein expressed by neurons, glial cells and several other cells in the body. Human TSEs include: CJD), Gerstmann-Sträussler-Scheinker (GSS) syndrome, kuru and fatal familial insomnia. TSEs are characterized by neuroamyloid formation and dementia and neuropathological changes including spongiform degeneration in the brain (Brown, 2005; Wong, Wang, Brown, & Jones, 1999).

CJD is an unusual but deadly disorder that causes swift-developing and progressive (Iwasaki, Mori, & Ito, 2012). CJD is caused by the transmissible proteinaceous infectious particle called "prion". This pathogen consists of a protein which transforms normal protein molecules into infectious molecules. CJD signs include rapidly declining memory, behavioral changes and ataxia (Iwasaki et al., 2012).

Similar to several NDs, the footprints of oxidative damage are readily identified throughout the brain of the affected patients (Brown, 2005; Elmallah, Borgmeyer, Betzel, & Redecke, 2013; Pamplona et al., 2008; Yun, Gerlach, Riederer, & Klein, 2006).

Methionine Oxidation of Prion Protein

The succinct underlying mechanisms in TSEs are unclear; however the footprints of oxidative stress exist in the aspect of pathophysiology of the disease that is understood. The chief event in the pathophysiology of TSEs is post-translational alteration of normal cellular prion protein (PrP^c) into an abnormal infectious and misfolded isoform known as scrapie (PrP^{sc}) (Elmallah et al., 2013). Scrapie is protease resistance and can be passed between individuals (Brown, 2005; Elmallah et al., 2013). In sporadic TSEs, PrP^c is converted to the abnormal misfolded infectious isoform (scrapie) via a mechanism triggered by methionine oxidation (Elmallah et al., 2013; Younan, Nadal, Davies, Brown, & Viles, 2012).

Human PrP^c is soluble and has significant susceptibility to proteinase K (PK) digestion whereas PrP^{sc} is an insoluble aggregated protein multimer with an enhanced PK resistance (Brown, 20050. Proteins with cysteine and methionine residues are the most vulnerable to oxidation by free radicals. This makes the mammalian PrP^c exceptionally prone to oxidation as a result of its high number of Met residues (Canello et al., 2008). Met-sulfoxide, the product of methionine oxidation, is often identified in brain deposited PrP^{sc} isoforms and is deemed to be a specific marker for pathogenic prion proteins (Canello et al., 2008). Little alterations PrP polypeptide can initiate a misfolding cascade of PrP^c into scrapie. Methionine oxidation into methionine sulfoxide is a trigger that begins a series of events leading to PrP misfolding. In TSEs methionine oxidation is seen as the initial trigger signal in the conversion of PrP^c into scrapie and formation of toxic species—the main pathophysiology of sporadic TSEs. Oxidation of methionine within PrPC disturbs its hydrophobic core and leads to the formation of monomeric molton-like species with high aggregation capabilities (Colombo, Meli, Morra, Gabizon, & Gasset, 2009)

Loss of Antioxidant Defense in Transmissible Spongiform Encephalopathies

Oxidative damage accounts for a majority of the prion diseases which arises sporadically (Elmallah et al., 2013). Markers of oxidative stress are even observed in early preclinical stages of scrapie infection (Yun et al., 2006). In prion diseases, there is a significant loss of antioxidant defense. As PrP^{Sc} accumulates in the brain, an increase in oxidative damage end-products and a reduction in antioxidant defense is observed. Conversion of the normal cellular prion protein which has antioxidant properties to its protease resistant isoform leads to a loss of this antioxidant activity. This process is facilitated by oxidation of methionine by H_2O_2 to methionine sulphoxide (Younan et al., 2012).

There is now a preponderance of evidence that suggests that normal cellular prion protein (PrP^{C}) is associated with cellular response to oxidative stress and that oxidative, glycoxidative, lipoxidative and nitrative protein damage to it is observed in PrD (Brazier, Doctrow, Masters, & Collins, 2008; Brown, Schmidt, & Kretzschmar, 1998; Freixes et al., 2006; Pamplona et al., 2008; Wong et al., 1999). PrP^C itself exerts anti-oxidant properties by undergoing β -cleavage (cleavage of copper-binding octapeptide repeats in PrP^C by reactive oxygen species) to protect the cell from damage from free radicals (Wong et al., 1999). Therefore β -cleavage of PrP^c is an important step in the mechanisms by which PrP defends cells from oxidative stress (Watt et al., 2005). Antioxidant capacity of purified prion protein diminishes by up to 85% in the most common human prion disease, sporadic CJD (sCJD), and correlates to elevated of markers oxidative stress in the brains of sCJD patients. Cultured astrocytes and neurons from PrP^C-deficient mice are very vulnerable to oxidative damage (Brown, Schulz-Schaeffer, Schmidt, & Kretzschmar, 1997). Immunohistochemical studies of brain tissues of prion-infected mice indicate an extensive increase in neuronal markers for lipid oxidation and other neuronal oxidative stress indicators such heme oxygenase-1 and nitrotyrosine (Guentchev, Voigtländer, Haberler, Groschup, & Budka, 2000). This supports the assertion that, ROS-mediated neuronal damage is present in the neurodegeneration observed in TSEs (Rizzardini et al., 1997; Wong et al., 2001).

The exposure to neurotoxic peptide derivatives of prion protein induced elevation of oxidative stress markers cultured astrocytes (Rizzardini et al., 1997). In a human TSE experiment, nitrotyrosine positive neurons were found throughout the brain signifying that the occurrence of oxidative stress is widespread in TSEs and also affects most neurons in the CNS (Guentchev, Groschup, Kordek, Liberski, & Budka, 1998). Free radicals-instigated oxidative stress is linked to iron accumulation and both total iron and Fe³⁺ are substantially elevated in the cerebral cortex, striatum, and brainstem of PrP^{sc} -infected mice in a scrapie model in addition to elevated malondialdehyde concentrations (Kim et al., 2000), thus implicating free radicals in TSEs.

Even though the *in vivo* function of normal prion protein still remains unknown (Sarnataro et al., 2017), it is now accepted that normal (PrP^{C}) has SOD-like activity when bound to Copper ions (Cu^{2+}) and thus may even be a marker of brain oxidative stress as it has significant antioxidant activity (Brown et al., 1999; Milhavet & Lehmann, 2002). That is why oxidative stress, due to impaired Cu^{2+} homeostasis may be a risk factor in the development of sporadic prion diseases (Requena et al., 2001). The conversion of PrP^{C} to $PrP^{S_{C}}$ leads to a loss of this SOD-like activity and an accumulation of products of oxidative damage. PrP^{C} knock-out mice also show diminutive SOD activity (Brown, Schmidt, & Kretzschmar, 1997). These evidences augment the suggestion that oxidative stress is a significant contributor of neurodegeneration in prion diseases.

Spinal Muscular Atrophy

Spinal muscular atrophy (SMA) is a common neuromuscular disorder characterized by muscle weakness and paralysis. It is caused by a deficiency in the survival of the neuron gene (*SMN1*) that leads to a reduction in the levels of functional SMN proteins (Wan, Ottinger, Cho, & Dreyfuss, 2008). The SMN complex consists of SMN and other proteins called Germins (Wan et al., 2008; Zhang et al., 2008) required for the biosynthesis of small nuclear ribonucleoprotein particles (SnRNPs) (Fischer, Liu, & Dreyfuss, 1997; Yong, Wan, & Dreyfuss, 2004). SnRNPs play essential roles in processing pre-mRNA to mRNA (Yong et al., 2004). Even though the disease has been mainly classified as a motor neuron

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disease, the SMN complex plays a key role in splicing regulation and thus making SMA splicing disease (Zhang et al., 2008).

ROS have been shown to inhibit the SMN complex (Wan et al., 2008). β -lapachone, a potent producer of ROS, along with other agents such as hydrogen peroxide, cumene hydroperoxide and environmental toxins which produce free radicals inhibit the effects of the SMN complex. This inhibition was shown to be dose dependent and the effect of the inhibition was shown to be of a functional deficiency equivalent to what happens in SMN protein deficiency. Thus, the SMN complex is very vulnerable to ROS (Wan et al., 2008).

Even though the direct role of mitochondria in the pathogenesis of SMA has not been fully elucidated, it has been shown in pre-clinical studies that, free radicals generation increases after SMN knockdown (Acsadi et al., 2009). SMN small interfering Ribonucleic acid (siRNA) transfection led to an increase in free radical production as compared to control at both 48 h and 72 h after SMN knockdown. This increase in free radicals may lead to mitochondrial DNA mutations and deletions (Acsadi et al., 2009). It was also realized that changes in mitochondrial function (which were observed as activity of caspase-3 and ATP levels) were observed before cell injury after SMN knockdown (Acsadi et al., 2009).

Traumatic Brain Injury

Traumatic brain injury (TBI) has been shown to be a major cause of death in young adults with one of its major causes being road traffic accidents (Ma et al., 2017). It has been projected by the World Health Organization that by 2020, road traffic accidents will be the third leading cause of death and disability worldwide (Finfer & Cohen, 2001) suggesting the concurrent increase in the incidence of TBI which will occur. However, in countries where there are low incidences of road traffic accidents due to safer roads and strict traffic regulations, TBIs are still reported due to falls (in the elderly) and violence suggesting that safer roads are not the only way to combat TBI (Maas, Stocchetti, & Bullock, 2008).

TBI refers to brain damage from a mechanical force (Choi et al., 2012; Maas et al., 2008) which can occur with or without loss of consciousness. Even though, some of the earlier manifestations of TBI have been managed effectively (thereby improving survival rate), most patients suffer from delayed neuronal death and cognitive impairment (Choi et al., 2012).

The initial damage from TBI is as a result of the mechanical injury which occurs at the time of impact; however secondary changes also develop later (Zhang et al., 2012). The initial injury results in skull fracture, cerebral contusions and epidural or subdural hematomas (Chakraborty, Skolnick, & Narayan, 2016). It has been shown that oxidative stress is important in the development of cerebral edema which is associated with TBI, the disruption of the blood brain barrier (BBB) and the resulting neuronal damage which occurs post-TBI (Ma et al., 2017). O_2^{-} , produced when an oxygen molecule obtains an electron from another molecule is the most common cellular free radical and the source of other ROS that leads to lipid peroxidation (Dohi et al., 2010). Excess O_2^{-} , causes tissue damage primarily through the formation of hydroxyl radical and peroxynitrite (Zhang et al., 2012). The reaction to form peroxynitrite is a diffusion-limited reaction with no requirements for enzymes. Both the •OH and ONOO• are powerful oxidants but with different biological implications as the hydroxyl radical is more promiscuous than the peroxynitrite in terms of substrate specificity (Pacher, Beckman, & Liaudet, 2007). These ROS can cause DNA damage and lipid peroxidation which could lead to inflammation in nearby tissues thus promoting apoptosis (Chakraborty et al., 2016). These reactive species can also oxidize polyunsaturated fatty acids in the cell membrane which leads to the formation of various reactive molecules including HNE. HNE is very reactive and an increase in its levels is used as a specific marker of oxidative stress (Romano et al., 2017).

The mitochondria have been known to be the major source of superoxide anion after brain injury, however it has been recently shown that NOX also plays a role. NOX usually refers to the NOX2 isoform which was characterized first. NOX2 has been shown to be highly co-localized in neurons at 1 h after TBI suggesting its significance in this disorder (Zhang et al., 2012). NOX2 has been shown to be localized in the cerebral cortex and hippocampal CA1 region and its activation has been shown to be dependent upon formation of an active complex with several phox subunits (p47phox, p67phox, p40phox) and activated Rac1, which activated and translocated to the cytoplasm from the membrane to form an active enzyme complex (Zhang et al., 2012). NOX2 has also been found to be localized in microglia and hence can contribute to neuroinflammation (Zhang et al., 2012).

FREE RADICALS AND OXIDATIVE STRESS AS TARGETS OF DRUG THERAPY IN NEURODEGENERATIVE DISORDERS

Mechanism of Synthetic Drugs and Natural Products in Reducing Free Radicals and Combating Oxidative Stress

Neurodegenerative disorders are different in their manifestations but a single common underlying factor is the induction of oxidative stress by free radicals. Different therapies have been postulated to manage these disorders including the use of agents which scavenge free radicals in an attempt to rid the body of such chemicals. To slow the progression of neurodegenerative disorders, approaches utilized include preventive therapy, disease modifying therapy and hormone therapy. Preventive therapy aims at improving the redox status (in order to ensure homeostasis) as well as preventing the resulting sequelae from the primary insult (Losada-Barreiro & Bravo-Díaz, 2017). However, most of these remedies have been shown to be ineffective in clinical trials even though positive results were obtained in pre-clinical trials (Ma et al., 2017). Mechanisms which have been exploited as potential targets to combat the various neurodegenerative disorders are elaborated on below.

NADPH Oxidase Inhibition in Traumatic Brain Injury

Because initial injury cannot be reversed in traumatic brain injury, significant research targets preventing or minimizing secondary injury (Chakraborty et al., 2016). ROS generation has been implicated in impairment brain functioning neuronal death after TBI. Both prophylactic and curative treatment with apocynin, a NADPH oxidase (NOX) inhibitor, originally isolated from the roots of *Picrorhiza kurroa* (Maraldi, 2013), significantly decreased oxidative damage in the cortex and the hippocampus in rodents (Zhang et al., 2012). Pretreatment with a specific NOX2 inhibitor, *gp91ds-tat*, also greatly reduced the neuronal damage and edema after TBI (Zhang et al., 2012). Further evidence that NADPH oxidase is an important target to combat neurodegeneration by reducing oxidative stress is adduced by the observation that *gp91phos* (NOX2) gene deficient-mice exhibited reduced primary cortical damage and ROS levels at the site injury (Dohi et al., 2010). Also, pretreatment with intraperitoneal injection of apocynin led to reduced TBI-initiated oxidative produced neuroprotective effects as observed by decreased atrophy and neuronal loss in the CA3 region of the hippocampus in rats (Choi et al., 2012). The neuroprotective

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plant extract, *Ginkgo biloba*, has also been shown to be an NADPH oxidase inhibitor which was able to reduce the activity of neutrophil-containing myeloperoxidase (Maraldi, 2013).

Oxygen Free Radical Scavengers as Neuroprotective Agents

Antioxidants produce their effects by donating their own electrons in order to counteract their effects of ROS. Generally their effects as classified by Losada-Barreiro & Bravo-Díaz (2017) may be to maintain the production of ROS to the minimum, scavenge produced ROS or restore damage ROS-target molecules.

Transcranial injection of the antioxidant glutathione, the powerful tripeptide antioxidant that efficiently scavenges the hydroxyl radicals as well as detoxifies H_2O_2 , after head injury significantly reduced inflammation and meningeal cell death (Danta & Piplani, 2014). Also, administration of N-acetylcysteine (precursor to glutathione) both in patients and rodents have shown positive outcomes in mild and moderate traumatic brain injury (Corps, Roth, & McGavern, 2015). This is one of the few successful clinical trials involving antioxidants as therapy for a neurodegenerative disorder. It is important however to note that, the likely key in using antioxidants to management TBI is to initiate therapy as soon as possible so as to avoid lesion expansion and the subsequent sequelae which results (Corps, Roth, & McGavern, 2015).

Another antioxidant with positive outcomes in both pre-clinical and clinical trials is the potent antioxidant, vitamin E. In randomized controlled, double-blind clinical trials with vitamin E in moderately impaired AD patients, the progression of functional impairment was delayed. α -Tochopherol also slows the advancement of PD in both clinical and preclinical trials (Danta & Piplani, 2014). Natural products such as *Gingko biloba*, flavonoids, soybean isoflavones and nicotine have been shown to reduce the neurotoxicity that is predominant in Alzheimer's disease by reducing oxidative stress (Velusamy et al., 2017). Other natural products including luteolin derivatives, *Calendula officinalis* flowers, olive oil and melatonin have shown possible neuroprotective effects in Huntington disease (HD) models all due to their antioxidant properties (Velusamy et al., 2017). For example, *Ginkgo biloba* has been shown to reduce the rate of 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MPTP)-induced neurodegeneration of dopaminergic neurons (Wąsik & Antkiewicz-Michaluk, 2017). This beneficial effect is due to its ability to scavenge superoxide free radicals. Additionally, *Ginkgo biloba* regulate glutathione reductase and other antioxidant systems, superoxide dismutase activity as well as its neurotrophic effects (Rojas et al., 2004). All these antioxidant therapies however are not effective as sole treatment but are effective against oxidative stress and slows the progression of HD (Manoharan et al., 2016)

Neuroprotective Steroids

Neuroprotective steroids are either synthetic or natural steroids that mainly protect the CNS or peripheral nervous system (PNS) from neurodegeneration. They include dehydroepiandrosterone, testosterone, estradiol, pregnenolone and progesterone. These agents act by mainly protecting neurons from oxidative stress, excitotoxicity and inflammation while promoting repair processes such as neurogenesis among others (Bansal & Singh, 2017). In AD, estradiol is known to regulate A β accumulation in the brain as well as protect neuronal cells from the resulting damage. It is also able to prevent the hyperphosphorylation of tau protein which is a hallmark of AD (Bansal & Singh, 2017).

Efficacy of Combatting Free Radicals in the Management of Neurodegenerative Disorders

The effectiveness of free radical scavenging agents as therapeutic agents is fraught with challenges. Thus the efficacy of some of these agents including apocynin in pre-clinical trials have not been translated in most clinical trials. Generally, this may be due to the inability of the agents to permeate the BBB or the fact that most of these agents do not target a specific oxidative mechanism (Ma et al., 2017).

One of the main challenges is the burden of off-target effects. For instance, diphenyliodonium (DPI) a very potent NOX inhibitor also has effects on cholinesterases, xanthine oxidase and calcium pump which may account for some of its adverse effects (Maraldi, 2013). Also, it has been shown that the use of NOX inhibitors targets both physiological and pathological effects of NOX and might lead to undesirable side effects. This will require the use of specific inhibitors which will help to reduce off-target effects as well as the need to selectively target only pathological NOX signaling (Ma et al., 2017).

Popular antioxidants like ascorbic acid and flavonoids as monotherapy or in combination have also not shown very positive results in human studies for the treatment of all neurodegenerative disorders (Ma et al., 2017). In certain conditions, where they prove to be beneficial, the nature of the formulation becomes a hindrance to their effective use. A perfect example is that of flavonoids which are highly polar molecules and hence are unable to cross the blood brain barrier effectively. Thus, for an antioxidant to be efficacious in ND management, it should have the following basic characteristics as classified by (Danta & Piplani, 2014):

- 1. The molecule must be able to accept electrons from reactive molecules and remain stable upon receipt of the electron
- 2. Be fairly lipophilic to facilitate crossing of the blood brain barrier and
- 3. Be fairly selective in order to prevent off-target effects

Future Research Directions Towards a Holistic Treatment of Neurodegenerative Disorders

Setbacks

There are currently no treatments for the complete amelioration of neurodegenerative diseases even though symptoms of some NDs can be partially assuaged using antioxidants. Positive results obtained in pre-clinical trials have not always been successfully replicated in humans during clinical trials (Ma et al., 2017). Oxygen-free radical scavengers have been used in clinical trials to manage several neuro-degenerative disorders but mostly with no positive outcomes (Ma et al., 2017).

With the important role played by SOD in the pathology of several neurodegenerative disorders it was expected that SOD would ameliorate several neurodegenerative disorders (Chakraborty et al., 2016). However, upon administering pegorgotein, a conjugated form of superoxide dismutase with reduced antigenicity and prolonged half-life, there was no statistically significant difference in neurologic outcome or mortality between pegorgotein and placebo-treated groups in a traumatic brain injury clinical trials (Young et al., 1996). The effects of tirilazad, an amino steroid with antioxidant and lipid peroxidation inhibition properties was also studied in clinical trials and it was observed that there was no difference in favorable outcomes or mortality between treatment and placebo groups (Chakraborty et al., 2016).

Prospects

It has become imperative to look at other approaches for mitigating oxidative stress beyond the routine antioxidant approach recognizing the various failures to translate observations in preclinical data to clinical trials.

The Endocannabinoid System

It is now believed that targeting the endocannabinoid system to regulate oxidative stress-induced cell death may hold therapeutic potential in NDs from traumatic brain injury through AD and PD to amyotrophic disorders and HD. This is backed by the evidence from effects of interaction at both cannabinoid receptors (CB1 and CB2) (Paloczi, Varga, Hasko, & Pacher, 2017). CB2 expression in microglia appears upregulated in oxidative stress related NDs, such as AD, MS, or PD although how free radicals induce CB2 receptor expression is still elusive while CB2 agonists have exerted neuroprotective effects and attenuated of neuronal cell damage (Paloczi, Varga, Hasko, & Pacher, 2017).

Additionally, the neuroprotective role of CB1 nonselective receptor agonists (WIN55, 212-2 and HU210) has been reported in a murine PD model. Interrupting CB1 signaling led to reduced ROS production and suppression of NOX which culminated in increased survival of nigrostriatal dopaminergic neurons in the striatum (Chung et al., 2011). It has also be shown that the activation of CB1 receptor by anandamide protects hippocampal neurons from oxidative injury by decreasing intracellular ROS and lowering the expression of type 2 NADPH oxidase —effects that are abolished in the presence of CB1 antagonist AM251 or CB1-siRNA (Jia et al., 2014). Moreover, the non-psychoactive phytocannabinoid, cannabidiol, shows neuroprotection by diminishing ROS levels and lipid peroxidation products in vitro and in vivo in Alzheimer's disease models (Iuvone et al., 2004). With regard to traumatic brain injury, a positive correlation occurs between tetrahydrocannabinol intake and decreased mortality in adult TBI patients (Nguyen et al., 2014) while anandamide-CB1 receptor signaling reduces the progression of clinical symptoms in G93A-SOD1 transgenic mice model of ALS (Bilsland et al., 2006).

On the contrary, other studies have also demonstrated that CB1 receptor activation may promote ROS generation in other body systems and therefore the contribution of CB1 signaling may be cell-type dependent or an indirect consequence of various processes (Steffens & Pacher, 2015). This situation warrants further studies to evaluate the interaction of cannabinoids and ROS production in the disease progression of the various neurodegenerative disorders in order to understand the complex interactions and ultimately provide more efficacious cannabinoid-based therapies with minimal offset effects for neurodegenerative disorders in future

Mitochondria Targeted Antioxidant Therapy

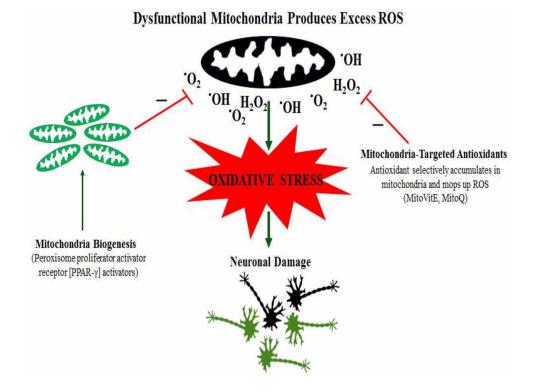
The mitochondria's physiological actions include a variety of important cellular regulatory processes including ROS generation and detoxification. With the failure to translate pre-clinical success in antioxidant therapies to clinical trials, interest in mitochondria-targeted antioxidants therapy is increasing as a potential option to circumvent this challenge. The mitochondrion is an attractive target for drug-delivery strategies it is an important source of ROS.

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This has led to the development of several investigational agents such as MitoVitE and MitoQ which are vitamins E and ubiquinone moiety of coenzyme Q10 derivative conjugated to the lipophilic cation triphylphosphonium respectively with the ability to cross the mitochondrial phospholipid bilayer, selectively accumulate in the mictochondria and mop up free radicals at their site of generation (Oyewole & Birch-Machin, 2015). The efficacy and toxicity of MitoQ have been evaluated in both pre-clinical and clinical trial for Parkinson's disease (Snow et al., 2010). MitoQ shows protection from MPTP-induced loss of behavioural activities and degeneration of dopaminergic neurons and terminals and demonstrated appreciable effects in phase I and II clinical trials (Jin et al., 2014; Snow et al., 2010). It has also been effective in mice models of Alzheimer's disease on mouse neuroblastoma (N2a) cells cultured with amyloid- β (A β) peptide (Manczak et al., 2010)

Another strategy of new interest is increasing mitochondria biogenesis. It is acknowledged that increases in neuronal mitochondrial numbers could compensate for bioenergetic dysfunction in neurodegeneration (Figure 1). Therefore, there is currently significant increase interest in peroxisome proliferator activator receptor (PPAR- γ) activators such as resveratrol, pioglitazone and rosiglitazone and the potential use to upregulate mitochondrial biogenesis and mitigate the increased oxidative burden seen in neurodegenerative disorders (Schapira, Olanow, Greenamyre, & Bezard, 2014).

Figure 1. Methods for mediating oxidative stress via reducing excess production of reactive oxygen species by dysfunctional mitochondria



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CONCLUSION

In the battle to effectively manage NDs, it has been shown that free radical-induced oxidative stress is a major player hence this chapter provides an overview of the multiple roles played by ROS in different NDs. The various mechanisms which target the generation or end effects of ROS and are being used to alleviate the sequelae resulting from these diseases have been discussed. Antioxidant systems such as the enzymes and low molecular weight reductants are reviewed with their possible mechanisms of action in certain NDs highlighted. The interplay of diet in aging and oxidative stress is also highlighted. Mechanisms of synthetic drugs and natural products in combatting ROS are reviewed with emphasis on their effectiveness in clinical trials and associated challenges. Lastly, the chapter examines novel targets requiring further research such as the endocannabinoid pathway and the mitochondria-targeted antioxidants. These have been outlined to stimulate research into novel management strategies of NDs. Great strides have been made in understanding the mechanisms of NDs, however, there is still a knowledge void that needs to be filled. Current therapeutic agents cannot adequately to provide the desired therapeutic effect, hence additional research is required to deliver efficacious therapeutic agents with minimal off-target effects.

REFERENCES

Acsadi, G., Lee, I., Li, X., Khaidakov, M., Pecinova, A., Parker, G. C., & Hüttemann, M. (2009). Mitochondrial dysfunction in a neural cell model of spinal muscular atrophy. *Journal of Neuroscience Research*, 87(12), 2748–2756. doi:10.1002/jnr.22106 PMID:19437551

Aguirre, T., van Den Bosch, L., Goetschalckx, K., Tilkin, P., Mathijis, G., Cassiman, J.-J., & Robberecht, W. (1998). Increased sensitivity of fibroblasts from amyotrophic lateral sclerosis patients to oxidative stress. *Annals of Neurology*, *43*(4), 452–457. doi:10.1002/ana.410430407 PMID:9546325

Alam, Z., Jenner, A., Daniel, S., Lees, A., Cairns, N., Marsden, C., ... Halliwell, B. (1997). Oxidative DNA damage in the parkinsonian brain: An apparent selective increase in 8-hydroxyguanine levels in substantianigra. *Journal of Neurochemistry*, 69(3), 1196–1203. doi:10.1046/j.1471-4159.1997.69031196.x PMID:9282943

Arlt, S., Beisiegel, U., & Kontush, A. (2002). Lipid peroxidation in neurodegeneration: New insights into Alzheimer's disease. *Current Opinion in Lipidology*, *13*(3), 289–294. doi:10.1097/00041433-200206000-00009 PMID:12045399

Bansal, R., & Singh, R. (2017). Exploring the potential of natural and synthetic neuroprotective steroids against neurodegenerative disorders: A literature review. *Medicinal Research Reviews*, 1098–1128. PMID:28697282

Bäumer, D., Talbot, K., & Turner, M. R. (2014). Advances in motor neurone disease. *Journal of the Royal Society of Medicine*, *107*(1), 14–21. doi:10.1177/0141076813511451 PMID:24399773

Bayram, B., Esatbeyoglu, T., Schulze, N., Ozcelik, B., Frank, J., & Rimbach, G. (2012). Comprehensive analysis of polyphenols in 55 extra virgin olive oils by HPLC-ECD and their correlation with antioxidant activities. *Plant Foods for Human Nutrition (Dordrecht, Netherlands)*, 67(4), 326–336. doi:10.100711130-012-0315-z PMID:23070730

Bhat, A. H., Dar, K. B., Anees, S., Zargar, M. A., Masood, A., Sofi, M. A., & Ganie, S. A. (2015). Oxidative stress, mitochondrial dysfunction and neurodegenerative diseases; a mechanistic insight. *Biomedicine and Pharmacotherapy*, *74*, 101–110. doi:10.1016/j.biopha.2015.07.025 PMID:26349970

Bickford, P. C., Flowers, A., & Grimmig, B. (2017). Aging leads to altered microglial function that reduces brain resiliency increasing vulnerability to neurodegenerative diseases. *Experimental Gerontology*, *94*, 4–8. doi:10.1016/j.exger.2017.01.027 PMID:28163132

Bilsland, L. G., Dick, J. R., Pryce, G., Petrosino, S., Di Marzo, V., Baker, D., & Greensmith, L. (2006). Increasing cannabinoid levels by pharmacological and genetic manipulation delay disease progression in SOD1 mice. *The FASEB Journal*, *20*(7), 1003–1005. doi:10.1096/fj.05-4743fje PMID:16571781

Birben, E., Sahiner, U., Sackesen, C., Erzurum, S., & Kalayci, O. (2012). Oxidative stress and antioxidant defense. *The World Allergy Organization Journal*, *5*(1), 9–19. doi:10.1097/WOX.0b013e3182439613 PMID:23268465

Brazier, M. W., Doctrow, S. R., Masters, C. L., & Collins, S. J. (2008). A manganese-superoxide dismutase/ catalase mimetic extends survival in a mouse model of human prion disease. *Free Radical Biology & Medicine*, 45(2), 184–192. doi:10.1016/j.freeradbiomed.2008.04.006 PMID:18455516

Brown, D. R. (2005). ORGINAL ARTICLE Neurodegeneration and oxidative stress: Prion disease results from loss of antioxidant defence. *Folia Neuropathologica*, 43(4), 229–243. PMID:16416388

Brown, D. R., Boon-Seng, W., Hafiz, F., & Clive, C. (1999). Normal prion protein has an activity like that of superoxide dismutase. *The Biochemical Journal*, 344(1), 1–5. doi:10.1042/bj3440001 PMID:10548526

Brown, D. R., Schmidt, B., & Kretzschmar, H. A. (1997). Effects of oxidative stress on prion protein expression in PC12 cells. *International Journal of Developmental Neuroscience*, *15*(8), 961–972. doi:10.1016/S0736-5748(97)00042-7 PMID:9641527

Brown, D. R., Schmidt, B., & Kretzschmar, H. A. (1998). A prion protein fragment primes type 1 astrocytes to proliferation signals from microglia. *Neurobiology of Disease*, *4*(6), 410–422. doi:10.1006/nbdi.1998.0169 PMID:9666480

Brown, D. R., Schulz-Schaeffer, W. J., Schmidt, B., & Kretzschmar, H. A. (1997). Prion Protein-Deficient Cells Show Altered Response to Oxidative Stress Due to Decreased SOD-1 Activity. *Experimental Neurology*, *146*(1), 104–112. doi:10.1006/exnr.1997.6505 PMID:9225743

Butterfield, D. A., & Boyd-Kimball, D. (2004). Amyloid β-peptide (1-42) contributes to the oxidative stress and neurodegeneration found in Alzheimer disease Brain. *Brain Pathology (Zurich, Switzerland)*, *14*(4), 426–432. doi:10.1111/j.1750-3639.2004.tb00087.x PMID:15605990

Free Radicals in Oxidative Stress, Aging, and Neurodegenerative Disorders

Butterfield, D. A., Castegna, A., Lauderback, C. M., & Drake, J. (2002). Evidence that amyloid betapeptide-induced lipid peroxidation and its sequelae in Alzheimer's disease brain contribute to neuronal death. *Neurobiology of Aging*, 23(5), 655–664. doi:10.1016/S0197-4580(01)00340-2 PMID:12392766

Cabezas, R., Ávila, M., Gonzalez, J., El-Bachá, R. S., Báez, E., García-Segura, L. M., ... Barreto, G. E. (2014). Astrocytic modulation of blood brain barrier: Perspectives on Parkinson's disease. *Frontiers in Cellular Neuroscience*, *8*, 211. doi:10.3389/fncel.2014.00211 PMID:25136294

Canello, T., Engelstein, R., Moshel, O., Xanthopoulos, K., Juanes, M. E., Langeveld, J., ... Gabizon, R. (2008). Methionine sulfoxides on PrPSc: A prion-specific covalent signature. *Biochemistry*, 47(34), 8866–8873. doi:10.1021/bi800801f PMID:18680312

Carrì, M. T., Valle, C., Bozzo, F., & Cozzolino, M. (2015). Oxidative stress and mitochondrial damage: Importance in non-SOD1 ALS. *Frontiers in Cellular Neuroscience*, *9*, 4. PMID:25741238

Carvalho, A. N., Lim, J. L., Nijland, P. G., Witte, M. E., & Van Horssen, J. (2014). Glutathione in multiple sclerosis: More than just an antioxidant? *Multiple Sclerosis Journal*, 20(11), 1425–1431. doi:10.1177/1352458514533400 PMID:24842957

Chakraborty, S., Skolnick, B., & Narayan, R. K. (2016). Neuroprotection trials in traumatic brain injury. *Current Neurology and Neuroscience Reports*, *16*(4), 29. doi:10.100711910-016-0625-x PMID:26883431

Choi, B. Y., Jang, B. G., Kim, J. H., Lee, B. E., Sohn, M., Song, H. K., & Suh, S. W. (2012). Prevention of traumatic brain injury-induced neuronal death by inhibition of NADPH oxidase activation. *Brain Research*, *1481*, 49–58. doi:10.1016/j.brainres.2012.08.032 PMID:22975130

Chung, Y. C., Bok, E., Huh, S. H., Park, J.-Y., Yoon, S.-H., Kim, S. R., ... Jin, B. K. (2011). Cannabinoid receptor type 1 protects nigrostriatal dopaminergic neurons against MPTP neurotoxicity by inhibiting microglial activation. *Journal of Immunology (Baltimore, Md.: 1950)*, *187*(12), 6508–6517. doi:10.4049/ jimmunol.1102435 PMID:22079984

Coelho, R. C. L. A., Hermsdorff, H. H. M., & Bressan, J. (2013). Anti-inflammatory properties of orange juice: Possible favorable molecular and metabolic effects. *Plant Foods for Human Nutrition (Dordrecht, Netherlands)*, 68(1), 1–10. doi:10.100711130-013-0343-3 PMID:23417730

Colombo, G., Meli, M., Morra, G., Gabizon, R., & Gasset, M. (2009). Methionine sulfoxides on prion protein helix-3 switch on the α -fold destabilization required for conversion. *PLoS One*, 4(1), e4296. doi:10.1371/journal.pone.0004296 PMID:19172188

Cookson, M. R., & Shaw, P. J. (1999). Oxidative stress and motor neurone disease. *Brain Pathology* (*Zurich, Switzerland*), 9(1), 165–186. doi:10.1111/j.1750-3639.1999.tb00217.x PMID:9989458

Corps, K. N., Roth, T. L., & McGavern, D. B. (2015). Inflammation and neuroprotection in traumatic brain injury. *JAMA Neurology*, 72(3), 355–362. doi:10.1001/jamaneurol.2014.3558 PMID:25599342

Danta, C. C., & Piplani, P. (2014). The discovery and development of new potential antioxidant agents for the treatment of neurodegenerative diseases. *Expert Opinion on Drug Discovery*, 9(10), 1205–1222. doi:10.1517/17460441.2014.942218 PMID:25056182

Dexter, D., Carter, C., Wells, F., Javoy-Agid, F., Agid, Y., Lees, A., ... Marsden, C. D. (1989). Basal lipid peroxidation in substantia nigra is increased in Parkinson's disease. *Journal of Neurochemistry*, *52*(2), 381–389. doi:10.1111/j.1471-4159.1989.tb09133.x PMID:2911023

Dias, V., Junn, E., & Mouradian, M. M. (2013). The role of oxidative stress in Parkinson's disease. *Journal of Parkinson's Disease*, *3*(4), 461–491. PMID:24252804

Diehl, J., Grimmer, T., Drzezga, A., Riemenschneider, M., Förstl, H., & Kurz, A. (2004). Cerebral metabolic patterns at early stages of frontotemporal dementia and semantic dementia. A PET study. *Neurobiology of Aging*, *25*(8), 1051–1056. doi:10.1016/j.neurobiolaging.2003.10.007 PMID:15212830

Dixit, S., Bernardo, A., Walker, J. M., Kennard, J. A., Kim, G. Y., Kessler, E. S., & Harrison, F. E. (2015). Vitamin C deficiency in the brain impairs cognition, increases amyloid accumulation and deposition, and oxidative stress in APP/PSEN1 and normally aging mice. *ACS Chemical Neuroscience*, *6*(4), 570–581. doi:10.1021/cn500308h PMID:25642732

Dohi, K., Ohtaki, H., Nakamachi, T., Yofu, S., Satoh, K., Miyamoto, K., ... Aruga, T. (2010). Gp91 phox (NOX2) in classically activated microglia exacerbates traumatic brain injury. *Journal of Neuroinflammation*, *7*(1), 41. doi:10.1186/1742-2094-7-41 PMID:20659322

Dröge, W. (2002). Free radicals in the physiological control of cell function. *Physiological Reviews*, 82(1), 47–95. doi:10.1152/physrev.00018.2001 PMID:11773609

Elmallah, M. I., Borgmeyer, U., Betzel, C., & Redecke, L. (2013). Impact of methionine oxidation as an initial event on the pathway of human prion protein conversion. *Prion*, 7(5), 404–411. doi:10.4161/pri.26745 PMID:24121542

Fenn, A. M., Henry, C. J., Huang, Y., Dugan, A., & Godbout, J. P. (2012). Lipopolysaccharide-induced interleukin (IL)-4 receptor-alpha expression and corresponding sensitivity to the M2 promoting effects of IL-4 are impaired in microglia of aged mice. *Brain, Behavior, and Immunity*, 26(5), 766–777. doi:10.1016/j.bbi.2011.10.003 PMID:22024136

Ferreira, M. E., de Vasconcelos, A. S., da Costa Vilhena, T., da Silva, T. L., da Silva Barbosa, A., Gomes, A. R., ... Percário, S. (2015). Oxidative Stress in Alzheimer's Disease: Should We Keep Trying Antioxidant Therapies? *Cellular and Molecular Neurobiology*, *35*(5), 595–614. doi:10.100710571-015-0157-y PMID:25616523

Finfer, S. R., & Cohen, J. (2001). Severe traumatic brain injury. *Resuscitation*, 48(1), 77–90. doi:10.1016/S0300-9572(00)00321-X PMID:11162885

Fischer, U., Liu, Q., & Dreyfuss, G. (1997). The SMN–SIP1 complex has an essential role in spliceosomal snRNP biogenesis. *Cell*, *90*(6), 1023–1029. doi:10.1016/S0092-8674(00)80368-2 PMID:9323130

Freixes, M., Rodríguez, A., Dalfó, E., & Ferrer, I. (2006). Oxidation, glycoxidation, lipoxidation, nitration, and responses to oxidative stress in the cerebral cortex in Creutzfeldt–Jakob disease. *Neurobiology* of Aging, 27(12), 1807–1815. doi:10.1016/j.neurobiolaging.2005.10.006 PMID:16310893 Gabbita, S. P., Lovell, M. A., & Markesbery, W. R. (1998). Increased nuclear DNA oxidation in the brain in Alzheimer's disease. *Journal of Neurochemistry*, 71(5), 2034–2040. doi:10.1046/j.1471-4159.1998.71052034.x PMID:9798928

Garcia-Ruiz, P. J., Chaudhuri, K. R., & Martinez-Martin, P. (2014). Non-motor symptoms of Parkinson's disease A review... from the past. *Journal of the Neurological Sciences*, *338*(1), 30–33. doi:10.1016/j. jns.2014.01.002 PMID:24433931

Gerlach, M., Ben-Shachar, D., Riederer, P., & Youdim, M. B. (1994). Altered brain metabolism of iron as a cause of neurodegenerative diseases? *Journal of Neurochemistry*, *63*(3), 793–807. doi:10.1046/j.1471-4159.1994.63030793.x PMID:7519659

Ghadge, G. D., Lee, J. P., Bindokas, V. P., Jordan, J., Ma, L., Miller, R. J., & Roos, R. P. (1997). Mutant Superoxide Dismutase-1-Linked Familial Amyotrophic Lateral Sclerosis: Molecular Mechanisms of Neuronal Death and Protection. *The Journal of Neuroscience*, *17*(22), 8756–8766. PMID:9348345

Gomberg, M. (1900). An instance of trivalent carbon: Triphenylmethyl. *Journal of the American Chemical Society*, 22(11), 757–771. doi:10.1021/ja02049a006

Gomperts, S. N., Locascio, J. J., Marquie, M., Santarlasci, A. L., Rentz, D. M., Maye, J., ... Growdon, J. H. (2012). Brain amyloid and cognition in Lewy body diseases. *Movement Disorders*, *27*(8), 965–973. doi:10.1002/mds.25048 PMID:22693110

Guentchev, M., Groschup, M. H., Kordek, R., Liberski, P. P., & Budka, H. (1998). Severe, early and selective loss of a subpopulation of GABAergic inhibitory neurons in experimental transmissible spongiform encephalopathies. *Brain Pathology (Zurich, Switzerland)*, 8(4), 615–623. doi:10.1111/j.1750-3639.1998. tb00188.x PMID:9804371

Guentchev, M., Voigtländer, T., Haberler, C., Groschup, M. H., & Budka, H. (2000). Evidence for oxidative stress in experimental prion disease. *Neurobiology of Disease*, 7(4), 270–273. doi:10.1006/nbdi.2000.0290 PMID:10964599

Halliwell, B. (1992). Reactive Oxygen Species and the Central Nervous System. *Journal of Neurochemistry*, *59*(5), 1609–1623. doi:10.1111/j.1471-4159.1992.tb10990.x PMID:1402908

Halliwell, B. (2007). Biochemistry of oxidative stress. *Biochemistry Society Transanctions*, 35(5), 1147–1150. doi:10.1042/BST0351147 PMID:17956298

Halliwell, B., & Gutteridge, J. M. (1990). Role of free radicals and catalytic metal ions in human disease: An overview. *Methods in Enzymology*, *186*, 1–85. doi:10.1016/0076-6879(90)86093-B PMID:2172697

Harman, D. (1956). Aging: A theory based on free radical and radiation chemistry. *Journal of Gerontology*, *11*(3), 298–300. doi:10.1093/geronj/11.3.298 PMID:13332224

Hockenbery, D. M., Oltvai, Z. N., Yin, X.-M., Milliman, C. L., & Korsmeyer, S. J. (1993). Bcl-2 functions in an antioxidant pathway to prevent apoptosis. *Cell*, 75(2), 241–251. doi:10.1016/0092-8674(93)80066-N PMID:7503812

Huang, W. J., Zhang, X., & Chen, W. W. (2016). Role of oxidative stress in Alzheimer's disease. *Biomedical Reports*, 4(5), 519–522. doi:10.3892/br.2016.630 PMID:27123241 Iuvone, T., Esposito, G., Esposito, R., Santamaria, R., Di Rosa, M., & Izzo, A. A. (2004). Neuroprotective effect of cannabidiol, a non-psychoactive component from Cannabis sativa, on β-amyloid-induced toxicity in PC12 cells. *Journal of Neurochemistry*, *89*(1), 134–141. doi:10.1111/j.1471-4159.2003.02327.x PMID:15030397

Iwasaki, Y., Mori, K., & Ito, M. (2012). Investigation of the clinical course and treatment of prion disease patients in the akinetic mutism state in Japan. *Clinical Neurology*, *52*(5), 314-319.

Jia, J., Ma, L., Wu, M., Zhang, L., Zhang, X., Zhai, Q., ... Xiong, L. (2014). Anandamide protects HT22 cells exposed to hydrogen peroxide by inhibiting CB1 receptor-mediated type 2 NADPH oxidase. *Oxida-tive Medicine and Cellular Longevity*. PMID:25136404

Jin, H., Kanthasamy, A., Ghosh, A., Anantharam, V., Kalyanaraman, B., & Kanthasamy, A. G. (2014). Mitochondria-targeted antioxidants for treatment of Parkinson's disease: Preclinical and clinical outcomes. *Molecular Basis of Disease*, *1842*(8), 1282–1294. doi:10.1016/j.bbadis.2013.09.007 PMID:24060637

Johnson, W. M., Wilson-Delfosse, A. L., & Mieyal, J. J. (2012). Dysregulation of glutathione homeostasis in neurodegenerative diseases. *Nutrients*, 4(10), 1399–1440. doi:10.3390/nu4101399 PMID:23201762

Joshi, Y. B., & Praticò, D. (2014). Lipid peroxidation in psychiatric illness: Overview of clinical evidence. *Oxidative Medicine and Cellular Longevity*. PMID:24868318

Khan, T., Hassan, I., Ahmad, A., Perveen, A., Aman, S., Quddusi, S., ... Aliev, G. (2016). Recent updates on the dynamic association between oxidative stress and neurodegenerative disorders. *CNS & Neurological Disorders - Drug Targets*, *15*(3), 310–320. doi:10.2174/1871527315666160202124518 PMID:26831262

Kim, N.-H., Park, S.-J., Jin, J.-K., Kwon, M.-S., Choi, E.-K., Carp, R. I., & Kim, Y.-S. (2000). Increased ferric iron content and iron-induced oxidative stress in the brains of scrapie-infected mice. *Brain Research*, 884(1), 98–103. doi:10.1016/S0006-8993(00)02907-3 PMID:11082491

Koellhoffer, E., McCullough, L., & Ritzel, R. (2017). Old maids: Aging and its impact on microglia function. *International Journal of Molecular Sciences*, *18*(4), 769. doi:10.3390/ijms18040769 PMID:28379162

Lee, D.-H., Gold, R., & Linker, R. A. (2012). Mechanisms of oxidative damage in multiple sclerosis and neurodegenerative diseases: Therapeutic modulation via fumaric acid esters. *International Journal of Molecular Sciences*, *13*(9), 11783–11803. doi:10.3390/ijms130911783 PMID:23109883

Lee, J., Kosaras, B., Del Signore, S. J., Cormier, K., McKee, A., Ratan, R. R., ... Ryu, H. (2011). Modulation of lipid peroxidation and mitochondrial function improves neuropathology in Huntington's disease mice. *Acta Neuropathologica*, *121*(4), 487–498. doi:10.100700401-010-0788-5 PMID:21161248

Leigh, P. N., & Ray-Chaudhuri, K. (1994). Motor neuron disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 57(8), 886–896. doi:10.1136/jnnp.57.8.886 PMID:8057109

Logroscino, G., Traynor, B. J., Hardiman, O., Chio, A., Couratier, P., Mitchell, J. D., ... Beghi, E. (2008). Descriptive epidemiology of amyotrophic lateral sclerosis: New evidence and unsolved issues. *Journal of Neurology, Neurosurgery, and Psychiatry*, 79(1), 6–11. doi:10.1136/jnnp.2006.104828 PMID:18079297

Free Radicals in Oxidative Stress, Aging, and Neurodegenerative Disorders

Losada-Barreiro, S., & Bravo-Díaz, C. (2017). Free radicals and polyphenols: The redox chemistry of neurodegenerative diseases. *European Journal of Medicinal Chemistry*, *133*, 379–402. doi:10.1016/j. ejmech.2017.03.061 PMID:28415050

Lushchak, V. I. (2014). Free radicals, reactive oxygen species, oxidative stress and its classification. *Chemico-Biological Interactions*, 224, 164–175. doi:10.1016/j.cbi.2014.10.016 PMID:25452175

Ma, M. W., Wang, J., Zhang, Q., Wang, R., Dhandapani, K. M., Vadlamudi, R. K., & Brann, D. W. (2017). NADPH oxidase in brain injury and neurodegenerative disorders. *Molecular Neurodegeneration*, *12*(1), 7. doi:10.118613024-017-0150-7 PMID:28095923

Maas, A. I., Stocchetti, N., & Bullock, R. (2008). Moderate and severe traumatic brain injury in adults. *Lancet Neurology*, 7(8), 728–741. doi:10.1016/S1474-4422(08)70164-9 PMID:18635021

Magrassi, L., Leto, K., & Rossi, F. (2013). Lifespan of neurons is uncoupled from organismal lifespan. *Proceedings of the National Academy of Sciences of the United States of America*, *110*(11), 4374–4379. doi:10.1073/pnas.1217505110 PMID:23440189

Manczak, M., Mao, P., Calkins, M. J., Cornea, A., Reddy, A. P., Murphy, M. P., ... Reddy, P. H. (2010). Mitochondria-targeted antioxidants protect against amyloid-β toxicity in Alzheimer's disease neurons. *Journal of Alzheimer's Disease*, 20(2), S609–S631. doi:10.3233/JAD-2010-100564 PMID:20463406

Manoharan, S., Guillemin, G. J., Abiramasundari, R. S., Essa, M. M., Akbar, M., & Akbar, M. D. (2016). The Role of Reactive Oxygen Species in the Pathogenesis of Alzheimer's Disease, Parkinson's Disease, and Huntington's Disease: A Mini Review. *Oxidative Medicine and Cellular Longevity*, 2016, 8590578. doi:10.1155/2016/8590578 PMID:28116038

Maraldi, T. (2013). Natural compounds as modulators of NADPH oxidases. *Oxidative Medicine and Cellular Longevity*. PMID:24381714

Mendez, M. F., & Shapira, J. S. (2011). Loss of emotional insight in behavioral variant frontotemporal dementia or "frontal anosodiaphoria". *Consciousness and Cognition*, 20(4), 1690–1696. doi:10.1016/j. concog.2011.09.005 PMID:21959203

Milhavet, O., & Lehmann, S. (2002). Oxidative stress and the prion protein in transmissible spongiform encephalopathies. *Brain Research. Brain Research Reviews*, *38*(3), 328–339. doi:10.1016/S0165-0173(01)00150-3 PMID:11890980

Moll, L., El-Ami, T., & Cohen, E. (2014). Selective manipulation of aging: A novel strategy for the treatment of neurodegenerative disorders. *Swiss Medical Weekly*, *144*, w13917. PMID:24526357

Nguyen, B. M., Kim, D., Bricker, S., Bongard, F., Neville, A., Putnam, B., ... Plurad, D. (2014). Effect of marijuana use on outcomes in traumatic brain injury. *The American Surgeon*, *80*(10), 979–983. PMID:25264643

Ohta, Y., Yashiro, K., Ohashi, K., & Imai, Y. (2012). Disruption of non-enzymatic antioxidant defense systems in the brain of rats with water-immersion restraint stress. *Journal of Clinical Biochemistry and Nutrition*, *51*(2), 136–142. doi:10.3164/jcbn.11-14 PMID:22962533

Olkowski, Z. L. (1998). Mutant AP endonuclease in patients with amyotrophic lateral sclerosis. *Neuroreport*, 9(2), 239–242. doi:10.1097/00001756-199801260-00012 PMID:9507962

Oyewole, A. O., & Birch-Machin, M. A. (2015). Mitochondria-targeted antioxidants. *The FASEB Journal*, 29(12), 4766–4771. doi:10.1096/fj.15-275404 PMID:26253366

Pacher, P., Beckman, J. S., & Liaudet, L. (2007). Nitric oxide and peroxynitrite in health and disease. *Physiological Reviews*, 87(1), 315–424. doi:10.1152/physrev.00029.2006 PMID:17237348

Paloczi, J., Varga, Z. V., Hasko, G., & Pacher, P. (in press). Neuroprotection in Oxidative Stress-Related Neurodegenerative Diseases: Role of Endocannabinoid System Modulation. *Antioxidants & Redox Signalling*. doi:10.1089/ars.2017.7144

Pamplona, R., Naudí, A., Gavín, R., Pastrana, M. A., Sajnani, G., Ilieva, E. V., ... Requena, J. R. (2008). Increased oxidation, glycoxidation, and lipoxidation of brain proteins in prion disease. *Free Radical Biology & Medicine*, 45(8), 1159–1166. doi:10.1016/j.freeradbiomed.2008.07.009 PMID:18703134

Panday, A., Sahoo, M. K., Osorio, D., & Batra, S. (2015). NADPH oxidases: An overview from structure to innate immunity-associated pathologies. *Cellular & Molecular Immunology*, *12*(1), 5–23. doi:10.1038/cmi.2014.89 PMID:25263488

Pedersen, W. A., Fu, W., Keller, J. N., Markesbery, W. R., Appel, S., Smith, R. G., ... Mattson, M. P. (1998). Protein modification by the lipid peroxidation product 4-hydroxynonenal in the spinal cords of amyotrophic lateral sclerosis patients. *Annals of Neurology*, *44*(5), 819–824. doi:10.1002/ana.410440518 PMID:9818940

Postuma, R. B., Gagnon, J. F., Pelletier, A., & Montplaisir, J. (2013). Prodromal autonomic symptoms and signs in Parkinson's disease and dementia with Lewy bodies. *Movement Disorders*, 28(5), 597–604. doi:10.1002/mds.25445 PMID:23554107

Pringsheim, T., Jette, N., Frolkis, A., & Steeves, T. D. L. (2014). The prevalence of Parkinson's disease: A systematic review and meta-analysis. *Movement Disorders*, 29(13), 1583–1590. doi:10.1002/mds.25945 PMID:24976103

Prusiner, S. B. (1991). Molecular biology of prion diseases. *Science*, 252(5012), 1515–1522. doi:10.1126cience.1675487 PMID:1675487

Rademakers, R., Neumann, M., & Mackenzie, I. R. (2012). Advances in understanding the molecular basis of frontotemporal dementia. *Nature Reviews. Neurology*, 8(8), 423–434. doi:10.1038/nrneurol.2012.117 PMID:22732773

Requena, J. R., Groth, D., Legname, G., Stadtman, E. R., Prusiner, S. B., & Levine, R. L. (2001). Copper-catalyzed oxidation of the recombinant SHa (29–231) prion protein. *Proceedings of the National Academy of Sciences of the United States of America*, *98*(13), 7170–7175. doi:10.1073/pnas.121190898 PMID:11404462

Rinaldi, P., Polidori, M. C., Metastasio, A., Mariani, E., Mattioli, P., Cherubini, A., ... Mecocci, P. (2003). Plasma antioxidants are similarly depleted in mild cognitive impairment and in Alzheimer's disease. *Neurobiology of Aging*, *24*(7), 915–919. doi:10.1016/S0197-4580(03)00031-9 PMID:12928050

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Rizzardini, M., Chiesa, R., Angeretti, N., Lucca, E., Salmona, M., Forloni, G., & Cantoni, L. (1997). Prion Protein Fragment 106–126 Differentially Induces Heme Oxygenase-1 mRNA in Cultured Neurons and Astroglial Cells. *Journal of Neurochemistry*, 68(2), 715–720. doi:10.1046/j.1471-4159.1997.68020715.x PMID:9003061

Rojas, P., Rojas, C., Ebadi, M., Montes, S., Monroy-Noyola, A., & Serrano-García, N. (2004). EGb761 pretreatment reduces monoamine oxidase activity in mouse corpus striatum during 1-methyl-4-phenylpyridinium neurotoxicity. *Neurochemical Research*, *29*(7), 1417–1423. doi:10.1023/ B:NERE.0000026406.64547.93 PMID:15202774

Romano, A., Serviddio, G., Calcagnini, S., Villani, R., Giudetti, A. M., Cassano, T., & Gaetani, S. (2017). Linking lipid peroxidation and neuropsychiatric disorders: Focus on 4-hydroxy-2-nonenal. *Free Radical Biology & Medicine*, *111*, 281–293. doi:10.1016/j.freeradbiomed.2016.12.046 PMID:28063940

Rosen, D. R., Siddiquef, T., Patterson, D., Figlewicz, D. A., Sapp, P., Hentatif, A., ... Cayabyabi, A. (1993). Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic. *Nature*, *362*(6415), 4. doi:10.1038/362059a0 PMID:8446170

Sarnataro, D., Pepe, A., & Zurzolo, C. (in press). Cell Biology of Prion Protein. *Progress in Molecular Biology and Translational Science*. PMID:28838675

Sayre, L. M., Smith, M. A., & Perry, G. (2001). Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Current Medicinal Chemistry*, 8(7), 721–738. doi:10.2174/0929867013372922 PMID:11375746

Schapira, A. H. V., Olanow, C. W., Greenamyre, J. T., & Bezard, E. (2014). Slowing of neurodegeneration in Parkinson's disease and Huntington's disease: Future therapeutic perspectives. *Lancet*, *384*(9942), 545–555. doi:10.1016/S0140-6736(14)61010-2 PMID:24954676

Schneider, S. A., & Obeso, J. A. (2014). Clinical and pathological features of Parkinson's disease. In Behavioral Neurobiology of Huntington's Disease and Parkinson's Disease. Springer Berlin Heidelberg. doi:10.1007/7854_2014_317

Selley, M., Close, D., & Stern, S. (2002). The effect of increased concentrations of homocysteine on the concentration of (E)-4-hydroxy-2-nonenal in the plasma and cerebrospinal fluid of patients with Alzheimer's disease. *Neurobiology of Aging*, *23*(3), 383–388. doi:10.1016/S0197-4580(01)00327-X PMID:11959400

Shaw, P. J. (1999). Motor neurone disease. *British Medical Journal*, *318*(7191), 1118–1121. doi:10.1136/ bmj.318.7191.1118 PMID:10213726

Shaw, P. J., Chinnery, R. M., Thagesen, H., Borthwick, G. M., & Ince, P. G. (1997). Immunocytochemical study of the distribution of the free radical scavenging enzymes Cu/Zn superoxide dismutase (SOD1); MN superoxide dismutase (MN SOD) and catalase in the normal human spinal cord and in motor neuron disease. *Journal of the Neurological Sciences*, *147*(2), 115–125. doi:10.1016/S0022-510X(96)05316-6 PMID:9106116

Shibata, N., Hirano, A., Yamamoto, T., Kato, Y., & Kobayashi, M. (2000). Superoxide dismutase-1 mutation-related neurotoxicity in familial amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, 1(3), 143–161. doi:10.1080/14660820050515151 PMID:11464949

Sies, H. (1997). Oxidative stress: Oxidants and antioxidants. *Experimental Physiology*, 82(2), 291–295. doi:10.1113/expphysiol.1997.sp004024 PMID:9129943

Sinclair, A. J., Bayer, A. J., Johnston, J., Warner, C., & Maxwell, S. R. (1998). Altered plasma antioxidant status in subjects with Alzheimer's disease and vascular dementia. *International Journal of Geriatric Psychiatry*, *13*(12), 840–845. doi: PMID:9884908

Smith, D. G., Cappai, R., & Barnham, K. J. (2007). The redox chemistry of the Alzheimer's disease amyloid β peptide. *Biochimica et Biophysica Acta (BBA)-. Biomembranes*, *1768*(8), 1976–1990. doi:10.1016/j.bbamem.2007.02.002

Snow, B. J., Rolfe, F. L., Lockhart, M. M., Frampton, C. M., O'Sullivan, J. D., Fung, V., ... Taylor, K. M. (2010). A double-blind, placebo-controlled study to assess the mitochondria-targeted antioxidant MitoQ as a disease-modifying therapy in Parkinson's disease. *Movement Disorders*, 25(11), 1670–1674. doi:10.1002/mds.23148 PMID:20568096

Sosa-Ortiz, A. L., Acosta-Castillo, I., & Prince, M. J. (2012). Epidemiology of dementias and Alzheimer's disease. *Archives of Medical Research*, *43*(8), 600–608. doi:10.1016/j.arcmed.2012.11.003 PMID:23159715

Steffens, S., & Pacher, P. (2015). The activated endocannabinoid system in atherosclerosis: Driving force or protective mechanism? *Current Drug Targets*, *16*(4), 334–341. doi:10.2174/13894501156661 41202113225 PMID:25469884

Sultana, R., Perluigi, M., & Butterfield, D. A. (2013). Lipid peroxidation triggers neurodegeneration: A redox proteomics view into the Alzheimer disease brain. *Free Radical Biology & Medicine*, 62, 157–169. doi:10.1016/j.freeradbiomed.2012.09.027 PMID:23044265

Turner, M. R., Bowser, R., Bruijn, L., Dupuis, L., Ludolph, A., McGrath, M., . . . Pullman, S. L. (2013). Mechanisms, models and biomarkers in amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration*, *14*(sup1), 19-32.

Valencia, A., Sapp, E., Kimm, J. S., McClory, H., Reeves, P. B., Alexander, J., ... Kegel, K. B. (2012). Elevated NADPH oxidase activity contributes to oxidative stress and cell death in Huntington's disease. *Human Molecular Genetics*, 22(6), 1112–1131. doi:10.1093/hmg/dds516 PMID:23223017

Velusamy, T., Panneerselvam, A. S., Purushottam, M., Anusuyadevi, M., Pal, P. K., Jain, S., ... Kandasamy, M. (2017). Protective Effect of Antioxidants on Neuronal Dysfunction and Plasticity in Huntington's Disease. *Oxidative Medicine and Cellular Longevity*. PMID:28168008

Wan, L., Ottinger, E., Cho, S., & Dreyfuss, G. (2008). Inactivation of the SMN complex by oxidative stress. *Molecular Cell*, *31*(2), 244–254. doi:10.1016/j.molcel.2008.06.004 PMID:18657506

Wąsik, A., & Antkiewicz-Michaluk, L. (2017). The mechanism of neuroprotective action of natural compounds. *Pharmacological Reports*, 69(5), 851–860. doi:10.1016/j.pharep.2017.03.018 PMID:28623709

Watt, N. T., Taylor, D. R., Gillott, A., Thomas, D. A., Perera, W. S. S., & Hooper, N. M. (2005). Reactive oxygen species-mediated β -cleavage of the prion protein in the cellular response to oxidative stress. *The Journal of Biological Chemistry*, 280(43), 35914–35921. doi:10.1074/jbc.M507327200 PMID:16120605

Wijesekera, L. C., & Leigh, P. N. (2009). Amyotrophic lateral sclerosis. Orphanet Journal of Rare Diseases, 4(1), 3. doi:10.1186/1750-1172-4-3 PMID:19192301

Wong, B. S., Brown, D. R., Pan, T., Whiteman, M., Liu, T., Bu, X., ... Rubenstein, R. (2001). Oxidative impairment in scrapie-infected mice is associated with brain metals perturbations and altered antioxidant activities. *Journal of Neurochemistry*, 79(3), 689–698. doi:10.1046/j.1471-4159.2001.00625.x PMID:11701772

Wong, B.-S., Wang, H., Brown, D. R., & Jones, I. M. (1999). Selective oxidation of methionine residues in prion proteins. *Biochemical and Biophysical Research Communications*, 259(2), 352–355. doi:10.1006/bbrc.1999.0802 PMID:10362513

Yong, J., Wan, L., & Dreyfuss, G. (2004). Why do cells need an assembly machine for RNA–protein complexes? *Trends in Cell Biology*, *14*(5), 226–232. doi:10.1016/j.tcb.2004.03.010 PMID:15130578

Younan, N. D., Nadal, R. C., Davies, P., Brown, D. R., & Viles, J. H. (2012). Methionine oxidation perturbs the structural core of the prion protein and suggests a generic misfolding pathway. *The Journal of Biological Chemistry*, 287(34), 28263–28275. doi:10.1074/jbc.M112.354779 PMID:22654104

Young, B., Runge, J. W., Waxman, K. S., Harrington, T., Wilberger, J., Muizelaar, J. P., ... Kupiec, J. W. (1996). Effects of pegorgotein on neurologic outcome of patients with severe head injury: A multicenter, randomized controlled trial. *Journal of the American Medical Association*, 276(7), 538–543. doi:10.1001/jama.1996.03540070034027 PMID:8709402

Yun, S.-W., Gerlach, M., Riederer, P., & Klein, M. A. (2006). Oxidative stress in the brain at early preclinical stages of mouse scrapie. *Experimental Neurology*, 201(1), 90–98. doi:10.1016/j.expneurol.2006.03.025 PMID:16806186

Zhang, Q.-G., Laird, M. D., Han, D., Nguyen, K., Scott, E., Dong, Y., ... Brann, D. W. (2012). Critical role of NADPH oxidase in neuronal oxidative damage and microglia activation following traumatic brain injury. *PLoS One*, *7*(4), e34504. doi:10.1371/journal.pone.0034504 PMID:22485176

Zhang, Z., Lotti, F., Dittmar, K., Younis, I., Wan, L., Kasim, M., & Dreyfuss, G. (2008). SMN deficiency causes tissue-specific perturbations in the repertoire of snRNAs and widespread defects in splicing. *Cell*, *133*(4), 585–600. doi:10.1016/j.cell.2008.03.031 PMID:18485868

Zorov, D. B., Juhaszova, M., & Sollott, S. J. (2014). Mitochondrial reactive oxygen species (ROS) and ROS-induced ROS release. *Physiological Reviews*, *94*(3), 909–950. doi:10.1152/physrev.00026.2013 PMID:24987008

Chapter 4 Mitochondrial Dysfunction in Aging and Neurodegeneration

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ABSTRACT

Mitochondria are a dynamic organelle of the cell involved in the various biological processes. Mitochondria are the site of the adenosine triphosphate (ATP) production, electron transport chain (ETC), oxidation of fatty acids, tricarboxylic acid (TCA), and cellular apoptosis. Besides these, mitochondria are the site of production of reactive oxygen species (ROS), which further disrupts the normal functioning of this organelle also making mitochondria itself as an important target of oxidative stress. Thus, mitochondria serve as an important target in the process of neurodegeneration. In the present chapter, the authors describe mitochondria and its functioning, dynamics, and the mitochondrial dysfunction in aging and neurodegenerative disorders (NDs).

INTRODUCTION

Mitochondria are the main organelle of the cell which plays key role in the various processes including oxidative phosphorylation, calcium signaling (Rizzuto et al, 2012), stress responses (Pellegrino & Haynes, 2015) and cellular apoptosis (Bratic & Larsson, 2013). The primary role of mitochondria is the production of ATP by the process of oxidative phosphorylation. During this process some of the electrons eventually leaks and this results in the generation of free radicals or reactive oxygen species (ROS) (Payne & Chinnery, 2015). The production of ROS is known to cause the oxidative damage to various components of mitochondrial electron transport chain (ETC) resulting in the mitochondrial dysfunction. Mitochondria dysfunctional is further characterized by decreased activity of ETC complexes, decreased rate of electron transport in complex-I and IV, reduced capacity of oxidative phosphorylation, decreased ATP production, increased water permeability in brain mitochondria, decreased ATP-synthase activity, increased accumulation of oxidative products, decreased mitochondrial membrane potential, increased

DOI: 10.4018/978-1-5225-5282-6.ch004

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mitochondrial size and fragility. Further all these factors are the determining factors of brain aging (Navarro & Boveris, 2010). Further inactivation of complex-1 has been recognized as characteristic of aging and neurodegeneration in brain. Mitochondrial dysfunction is more prominent in the areas of brain including hippocampus, frontal cortex and substantia nigra (Navarro & Boveris, 2010). In the present work author discuss different causes and features of mitochondrial dysfunction and demonstrate the role of mitochondrial dysfunction in the neurodegenerative pathologies.

BACKGROUND

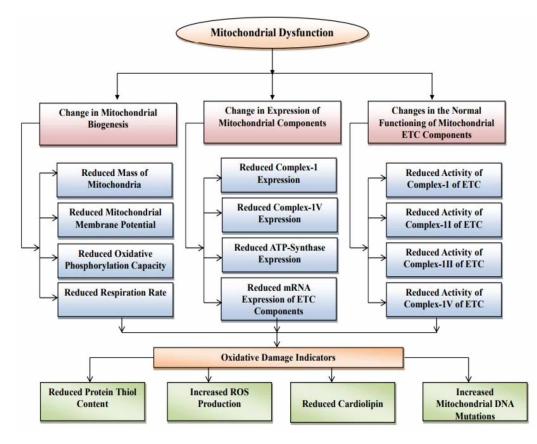
Mitochondria are the organelle linked with the production of ATP and dysfunction of this organelle thus led to ATP deficit. Besides the synthesis or production of ATP, mitochondria are the important site of various other physiological processes including sequestration of calcium within neurons (Trevelyan et al, 2010), misfolded/altered/damaged protein clearance pathways (Emmanouilidou et al, 2010; Jana, 2012; Li & Li, 2011; Rubinsztein et al, 2011), site of ROS production (Cui et al, 2012) and production of apoptogenic factors involved in the process of cellular apoptosis (Tsujimoto, 2000). The production of ROS increases with the increase in the age and aging is characterized by the increased generation of ROS which evokes oxidative stress and affects normal functioning of various cellular molecules (Gerschaman et al, 1954; Harman, 1956). Mitochondrial ROS production mediated damaged to cellular molecules led to the accumulation of oxidative and damaged products (Beckman & Ames, 1998; Harman, 2006; Vina et el, 2003). Further mitochondria derived from the brain of aged individual showed increased membrane permeability (Navarro & Boveris, 2004); reduced membrane potential and increased degradation (Beckman & Ames, 1998; Navarro & Boveris, 2004, 2007b) and decreased ATP-synthase activity (Lam et el, 2009). Dysfunctional mitochondria results in the prolonged elevation of intracellular calcium that results in the neuronal dysfunction observed in PD patients (Trevelyan et al, 2010). Reduced activities of mitochondrial protein clearance pathways including UPS system and autophagy (Emmanouilidou et al, 2010; Jana, 2012; Li & Li, 2011; Rubinsztein et al, 2011) has been observed in the brain of the PD patients SN region (Bedford et al, 2008; McNaught et al, 2003; McNaught & Jenner, 2001). Also the activities of these pathways declines with the increase in the age (Emmanouilidou et al, 2010; Jana, 2012; Li & Li, 2011; McNaught et al, 2003; McNaught & Jenner, 2001; Rubinsztein et al, 2011) suggesting aging increased the risk of PD onset. Also it has been reported that the risk of developing PD increases after the age of 65 years and the PD generally affects the individuals in the later stages of life (de Lau & Breteler, 2006; Fearnley & Lees, 1991; Ma et al, 1999b; Nussbaum & Ellis, 2003; Wood-Kaczmar et al, 2006). The different events that evolved in context of mitochondrial dysfunction have been shown in Figure 1.

MITOCHONDRIA AND AGING

Mitochondria consist of 2 membranes; outer membrane is porous while the inner membrane is impermeable. It is suggested that Bcl-2 class proteins (including proapoptotic proteins i.e. Bax, Bak, Bok, Bid, Bad, and Puma and antiapoptotic proteins i.e. Bcl-2, Bcl- x_L , and Mcl-1) (Adams & Cory, 1998) maintains the integrity of the outer membrane of the mitochondria (Chipuk et al, 2010). The space between the 2 membranes is called inter membranous pace in which cytochrome c (cyt-c) is present (Tait & Green,

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Figure 1. Mitochondrial dysfunction and the events involved. Mitochondrial dysfunction mainly arises due to impaired mitochondrial biogenesis, impaired expression of the mitochondrial components and the impaired functioning of the mitochondrial components. These events results in the oxidative damages which is further characterized by the increased ROS production, reduced protein thiol content, reduced cardiolipin and increased mitochondrial DNA mutation. These events further results in the neurodegeneration (Balaban et al, 2005; Hu & Wang, 2016).



2010). Bcl-2-family proteins possess ion channel activity and render membranes permeable to cyt-c (Korsmeyer et al, 2000, Kuwana et al, 2002). The genetic material present inside the mitochondria are double stranded circular DNA (Taylor & Turnbull, 2005) known as mitochondrial DNA (mit-DNA). mit-DNA is not protected by histone proteins therefore highly mutagenic in nature (Cheng et al, 1992; Kuchino et al, 1987; Yakes & Van Houten, 1997); lack DNA repair mechanisms (Stuart & Brown, 2006) and containing oxidized bases (Ames 1989; Richter et al, 1988) making it prone to oxidative damage (Mecocci et al, 1997). mit-DNA encodes for the several components of ETC, thus any damage to mit-DNA affects the functioning of ETC (Alexeyev et al, 2004) which further affects the production of ATP and ROS (Indo et al, 2007). mit-DNA mutations arise due to the consequence of unrepaired DNA damage (Larsson, 2010) and its frequency increases during aging (Corral-Debrinski et al, 1992; Fayet et al, 2002; Yen et al, 1991) resulting in increased ROS production, reduced oxidative phosphorylation, and increased cell death (Allen & Coombs, 1980; Niranjan et al, 1982; Rossi et al, 1988; Wunderlich et al, 1970). mit-DNA deletion also increases with the increase in age and their levels were found highest in

the including basal ganglia, substantia nigra (SN) and cortex of the brain (Corral-Debrinski et al, 1992; Soong et al, 1992). Aging is characterized by the increased generation of ROS (Gerschaman et al, 1954; Harman, 1956) and mitochondria are the site of ROS production, therefore the theory of aging centers around the mitochondria (Beckman & Ames, 1998; Harman, 2006; Vina et el, 2003). Further mitochondrial ROS production is considered as a main characteristic of aging (Beckman & Ames, 1998; Harman, 2006; Vina et el, 2003) and age related neuronal degeneration (Harman, 1972). Aging is also accompanied by the reduction in DNA repair mechanism (Imam et al, 2006) suggesting age related deficits in mit-DNA repair enzymes potentiate the process of neurodegeneration (Jeppesen et al, 2011; Zhang et al, 2010). Aging brain is characterized by the accumulation of defective or dysfunctional mitochondria (Boveris & Navarro, 2008; Navarro & Boveris, 2007a, b) suggesting age-related mitochondrial dysfunction contributes to neuronal degeneration (Balaban et al, 2005). Different features of mitochondrial dysfunction are shown in Table 1.

MITOCHONDRIA AS A SITE OF REACTIVE OXYGEN SPECIES PRODUCTION

Mitochondria are a major site of ROS production and the production of ROS increases with the increase in the age of the individual (Sohal et al, 1994; Sohal & Sohal, 1991). ROS are produced by the mitochondria due to the defects in the oxidative phosphorylation mechanism (Murphy, 2009). ROS also produced in the body during normal conditions (Wei et al, 2001) and their production requires the molecular oxygen (Smith et al, 2007). The leakage of the electron from the ETC combines with the molecular oxygen to form the superoxide radical (O_2^{\bullet}) (Finkel & Holbrook, 2000) which is highly toxic in nature and therefore converted by the enzyme superoxide dismutase into hydrogen peroxides (H_2O_2) which is further toxic in nature and in the presence of Fe²⁺, H_2O_2 is further converted into hydroxyl radical (OH[•]) (Valko et al,

S. No.	Parameters	Levels/Activity/Expression
1.	Oxidative parameters [thiobarbituric acid reactive substances (TBARS) or <i>malondialdehyde (MDA)</i>]	Increased levels of TBARS or MDA.
2.	Mitochondrial enzymes	Decreased activity of enzymes such as ATP synthase, DNA glycosylase
3.	Mitochondrial proteins	Decreased expression and levels of mitochondrial proteins (such as cardiolipin)
4.	Mitochondrial DNA	Increased number and rate of mutations and deletion of mitochondrial DNA
5.	Mitochondrial mRNA	Decreased mitochondrial mRNA expression
6.	Mitochondrial mass	Decreased mitochondrial mass
7.	Mitochondrial oxidative phosphorylation	Decreased mitochondrial oxidative phosphorylation
8.	Mitochondrial membrane potential	Decreased mitochondrial membrane potential
9.	Mitochondrial electron transport chain complexes	Decreased activity of complex-I, II, III and IV of mitochondrial electron transport chain
10.	Mitochondrial ATP synthesis	Decreased ATP synthesis

Table 1. Different features of the mitochondrial dysfunction in age related neurodegeneration.

2007). ROS initially functions to compensate the damage by the mitochondrial biogenesis (Chakrabarti et el, 2011). O¹ and H₂O₂ exerted the oxidative damage to cellular macromolecules (Balaban et al, 2005) and OH[•] damage macromolecules within mitochondria (Van Houten et al, 2006). Further the production of ROS increases with the increase in the age of the individual (Sohal et al, 1994; Sohal & Sohal, 1991) and the increased production of ROS is the main cause for age neuronal damage and degeneration (Harman, 1972). Exposure to the ROS inactivate complex-I, II, and III of ETC of mitochondria (Ghezzi & Zeviani, 2012). Further the reduced activity of complex I of ETC has been reported in patients of PD (Parker et al, 1989; Schapira et al, 1990; Swerdlow et al, 1996). ROS contribute to the development of vascular inflammation in aging (Ungvari et al, 2007) and accelerates senescence in endothelial cells in aging (Csiszar et al, 2004; Pearson et al, 2008). ROS promote the mitochondrial outer membrane permeabilization (MOMP) (Garcia-Perez et al, 2012) which further results in the release of cyt-c in to the cytoplasm further resulting in the activation of caspase responsible for cell death (Newmeyer & Ferguson-Miller, 2003). Further the high levels of the ROS activate p53-mediated apoptosis (Liu et al, 2008). Thus the raised levels and accumulation of ROS in the aged individuals (Rebrin & Sohal, 2008) results in the neuronal cell damage (Du et al, 2003).

MITOCHONDRIAL DYSFUNCTION IN NEURODEGENERATIVE DISORDERS

The neuronal degeneration arises due to the mitochondrial dysfunction is further linked with the genetic mutations (Table 2).

tion (Ebrahimi-Fakhari et al, 2012; Klein & Westenberger, 2012; Miklya et al, 2014; Sterky et al, 2011; West et al, 2007).					
Sr. No.	Gene	Mutation and its Effect			
		Mutations cause structural changes in mitochondria oxidative stress reduced			

Table 2. Genes mutation responsible for the mitochondrial dysfunction responsible for neurodegenera-
tion (Ebrahimi-Fakhari et al, 2012; Klein & Westenberger, 2012; Miklya et al, 2014; Sterky et al, 2011;
West et al, 2007).

Sr. No.	Gene	Mutation and its Effect
1	SNCA	Mutations cause structural changes in mitochondria, oxidative stress, reduced complex-I activity, reduced generation of ATP, increase mitochondrial fragmentation and degeneration, altered mitochondrial membrane potential and MOMP leading to the leakage of pro-apoptotic molecules responsible for neuronal cell death.
2	Parkin	Mutation accounts for the reduced activity of complex- I and IV, reduced mitochondrial integrity, reduced mitochondrial enzyme activity and reduced mitophagy.
3	PINK1	<i>M</i> utation led to the reduced enzyme activity, reduced complex-1 activity, reduced ATP synthesis and reduced mitophagy.
4	LRRK2	Mutations induced mitochondrial fragmentation, increased the level of ROS and reduced ATP production.
5	DJ-1	Mutation results in the reduced activity of complex-1 and II, reduced ATP production, reduced oxygen consumption, reduced mitochondrial membrane potential and disruption of the mitochondrial integrity.
6	APP	Mutations induces the production and deposition of plaques of amyloid β (A β) peptide inhibit mitochondrial respiration; increased production of ROS, reduced production of ATP, increased entry of Ca ²⁺ entry into neurons, opening of mtPTP, and trigger cell death in neurons.
7	PSEN1	Mutations increase the length of $A\beta$ peptides responsible for the aggregation, increase ROS production and neurotoxicity.

Mitochondrial Dysfunction in Parkinson's Disease

Mitochondrial dysfunction plays a key role in the PD pathogenesis. PD is further characterized by the higher levels of the mit-DNA deletions in the SN region of the brain suggesting the involvement of the mitochondrial dysfunctioning in the brain of the PD patients (Hu & Wang, 2016). α -synuclein is synaptic protein (Bendor et al, 2013), important for synaptic vesicle recycling (Cheng et al, 2011; Murphy et al. 2000) aggregate to form protofibrils in response to oxidative stress (Bendor et al. 2013; Brevdo et al, 2012; Fujiwara et al, 2002; Giasson et al, 2000). Mutations in α -synuclein gene or SNCA gene cause structural changes in mitochondria (Botella et al, 2008). α -synuclein aggregation mediated oxidative stress (Bendor et al, 2013; Breydo et al, 2012; Fujiwara et al, 2002; Giasson et al, 2000), structural changes in mitochondria (Botella et al, 2008) reduced complex-I activity of ETC and ROS formation (Devi et al, 2008; Parihar et al, 2008, 2009) reduce the generation of ATP (Rowland & Voeltz, 2012), increase mitochondrial fragmentation and degeneration (Martin et al, 2006; Plotegher et al, 2014), reduce mitochondrial Ca²⁺ retention (Calì et al, 2012) altered mitochondrial membrane potential (Luth et al, 2014) and MOMP leading to the leakage of pro-apoptotic molecules (Bandopadhyay & de Belleroche, 2010) responsible for the neuronal cell death. Previous reports suggested that *Parkin* plays a key role in the clearance of misfolded proteins. Parkin serves as an ubiquitin E3 ligase that degrades misfolded proteins (Yoshii et al. 2011; Zhang et al. 2000) and the *Parkin* mediated ubiquitination serve as signal for the degradation of ubiquitinated substance by UPS (Dawson & Dawson, 2010). Parkin maintain the proper functioning of mitochondria and mit-DNA (Kuroda et al, 2006), inhibits ROS formation (Kuroda et al, 2006; Temme et el, 2009), protects mit-DNA from damage (Rothfuss et al, 2009, Watson et al, 2004), regulates mitochondrial fission and fusion mechanism (Glauser et el, 2011; Narendra et el, 2008) and mutation in the gene encodes for *Parkin* accounts for the familial and sporadic cases of PD (Ebrahimi-Fakhari et al, 2012; Miklya et al, 2014; West et al, 2007) and its early onset (Klein & Westenberger, 2012). *PINK1* is responsible for the normal functioning of mitochondria and it is reported that the loss of mitochondrial membrane potential and increase accumulation of misfolded/altered protein led to the stabilization of *PINK1* on outer mitochondrial membrane, which is the sign of elimination of mitochondria (Pickrell & Youle, 2015). The ultimate target of *PINK1* is *Parkin* present in the cytoplasm. PINK1 phosphorylate mitofusin2 (Mfn2) for the binding of Parkin to the mitochondria by the process of mitophagy (Pickrell & Youle, 2015). PINK1 also phosphorylates Parkin at Ser65 position responsible for the activation of *Parkin* (Truban et al. 2016). *PINK1-Parkin* signaling plays a key role in the mitophagy (Narendra et al, 2010; Schapira, 2012) and the alteration in this signaling pathway is responsible for mitochondrial dysfunction (Sterky et al, 2011). Mitophagy is defined as the removal of the targeted or specific or damaged or excess of mitochondria by the process of autophagy (Lemasters, 2005). Mitophagy removes the mitochondria which becomes dysfunctional or produces excess ROS (Kim et al, 2007). High population of enlarged and swollen mitochondria has been reported in the sever models of PD (Martin et al, 2006; Poole et al, 2008; Yang et al, 2006) suggesting the role of the mitochondrial dysfunctioning in the pathogenesis of PD. LRRK2 belongs to the Roco family proteins (Bosgraaf & Van Haastert, 2003; Marín et al, 2008). In the neurons LRRK2 colocalizes with Dynamin like protein 1 (DLP1) and its expression induced mitochondrial fragmentation and increased the level of ROS (Niu et al, 2012; West et al, 2005). LRRK2 is involved in the various cellular functions including vesicular trafficking, proteolysis, regulation of neuritic outgrowth and neuritic morphology (Li et al, 2014). LRRK2 is known to regulate NF-KB activity (Gardet et al, 2010) which is further responsible for the induction of TNF mediated extrinsic pathway of the cell death and the inhibition of this pathway might reduce the neuronal death in the PD (Dauer & Przedborski, 2003; Hayley et al, 2004; McCoy et al, 2006). LRRK2 gene mutations results in the onset of autosomal dominant PD in the later ages most frequently (Klein & Westenberger, 2012). DJ-1 is a predominantly cytosolic, homodimeric protein, ubiquitously expressed in both brain and provides the neuroprotection by protecting cells against reactive oxygen species (ROS) (Björkblom et al, 2013). DJ-1 contains 3 cysteine residues of which only C106 is highly susceptible to oxidative damage and it is reported that the alterations in the normal functioning of this residue led to the loss of the DJ1 function completely. DJ-1 quenches ROS (Ariga et al, 2013) inhibit p53 and stimulates SOD expression (Ariga et al, 2013). Further DJ-1 mutations are responsible for the development of autosomal recessive form of PD but found in rare cases (Klein & Westenberger, 2012). The reports from the preclinical studies suggested that mitochondria are the potential target of the toxin used for the induction of neurodegeneration in the preclinical models (Greenamyre & Hastings, 2004). Toxin, for example MPTP induces PD in the preclinical animal models (Davis et al, 1979; Langston et al, 1983). MPTP rapidly crosses the BBB (Riachi et al, 1989) and is oxidized to 1-methyl-4-phenylpyridinium (MPP+) (Chiba et al, 1984) which is taken up by the dopaminergic neurons (Javitch et al, 1985), resulting in the inhibition of complex-I of ETC (Nicklas et al, 1987) leading to the production of ROS (Keeney et al, 2006; Perier et al 2005; Ramsay et al, 1987) and depletion of the ATP level (Chan et al, 1991; Davey & Clark 1996; Di Monte et al, 1986; Scotcher et al, 1990). The increased the production of ROS following the inhibition of the complex-1 is responsible for the death of the dopaminergic neurons (Jackson-Lewis et al, 1995; Tatton & Kish, 1997). ROS also damages lysosomal membranes leading impairment in the autophagic mechanism seen in the PD patients (Dehay et al, 2010). Another agent, 6-hydroxy dopamine (6-OHDA) a hydroxylated analogue of dopamine capable damaged the dopaminergic neurons in the SN region of brain (Ungerstedt, 1968). 6-OHDA following administration, uptake by the dopaminergic neurons, and inside the neurons it oxidizes to form ROS (Mazzio et al, 2004), reduced the antioxidants enzymes (Kunikowska & Jenner, 2001; Perumal et al, 1992), raised the levels of iron (Oestreicher et al, 1994), inhibits the activities of complexes I and IV of ETC (Glinka et al, 1997) and thus induced neurodegeneration. Paraquat is another agent uptake via the transporter in the brain (Shimizu et al, 2001) inside the neurons inhibits complex-I (Miller, 2007) and complex-III (Zhang et al, 2003) of ETC and thus induces neuronal degeneration by targeting mitochondria (Ascherio et al, 2006; Costello et al, 2009). Further rotenone, is a lipophilic toxin that crosses the BBB, enters into neurons inhibits the activity of the complex-I of ETC mitochondria, increased the production of ROS and reduced the levels of glutathione (Sherer et al, 2003a). Further rotenone in the brain activates the microglia cells (Sherer et al, 2003c) and inhibits proteasomal activity (Wang et al, 2006) and caused the oxidative damage in the striatum and cortex (Alam et al, 1997; Sherer et al, 2003a, b). Thus the blockade of mitochondrial ETC by inhibiting the oxidative phosphorylation and led to the degeneration of the dopaminergic neurons (Betarbet et al, 2000; Dauer & Przedborski 2003) while promoting the mitochondrial ETC protects the neurons against neurodegeneration (Tieu et al, 2003).

Mitochondrial Dysfunction in Alzheimer's Disease

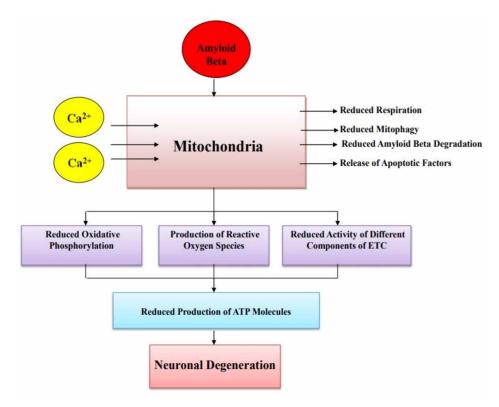
Alzheimer's disease (AD) is a NDs in which the patients suffers from the memory and behavioral deficits (Mattson, 2004; Reddy, 2008; Reddy & Beal, 2008). Several hypotheses have been formulated to explain the process of neurodegeneration in AD, out of which amyloid hypothesis is most acceptable (Hardy & Higgins, 1992). According to amyloid hypothesis accumulation or the reduced clearance of amyloid β (A β) peptide is responsible for neurodegeneration in AD (Hardy & Selkoe, 2002). Deposition of A β

peptide inhibit mitochondrial respiration (Casley et al, 2002); induces the increased entry of Ca^{2+} entry into neurons (Sanz-Blasco et al, 2008), opening of mtPTP (Reddy and Beal, 2008), trigger cell death in neurons (Giacomello et al, 2007). Previous reports suggested increased cyt-c oxidase, and reduced energy metabolism in the brain of AD patients prior to A β plaque formation (Hirai et al, 2001; Lin & Beal, 2006). Also the AD patients showed the down-regulation of complex-I and up-regulation of complex-III and IV genes (Manczak et al, 2004). A β results in the dysfunctioning of mitochondrial dysfunction and thus augmented ROS levels (Belkacemi & Ramassamy, 2012). Further the mitochondria-derived ROS increased the production of A β (Leuner et al, 2012). The cycle keeps on repeating and so is the neuronal degeneration in the AD patients suggesting mitochondrial dysfunction play a key role in the process of neurodegeneration in AD (Fukui & Moraes, 2008; Picone et al, 2014; Swerdlow et al, 2014) (shown in Figure 2). The dysfunctioning of mitochondria in AD is characterized by the reduced metabolism and utilization of glucose (Mosconi et al, 2011; Reiman et al, 1996), decreased activity of the cytochrome oxidase, pyruvate dehydrogenase and α -ketodehydrogenase (Reddy, 2008), increased levels and accumulation of the mit-DNA changes (Coskun et al, 2004; Lin et al, 2002), increased generation of ROS, oxidative damage and reduced ATP production (Butterfield et al, 2001; Devi et al, 2006; Maurer et al, 2000; Smith et al, 1996). Further the mitochondria isolated from the neurons of AD subjects showed significant reduction in mitochondrial length and increased width with a significant increased overall size consistent with unopposed fission suggesting alterations of mitochondrial dynamics (Wang et al. 2008). Further AD patients showed impaired mitochondrial biogenesis as compared to age matched control (Hirai et al, 2001; Qin et al, 2009; Sheng et al, 2012). The overproduction of A β overproduction further increased the number of fragmented mitochondria (Wang et al, 2008) mtDNA mutations (Hamblet et al, 2006) and decrease in oxidative phosphorylation and ATP synthesis (Chandrasekaran et al, 1996), suggesting Aβ affect the dynamics of mitochondria also (Calkins et al, 2011; Manczak et al, 2011; Wang et al, 2008). Parkin plays a key role in the clearance of the altered or misfolded proteins, and thus ubiquitinate A β for the proteosomal degradation and thus reduced the intracellular A β levels (Burns et al, 2009). Parkin reverses intracellular Aβ accumulation and its negative effects on proteasome function (Rosen et al, 2010). Parkin-induced autophagy facilitated clearance of vesicles containing debris and defective mitochondria counteracting oxidative stress and preventing mitochondrial dysfunction (Khandelwal et al, 2011). Further the reduced activity of *Parkin* has been observed in the cortex of AD brains (Rosen et al, 2010).

FUTURE RESEARCH DIRECTIONS

The future work should involve the determination of the exact mitochondrial population in the neurons of normal individuals and the aging individuals and the patients suffering from AD and PD. The research should focus on targeting the healthier mitochondria in the neurons. Since the neuronal cell death is preceded by the opening of the mitochondrial membrane transition pore, therefore inhibition of the opening of the pore might be beneficial. Molecules for e.g., olesoxime inhibits the opening of mitochondrial transition pore in response to oxidative stress (Bordet et al, 2007) and can protect the cells from cell death (Gouarne et al, 2015).

Figure 2. Mitochondrial dysfunction in the pathogenesis of Alzheimer's disease. Amyloid beta $(A\beta)$ peptide deposition on neurons results in mitochondrial dysfunction characterized by the reduced respiration, reduced mitophagy, reduced degradation of $A\beta$ and release of apoptotic factors responsible for the decreased production of ATP and neurodegeneration (Fukui & Moraes, 2008; Picone et al, 2014; Swerdlow et al, 2014).



CONCLUSION

Mitochondria are the dynamic organelle of the cell responsible for the various neuronal functions. Aging is an important factor that contributes to the mitochondrial dysfunction. Further aging increased the risk of various NDs. It is suggested that the aging induces the production of ROS which is responsible for the reduced functioning of the various components of the mitochondrial ETC suggesting the dysfunction of the ETC components as a cause of the neuronal degeneration. Therefore targeting mitochondria for the drug development in the treatment of the neurodegenerative disorder may provide a new direction for the treatment.

REFERENCES

Adams, J. M., & Cory, S. (1998). The Bcl-2 protein family: Arbiters of cell survival. *Science*, 281(5381), 1322–1326. doi:10.1126cience.281.5381.1322 PMID:9735050

Mitochondrial Dysfunction in Aging and Neurodegeneration

Alam, Z. I., Daniel, S. E., Lees, A. J., Marsden, D. C., Jenner, P., & Halliwell, B. (1997). A generalised increase in protein carbonyls in the brain in Parkinson's but not incidental Lewy body disease. *Journal of Neurochemistry*, *69*(3), 1326–1329. doi:10.1046/j.1471-4159.1997.69031326.x PMID:9282961

Alexeyev, M. F., LeDOUX, S. P., & Wilson, G. L. (2004). Mitochondrial DNA and aging. *Clinical Science*, *107*(4), 355–364. doi:10.1042/CS20040148 PMID:15279618

Allen, J. A., & Coombs, M. M. (1980). Covalent binding of polycyclic aromatic compounds to mitochondrial and nuclear DNA. *Nature*, 287(5779), 244–245. doi:10.1038/287244a0 PMID:7432460

Ames, B. N. (1989). Endogenous oxidative DNA damage, aging, and cancer. *Free Radical Research Communications*, 7(3-6), 121–128. doi:10.3109/10715768909087933 PMID:2684796

Ariga, H., Takahashi-Niki, K., Kato, I., Maita, H., Niki, T., & Iguchi-Ariga, S. M. M. (2013). Neuroprotective Function of DJ-1 in Parkinson's Disease. *Oxidative Medicine and Cellular Longevity*, 2013, 683920. doi:10.1155/2013/683920 PMID:23766857

Ascherio, A., Chen, H., Weisskopf, M. G., O'Reilly, E., McCullough, M. L., Calle, E. E., ... Thun, M. J. (2006). Pesticide exposure and risk for Parkinson's disease. *Annals of Neurology*, *60*(2), 197–203. doi:10.1002/ana.20904 PMID:16802290

Balaban, R. S., Nemoto, S., & Finkel, T. (2005). Mitochondria, oxidants, and aging. *Cell*, *120*(4), 483–495. doi:10.1016/j.cell.2005.02.001 PMID:15734681

Bandopadhyay, R., & de Belleroche, J. (2010). Pathogenesis of Parkinson's disease: Emerging role of molecular chaperones. *Trends in Molecular Medicine*, *16*(1), 27–36. doi:10.1016/j.molmed.2009.11.004 PMID:20036196

Beckman, K. B., & Ames, B. N. (1998). The free radical theory of aging matures. *Physiological Reviews*, 78(2), 547–581. doi:10.1152/physrev.1998.78.2.547 PMID:9562038

Bedford, L., Hay, D., Devoy, A., Paine, S., Powe, D. G., Seth, R., ... Mee, M. (2008). Depletion of 26S proteasomes in mouse brain neurons causes neurodegeneration and Lewy-like inclusions resembling human pale bodies. *The Journal of Neuroscience*, 28(33), 8189–8198. doi:10.1523/JNEUROSCI.2218-08.2008 PMID:18701681

Belkacemi, A., & Ramassamy, C. (2012). Time sequence of oxidative stress in the brain from transgenic mouse models of Alzheimer's disease related to the amyloid-β cascade. *Free Radical Biology & Medicine*, *52*(3), 593–600. doi:10.1016/j.freeradbiomed.2011.11.020 PMID:22172527

Bendor, J. T., Logan, T. P., & Edwards, R. H. (2013). The function of alpha-synuclein. *Neuron*, 79(6), 1044–1066. doi:10.1016/j.neuron.2013.09.004 PMID:24050397

Betarbet, R., Sherer, T. B., MacKenzie, G., Garcia-Osuna, M., Panov, A. V., & Greenamyre, J. T. (2000). Chronic systemic pesticide exposure reproduces features of Parkinson's disease. *Nature Neuroscience*, *3*(12), 1301–1306. doi:10.1038/81834 PMID:11100151

Björkblom, B., Adilbayeva, A., Maple-Grødem, J., Piston, D., Ökvist, M., Xu, X. M., ... Møller, S. G. (2013). Parkinson Disease Protein DJ-1 Binds Metals and Protects against Metal-induced Cytotoxicity. *The Journal of Biological Chemistry*, 288(31), 22809–22820. doi:10.1074/jbc.M113.482091 PMID:23792957

Bordet, T., Buisson, B., Michaud, M., Drouot, C., Galea, P., Delaage, P., ... Lacapere, J. J. (2007). Identification and characterization of cholest-4-en-3-one, oxime (TRO19622), a novel drug candidate for amyotrophic lateral sclerosis. *The Journal of Pharmacology and Experimental Therapeutics*, *322*(2), 709–720. doi:10.1124/jpet.107.123000 PMID:17496168

Bosgraaf, L., & Van Haastert, P. J. (2003). Roc, a Ras/GTPase domain in complex proteins. *Biochimica et Biophysica Acta (BBA)-. Molecular Cell Research*, *1643*(1), 5–10.

Botella, J. A., Bayersdorfer, F., & Schneuwly, S. (2008). Superoxide dismutase overexpression protects dopaminergic neurons in a Drosophila model of Parkinson's disease. *Neurobiology of Disease*, *30*(1), 65–73. doi:10.1016/j.nbd.2007.11.013 PMID:18243716

Boveris, A., & Navarro, A. (2008). Brain mitochondrial dysfunction in aging. *IUBMB Life*, 60(5), 308–314. doi:10.1002/iub.46 PMID:18421773

Bratic, A., & Larsson, N. G. (2013). The role of mitochondria in aging. *The Journal of Clinical Investigation*, *123*(3), 951–957. doi:10.1172/JCI64125 PMID:23454757

Breydo, L., Wu, J. W., & Uversky, V. N. (2012). α-Synuclein misfolding and Parkinson's disease. *Biochimica et Biophysica Acta (BBA)-. Molecular Basis of Disease*, 1822(2), 261–285. doi:10.1016/j. bbadis.2011.10.002

Burns, M. P., Zhang, L., Rebeck, G. W., Querfurth, H. W., & Moussa, C. E. H. (2009). Parkin promotes intracellular Aβ 1–42 clearance. *Human Molecular Genetics*, *18*(17), 3206–3216. doi:10.1093/hmg/ddp258 PMID:19483198

Butterfield, D. A., Drake, J., Pocernich, C., & Castegna, A. (2001). Evidence of oxidative damage in Alzheimer's disease brain: Central role for amyloid β -peptide. *Trends in Molecular Medicine*, 7(12), 548–554. doi:10.1016/S1471-4914(01)02173-6 PMID:11733217

Calì, T., Ottolini, D., Negro, A., & Brini, M. (2012). α-Synuclein controls mitochondrial calcium homeostasis by enhancing endoplasmic reticulum-mitochondria interactions. *The Journal of Biological Chemistry*, 287(22), 17914–17929. doi:10.1074/jbc.M111.302794 PMID:22453917

Calkins, M. J., Manczak, M., Mao, P., Shirendeb, U., & Reddy, P. H. (2011). Impaired mitochondrial biogenesis, defective axonal transport of mitochondria, abnormal mitochondrial dynamics and synaptic degeneration in a mouse model of Alzheimer's disease. *Human Molecular Genetics*, 20(23), 4515–4529. doi:10.1093/hmg/ddr381 PMID:21873260

Casley, C. S., Canevari, L., Land, J. M., Clark, J. B., & Sharpe, M. A. (2002). β-Amyloid inhibits integrated mitochondrial respiration and key enzyme activities. *Journal of Neurochemistry*, 80(1), 91–100. doi:10.1046/j.0022-3042.2001.00681.x PMID:11796747

Chakrabarti, S., Munshi, S., Banerjee, K., Thakurta, I. G., Sinha, M., & Bagh, M. B. (2011). Mitochondrial Dysfunction during Brain Aging: Role of Oxidative Stress and Modulation by Antioxidant Supplementation. *Aging and Disease*, 2(3), 242–256. PMID:22396876

Mitochondrial Dysfunction in Aging and Neurodegeneration

Chan, P., DeLanney, L. E., Irwin, I., Langston, J. W., & Monte, D. (1991). Rapid ATP Loss Caused by 1-Methyl-4-Phenyl-1, 2, 3, 6-Tetrahydropyridine in Mouse Brain. *Journal of Neurochemistry*, *57*(1), 348–351. doi:10.1111/j.1471-4159.1991.tb02134.x PMID:2051170

Chandrasekaran, K., Hatanpää, K., Brady, D. R., & Rapoport, S. I. (1996). Evidence for physiological down-regulation of brain oxidative phosphorylation in Alzheimer's disease. *Experimental Neurology*, *142*(1), 80–88. doi:10.1006/exnr.1996.0180 PMID:8912900

Cheng, F., Vivacqua, G., & Yu, S. (2011). The role of alpha-synuclein in neurotransmission and synaptic plasticity. *Journal of Chemical Neuroanatomy*, 42(4), 242–248. doi:10.1016/j.jchemneu.2010.12.001 PMID:21167933

Cheng, K. C., Cahill, D. S., Kasai, H., Nishimura, S., & Loeb, L. A. (1992). 8-Hydroxyguanine, an abundant form of oxidative DNA damage, causes G----T and A----C substitutions. *The Journal of Biological Chemistry*, 267(1), 166–172. PMID:1730583

Chiba, K., Trevor, A., & Castagnoli, N. Jr. (1984). Metabolism of the neurotoxic tertiary amine, MPTP, by brain monoamine oxidase. *Biochemical and Biophysical Research Communications*, *120*(2), 574–578. doi:10.1016/0006-291X(84)91293-2 PMID:6428396

Chipuk, J. E., Moldoveanu, T., Llambi, F., Parsons, M. J., & Green, D. R. (2010). The BCL-2 family reunion. *Molecular Cell*, *37*(3), 299–310. doi:10.1016/j.molcel.2010.01.025 PMID:20159550

Corral-Debrinski, M., Horton, T., Lott, M. T., Shoffner, J. M., Beal, M. F., & Wallace, D. C. (1992). Mitochondrial DNA deletions in human brain: Regional variability and increase with advanced age. *Nature Genetics*, 2(4), 324–329. doi:10.1038/ng1292-324 PMID:1303288

Coskun, P. E., Beal, M. F., & Wallace, D. C. (2004). Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proceedings of the National Academy of Sciences of the United States of America*, *101*(29), 10726–10731. doi:10.1073/ pnas.0403649101 PMID:15247418

Costello, S., Cockburn, M., Bronstein, J., Zhang, X., & Ritz, B. (2009). Parkinson's disease and residential exposure to maneb and paraquat from agricultural applications in the central valley of California. *American Journal of Epidemiology*, *169*(8), 919–926. doi:10.1093/aje/kwp006 PMID:19270050

Csiszar, A., Ungvari, Z., Koller, A., Edwards, J. G., & Kaley, G. (2004). Proinflammatory phenotype of coronary arteries promotes endothelial apoptosis in aging. *Physiological Genomics*, *17*(1), 21–30. doi:10.1152/physiolgenomics.00136.2003 PMID:15020720

Cui, H., Kong, Y., & Zhang, H. (2012). Oxidative Stress, Mitochondrial Dysfunction, and Aging. *Journal of Signal Transduction*, 2012, 646354. doi:10.1155/2012/646354 PMID:21977319

Dauer, W., & Przedborski, S. (2003). Parkinson's Disease, Mechanisms and Models. *Neuron*, 39(6), 889–909. doi:10.1016/S0896-6273(03)00568-3 PMID:12971891

Davey, G. P., & Clark, J. B. (1996). Threshold effects and control of oxidative phosphorylation in nonsynaptic rat brain mitochondria. *Journal of Neurochemistry*, *66*(4), 1617–1624. doi:10.1046/j.1471-4159.1996.66041617.x PMID:8627318 Davis, G. C., Williams, A. C., Markey, S. P., Ebert, M. H., Caine, E. D., Reichert, C. M., & Kopin, I. J. (1979). Chronic parkinsonism secondary to intravenous injection of meperidine analogues. *Psychiatry Research*, *1*(3), 249–254. doi:10.1016/0165-1781(79)90006-4 PMID:298352

Dawson, T. M., & Dawson, V. L. (2010). The Role of Parkin in Familial and Sporadic Parkinson's Disease. *Movement Disorders : Official Journal of the Movement Disorder Society*, 25(1), S32-S39.

De Lau, L. M., & Breteler, M. M. (2006). Epidemiology of Parkinson's disease. *Lancet Neurology*, 5(6), 525–535. doi:10.1016/S1474-4422(06)70471-9 PMID:16713924

Dehay, B., Bové, J., Rodríguez-Muela, N., Perier, C., Recasens, A., Boya, P., & Vila, M. (2010). Pathogenic lysosomal depletion in Parkinson's disease. *The Journal of Neuroscience*, *30*(37), 12535–12544. doi:10.1523/JNEUROSCI.1920-10.2010 PMID:20844148

Deng, H., Dodson, M. W., Huang, H., & Guo, M. (2008). The Parkinson's disease genes pink1 and parkin promote mitochondrial fission and/or inhibit fusion in Drosophila. *Proceedings of the National Academy of Sciences of the United States of America*, *105*(38), 14503–14508. doi:10.1073/pnas.0803998105 PMID:18799731

Devi, L., Prabhu, B. M., Galati, D. F., Avadhani, N. G., & Anandatheerthavarada, H. K. (2006). Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheimer's disease brain is associated with mitochondrial dysfunction. *The Journal of Neuroscience*, *26*(35), 9057–9068. doi:10.1523/JNEUROSCI.1469-06.2006 PMID:16943564

Devi, L., Raghavendran, V., Prabhu, B. M., Avadhani, N. G., & Anandatheerthavarada, H. K. (2008). Mitochondrial import and accumulation of α-synuclein impair complex I in human dopaminergic neuronal cultures and Parkinson disease brain. *The Journal of Biological Chemistry*, 283(14), 9089–9100. doi:10.1074/jbc.M710012200 PMID:18245082

Di Monte, D., Jewell, S. A., Ekström, G., Sandy, M. S., & Smith, M. T. (1986). 1-Methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MPTP) and 1-methyl-4-phenylpyridine (MPP+) cause rapid ATP depletion in isolated hepatocytes. *Biochemical and Biophysical Research Communications*, *137*(1), 310–315. doi:10.1016/0006-291X(86)91211-8 PMID:3487319

Du, L., Zhang, X., Han, Y. Y., Burke, N. A., Kochanek, P. M., Watkins, S. C., ... Clark, R. S. (2003). Intra-mitochondrial poly (ADP-ribosylation) contributes to NAD+ depletion and cell death induced by oxidative stress. *The Journal of Biological Chemistry*, 278(20), 18426–18433. doi:10.1074/jbc. M301295200 PMID:12626504

Ebrahimi-Fakhari, D., Wahlster, L., & McLean, P. J. (2012). Protein degradation pathways in Parkinson's disease: Curse or blessing. *Acta Neuropathologica*, *124*(2), 153–172. doi:10.100700401-012-1004-6 PMID:22744791

Emmanouilidou, E., Stefanis, L., & Vekrellis, K. (2010). Cell-produced α -synuclein oligomers are targeted to, and impair, the 26S proteasome. *Neurobiology of Aging*, *31*(6), 953–968. doi:10.1016/j. neurobiolaging.2008.07.008 PMID:18715677

Mitochondrial Dysfunction in Aging and Neurodegeneration

Fayet, G., Jansson, M., Sternberg, D., Moslemi, A. R., Blondy, P., Lombès, A., ... Oldfors, A. (2002). Ageing muscle: Clonal expansions of mitochondrial DNA point mutations and deletions cause focal impairment of mitochondrial function. *Neuromuscular Disorders*, *12*(5), 484–493. doi:10.1016/S0960-8966(01)00332-7 PMID:12031622

Fearnley, J. M., & Lees, A. J. (1991). Ageing and Parkinson's disease: Substantia nigra regional selectivity. *Brain*, *114*(5), 2283–2301. doi:10.1093/brain/114.5.2283 PMID:1933245

Finkel, T., & Holbrook, N. J. (2000). Oxidants, oxidative stress and the biology of ageing. *Nature*, 408(6809), 239–247. doi:10.1038/35041687 PMID:11089981

Fujiwara, H., Hasegawa, M., Dohmae, N., Kawashima, A., Masliah, E., Goldberg, M. S., ... Iwatsubo, T. (2002). [alpha]-Synuclein is phosphorylated in synucleinopathy lesions. *Nature Cell Biology*, *4*(2), 160–164. doi:10.1038/ncb748 PMID:11813001

Fukui, H., & Moraes, C. T. (2008). The mitochondrial impairment, oxidative stress and neurodegeneration connection: Reality or just an attractive hypothesis? *Trends in Neurosciences*, *31*(5), 251–256. doi:10.1016/j.tins.2008.02.008 PMID:18403030

Garcia-Perez, C., Roy, S. S., Naghdi, S., Lin, X., Davies, E., & Hajnóczky, G. (2012). Bid-induced mitochondrial membrane permeabilization waves propagated by local reactive oxygen species (ROS) signaling. *Proceedings of the National Academy of Sciences of the United States of America*, *109*(12), 4497–4502. doi:10.1073/pnas.1118244109 PMID:22393005

Gardet, A., Benita, Y., Li, C., Sands, B. E., Ballester, I., Stevens, C., ... Podolsky, D. K. (2010). LRRK2 is involved in the IFN-γ response and host response to pathogens. *Journal of Immunology (Baltimore, Md.: 1950)*, *185*(9), 5577–5585. doi:10.4049/jimmunol.1000548 PMID:20921534

Gerschman, R., Gilbert, D. L., Nye, S. W., Dwyer, P., & Fenn, W. O. (1954). Oxygen poisoning and xirradiation: A mechanism in common. *Science*, *119*(3097), 623–626. doi:10.1126cience.119.3097.623 PMID:13156638

Ghezzi, D., & Zeviani, M. (2012). Assembly factors of human mitochondrial respiratory chain complexes: physiology and pathophysiology. In *Mitochondrial Oxidative Phosphorylation* (pp. 65–106). Springer New York. doi:10.1007/978-1-4614-3573-0_4

Giacomello, M., Drago, I., Pizzo, P., & Pozzan, T. (2007). Mitochondrial Ca2+ as a key regulator of cell life and death. *Cell Death and Differentiation*, *14*(7), 1267–1274. doi:10.1038j.cdd.4402147 PMID:17431419

Giasson, B. I., Duda, J. E., Murray, I. V., Chen, Q., Souza, J. M., Hurtig, H. I., ... Lee, V. M. Y. (2000). Oxidative damage linked to neurodegeneration by selective α -synuclein nitration in synucleinopathy lesions. *Science*, 290(5493), 985–989. doi:10.1126cience.290.5493.985 PMID:11062131

Glauser, L., Sonnay, S., Stafa, K., & Moore, D. J. (2011). Parkin promotes the ubiquitination and degradation of the mitochondrial fusion factor mitofusin 1. *Journal of Neurochemistry*, *118*(4), 636–645. doi:10.1111/j.1471-4159.2011.07318.x PMID:21615408

Glinka, Y., Gassen, M., & Youdim, M. B. H. (1997). Mechanism of 6-hydroxydopamine neurotoxicity. In *Advances in Research on Neurodegeneration* (pp. 55–66). Vienna: Springer. doi:10.1007/978-3-7091-6842-4_7

Gouarné, C., Tracz, J., Paoli, M. G., Deluca, V., Seimandi, M., Tardif, G., ... Pruss, R. M. (2015). Protective role of olesoxime against wild-type α-synuclein-induced toxicity in human neuronally differentiated SHSY-5Y cells. *British Journal of Pharmacology*, *172*(1), 235–245. doi:10.1111/bph.12939 PMID:25220617

Greenamyre, J. T., & Hastings, T. G. (2004). Parkinson's--divergent causes, convergent mechanisms. *Science*, *304*(5674), 1120–1122. doi:10.1126cience.1098966 PMID:15155938

Hamblet, N. S., Ragland, B., Ali, M., Conyers, B., & Castora, F. J. (2006). Mutations in mitochondrial-encoded cytochrome c oxidase subunits I, II, and III genes detected in Alzheimer's disease using single-strand conformation polymorphism. *Electrophoresis*, 27(2), 398–408. doi:10.1002/elps.200500420 PMID:16358358

Hardy, J., & Selkoe, D. J. (2002). The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science*, 297(5580), 353-356.

Hardy, J. A., & Higgins, G. A. (1992). Alzheimer's disease: The amyloid cascade hypothesis. *Science*, 256(5054), 184–185. doi:10.1126cience.1566067 PMID:1566067

Harman, D. (1972). The biologic clock: The mitochondria? *Journal of the American Geriatrics Society*, 20(4), 145–147. doi:10.1111/j.1532-5415.1972.tb00787.x PMID:5016631

Harman, D. (2006). Free radical theory of aging: An update. *Annals of the New York Academy of Sciences*, *1067*(1), 10–21. doi:10.1196/annals.1354.003 PMID:16803965

Harraan, D. (1956). Aging: A theory based on free radical and radiation chemistry. *Journal of Gerontology*, *11*(3), 298–300. doi:10.1093/geronj/11.3.298 PMID:13332224

Hayley, S., Crocker, S. J., Smith, P. D., Shree, T., Jackson-Lewis, V., Przedborski, S., ... Park, D. S. (2004). Regulation of dopaminergic loss by Fas in a 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine model of Parkinson's disease. *The Journal of Neuroscience*, *24*(8), 2045–2053. doi:10.1523/JNEURO-SCI.4564-03.2004 PMID:14985447

Hirai, K., Aliev, G., Nunomura, A., Fujioka, H., Russell, R. L., Atwood, C. S., ... Shimohama, S. (2001). Mitochondrial abnormalities in Alzheimer's disease. *The Journal of Neuroscience*, *21*(9), 3017–3023. PMID:11312286

Hu, Q., & Wang, G. (2016). Mitochondrial dysfunction in Parkinson's disease. *Translational Neurode-generation*, *5*(1), 14. doi:10.118640035-016-0060-6 PMID:27453777

Imam, S. Z., Karahalil, B., Hogue, B. A., Souza-Pinto, N. C., & Bohr, V. A. (2006). Mitochondrial and nuclear DNA-repair capacity of various brain regions in mouse is altered in an age-dependent manner. *Neurobiology of Aging*, *27*(8), 1129–1136. doi:10.1016/j.neurobiolaging.2005.06.002 PMID:16005114

Mitochondrial Dysfunction in Aging and Neurodegeneration

Indo, H. P., Davidson, M., Yen, H. C., Suenaga, S., Tomita, K., Nishii, T., ... Majima, H. J. (2007). Evidence of ROS generation by mitochondria in cells with impaired electron transport chain and mitochondrial DNA damage. *Mitochondrion*, 7(1), 106–118. doi:10.1016/j.mito.2006.11.026 PMID:17307400

Jackson-Lewis, V., Jakowec, M., Burke, R. E., & Przedborski, S. (1995). Time course and morphology of dopaminergic neuronal death caused by the neurotoxin 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine. *Neurodegeneration*, 4(3), 257–269. doi:10.1016/1055-8330(95)90015-2 PMID:8581558

Jana, N. R. (2012). Protein homeostasis and aging: Role of ubiquitin protein ligases. *Neurochemistry International*, *60*(5), 443–447. doi:10.1016/j.neuint.2012.02.009 PMID:22353631

Javitch, J. A., D'Amato, R. J., Strittmatter, S. M., & Snyder, S. H. (1985). Parkinsonism-inducing neurotoxin, N-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine: Uptake of the metabolite N-methyl-4-phenylpyridine by dopamine neurons explains selective toxicity. *Proceedings of the National Academy of Sciences of the United States of America*, 82(7), 2173–2177. doi:10.1073/pnas.82.7.2173 PMID:3872460

Jeppesen, D. K., Bohr, V. A., & Stevnsner, T. (2011). DNA repair deficiency in neurodegeneration. *Progress in Neurobiology*, *94*(2), 166–200. doi:10.1016/j.pneurobio.2011.04.013 PMID:21550379

Keeney, P. M., Xie, J., Capaldi, R. A., & Bennett, J. P. (2006). Parkinson's disease brain mitochondrial complex I has oxidatively damaged subunits and is functionally impaired and misassembled. *The Journal of Neuroscience*, *26*(19), 5256–5264. doi:10.1523/JNEUROSCI.0984-06.2006 PMID:16687518

Khandelwal, P. J., Herman, A. M., Hoe, H. S., Rebeck, G. W., & Moussa, C. E. H. (2011). Parkin mediates beclin-dependent autophagic clearance of defective mitochondria and ubiquitinated A β in AD models. *Human Molecular Genetics*, 20(11), 2091–2102. doi:10.1093/hmg/ddr091 PMID:21378096

Kim, I., Rodriguez-Enriquez, S., & Lemasters, J. J. (2007). Selective degradation of mitochondria by mitophagy. *Archives of Biochemistry and Biophysics*, *462*(2), 245–253. doi:10.1016/j.abb.2007.03.034 PMID:17475204

Klein, C., & Westenberger, A. (2012). Genetics of Parkinson's Disease. *Cold Spring Harbor Perspectives in Medicine*, 2(1), a008888. doi:10.1101/cshperspect.a008888 PMID:22315721

Korsmeyer, S. J., Wei, M. C., Saito, M. T., Weiler, S., Oh, K. J., & Schlesinger, P. H. (2000). Pro-apoptotic cascade activates BID, which oligomerizes BAK or BAX into pores that result in the release of cyto-chrome c. *Cell Death and Differentiation*, 7(12), 1166–1173. doi:10.1038j.cdd.4400783 PMID:11175253

Kuchino, Y., Mori, F., Kasai, H., Inoue, H., Iwai, S., Miura, K., ... Nishimura, S. (1987). Misreading of DNA templates containing 8-hydroxydeoxyguanosine at the modified base and at adjacent residues. *Nature*, *327*(6117), 77–79. doi:10.1038/327077a0 PMID:3574469

Kunikowska, G., & Jenner, P. (2001). 6-Hydroxydopamine-lesioning of the nigrostriatal pathway in rats alters basal ganglia mRNA for copper, zinc-and manganese-superoxide dismutase, but not glutathione peroxidase. *Brain Research*, *922*(1), 51–64. doi:10.1016/S0006-8993(01)03149-3 PMID:11730701

Kuroda, Y., Mitsui, T., Kunishige, M., Shono, M., Akaike, M., Azuma, H., & Matsumoto, T. (2006). Parkin enhances mitochondrial biogenesis in proliferating cells. *Human Molecular Genetics*, *15*(6), 883–895. doi:10.1093/hmg/ddl006 PMID:16449237

Kuwana, T., Mackey, M. R., Perkins, G., Ellisman, M. H., Latterich, M., Schneiter, R., ... Newmeyer, D. D. (2002). Bid, Bax, and lipids cooperate to form supramolecular openings in the outer mitochondrial membrane. *Cell*, *111*(3), 331–342. doi:10.1016/S0092-8674(02)01036-X PMID:12419244

Lam, P. Y., Yin, F., Hamilton, R. T., Boveris, A., & Cadenas, E. (2009). Elevated neuronal nitric oxide synthase expression during ageing and mitochondrial energy production. *Free Radical Research*, *43*(5), 431–439. doi:10.1080/10715760902849813 PMID:19347761

Langston, J. W., Ballard, P., Tetrud, J. W., & Irwin, I. (1983). Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. *Science*, *219*(4587), 979–980. doi:10.1126cience.6823561 PMID:6823561

Larsson, N. G. (2010). Somatic mitochondrial DNA mutations in mammalian aging. *Annual Review of Biochemistry*, 79(1), 683–706. doi:10.1146/annurev-biochem-060408-093701 PMID:20350166

Lemasters, J. J. (2005). Selective mitochondrial autophagy, or mitophagy, as a targeted defense against oxidative stress, mitochondrial dysfunction, and aging. *Rejuvenation Research*, 8(1), 3–5. doi:10.1089/ rej.2005.8.3 PMID:15798367

Leuner, K., Schütt, T., Kurz, C., Eckert, S. H., Schiller, C., Occhipinti, A., ... Palmiter, R. D. (2012). Mitochondrion-derived reactive oxygen species lead to enhanced amyloid beta formation. *Antioxidants & Redox Signalling*, *16*(12), 1421–1433. doi:10.1089/ars.2011.4173 PMID:22229260

Li, J.-Q., Tan, L., & Yu, J.-T. (2014). The role of the LRRK2 gene in Parkinsonism. *Molecular Neuro*degeneration, 9(1), 47. doi:10.1186/1750-1326-9-47 PMID:25391693

Li, X. J., & Li, S. (2011). Proteasomal dysfunction in aging and Huntington disease. *Neurobiology of Disease*, 43(1), 4–8. doi:10.1016/j.nbd.2010.11.018 PMID:21145396

Lin, M. T., & Beal, M. F. (2006). Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature*, 443(7113), 787–795. doi:10.1038/nature05292 PMID:17051205

Lin, M. T., Simon, D. K., Ahn, C. H., Kim, L. M., & Beal, M. F. (2002). High aggregate burden of somatic mtDNA point mutations in aging and Alzheimer's disease brain. *Human Molecular Genetics*, *11*(2), 133–145. doi:10.1093/hmg/11.2.133 PMID:11809722

Liu, B., Chen, Y., & Clair, D. K. S. (2008). ROS and p53: A versatile partnership. *Free Radical Biology* & *Medicine*, 44(8), 1529–1535. doi:10.1016/j.freeradbiomed.2008.01.011 PMID:18275858

Luth, E. S., Stavrovskaya, I. G., Bartels, T., Kristal, B. S., & Selkoe, D. J. (2014). Soluble, prefibrillar α -synuclein oligomers promote complex I-dependent, Ca2+-induced mitochondrial dysfunction. *The Journal of Biological Chemistry*, 289(31), 21490–21507. doi:10.1074/jbc.M113.545749 PMID:24942732

Ma, S. Y., Röytt, M., Collan, Y., & Rinne, J. O. (1999b). Unbiased morphometrical measurements show loss of pigmented nigral neurones with ageing. *Neuropathology and Applied Neurobiology*, 25(5), 394–399. doi:10.1046/j.1365-2990.1999.00202.x PMID:10564529

Manczak, M., Calkins, M. J., & Reddy, P. H. (2011). Impaired mitochondrial dynamics and abnormal interaction of amyloid beta with mitochondrial protein Drp1 in neurons from patients with Alzheimer's disease: Implications for neuronal damage. *Human Molecular Genetics*, 20(13), 2495–2509. doi:10.1093/hmg/ddr139 PMID:21459773

Manczak, M., Park, B. S., Jung, Y., & Reddy, P. H. (2004). Differential expression of oxidative phosphorylation genes in patients with Alzheimer's disease. *Neuromolecular Medicine*, *5*(2), 147–162. doi:10.1385/NMM:5:2:147 PMID:15075441

Marín, I., van Egmond, W. N., & van Haastert, P. J. (2008). The Roco protein family: A functional perspective. *The FASEB Journal*, 22(9), 3103–3110. doi:10.1096/fj.08-111310 PMID:18523161

Martin, L. J., Pan, Y., Price, A. C., Sterling, W., Copeland, N. G., Jenkins, N. A., ... Lee, M. K. (2006). Parkinson's disease α -synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death. *The Journal of Neuroscience*, 26(1), 41–50. doi:10.1523/JNEUROSCI.4308-05.2006 PMID:16399671

Mattson, M. P. (2004). Pathways towards and away from Alzheimer's disease. *Nature*, 430(7000), 631–639. doi:10.1038/nature02621 PMID:15295589

Maurer, I., Zierz, S., & Möller, H. J. (2000). A selective defect of cytochrome c oxidase is present in brain of Alzheimer disease patients. *Neurobiology of Aging*, 21(3), 455–462. doi:10.1016/S0197-4580(00)00112-3 PMID:10858595

Mazzio, E. A., Reams, R. R., & Soliman, K. F. (2004). The role of oxidative stress, impaired glycolysis and mitochondrial respiratory redox failure in the cytotoxic effects of 6-hydroxydopamine in vitro. *Brain Research*, *1004*(1), 29–44. doi:10.1016/j.brainres.2003.12.034 PMID:15033417

McCoy, M. K., Martinez, T. N., Ruhn, K. A., Szymkowski, D. E., Smith, C. G., Botterman, B. R., ... Tansey, M. G. (2006). Blocking soluble tumor necrosis factor signaling with dominant-negative tumor necrosis factor inhibitor attenuates loss of dopaminergic neurons in models of Parkinson's disease. *The Journal of Neuroscience*, *26*(37), 9365–9375. doi:10.1523/JNEUROSCI.1504-06.2006 PMID:16971520

McNaught, K. S. P., Belizaire, R., Isacson, O., Jenner, P., & Olanow, C. W. (2003). Altered proteasomal function in sporadic Parkinson's disease. *Experimental Neurology*, *179*(1), 38–46. doi:10.1006/ exnr.2002.8050 PMID:12504866

McNaught, K. S. P., & Jenner, P. (2001). Proteasomal function is impaired in substantia nigra in Parkinson's disease. *Neuroscience Letters*, 297(3), 191–194. doi:10.1016/S0304-3940(00)01701-8 PMID:11137760

Mecocci, P., Beal, M. F., Cecchetti, R., Polidori, M. C., Cherubini, A., Chionne, F., ... Senin, U. (1997). Mitochondrial membrane fluidity and oxidative damage to mitochondrial DNA in aged and AD human brain. *Molecular and Chemical Neuropathology*, *31*(1), 53–64. doi:10.1007/BF02815160 PMID:9271005

Miklya, I., Göltl, P., Hafenscher, F., & Pencz, N. (2014). The role of parkin in Parkinson's disease. *Neuropsychopharmacologia Hungarica: a Magyar Pszichofarmakologiai Egyesulet lapja= official journal of the Hungarian Association of Psychopharmacology, 16*(2), 67-76.

Miller, G. W. (2007). Paraquat: The red herring of Parkinson's disease research. *Toxicological Sciences*, *100*(1), 1–2. doi:10.1093/toxsci/kfm223 PMID:17934192

Mosconi, L., de Leon, M., Murray, J., Lu, J., Javier, E., McHugh, P., & Swerdlow, R. H. (2011). Reduced mitochondria cytochrome oxidase activity in adult children of mothers with Alzheimer's disease. *Journal of Alzheimer's Disease*, 27(3), 483–490. PMID:21841246

Murphy, D. D., Rueter, S. M., Trojanowski, J. Q., & Lee, V. M. Y. (2000). Synucleins are developmentally expressed, and α -synuclein regulates the size of the presynaptic vesicular pool in primary hippocampal neurons. *The Journal of Neuroscience*, 20(9), 3214–3220. PMID:10777786

Murphy, M. P. (2009). How mitochondria produce reactive oxygen species. *The Biochemical Journal*, *417*(1), 1–13. doi:10.1042/BJ20081386 PMID:19061483

Narendra, D., Tanaka, A., Suen, D. F., & Youle, R. J. (2008). Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. *The Journal of Cell Biology*, *183*(5), 795–803. doi:10.1083/jcb.200809125 PMID:19029340

Narendra, D. P., Kane, L. A., Hauser, D. N., Fearnley, I. M., & Youle, R. J. (2010). p62/SQSTM1 is required for Parkin-induced mitochondrial clustering but not mitophagy; VDAC1 is dispensable for both. *Autophagy*, *6*(8), 1090–1106. doi:10.4161/auto.6.8.13426 PMID:20890124

Navarro, A., & Boveris, A. (2007a). Brain mitochondrial dysfunction in aging: conditions that improve survival, neurological performance and mitochondrial function. *Frontiers in Bioscience: A Journal & Virtual Library, 12*, 1154-1163.

Navarro, A., & Boveris, A. (2007b). The mitochondrial energy transduction system and the aging process. *American Journal of Physiology. Cell Physiology*, 292(2), C670–C686. doi:10.1152/ajpcell.00213.2006 PMID:17020935

Navarro, A., & Boveris, A. (2010). Brain Mitochondrial Dysfunction in Aging, Neurodegeneration, and Parkinson's Disease. *Frontiers in Aging Neuroscience*, *2*, 34. PMID:20890446

Navarro, A., Gomez, C., López-Cepero, J. M., & Boveris, A. (2004). Beneficial effects of moderate exercise on mice aging: Survival, behavior, oxidative stress, and mitochondrial electron transfer. *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology*, 286(3), R505–R511. doi:10.1152/ajpregu.00208.2003 PMID:14615275

Newmeyer, D. D., & Ferguson-Miller, S. (2003). Mitochondria: Releasing power for life and unleashing the machineries of death. *Cell*, *112*(4), 481–490. doi:10.1016/S0092-8674(03)00116-8 PMID:12600312

Nicklas, W. J., Youngster, S. K., Kindt, M. V., & Heikkila, R. E. (1987). IV. MPTP, MPP+ and mitochondrial function. *Life Sciences*, 40(8), 721–729. doi:10.1016/0024-3205(87)90299-2 PMID:3100899

Niranjan, B. G., Bhat, N. K., & Avadhani, N. G. (1982). Preferential attack of mitochondrial DNA by aflatoxin B1 during hepatocarcinogenesis. *Science*, *215*(4528), 73–75. doi:10.1126cience.6797067 PMID:6797067

Niu, J., Yu, M., Wang, C., & Xu, Z. (2012). Leucine-rich repeat kinase 2 disturbs mitochondrial dynamics via Dynamin-like protein. *Journal of Neurochemistry*, *122*(3), 650–658. doi:10.1111/j.1471-4159.2012.07809.x PMID:22639965 Nussbaum, R. L., & Ellis, C. E. (2003). Alzheimer's disease and Parkinson's disease. *The New England Journal of Medicine*, *348*(14), 1356–1364. doi:10.1056/NEJM2003ra020003 PMID:12672864

Oestreicher, E., Sengstock, G. J., Riederer, P., Olanow, C. W., Dunn, A. J., & Arendash, G. W. (1994). Degeneration of nigrostriatal dopaminergic neurons increases iron within the substantia nigra: A histochemical and neurochemical study. *Brain Research*, *660*(1), 8–18. doi:10.1016/0006-8993(94)90833-8 PMID:7828004

Parihar, M. S., Parihar, A., Fujita, M., Hashimoto, M., & Ghafourifar, P. (2008). Mitochondrial association of alpha-synuclein causes oxidative stress. *Cellular and Molecular Life Sciences*, 65(7), 1272–1284. doi:10.100700018-008-7589-1 PMID:18322646

Parihar, M. S., Parihar, A., Fujita, M., Hashimoto, M., & Ghafourifar, P. (2009). Alpha-synuclein overexpression and aggregation exacerbates impairment of mitochondrial functions by augmenting oxidative stress in human neuroblastoma cells. *The International Journal of Biochemistry & Cell Biology*, *41*(10), 2015–2024. doi:10.1016/j.biocel.2009.05.008 PMID:19460457

Parker, W. D., Boyson, S. J., & Parks, J. K. (1989). Abnormalities of the electron transport chain in idiopathic Parkinson's disease. *Annals of Neurology*, *26*(6), 719–723. doi:10.1002/ana.410260606 PMID:2557792

Payne, B. A. I., & Chinnery, P. F. (2015). Mitochondrial dysfunction in aging: Much progress but many unresolved questions. *Biochimica et Biophysica Acta*, *1847*(11), 1347–1353. doi:10.1016/j.bba-bio.2015.05.022 PMID:26050973

Pearson, K. J., Baur, J. A., Lewis, K. N., Peshkin, L., Price, N. L., Labinskyy, N., ... Jamieson, H. A. (2008). Resveratrol delays age-related deterioration and mimics transcriptional aspects of dietary restriction without extending life span. *Cell Metabolism*, 8(2), 157–168. doi:10.1016/j.cmet.2008.06.011 PMID:18599363

Pellegrino, M. W., & Haynes, C. M. (2015). Mitophagy and the mitochondrial unfolded protein response in neurodegeneration and bacterial infection. *BMC Biology*, *13*(1), 22. doi:10.118612915-015-0129-1 PMID:25857750

Perier, C., Tieu, K., Guégan, C., Caspersen, C., Jackson-Lewis, V., Carelli, V., ... Vila, M. (2005). Complex I deficiency primes Bax-dependent neuronal apoptosis through mitochondrial oxidative damage. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(52), 19126–19131. doi:10.1073/pnas.0508215102 PMID:16365298

Perumal, A. S., Gopal, V. B., Tordzro, W. K., Cooper, T. B., & Cadet, J. L. (1992). Vitamin E attenuates the toxic effects of 6-hydroxydopamine on free radical scavenging systems in rat brain. *Brain Research Bulletin*, 29(5), 699–701. doi:10.1016/0361-9230(92)90142-K PMID:1422867

Pickrell, A. M., & Youle, R. J. (2015). The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease. *Neuron*, 85(2), 257–273. doi:10.1016/j.neuron.2014.12.007 PMID:25611507

Picone, P., Nuzzo, D., Caruana, L., Scafidi, V., & Di Carlo, M. (2014). Mitochondrial dysfunction: Different routes to Alzheimer's disease therapy. *Oxidative Medicine and Cellular Longevity*. PMID:25221640

Plotegher, N., Gratton, E., & Bubacco, L. (2014). Number and Brightness analysis of alpha-synuclein oligomerization and the associated mitochondrial morphology alterations in live cells. *Biochimica et Biophysica Acta (BBA)-General Subjects, 1840*(6).

Poole, A. C., Thomas, R. E., Andrews, L. A., McBride, H. M., Whitworth, A. J., & Pallanck, L. J. (2008). The PINK1/Parkin pathway regulates mitochondrial morphology. *Proceedings of the National Academy of Sciences of the United States of America*, *105*(5), 1638–1643. doi:10.1073/pnas.0709336105 PMID:18230723

Qin, W., Haroutunian, V., Katsel, P., Cardozo, C. P., Ho, L., Buxbaum, J. D., & Pasinetti, G. M. (2009). PGC-1α expression decreases in the Alzheimer disease brain as a function of dementia. *Archives of Neurology*, *66*(3), 352–361. doi:10.1001/archneurol.2008.588 PMID:19273754

Ramsay, R. R., Kowal, A. T., Johnson, M. K., Salach, J. I., & Singer, T. P. (1987). The inhibition site of MPP+, the neurotoxic bioactivation product of 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine is near the Q-binding site of NADH dehydrogenase. *Archives of Biochemistry and Biophysics*, 259(2), 645–649. doi:10.1016/0003-9861(87)90531-5 PMID:2827583

Rebrin, I., & Sohal, R. S. (2008). Pro-oxidant shift in glutathione redox state during aging. *Advanced Drug Delivery Reviews*, 60(13), 1545–1552. doi:10.1016/j.addr.2008.06.001 PMID:18652861

Reddy, P. H. (2008). Mitochondrial medicine for aging and neurodegenerative diseases. *Neuromolecular Medicine*, *10*(4), 291–315. doi:10.100712017-008-8044-z PMID:18566920

Reddy, P. H., & Beal, M. F. (2008). Amyloid beta, mitochondrial dysfunction and synaptic damage: Implications for cognitive decline in aging and Alzheimer's disease. *Trends in Molecular Medicine*, *14*(2), 45–53. doi:10.1016/j.molmed.2007.12.002 PMID:18218341

Reiman, E. M., Caselli, R. J., Yun, L. S., Chen, K., Bandy, D., Minoshima, S., ... Osborne, D. (1996). Preclinical evidence of Alzheimer's disease in persons homozygous for the ε4 allele for apolipoprotein E. *The New England Journal of Medicine*, *334*(12), 752–758. doi:10.1056/NEJM199603213341202 PMID:8592548

Riachi, N. J., LaManna, J. C., & Harik, S. I. (1989). Entry of 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine into the rat brain. *The Journal of Pharmacology and Experimental Therapeutics*, 249(3), 744–748. PMID:2786562

Richter, C., Park, J. W., & Ames, B. N. (1988). Normal oxidative damage to mitochondrial and nuclear DNA is extensive. *Proceedings of the National Academy of Sciences of the United States of America*, 85(17), 6465–6467. doi:10.1073/pnas.85.17.6465 PMID:3413108

Rizzuto, R., De Stefani, D., Raffaello, A., & Mammucari, C. (2012). Mitochondria as sensors and regulators of calcium signalling. *Nature Reviews. Molecular Cell Biology*, *13*(9), 566–578. doi:10.1038/nrm3412 PMID:22850819

Rosen, K. M., Moussa, C. E. H., Lee, H. K., Kumar, P., Kitada, T., Qin, G., ... Querfurth, H. W. (2010). Parkin reverses intracellular β -amyloid accumulation and its negative effects on proteasome function. *Journal of Neuroscience Research*, 88(1), 167–178. doi:10.1002/jnr.22178 PMID:19610108

Mitochondrial Dysfunction in Aging and Neurodegeneration

Rossi, S. C., Gorman, N., & Wetterhahn, K. E. (1988). Mitochondrial reduction of the carcinogen chromate: Formation of chromium (V). *Chemical Research in Toxicology*, *1*(2), 101–107. doi:10.1021/tx00002a003 PMID:2979716

Rothfuss, O., Fischer, H., Hasegawa, T., Maisel, M., Leitner, P., Miesel, F., ... Patenge, N. (2009). Parkin protects mitochondrial genome integrity and supports mitochondrial DNA repair. *Human Molecular Genetics*, *18*(20), 3832–3850. doi:10.1093/hmg/ddp327 PMID:19617636

Rowland, A. A., & Voeltz, G. K. (2012). Endoplasmic reticulum-mitochondria contacts: Function of the junction. *Nature Reviews. Molecular Cell Biology*, *13*(10), 607–625. doi:10.1038/nrm3440 PMID:22992592

Rubinsztein, D. C., Mariño, G., & Kroemer, G. (2011). Autophagy and aging. *Cell*, *146*(5), 682–695. doi:10.1016/j.cell.2011.07.030 PMID:21884931

Sanz-Blasco, S., Valero, R. A., Rodríguez-Crespo, I., Villalobos, C., & Núñez, L. (2008). Mitochondrial Ca2+ overload underlies Aβ oligomers neurotoxicity providing an unexpected mechanism of neuroprotection by NSAIDs. *PLoS One*, *3*(7), e2718. doi:10.1371/journal.pone.0002718 PMID:18648507

Schapira, A. H. V. (2012). Targeting mitochondria for neuroprotection in Parkinson's disease. *Antioxidants & Redox Signalling*, *16*(9), 965–973. doi:10.1089/ars.2011.4419 PMID:22229791

Schapira, A. H. V., Cooper, J. M., Dexter, D., Clark, J. B., Jenner, P., & Marsden, C. D. (1990). Mitochondrial complex I deficiency in Parkinson's disease. *Journal of Neurochemistry*, *54*(3), 823–827. doi:10.1111/j.1471-4159.1990.tb02325.x PMID:2154550

Scotcher, K. P., Irwin, I., DeLanney, L. E., Langston, J. W., & Monte, D. (1990). Effects of 1-Methyl-4-Phenyl-1, 2, 3, 6-Tetrahydropyridine and 1-Methyl-4-Phenylpyridinium Ion on ATP Levels of Mouse Brain Synaptosomes. *Journal of Neurochemistry*, *54*(4), 1295–1301. doi:10.1111/j.1471-4159.1990. tb01962.x PMID:2313288

Sheng, B., Wang, X., Su, B., Lee, H. G., Casadesus, G., Perry, G., & Zhu, X. (2012). Impaired mitochondrial biogenesis contributes to mitochondrial dysfunction in Alzheimer's disease. *Journal of Neurochemistry*, *120*(3), 419–429. doi:10.1111/j.1471-4159.2011.07581.x PMID:22077634

Sherer, T. B., Betarbet, R., Kim, J. H., & Greenamyre, J. T. (2003c). Selective microglial activation in the rat rotenone model of Parkinson's disease. *Neuroscience Letters*, *341*(2), 87–90. doi:10.1016/S0304-3940(03)00172-1 PMID:12686372

Sherer, T. B., Betarbet, R., Testa, C. M., Seo, B. B., Richardson, J. R., Kim, J. H., ... Greenamyre, J. T. (2003a). Mechanism of toxicity in rotenone models of Parkinson's disease. *The Journal of Neuroscience*, 23(34), 10756–10764. PMID:14645467

Sherer, T. B., Kim, J. H., Betarbet, R., & Greenamyre, J. T. (2003b). Subcutaneous rotenone exposure causes highly selective dopaminergic degeneration and α -synuclein aggregation. *Experimental Neurology*, *179*(1), 9–16. doi:10.1006/exnr.2002.8072 PMID:12504863

Shimizu, K., Ohtaki, K., Matsubara, K., Aoyama, K., Uezono, T., Saito, O., ... Shiono, H. (2001). Carriermediated processes in blood–brain barrier penetration and neural uptake of paraquat. *Brain Research*, *906*(1), 135–142. doi:10.1016/S0006-8993(01)02577-X PMID:11430870

Smith, D. G., Cappai, R., & Barnham, K. J. (2007). The redox chemistry of the Alzheimer's disease amyloid β peptide. *Biochimica et Biophysica Acta (BBA)-. Biomembranes*, *1768*(8), 1976–1990. doi:10.1016/j.bbamem.2007.02.002

Smith, M. A., Richey, G. P. P., Sayre, L. M., Anderson, V. E., Beal, M. F., & Kowal, N. (1996). Test for oxidative damage in Alzheimer's. *Nature*, *382*(6587), 120–121. doi:10.1038/382120b0 PMID:8700201

Sohal, R. S., Ku, H. H., Agarwal, S., Forster, M. J., & Lal, H. (1994). Oxidative damage, mitochondrial oxidant generation and antioxidant defenses during aging and in response to food restriction in the mouse. *Mechanisms of Ageing and Development*, 74(1), 121–133. doi:10.1016/0047-6374(94)90104-X PMID:7934203

Sohal, R. S., & Sohal, B. H. (1991). Hydrogen peroxide release by mitochondria increases during aging. *Mechanisms of Ageing and Development*, 57(2), 187–202. doi:10.1016/0047-6374(91)90034-W PMID:1904965

Soong, N. W., Hinton, D. R., Cortopassi, G., & Arnheim, N. (1992). Mosaicism for a specific somatic mitochondrial DNA mutation in adult human brain. *Nature Genetics*, 2(4), 318–323. doi:10.1038/ng1292-318 PMID:1303287

Sterky, F. H., Lee, S., Wibom, R., Olson, L., & Larsson, N. G. (2011). Impaired mitochondrial transport and Parkin-independent degeneration of respiratory chain-deficient dopamine neurons in vivo. *Proceedings of the National Academy of Sciences of the United States of America*, *108*(31), 12937–12942. doi:10.1073/pnas.1103295108 PMID:21768369

Stuart, J. A., & Brown, M. F. (2006). Mitochondrial DNA maintenance and bioenergetics. *Biochimica et Biophysica Acta (BBA)-. Bioenergetics*, 1757(2), 79–89. doi:10.1016/j.bbabio.2006.01.003

Swerdlow, R. H., Burns, J. M., & Khan, S. M. (2014). The Alzheimer's disease mitochondrial cascade hypothesis: Progress and perspectives. *Biochimica et Biophysica Acta (BBA)-. Molecular Basis of Disease*, *1842*(8), 1219–1231. doi:10.1016/j.bbadis.2013.09.010 PMID:24071439

Swerdlow, R. H., Parks, J. K., Miller, S. W., Davis, R. E., Tuttle, J. B., Trimmer, P. A., ... Parker, W. D. (1996). Origin and functional consequences of the complex I defect in Parkinson's disease. *Annals of Neurology*, *40*(4), 663–671. doi:10.1002/ana.410400417 PMID:8871587

Tait, S. W., & Green, D. R. (2010). Mitochondria and cell death: Outer membrane permeabilization and beyond. *Nature Reviews. Molecular Cell Biology*, *11*(9), 621–632. doi:10.1038/nrm2952 PMID:20683470

Tatton, N. A., & Kish, S. J. (1997). In situ detection of apoptotic nuclei in the substantia nigra compacta of 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine-treated mice using terminal deoxynucleotidyl transferase labelling and acridine orange staining. *Neuroscience*, 77(4), 1037–1048. doi:10.1016/S0306-4522(96)00545-3 PMID:9130785

Taylor, R. W., & Turnbull, D. M. (2005). Mitochondrial DNA mutations in human disease. *Nature Reviews. Genetics*, 6(5), 389–402. doi:10.1038/nrg1606 PMID:15861210

Temme, C., Weissbach, R., Lilie, H., Wilson, C., Meinhart, A., Meyer, S., ... Wahle, E. (2009). The Drosophila melanogaster gene cg4930 encodes a high affinity inhibitor for endonuclease G. *The Journal of Biological Chemistry*, 284(13), 8337–8348. doi:10.1074/jbc.M808319200 PMID:19129189

Tieu, K., Perier, C., Caspersen, C., Teismann, P., Wu, D. C., Yan, S. D., ... Przedborski, S. (2003). D-β-Hydroxybutyrate rescues mitochondrial respiration and mitigates features of Parkinson disease. *The Journal of Clinical Investigation*, *112*(6), 892–901. doi:10.1172/JCI200318797 PMID:12975474

Trevelyan, A. J., Kirby, D. M., Smulders-Srinivasan, T. K., Nooteboom, M., Acin-Perez, R., Enriquez, J. A., ... Turnbull, D. M. (2010). Mitochondrial DNA mutations affect calcium handling in differentiated neurons. *Brain*, *133*(3), 787–796. doi:10.1093/brain/awq023 PMID:20207702

Truban, D., Hou, X., Caulfield, T. R., Fiesel, F. C., & Springer, W. (2016). PINK1, Parkin, and Mitochondrial Quality Control: What can we Learn about Parkinson's Disease Pathobiology? *Journal of Parkinson's Disease*, 7(1), 13–29. doi:10.3233/JPD-160989 PMID:27911343

Tsujimoto, Y. (2000). Mitochondria and cell death. *Cell Death and Differentiation*, 7(1), 134–135. doi:10.1038j.cdd.4400645

Ungerstedt, U. (1968). 6-Hydroxy-dopamine induced degeneration of central monoamine neurons. *European Journal of Pharmacology*, 5(1), 107–110. doi:10.1016/0014-2999(68)90164-7 PMID:5718510

Ungvari, Z., Orosz, Z., Labinskyy, N., Rivera, A., Xiangmin, Z., Smith, K., & Csiszar, A. (2007). Increased mitochondrial H 2 O 2 production promotes endothelial NF-κB activation in aged rat arteries. *American Journal of Physiology. Heart and Circulatory Physiology*, 293(1), H37–H47. doi:10.1152/ ajpheart.01346.2006 PMID:17416599

Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T., Mazur, M., & Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The International Journal of Biochemistry & Cell Biology*, *39*(1), 44–84. doi:10.1016/j.biocel.2006.07.001 PMID:16978905

Van Egmond, W. N., & van Haastert, P. J. (2010). Characterization of the Roco protein family in Dictyostelium discoideum. *Eukaryotic Cell*, *9*(5), 751–761. doi:10.1128/EC.00366-09 PMID:20348387

Van Houten, B., Woshner, V., & Santos, J. H. (2006). Role of mitochondrial DNA in toxic responses to oxidative stress. *DNA Repair*, 5(2), 145–152. doi:10.1016/j.dnarep.2005.03.002 PMID:15878696

Viña, J., Sastre, J., Pallardó, F., & Borrás, C. (2003). Mitochondrial theory of aging: Importance to explain why females live longer than males. *Antioxidants & Redox Signalling*, *5*(5), 549–556. doi:10.1089/152308603770310194 PMID:14580309

Wang, H., Lim, P. J., Karbowski, M., & Monteiro, M. J. (2008). Effects of overexpression of huntingtin proteins on mitochondrial integrity. *Human Molecular Genetics*, *18*(4), 737–752. doi:10.1093/hmg/ ddn404 PMID:19039036

Wang, X. F., Li, S., Chou, A. P., & Bronstein, J. M. (2006). Inhibitory effects of pesticides on proteasome activity: Implication in Parkinson's disease. *Neurobiology of Disease*, 23(1), 198–205. doi:10.1016/j. nbd.2006.02.012 PMID:16626962

Watson, R. E., McKim, J. M., Cockerell, G. L., & Goodman, J. I. (2004). The value of DNA methylation analysis in basic, initial toxicity assessments. *Toxicological Sciences*, 79(1), 178–188. doi:10.1093/toxsci/kfh099 PMID:15103049

Wei, M. C., Zong, W. X., Cheng, E. H. Y., Lindsten, T., Panoutsakopoulou, V., Ross, A. J., ... Korsmeyer, S. J. (2001). Proapoptotic BAX and BAK: A requisite gateway to mitochondrial dysfunction and death. *Science*, *292*(5517), 727–730. doi:10.1126cience.1059108 PMID:11326099

West, A. B., Dawson, V. L., & Dawson, T. M. (2007). The role of Parkin in Parkinson's disease. *Neuro-logical Disease & Therapy*, 83, 199.

West, A. B., Moore, D. J., Biskup, S., Bugayenko, A., Smith, W. W., Ross, C. A., ... Dawson, T. M. (2005). Parkinson's disease-associated mutations in leucine-rich repeat kinase 2 augment kinase activity. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(46), 16842–16847. doi:10.1073/pnas.0507360102 PMID:16269541

Wood-Kaczmar, A., Gandhi, S., & Wood, N. W. (2006). Understanding the molecular causes of Parkinson's disease. *Trends in Molecular Medicine*, *12*(11), 521–528. doi:10.1016/j.molmed.2006.09.007 PMID:17027339

Wunderlich, V., Schütt, M., Böttger, M., & Graffi, A. (1970). Preferential alkylation of mitochondrial deoxyribonucleic acid by N-methyl-N-nitrosourea. *The Biochemical Journal*, *118*(1), 99–109. doi:10.1042/bj1180099 PMID:5472159

Yakes, F. M., & Van Houten, B. (1997). Mitochondrial DNA damage is more extensive and persists longer than nuclear DNA damage in human cells following oxidative stress. *Proceedings of the National Academy of Sciences of the United States of America*, 94(2), 514–519. doi:10.1073/pnas.94.2.514 PMID:9012815

Yang, Y., Gehrke, S., Imai, Y., Huang, Z., Ouyang, Y., Wang, J. W., ... Lu, B. (2006). Mitochondrial pathology and muscle and dopaminergic neuron degeneration caused by inactivation of Drosophila Pink1 is rescued by Parkin. *Proceedings of the National Academy of Sciences of the United States of America*, *103*(28), 10793–10798. doi:10.1073/pnas.0602493103 PMID:16818890

Yen, T. C., Su, J. H., King, K. L., & Wei, Y. H. (1991). Ageing-associated 5 kb deletion in human liver mitochondrial DNA. *Biochemical and Biophysical Research Communications*, *178*(1), 124–131. doi:10.1016/0006-291X(91)91788-E PMID:2069552

Yoshii, S. R., Kishi, C., Ishihara, N., & Mizushima, N. (2011). Parkin mediates proteasome-dependent protein degradation and rupture of the outer mitochondrial membrane. *The Journal of Biological Chemistry*, 286(22), 19630–19640. doi:10.1074/jbc.M110.209338 PMID:21454557

Mitochondrial Dysfunction in Aging and Neurodegeneration

Zhang, J., Fitsanakis, V. A., Gu, G., Jing, D., Ao, M., Amarnath, V., & Montine, T. J. (2003). Manganese ethylene-bis-dithiocarbamate and selective dopaminergic neurodegeneration in rat: A link through mitochondrial dysfunction. *Journal of Neurochemistry*, 84(2), 336–346. doi:10.1046/j.1471-4159.2003.01525.x PMID:12558996

Zhang, Q., Raoof, M., Chen, Y., Sumi, Y., Sursal, T., Junger, W., ... Hauser, C. J. (2010). Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature*, *464*(7285), 104–107. doi:10.1038/ nature08780 PMID:20203610

Zhang, Y., Gao, J., Chung, K. K., Huang, H., Dawson, V. L., & Dawson, T. M. (2000). Parkin functions as an E2-dependent ubiquitin–protein ligase and promotes the degradation of the synaptic vesicle-associated protein, CDCrel-1. *Proceedings of the National Academy of Sciences of the United States of America*, 97(24), 13354–13359. doi:10.1073/pnas.240347797 PMID:11078524

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ABSTRACT

Neurodegenerative disorders (NDs) are characterized by dysfunction and loss of neurons associated with altered proteins that accumulate in the human brain and peripheral organs. Mitochondrial and Golgi apparatus (GA) dysfunctions are supposed to be responsible for various NDs. Damaged mitochondria do not produce sufficient adenosine triphosphate (ATP) and produce reactive oxygen species (ROS) and pro-apoptotic factors. Mitochondrial dysfunctions may be caused by various factors such as environmental causes, mutations in both nuclear or mitochondrial deoxyribonucleic acid (DNA), that code many mitochondrial components. Three factors that are mainly responsible for the morphological changes in GA are certain pathological conditions, drugs, and over expression of Golgi associated proteins. In this chapter, common aspects of mitochondrial and GA dysfunction concerned about NDs are summarized and described for Alzheimer's disease (AD), Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), and Huntington's disease (HD).

DOI: 10.4018/978-1-5225-5282-6.ch005

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INTRODUCTION

NDs are characterized by bioenergetics defect, oxidative stress (OS) and progressive loss of physiologically or anatomically related neuronal system in the human brain (Schapira, 2008; Malkus et al, 2009; Oliveira, 2010). Neurons are the fundamental structure and functional unit of the nervous system of living beings which includes the spinal cord and brain. The neuronal diseases are incurable conditions result in progressive degeneration of nerve cells and cause ataxias or dementias (Beal, 2005; Manczak et al, 2005). The examples of NDs are AD, PD, ALS, prion disease, motor neuron diseases (MND), HD, spinocerebellar ataxia (SCA) and spinal muscular atrophy (SMA). The exact molecular, cellular and pathological mechanism responsible for the progression of these diseases and neuronal cell death are not fully understood. Every cell is a complex communications of various organelles such as endoplasmic reticulum (ER), GA, mitochondria, nucleus, etc. carrying out several functions. These organelles exchange materials and signals to make the cell functional properly without producing and defects. Nowadays, advanced techniques are allowing researchers to understand the connecting link between these organelles. Among the cell organelles, defective connections between mitochondria and GA have been concerned in several NDs. Generation of ROS in mitochondria and GA has been identified as an important factor for the cell death (Swerdlow & Khan, 2004; Lin & Beal, 2006; Reddy & Beal, 2008; Reddy, 2008). Mitochondria are exceptional amongst other organelles, as they dispose their own, mitochondrial DNA (mtDNA), which is mainly inherited from the mother. Their number is very high in neurons, mainly in synaptic terminals, as they are the major energy producers through tricarboxylic acid cycle (TCA) and oxidative phosphorylation. They undergo continual fusion and fission leading to their elongation or fragmentation respectively. GA is associated with protein trafficking in a cell and plays an important role in the pathogenesis of NDs. The objective of the chapter is to focus on the impact of dysfunctional mitochondria and GA on various NDs.

BACKGROUND

Various NDs have been recognized for centuries and research into their causes and effects has been taking place for decades. NDs are characterized by progressive loss of anatomically related neuronal systems. As life expectancy continues in developed countries, the occurrence of these disorders also increases. In the literature, much knowledge is being available concerning the mechanisms of disease, but the causing factors of these problems are still not well known. To date, many mechanisms have been recommended for explanation of protein aggregation metabolism, and misfolding, protein neuronal function, and cell signaling, but still it is difficult to understand the mechanism clearly at the cellular and molecular level (Tan et al, 2014). To understand these diseases, we need to understand how these cells function, their response to local environment and effect of dysfunctional organelles on these diseases. In this book chapter, authors focused on the role of mitochondria and GA on the dysfunction of neurons that result in various NDs. Mitochondrial play an important role in Ca²⁺ homeostasis, ATP generation, ROS formation, and even apoptosis etc. and any dysfunction in these processes results in the dysfunction of neurons in a large number of NDs (Baloyannis, 2006). Functional or structural alterations of the Golgi pathology also recognized as a constant pathological characteristic of various NDs including PD, AD, HD, ALS, and prion diseases (Canet-Avilés et al, 2004). The neuropathological changes observed in these diseases can vary with the type of mutation in mtDNA, and level of Golgi dysfunction. GA marked as

fragmentation into disconnected cisternae, stacks, vesicles and tubules, and as atrophy (Canet-Avilés et al, 2004). These morphological changes in GA alternate the retrograde and anterograde transport in the secretary pathway in all the NDs. Mutations in the PD-associated genes such as α -synuclein (*SNCA*), *Pink1, Parkin, LRRK2, DJ1* etc. have been shown to affect both functioning of mitochondria and Golgi structure (Lee et al, 2004).

However, this field of NDs faces further challenges. It will be crucial to evaluate that whether mitochondrial dysfunction and Golgi pathology is causative, contributory, or homeostatic in NDs. In particular, it is crucial to understand whether both organelles in theses disease is restricted to the types of neuron that are affected such as dopaminergic neurons in PD, motor neurons in ALS, striatal neurons in HD. Furthermore, we will need to determine whether alterations in mitochondria and GA provide a possible explanation for the non-cell autonomous disease spread observed in various neurodegenerative diseases. Finally there is a need to engage growing knowledge on the mechanisms of involvement of these organelles in NDs be translated into earlier diagnosis and emergence of new therapies for these neurodegenerative disorders.

ROLE OF MITOCHONDRIA IN NEURODEGENERATIVE DISORDERS

Mitochondria are cytoplasmic organelles termed "powerhouse of the cell" essential for the production of most of the cell energy in the form of ATP (Pagliarini & Rutter, 2013). They also perform several other cellular functions, including: maintain calcium fluctuations; the release of proteins that known to activate the caspase family of proteases; modification of the reduction-oxidation potential of cells; biosynthesis of amino acids and steroids; β -oxidation of fatty acids and; production and modulation of ROS (Reddy, 2007; Sas et al, 2007). Mitochondria are compartmentalized into the outer and the inner mitochondrial lipids membrane composed of phospholipid bilayers and proteins. The outer membrane is porous and allows the passage of low molecular-weight substances between the intermembrane space and the cytosol through porins (Silhavy et al, 2010). Unlike outer membrane, the inner membrane does not contain porin so highly impermeable to all molecules. Mostly mitochondria are transmitted maternally, however, rarely; a recombination of mtDNA and paternal inheritance have been reported (Reddy & Beal, 2005). Mitochondria are controlled by both nuclear and mitochondrial genomes where mtDNA consists of a 16,571 base-pair, circular double-stranded DNA molecule (Anderson et al, 1981). It contains 2–10 copies of mtDNA (Reddy, 2008). mtDNA encodes 13 polypeptide genes that encode essential components of the electron transport chain (ETC). mtDNA also contains the 22 tRNA genes and 12S and 16S rRNA genes required for mitochondrial protein synthesis (Reddy & Beal, 2005). Nuclear genes encode the remaining mitochondrial proteins and transport into mitochondria, DNA and RNA polymerases, metabolic enzymes, mtDNA regulatory factors and ribosomal proteins.

SITES OF FREE RADICAL PRODUCTION IN THE MITOCHONDRIA

Mitochondrial ATP is generated via oxidative phosphorylation within the inner mitochondrial membrane and various free radicals are produced as a byproduct of this. In the respiratory chain, complexes I and III leak electrons to oxygen, producing superoxide radicals (Jastroch et al, 2010). The manganese superoxide dismutase (Mn-SOD) converts superoxide to hydrogen peroxide (H_2O_2) and oxygen (Figure

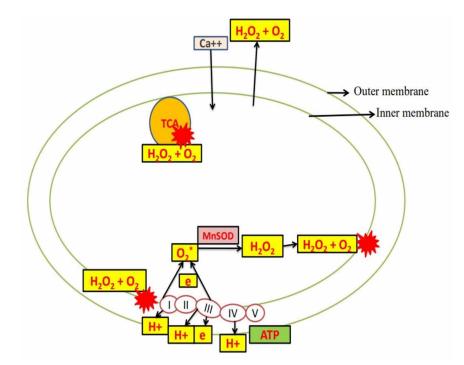
1). H_2O_2 and O_2 permeate by membranes and become a source of reactive hydroxyl radicals (Hroudova et al, 2014). Complex I generates electrons toward the mitochondrial matrix, whereas complex III generates toward both the matrix and the inter-membrane space (Brieger et al, 2012). The components of TCA i.e. α -ketoglutarate dehydrogenase, generate superoxide radicals in the matrix. These free radicals synthesized in mitochondria are carried to the cytoplasm via voltage-dependent anion channels, where they participate in lipid peroxidation, DNA and protein oxidation (Bolisetty & Jaimes, 2013).

Excitotoxicity and apoptosis are the two ways of neuronal cell death and the role of mitochondria is crucial in both the cases (Emerit et al, 2004; Davis & Williams, 2012). In apoptosis, the outer membrane of mitochondria is permeabilized, that leads to release of various apoptotic proteins like cyt *c*. There are many interrelated mitochondrial pathways that facilitate apoptosis or cell death:

- Opening of mitochondrial permeability transition pore (MPTP) lead to mitochondrial swelling and cell death through necrosis or apoptosis (Aronis et al, 2003);
- Increase in the permeability of the membrane causes leak of apoptotic factors i.e. second mitochondria-derived activator of caspases (SMAC) and cyt *c*, which trigger the caspase cascade and leading to apoptosis (Aronis et al, 2003); and
- Release of apoptosis-inducing factor (caspase-independent death effector) that triggers chromatin condensation, leading to DNA degradation (Aronis et al, 2003).

Mitochondria undergo breakdown or fragmentation during apoptosis before activation of caspases (Lu, 2009). Recently, attention is paid to the damage of ETC complex induced by ROS that is mediated by the oxidative and peroxidation damage of cardiolipin (Musatov & Robinson, 2012; Chaturvedi &

Figure 1. Different sites of free radical production in mitochondria

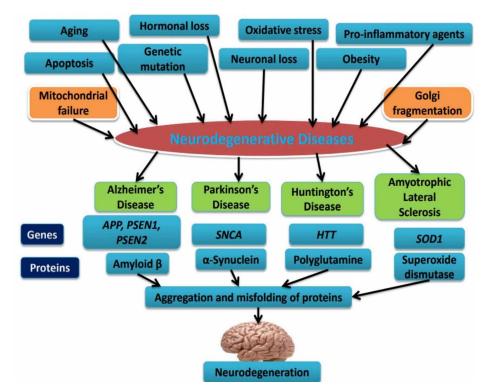


Beal, 2013). Cardiolipin, a membrane protein is required for both the stability of respiratory super complexes and as a diffusion microdomain for the ubiquinone (Paradies et al, 2014). It also plays an active role in mitochondrial mediated apoptosis, by oxidizing, and interacting with cyt c and Bcl-2 proteins (Yin & Zhu, 2012).

Mitochondria play an important role in the metabolism of all mammalian cells, including abnormality in mitochondrial structure and function and brain neuron that may lead to age-related NDs. The first evidence of participation of mitochondria in pathogenesis of neurodegenerative process was reported in platelet mitochondria of patients suffering from PD (Schapira et al, 1990). Later, scientists reported ETC deficiencies i.e. complex I and cytochrome *c* oxidase (complex IV, COX) in AD and complexes II and III in HD (Moran et al, 2012). Recently various cellular, molecular, biochemical, and animal model studies of NDs via mutant proteins associated (MPA) with mitochondria several (MS) revealed that mutant proteins such as amyloid beta in AD, mutant SOD1 in ALS, mutant HTT in HD, mutant DJ1, mutant Parkin and mutant SNCA in PD, and frataxin in Friedreich's ataxia (FRDA) are present on mitochondrial membranes (Figure 2), leading to a low production of cellular ATP due to the increased production of free radicals and that ultimately leads to cell death (Reddy, 2008).

Considering the nature of NDs and imperfect regenerative capacity of neurons, inappropriate functioning of mitochondria can have destructive effects on neuronal survival. There are various evidence of impair mitochondrial function as a cause rather than result of neurodegeneration.

Figure 2. Abnormal misfolding and aggregation of proteins due to gene mutations responsible for various neurodegenerative diseases



ABNORMAL MITOCHONDRIAL DYNAMICS IN NEURODEGENERATIVE DISORDERS

Alzheimer's Disease

AD is the most common neurodegenerative disorder of aging that lead to memory loss, disability in language, behavior and death (Lockrow et al, 2012). The disease is distinguished by a neuronal loss and deposition of 42-amino acid amyloid- β (A β) derivatives and presence of senile plaque and neurofibrillary tangles (Scheuner et al, 1996). Hardy & Higgins (1992) proposed "the amyloid cascade hypothesis" states that disturbed processing of amyloid precursor protein (APP) or change in A β results in imbalance between A β production and removal. Damage to the structure of mitochondria and increased OS are extensively reported in AD (Zhu et al, 2006). Dysfunctional mitochondria showed diminished respiratory capacity and deficiency in several key enzymes including α -ketoglutarate dehydrogenase complex (α KGDHC), pyruvate dehydrogenase complex (PHDC), two rats limiting enzymes of TCA and cytochrome oxidase (COX) (Maurer et al, 2000; Casley et al, 2002). In AD, cytoplasmic hybrids made from mitochondrial DNA and altered calcium homeostasis has also been reported in various studies (Khan et al, 2000). As compared to healthy organism, rearrangement of sporadic mtDNA or mutations is significantly increased in AD patients (Corral-Debrinski et al, 1994; Coskun et al, 2004). Lustbader and group (2004) demonstrated that blocking the association of A β and A β -binding alcohol dehydrogenase (A β -AD) can suppress the apoptosis and generation of free radicals in neurons (Crouch et al, 2005; Devi et al, 2006).

Studies on quantitative ultrastructural morphometric analysis revealed that as compared to age matched control group brains, AD brains contains lower percentage of normal mitochondria and higher percentage of mitochondria with broken cristae (Monte et al, 2000). This is further supported by the presence of longer mitochondria in fibroblasts, then those of age-matched normal human fibroblast (Wang et al, 2008).

Several studies reported that activity of complex I and gene expression of ND4 subunit of complex I get decreased in AD brains and in temporal cortex of AD patients (Chandrasekaran et al, 1996). Manczak and group (2005) observed differential expression of gene present in complex I, V and COX in AD brains. In the surviving neurons, gene expression of COX gets increased due to high oxidative damage and alteration of mitochondrial function (Manczak et al, 2004). Fusion and fission pathway also play an important role in relation to altered mtDNA as fusion of mitochondria enable exchange of mtDNA with other content. Mfn 2 knockout inhibit this process resulted in formation of mtDNA lacking mitochondria (Karbowski & Youle, 2003).

Disorder of Electron Transport Chain in Alzheimer's Disease

As the activity of COX was found to be very low in platelets, cortex and hippocampus of AD patients, indicates the anatomical specificity (Maurer et al, 2000). The mitochondrial deficiency was observed in the platelets of AD patients indicating reduction in COX and complex III activity (Valla et al, 2006). Reduction in acetylcholinesterase (AChE) indicates that it could increase the activity of A β (Fodero et al, 2004). Different mitochondrial malformation related with NDs result in oxidative stress, aberrant homeostasis of cytosolic calcium. Oxidative phosphorylation (OXPHOS) does not involve in thermodynamic equilibrium but increase a rate of uncoupling (Figure 3). Decrease in mitochondrial membrane potential ($\Delta \psi_m$) result in hydrolysis of ATP formed in cytoplasm, whereas increase in $\Delta \psi_m$ cause proton

leakage and result in increase in uncoupling. Overproduction of ROS, increase in mitochondrial permeability transition (MPT) and proton leakage result in apoptosis (Hroudova et al, 2014).

Parkinson's Disease

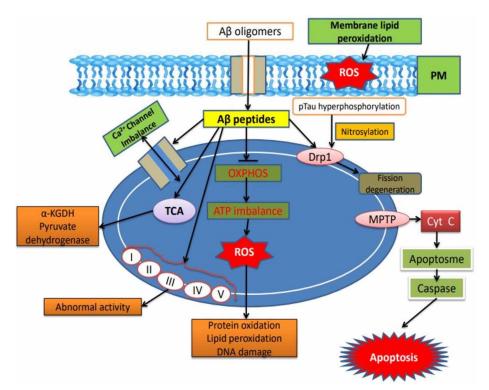
PD is the second common neurodegenerative disorder after AD. PD is a disorder of CNS that belongs to movement disorder and characterized by rigidity, loss of posture stability, resting tremor and bradykinesia (Ayano, 2016). The symptoms of PD arise due to loss of dopaminergic (DA) neurons in the substantia nigra. Surviving nigral neurons contains abnormal aggregates of proteins called Lewy bodies, an intracytoplasmic eosinophilic inclusion that is composed of fibrillar presynaptic α -synuclein protein (Spillantini et al, 1997). Various genetic studies in PD cases have identified mutation in five genes i.e. α -synuclein (*SNCA*), PTEN-induced putative kinse (*PINK-1*), *Parkin*, *DJ-1* and leucine rich repeat kinase 2 (*LRRK2*) (Abeliovich & Beal, 2006; Lesage & Brice, 2009).

α-Synuclein

Dysfunction of mitochondria plays a critical role in the pathogenesis of PD in transgenic mice, overexpressing nature of *SNCA* induces increase in OS, reduction in threshold for nigral breakdown and impaired

Figure 3. Mitochondrial damage in Alzheimer's disease

Where, $A\beta$: Amyloid β ; PM: Plasma membrane; ROS: Reactive oxygen species; TCA: Tricarboxylic acid; OXPHOS: Oxidative phosphorylation; DRP1: Dynamin related protein 1; MPTP: Mitochondrial permeability transition pore (Adopted and modified from Kumar and Singh, 2015).



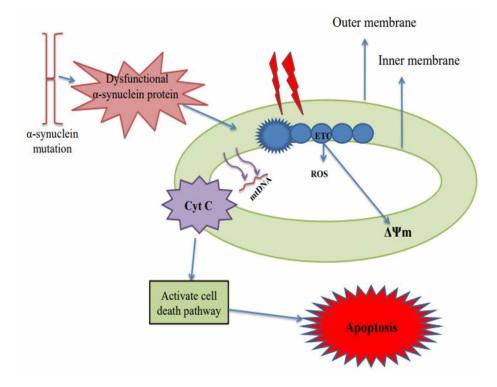
function of mitochondria (Figure 4). Martin et al (2006) undergoes α -synuclein immunostaining study in mice and evaluate that this protein affect mitochondria.

PINK1

The *PINK1* gene provides information for making a PTEN protein that induced putative kinase 1. *PINK1* gene encoding a 581 amino acid protein with N-terminal mitochondrial targeting sequence and a threonine/serine kinase domain (Mills et al, 2008). Mostly missense mutation impairs the kinase activity; that is essential for the neuroprotective nature of *PINK1*. Increased expression of *PINK1* provides protection from apoptotic death (Arena et al, 2013). Whereas loss of its function increase the vulnerability of cells to death induced by stress (Petit et al, 2005; Wood-Kaczmar et al, 2008). *PINK1* deficiency induced various effects on mitochondrial morphology and function such as decrease in $\Delta \psi_m$, activities of complex I and IV, mtDNA level, ATP production, increase in ROS and abnormal morphology (Deas et al, 2009). Gandhi et al (2009) observed that *PINK1* deficiency is responsible for the dysbalance of calcium homeostasis, where basal concentration of calcium in mitochondria is high and its overload disturb the Na⁺/Ca²⁺ exchanges present in inner mitochondrial membrane.

Figure 4. Effects of mutated α -synuclein on mitochondria

Mutated SNCA gene cause dysfunction and aggregation of protein at inner mitochondrial membrane causing dysfunction of complex I, that reduces ATP formation, increase ROS production and initiate cell death (Adapted from Mounsey & Teismann, 2011).



Parkin

In 1998, mutated Parkin gene was identified as a cause of autosomal recessive PD in Japanese families (Kitada et al, 1998). The *Parkin* gene encodes total 465 amino acids protein attached with ubiquitin-like domain (UBL) at N-terminus and Ring-Between- Ring (RBR) domains at C-terminus. Parkin protect the cell against various stresses such as excitotoxicity, endoplasmic reticulum stress, overexpression of α -synuclein, A β peptide, tau, mitochondrial dysfunction, proteosome inhibition, and expanded polyglutamine fragments (Doss-Pepe et al, 2005; Winklhofer, 2007). Biochemical and genetic studies showed that mutation induce a loss of *Parkin* function leading to its accumulation cause neurotoxicity, results in death of dopaminergic neurons. Darios et al. (2003) observed that Parkin prevent swelling of mitochondria and release of cyt c in ceramide-treated cells. Parkin and PINK1 have a direct role in various cells mitochondrial control pathways, including reduced membrane potential and mitophagy. The "MitoPark" mice are an excellent example of "mitochondrial hypothesis" of PD (Johri & Beal, 2012). In MitoPark mice, on the elimination of mitochondrial transcription factor (*Tfam*) gene, mitochondrial function gets disturbed in DA neurons (Ekstrand et al, 2007). The Tfam gene is present in the nuclear genome and it is always imported into mitochondria, where it binds to DNA for maintenance of mtDNA and transcription in mammals. Tfam stabilise mtDNA by regulating copy number and also required for biogenesis of mitochondria (Larsson et al, 1998). Cellular changes observed in this problem is similar to those observed in PD such as degeneration of dopaminergic neurons (DA) pathways, intracellular inclusions in DA neurons and loss of dopamine.

DJ-1

DJ-1 belongs to Thij/Pfpl family and has common structure with stress-inducible *E. coli* chaperone (HSp31) (Lee et al, 2003). DJ-1 gene encodes 189 amino acids proteins and mutation of this cause onset of autosomal recessive PD (Bonifati et al, 2003). Less expressions of DJ-1 make neuron cells more susceptible to oxidative injury whereas its over-expression protects cells from damage induced by OS (Park et al, 2005; Paterna et al, 2007). DJ-1 is converted into acidic pI variant against OS, on the formation of cysteine-sulfinic acid at cysteine 106 (Kinumi et al, 2004). Cysteine 106 is very important for the neuroprotective activity of DJ-1 (Yokota et al, 2003; Waak et al, 2009). Various reports showed the presence of DJ-1 in nucleus, cytosol and mitochondria, where OS disturb location of DJ-1 in mitochondria (Li et al, 2005; Lev et al, 2008). In response to OS, endogenous DJ-1 translocates to mitochondria and nucleus of human neuroblastoma cells (Junn et al, 2009).

In *DJ-1* knockout mice, isolated mitochondria show an increase in H_2O_2 accompanied by reduction in aconitase activity in mitochondria, indicating a problem in scavenging ROS (Andres-Mateos et al, 2007). From research on *Drosophila* model and human cell culturing it was observed that *DJ-1* does not function with *PINK1/Parkin* pathway (Exner et al, 2007).

LRRK2

LRRK2 gene mutation is most common cause of autosomal dominant PD associated with late-onset and sporadic PD (Paisan-Ruiz et al, 2005). *LRRK2* gene encodes various multidomain proteins of 2527 amino acids, such as a ROC domain, leucine rich repeats, WD40- repeat domain and COR domain (Biskup and West, 2009; Gandhi et al, 2009). *In vitro*, mutation increase the kinase activity of *LRRK2*, assessed

by phosphorylation or autophosphorylation of substrates that affects function mechanism (Greggio et al, 2006; Smith et al, 2006).

Amyotrophic Lateral Sclerosis

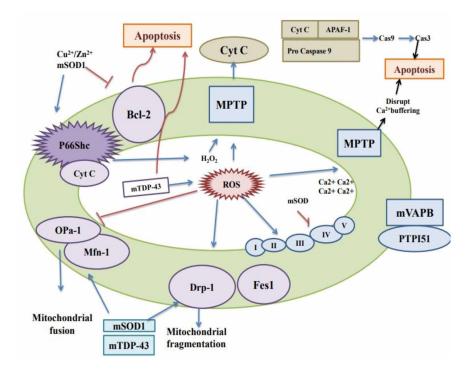
ALS is also known as Lau Gehrig's disease in humans, is a ND. ALS disease is characterized by a fatal degeneration of pyramidal neurons in the motor cortex with other nearby corticospinal tracts and motor neurons in the brain and spinal cord, resulting in atrophy, paralysis, muscle weakness, speech deficit and eventual death (Zarei et al, 2015). Sporadic in 90% of cases out of which 20% of the familial ALS cases are due to mutation in the copper-zinc superoxide dismutase (SOD 1) gene on chromosome 21 (Gros-louis et al, 2006). Mutation on SOD 1 gene is considered to be a toxic gain of function rather than a loss of SOD 1 activity (Rothstein, 2009; Su et al, 2010). Recently, mutation in two DNA/RNA binding proteins i.e. TAR DNA-binding protein-43 (TOP-43) and fused/translocated in liposarcoma (FUS/ TLS) have also been found and responsible for 4% of ALS cases (Kwiatkowski et al, 2009; Vance et al, 2009; Da Cruz & Cleveland, 2011). The mutation in transitional ER also known as valosin-containing protein gene was reported to be the known cause of 1-2% of familial ALS cases. This mutation effects on the regulation of mitochondrial calcium homeostasis. Mitochondrial morphological changes such as deformed cristae, swelling, defected respiratory chain activity and reduced mtDNA copy number are the preliminary sign of disease onset in organism with SOD 1 mutations (Nakano et al, 1987). Changes in mitochondrial ETC, in both neural and non-neural tissue have been noted by several groups from patient with ALS (Figure 5).

In patients with sporadic ALS, reduced activity of complex IV was observed whereas in patient with familial ALS, increased complex I activity was observed (Bowling et al, 1993). Deficiencies of complex I, II-III were observed in patients infected with familial ALS due to mutation in SOD 1 and also observed in SOD 1 transgenic mouse (Browne et al, 1998). In patients with sporadic ALS, both protein and lipid oxidation increased in glia and spinal cord motor neurons (Shibata, 2001). Mutant SOD1 aggregates at the outer membrane of mitochondria, inactivates Bcl-2 (antiapoptosis protein) resulting in release of cytochrome c. This results in intrinsic apoptosis.

Huntington's Disease

HD is a ND caused by the elongation of CAG repeats in first exon huntingtin (*HTT*) gene on chromosome 4 (Vonsattel & DiFiglia, 1998). This disease is characterized by intellectual decline, psychiatric illness, motor-impairment and personality change. There are many evidences for deficits and mitochondrial dysfunction in HD, like weight loss, increased lactate in basal ganglia and cerebral cortex, low activities of OXPHOS complexes II and III, membrane depolarisation of lymphoblasts and low aconitase activity in ganglia (Tabrizi et al, 1999). mtDNA deletion and damage were observed in animal models and HD patients (Acevedo-Torres et al, 2009). m*HTT* directly affect calcium in axon (Orr et al, 2008). It also inhibit mitochondrial fusion and reduced level of ATP, results in mitochondrial fragmentation (Wang et al, 2008) (Figure 6).

Figure 5. Amyotrophic lateral sclerosis linked mutant proteins mitochondrial malfunction Mutant SOD1 accumulate at outer membrane of mitochondria and inhibit the activity of Bcl-2, an anti-apoptotic protein results in the release in cytochrome-c. Mutant SOD 1 causes various mitochondrial dysfunctions such as impaired protein import and activity of ETC, aberrant morphology and increased oxidative stress, etc.



ABNORMAL GOLGI APPARATUS DYNAMICS IN NEURODEGENERATIVE DISORDERS

Morphology of Golgi Apparatus

GA is a secretory organelle with complex morphology and occupies an important position in this system. Structurally it is similar to smooth ER but relatively more compact (English and Voeltz, 2013). It consists of flattened, membrane-bound sacs called cisternae, which are involved in processing and sorting of luminal proteins together with a system of associated vesicles called Golgi vesicles. New cisternae are formed from buds derived from the smooth ER. Organization of GA is maintained by a proteinaceous matrix, inositol phospholipids, and cytoskeletal components. The post-translational modification of lipids and proteins mostly by glycosylation is carried out in GA therefore it has an important role to play in NDs (Ashaq et al, 2015).

The Golgi constitution is disrupted in a number of NDs, advocating a common mechanism and involvement of Golgi defects. Three factors that are mainly responsible for the morphological changes in GA are certain pathological conditions, drugs and over expression of Golgi associated proteins. Protein phosphatase inhibitor okadaic acid, fungal metabolite brefeldin A and nocodazole are some of the pharmacological agents that are responsible for collapsing of tubules and vesicles and shortening of cisternae or ministacks (Boe et al, 1991). GA fragmentation has been observed in an early, preclinical

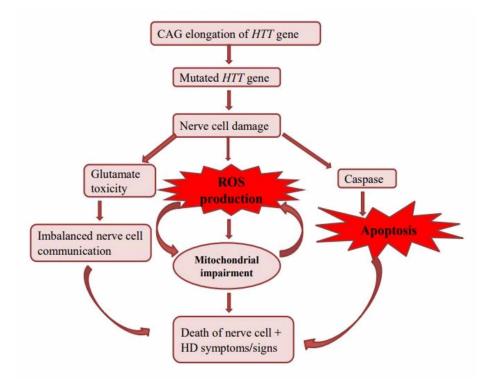


Figure 6. Schematic representation of mitochondrial dysfunction in Huntington's disease

stage of NDs therefore it can be used as a dependable indicator of activity of neurodegeneration. Following Table 1 shows the pathological implications of GA in NDs.

Neurodegenerative Diseases and Accumulation of Misfolded Proteins

NDs share some common histopathologic abnormalities such as neuronal failure, gliosis and the occurrence of inclusion bodies in both neurons and glial cells, which correspond to intracellular accumulation of misfolded or aberrant proteins (Choonara et al, 2009). Since GA is a central organelle involved in handing of proteins, therefore its role in NDs is being explored seriously. Mutated forms of SOD1, α -synuclein and tau are the key proteins that have a direct correlation with GA degeneration and its loss of function (Fan et al, 2008).

Mutant SOD1

ROS are molecules which are derived from oxygen that have accepted extra electrons and can oxidize other molecules. ROS includes singlet oxygen $({}^{1}O_{2})$, $H_{2}O_{2}$ and the highly reactive hydroxyl radical ($\blacksquare OH$) (Waris & Ahsan, 2006). According to an estimate as much as 1% of the total mitochondrial O_{2} consumption is used to produce superoxide (Jackson et al, 2016; Vyas et al, 2016). ROS have been implicated in many diseases. There are eight known sites that are capable of producing superoxide. ROS released

Disorder	Alterations in GA	Reference
Alzheimer's disease	• Mitochondrial alterations in the thalamus and the red nucleus showed fragmentation of the cisternae	Baloyannis et al, 2006
	 Phosphorylation of the GA stacking protein GRASP65 disrupted its function, resulting in GA fragmentation Perturbing GA structure affects trafficking, processing, and sorting of proteins essential for synaptic and dendritic integrity 	Joshi et al, 2015
	• GA stacks appeared disconnected and of reduced diameter, with the concomitant presence of vesicles in the vicinity of the stacks	Baloyannis, 2014
	• 20 genes are involved in lipid metabolism, inflammatory response and endocytosis constitute risk factors for late-onset AD	Giri et al, 2016
Multiple system atrophy	 Pathogenetic mechanisms causing inclusion body formation and abnormalities of the GA-trans-Golgi network. Lesions of the inferior olivary nucleus may not always reflect changes of trans synaptic degeneration secondary to Purkinje cell loss 	Sakurai et al, 2002
	• GA and trans-Golgi network of enlarged neurons lost the normal network, underwent fragmentation, reduction in number, and aggregation around nuclei	Takamine et al, 2000
Amyotrophic lateral sclerosis	• Golgi was reduced and fragmented appearing as disconnected punctate structures	Mourelatos et al, 1990; Gonatas et al, 1992
	• GA fragmentation is also present in spinal anterior horn cells in sporadic ALS patients with cytoplasmic mislocalization of WT TDP-43	Fujita et al, 1999, 2000, 2008
	• GA fragmentation was present in cells expressing ALS-linked mutant FUS, optineurin and vesicle-associated membrane protein B	Farg et al, 2012; Sundaramoorthy et al, 2015
Parkinson's disease	 GA fragmentation might trigger the aggregation of α-synuclein and the formation of inclusions, alterations in anterograde and retrograde transport between the ER and GA, and cytoskeleton damage Fragmentation is directly related with alterations in the levels of Rab1, 2 and 8 and the SNARE protein syntaxin 5 	Rendon et al, 2013
Corticobasal degeneration; Creutzfeldt-Jakob disease	 Ballooned neurons showed fragmentation of the GA Reduction in the number of fragmented GA elements by a unique perinuclear distribution 	Sakurai et al, 2000

Table 1. The pathological alterations of Golgi apparatus in neurodegenerative disorders.

from complex III are required for many biological processes including oxygen sensing, cell differentiation, and adaptive immunity.

The SOD family plays a vital physiological part in extenuating detrimental effects of ROS (Griess et al, 2017). SOD1 is an abundant intracellular enzyme with an indispensable role in antioxidant defense and its mutant form plays an important role in various NDs such as ALS (Cluskey & Ramsden, 2001).

The most frequently recognized mutations in SOD1 that affect protein activity are D90A, A4V and G93A. Mutated SOD1 leads to the accumulation of highly toxic hydroxyl radicals causing degradation of both nuclear and mitochondrial DNA and protein misfolding (Kaur et al, 2016). Misfolded protein aggregates, ubiquitin-proteasome system destruction and neuronal apoptosis carried out by receptor or mitochondrial-dependent pathways are implicated in mutant SOD1-induced toxicity (Fan et al, 2008). A third pathway of apoptosis has been proposed that is linked with ER stress. Neuronal fragmentation and degeneration may be due to aggregation of mutant SOD1 with the unfolded protein response components which are transported to ER (Turner & Atkin, 2006).

Key proteins involved in the anchorage of Golgi membranes with microtubules, could be early targets of mutated SOD1 (Karecla & Kreis, 1992). Motor neurons in the spinal cord and brain stem are mainly affected in ALS. This disease is familial (FALS) and inherited in an autosomal dominant manner. OS caused by human SOD1 mutations is supposed to play a significant role in the pathogenesis of familial ALS (FALS) and the FALS-like MND seen in the mutant SOD1 transgenic mice (Zhang et al, 1997). GA fragmentation has been observed in spinal cord motor neurons of transgenic mice expressing ALS-linked SOD1G93A and show neurodegeneration (Turner & Atkin, 2006). SOD insoluble protein complexes that disrupt both fast and slow components of axonal transport have been detected in cellular in motor neurons from ALS transgenic mice (Johnston et al, 2000). Mutant SOD1 toxicity may be due to its coprecipitation with Hsp25 because it deprives cells of the anti-apoptotic and other protective effects of Hsp25 (Strey et al, 2004). Overexpression of mutant SOD1 in ALS retards electron transport chain in mitochondria and decreases its calcium-loading capacity. SOD1 limits itself to the outer mitochondrial membrane, intermembrane space and matrix, targeting of mutant SOD1 to mitochondria causes cyte c release and apoptosis (Tan et al, 2014). Mutant SOD1 triggers anomalous mitochondrial ROS production and forms aggregates that may block the outer mitochondrial membrane protein importation machinery or bind and confiscate the antiapoptotic protein Bcl-2 (Guo et al, 2013).

α-Synuclein

 α -Synuclein is a 14 kD cytosolic neuronal, acidic lipid binding presynaptic protein whose exact function remains unknown. It has been intimately linked with the etiology of PD. A mutation was identified in the *SNCA* gene, which codes for a presynaptic protein thought to be involved in neuronal plasticity (Polymeropoulos et al, 1997). These pathological inclusions have been implicated in neurodegenerative disorders, called 'tauopathies' and 'synucleinopathies'. Evidence indicates that tauopathies and synucleinopathies may be linked (Lee et al, 2004). Mutated or over-expressed, α -synuclein has considerable impact on many membrane trafficking and stress pathways, including exocytosis, ER-to-Golgi transport, ER stress, Golgi homeostasis, endocytosis, autophagy, oxidative stress, and others (Wang & Hay, 2015).

Stathmin

Stathmin, also referred to as Op18, is a 17 KDa cytosolic phosphoprotein (Redeker et al, 2000). It is involved in a relay integrating diverse intracellular signaling pathways involved in the control of cell proliferation, differentiation etc. (Redeker et al, 2000). It interacts with several putative downstream target and/or partner proteins. It is required for the regulation of cytoskeleton, motility, division and cell cycle. Microtubule remodeling according to the requirement of the cell is controlled by it. A reduced amount of this protein has been observed in the neocortex of patients with AD, whereas it was augmented in some neurons with neurofibrillary tangles (Jin et al, 1996). Curmi et al, (1999) have reported that its phosphorylation reduces its affinity for tubulin and hence its action on microtubule dynamics.

Structural integrity of GA is maintained by microtubules. Stathmin is also recognized to be a probable factor in mutant SOD1-mediated toxicity, however, these claims need further verification because spinal cord sections from paralyzed mice seldom show co-localization of stathmin with SOD1 (Strey et al, 2004).

Tau

Tau is involved in the formation of microtubules during the growing of axon. Tau is known to inhibit kinesin-dependent transport of peroxisomes, neurofilaments, and GA derived vesicles into neurites. Loss of peroxisomes makes cells susceptible to OS leading to degeneration (Stamer et al, 2002). Fragmentation of GA has been directly linked with phosphorylation of tau which is associated with a transient dissociation of tau from the cytoskeleton and a decrease of the acetylated tubulin. Mutant human tau P301L has been implicated in development of neurofibrillary tangles an important feature observed in degenerating neurons (Iqbal et al, 2010). It can safely be assumed that GA fragmentation is not due to the process of programmed cell death (PCD) because apoptotic hallmarks such as apoptotic nuclei and activation of caspases-3 were not detected. Calnexin, a marker of the rough ER, was found to be intact but elimination of the Golgi marker protein MG160 was observed (Gonatas et al. 1989).

CONCLUSION

Damaged neurons do not have the capacity to replace themselves; therefore neurodegenerative diseases are not curable. Considerable research has been carried out during the last many years to understand pathological mechanism responsible for the progression of these diseases. Thus far, studies on mitochondrial genome have not investigated the relation between mtDNA change and mitochondrial dysfunction, although loss of mitochondrial genomic integrity has been occupied in many neurodegenerative diseases. GA occupies an important position as a secretary organelle. Certain pathological conditions and over expression of GA associated protein are responsible for its degradation. Mutated forms of SOD1, α -synuclein and tau are the key proteins are implicated in neurodegenerative diseases. Therefore, there is a need to understand the exact molecular, cellular, and pathological mechanism responsible for these diseases so that therapies can be designed to slow down or prevent neuronal loss.

REFERENCES

Abeliovich, A., & Flint Beal, M. (2006). Parkinsonism genes: Culprits and clues. *Journal of Neurochemistry*, *99*(4), 1062–1072. doi:10.1111/j.1471-4159.2006.04102.x PMID:16836655

Acevedo-Torres, K., Berríos, L., Rosario, N., Dufault, V., Skatchkov, S., Eaton, M. J., & Ayala-Torres, S. (2009). Mitochondrial DNA damage is a hallmark of chemically induced and the R6/2 transgenic model of Huntington's disease. *DNA Repair*, 8(1), 126–136. doi:10.1016/j.dnarep.2008.09.004 PMID:18935984

Anderson, S., Bankier, A. T., Barrell, B. G., De Bruijn, M. H., Coulson, A. R., Drouin, J., ... Schreier, P. H. (1981). Sequence and organization of the human mitochondrial genome. *Nature*, *290*(5806), 457–465. doi:10.1038/290457a0 PMID:7219534

Andres-Mateos, E., Perier, C., Zhang, L., Blanchard-Fillion, B., Greco, T. M., Thomas, B., & Dawson, T. M. (2007). DJ-1 gene deletion reveals that DJ-1 is an atypical peroxiredoxin-like peroxidase. *Proceedings of the National Academy of Sciences of the United States of America*, *104*(37), 14807–14812. doi:10.1073/pnas.0703219104 PMID:17766438

Arena, G., Gelmetti, V., Torosantucci, L., Vignone, D., Lamorte, G., De Rosa, P., ... Valente, E. M. (2013). PINK1 protects against cell death induced by mitochondrial depolarization, by phosphorylating Bcl-xL and impairing its pro-apoptotic cleavage. *Cell Death and Differentiation*, *20*(7), 920–930. doi:10.1038/ cdd.2013.19 PMID:23519076

Aronis, A., Melendez, J. A., Golan, O., Shilo, S., Dicter, N., & Tirosh, O. (2003). Potentiation of Fasmediated apoptosis by attenuated production of mitochondria-derived reactive oxygen species. *Cell Death and Differentiation*, *10*(3), 335–344. doi:10.1038j.cdd.4401150 PMID:12700633

Ashaq, I., Shajrul, A., Akbar, M., & Rashid, F. (2015). Protein Misfolding Diseases: In Perspective of Gain and Loss of Function. In Proteostasis and Chaperone Surveillance (pp. 105-118). Springer India.

Ayano, G. (2016). Parkinson's Disease: A Concise Overview of Etiology, Epidemiology, Diagnosis, Comorbidity and Management. *Journal of Neurological Disorders*, 4(6), 1–6. doi:10.4172/2329-6895.1000298

Baloyannis, S. J. (2006). Mitochondrial alterations in Alzheimer's disease. *Journal of Alzheimer's Disease*, 9(2), 119–126. doi:10.3233/JAD-2006-9204 PMID:16873959

Baloyannis, S. J. (2014). Golgi apparatus and protein trafficking in Alzheimer's disease. *Journal of Alzheimer's Disease*, 42(3), 153–162. PMID:24946873

Beal, M. F. (2005). Mitochondria take center stage in aging and neurodegeneration. *Annals of Neurology*, *58*(4), 495–505. doi:10.1002/ana.20624 PMID:16178023

Biskup, S., & West, A. B. (2009). Zeroing in on LRRK2-linked pathogenic mechanisms in Parkinson's disease. *Biochimica et Biophysica Acta (BBA)-. Molecular Basis of Disease*, 1792(7), 625–633. doi:10.1016/j.bbadis.2008.09.015 PMID:18973807

Bøe, R., Gjertsen, B. T., Vintermyr, O. K., Houge, G., Lanotte, M., & Døskeland, S. O. (1991). The protein phosphatase inhibitor okadaic acid induces morphological changes typical of apoptosis in mammalian cells. *Experimental Cell Research*, *195*(1), 237–246. doi:10.1016/0014-4827(91)90523-W PMID:1647324

Bolisetty, S., & Jaimes, E. A. (2013). Mitochondria and reactive oxygen species: Physiology and pathophysiology. *International Journal of Molecular Sciences*, *14*(3), 6306–6344. doi:10.3390/ijms14036306 PMID:23528859

Bonifati, V., Rizzu, P., Van Baren, M. J., Schaap, O., Breedveld, G. J., Krieger, E., & van Dongen, J. W. (2003). Mutations in the DJ-1 gene associated with autosomal recessive early-onset Parkinsonism. *Science*, *299*(5604), 256–259. doi:10.1126cience.1077209 PMID:12446870

Bowling, A. C., Schulz, J. B., Brown, R. H. Jr, & Beal, M. F. (1993). Superoxide dismutase activity, oxidative damage, and mitochondrial energy metabolism in familial and sporadic amyotrophic lateral sclerosis. *Journal of Neurochemistry*, *61*(6), 2322–2325. doi:10.1111/j.1471-4159.1993.tb07478.x PMID:8245985

Brieger, K., Schiavone, S., Miller, F. J. Jr, & Krause, K. H. (2012). Reactive oxygen species: From health to disease. *Swiss Medical Weekly*, *142*, w13659. PMID:22903797

Browne, S. E., Bowling, A. C., Baik, M. J., Gurney, M., Brown, R. H. Jr, & Beal, M. F. (1998). Metabolic dysfunction in familial, but not sporadic, amyotrophic lateral sclerosis. *Journal of Neurochemistry*, *71*(1), 281–287. doi:10.1046/j.1471-4159.1998.71010281.x PMID:9648876 Casley, C. S., Canevari, L., Land, J. M., Clark, J. B., & Sharpe, M. A. (2002). βAmyloid inhibits integrated mitochondrial respiration and key enzyme activities. *Journal of Neurochemistry*, 80(1), 91–100. doi:10.1046/j.0022-3042.2001.00681.x PMID:11796747

Chandra, S., Gallardo, G., Fernández-Chacón, R., Schlüter, O. M., & Südhof, T. C. (2005). α -synuclein cooperates with CSP α in preventing neurodegeneration. *Cell*, 123(3), 383–396. doi:10.1016/j. cell.2005.09.028 PMID:16269331

Chandrasekaran, K., Hatanpää, K., Brady, D. R., & Rapoport, S. I. (1996). Evidence for physiological down-regulation of brain oxidative phosphorylation in Alzheimer's disease. *Experimental Neurology*, *142*(1), 80–88. doi:10.1006/exnr.1996.0180 PMID:8912900

Choonara, Y. E., Pillay, V., Du Toit, L. C., Modi, G., Naidoo, D., Ndesendo, V. M., & Sibambo, S. R. (2009). Trends in the molecular pathogenesis and clinical therapeutics of common neurodegenerative disorders. *International Journal of Molecular Sciences*, *10*(6), 2510–2557. doi:10.3390/ijms10062510 PMID:19582217

Cluskey, S., & Ramsden, D. B. (2001). Mechanisms of neurodegeneration in amyotrophic lateral sclerosis. *Molecular Pathology*, *54*(6), 386. PMID:11724913

Corral-Debrinski, M., Horton, T., Lott, M. T., Shoffner, J. M., McKee, A. C., Beal, M. F., ... Wallace, D. C. (1994). Marked changes in mitochondrial DNA deletion levels in Alzheimer brains. *Genomics*, 23(2), 471–476. doi:10.1006/geno.1994.1525 PMID:7835898

Coskun, P. E., Beal, M. F., & Wallace, D. C. (2004). Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proceedings of the National Academy of Sciences of the United States of America*, *101*(29), 10726–10731. doi:10.1073/pnas.0403649101 PMID:15247418

Crouch, P. J., Blake, R., Duce, J. A., Ciccotosto, G. D., Li, Q. X., Barnham, K. J., & Masters, C. L. (2005). Copper-dependent inhibition of human cytochrome c oxidase by a dimeric conformer of amyloid- β 1-42. *The Journal of Neuroscience*, 25(3), 672–679. doi:10.1523/JNEUROSCI.4276-04.2005 PMID:15659604

Curmi, P. A., Gavet, O., Charbaut, E., Ozon, S., Lachkar-Colmerauer, S., Manceau, V., ... Sobel, A. (1999). Stathmin and its phosphoprotein family: General properties, biochemical and functional interaction with tubulin. *Cell Structure and Function*, 24(5), 345–357. doi:10.1247/csf.24.345 PMID:15216892

Da Cruz, S., & Cleveland, D. W. (2011). Understanding the role of TDP-43 and FUS/TLS in ALS and beyond. *Current Opinion in Neurobiology*, 21(6), 904–919. doi:10.1016/j.conb.2011.05.029 PMID:21813273

Darios, F., Corti, O., Lücking, C. B., Hampe, C., Muriel, M. P., Abbas, N., & Brice, A. (2003). Parkin prevents mitochondrial swelling and cytochrome c release in mitochondria-dependent cell death. *Human Molecular Genetics*, *12*(5), 517–526. doi:10.1093/hmg/ddg044 PMID:12588799

Davis, R. E., & Williams, M. (2012). Mitochondrial function and dysfunction: An update. *The Journal of Pharmacology and Experimental Therapeutics*, *342*(3), 598–607. doi:10.1124/jpet.112.192104 PMID:22700430

De Vos, K. J., Mórotz, G. M., Stoica, R., Tudor, E. L., Lau, K. F., Ackerley, S., & Miller, C. C. (2011). VAPB interacts with the mitochondrial protein PTPIP51 to regulate calcium homeostasis. *Human Molecular Genetics*, *21*(6), 1299–1311. doi:10.1093/hmg/ddr559 PMID:22131369

Deas, E., Plun-Favreau, H., & Wood, N. W. (2009). PINK1 function in health and disease. *EMBO Molecular Medicine*, 1(3), 152–165. doi:10.1002/emmm.200900024 PMID:20049715

Devi, L., Prabhu, B. M., Galati, D. F., Avadhani, N. G., & Anandatheerthavarada, H. K. (2006). Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheimer's disease brain is associated with mitochondrial dysfunction. *The Journal of Neuroscience*, *26*(35), 9057–9068. doi:10.1523/JNEUROSCI.1469-06.2006 PMID:16943564

Doss-Pepe, E. W., Chen, L., & Madura, K. (2005). *α-synuclein* and *Parkin* contribute to the assembly of ubiquitin lysine 63-linked multiubiquitin chains. *The Journal of Biological Chemistry*, 280(17), 16619–16624. doi:10.1074/jbc.M413591200 PMID:15718234

Ekstrand, M. I., Terzioglu, M., Galter, D., Zhu, S., Hofstetter, C., Lindqvist, E., & Hoffer, B. (2007). Progressive Parkinsonism in mice with respiratory-chain-deficient dopamine neurons. *Proceedings of the National Academy of Sciences of the United States of America*, *104*(4), 1325–1330. doi:10.1073/pnas.0605208103 PMID:17227870

Emerit, J., Edeas, M., & Bricaire, F. (2004). Neurodegenerative diseases and oxidative stress. *Biomedicine and Pharmacotherapy*, *58*(1), 39–46. doi:10.1016/j.biopha.2003.11.004 PMID:14739060

English, A. R., & Voeltz, G. K. (2013). Endoplasmic reticulum structure and interconnections with other organelles. *Cold Spring Harbor Perspectives in Biology*, *5*(4), a013227. doi:10.1101/cshperspect. a013227 PMID:23545422

Exner, N., Treske, B., Paquet, D., Holmström, K., Schiesling, C., Gispert, S., & Krüger, R. (2007). Loss-of-function of human PINK1 results in mitochondrial pathology and can be rescued by Parkin. *The Journal of Neuroscience*, *27*(45), 12413–12418. doi:10.1523/JNEUROSCI.0719-07.2007 PMID:17989306

Fan, J., Hu, Z., Zeng, L., Lu, W., Tang, X., Zhang, J., & Li, T. (2008). Golgi apparatus and neurodegenerative diseases. *International Journal of Developmental Neuroscience*, *26*(6), 523–534. doi:10.1016/j. ijdevneu.2008.05.006 PMID:18599251

Farg, M. A., Soo, K. Y., Warraich, S. T., Sundaramoorthy, V., Blair, I. P., & Atkin, J. D. (2012). Ataxin-2 interacts with FUS and intermediate-length polyglutamine expansions enhance FUS-related pathology in amyotrophic lateral sclerosis. *Human Molecular Genetics*, 22(4), 717–728. doi:10.1093/hmg/dds479 PMID:23172909

Fodero, L. R., Mok, S. S., Losic, D., Martin, L. L., Aguilar, M. I., Barrow, C. J., & Small, D. H. (2004). α 7-Nicotinic acetylcholine receptors mediate an A β 1–42-induced increase in the level of acetylcholinesterase in primary cortical neurones. *Journal of Neurochemistry*, 88(5), 1186–1193. doi:10.1046/j.1471-4159.2003.02296.x PMID:15009674

Fujita, Y., Mizuno, Y., Takatama, M., & Okamoto, K. (2008). Anterior horn cells with abnormal TDP-43 immuno reactivities show fragmentation of the Golgi apparatus in ALS. *Journal of the Neurological Sciences*, *269*(1), 30–34. doi:10.1016/j.jns.2007.12.016 PMID:18206910

Fujita, Y., Okamoto, K., Sakurai, A., Amari, M., Nakazato, Y., & Gonatas, N. K. (1999). Fragmentation of the Golgi apparatus of Betz cells in patients with amyotrophic lateral sclerosis. *Journal of the Neurological Sciences*, *163*(1), 81–85. doi:10.1016/S0022-510X(99)00014-3 PMID:10223416

Fujita, Y., Okamoto, K., Sakurai, A., Gonatas, N. K., & Hirano, A. (2000). Fragmentation of the Golgi apparatus of the anterior horn cells in patients with familial amyotrophic lateral sclerosis with SOD1 mutations and posterior column involvement. *Journal of the Neurological Sciences*, *174*(2), 137–140. doi:10.1016/S0022-510X(00)00265-3 PMID:10727699

Gandhi, P. N., Chen, S. G., & Wilson Delfosse, A. L. (2009). Leucine rich repeat kinase 2 (LRRK2): A key player in the pathogenesis of Parkinson's disease. *Journal of Neuroscience Research*, 87(6), 1283–1295. doi:10.1002/jnr.21949 PMID:19025767

Giri, M., Zhang, M., & Lü, Y. (2016). Genes associated with Alzheimer's disease: An overview and current status. *Clinical Interventions in Aging*, *11*, 665. doi:10.2147/CIA.S105769 PMID:27274215

Gonatas, J. O., Mezitis, S. G., Stieber, A., Fleischer, B., & Gonatas, N. K. (1989). MG-160. A novel sialoglycoprotein of the medial cisternae of the Golgi apparatus. *The Journal of Biological Chemistry*, 264(1), 646–653. PMID:2909545

Gonatas, N. K., Stieber, A., Mourelatos, Z., Chen, Y., Gonatas, J. O., Appel, S. H., ... Hauw, J. J. (1992). Fragmentation of the Golgi apparatus of motor neurons in amyotrophic lateral sclerosis. *American Journal of Pathology*, *140*(3), 731. PMID:1546747

Greggio, E., Jain, S., Kingsbury, A., Bandopadhyay, R., Lewis, P., Kaganovich, A., & Ahmad, R. (2006). Kinase activity is required for the toxic effects of mutant LRRK2/dardarin. *Neurobiology of Disease*, 23(2), 329–341. doi:10.1016/j.nbd.2006.04.001 PMID:16750377

Griess, B., Tom, E., Domann, F., & Teoh-Fitzgerald, M. (2017). Extracellular superoxide dismutase and its role in cancer. *Free Radical Biology & Medicine*, *112*, 464–479. doi:10.1016/j.freeradbiomed.2017.08.013 PMID:28842347

Gros-Louis, F., Gaspar, C., & Rouleau, G. A. (2006). Genetics of familial and sporadic amyotrophic lateral sclerosis. *Biochimica et Biophysica Acta (BBA)- Molecular Basis of Disease*, *1762*(11), 956–972. doi:10.1016/j.bbadis.2006.01.004

Guo, C., Sun, L., Chen, X., & Zhang, D. (2013). Oxidative stress, mitochondrial damage and neurodegenerative diseases. *Neural Regeneration Research*, 8(21), 2003. PMID:25206509

Hardy, J. A., & Higgins, G. A. (1992). Alzheimer's disease: The amyloid cascade hypothesis. *Science*, 256(5054), 184–185. doi:10.1126cience.1566067 PMID:1566067

Hroudová, J., Singh, N., & Fišar, Z. (2014). Mitochondrial dysfunctions in neurodegenerative diseases: Relevance to Alzheimer's disease. *BioMed Research International*, 2014, 1–9. doi:10.1155/2014/175062 PMID:24900954

Iqbal, K., Liu, F., Gong, C. X., & Grundke-Iqbal, I. (2010). Tau in Alzheimer disease and related tauopathies. *Current Alzheimer Research*, 7(8), 656–664. doi:10.2174/156720510793611592 PMID:20678074

Jackson, M. J., Vasilaki, A., & McArdle, A. (2016). Cellular mechanisms underlying oxidative stress in human exercise. *Free Radical Biology & Medicine*, *98*(13-17). PMID:26912036

Jastroch, M., Divakaruni, A. S., Mookerjee, S., Treberg, J. R., & Brand, M. D. (2010). Mitochondrial proton and electron leaks. *Essays in Biochemistry*, *47*, 53–67. doi:10.1042/bse0470053 PMID:20533900

Jin, L. W., Masliah, E., Iimoto, D., Deteresa, R., Mallory, M., Sundsmo, M., ... Saitoh, T. (1996). Neurofibrillary tangle-associated alteration of stathmin in Alzheimer's disease. *Neurobiology of Aging*, *17*(3), 331–341. doi:10.1016/0197-4580(96)00021-8 PMID:8725893

Johnston, J. A., Dalton, M. J., Gurney, M. E., & Kopito, R. R. (2000). Formation of high molecular weight complexes of mutant Cu, Zn-superoxide dismutase in a mouse model for familial amyotrophic lateral sclerosis. *Proceedings of the National Academy of Sciences of the United States of America*, 97(23), 12571–12576. doi:10.1073/pnas.220417997 PMID:11050163

Johri, A., & Beal, M. F. (2012). Mitochondrial dysfunction in neurodegenerative diseases. *The Journal of Pharmacology and Experimental Therapeutics*, *342*(3), 619–630. doi:10.1124/jpet.112.192138 PMID:22700435

Joshi, G., & Bekier, M. E., & II, Y. W. (2015). Golgi fragmentation in Alzheimer's disease. *Frontiers in Neuroscience*, 24(9), 340. PMID:26441511

Junn, E., Jang, W. H., Zhao, X., Jeong, B. S., & Mouradian, M. M. (2009). Mitochondrial localization of DJ-1 leads to enhanced neuroprotection. *Journal of Neuroscience Research*, 87(1), 123–129. doi:10.1002/jnr.21831 PMID:18711745

Karbowski, M., & Youle, R. J. (2003). Dynamics of mitochondrial morphology in healthy cells and during apoptosis. *Cell Death and Differentiation*, *10*(8), 870–880. doi:10.1038j.cdd.4401260 PMID:12867994

Karecla, P. I., & Kreis, T. E. (1992). Interaction of membranes of the Golgi complex with microtubules *in vitro*. *European Journal of Cell Biology*, *57*(2), 139–146. PMID:1387362

Kaur, S. J., McKeown, S. R., & Rashid, S. (2016). Mutant SOD1 mediated pathogenesis of amyotrophic lateral sclerosis. *Gene*, *577*(2), 109–118. doi:10.1016/j.gene.2015.11.049 PMID:26657039

Khan, S. M., Cassarino, D. S., Abramova, N. N., Keeney, P. M., Borland, M. K., Trimmer, P. A., ... Westaway, D. (2005). Hypersensitivity of DJ-1-deficient mice to 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyrindine (MPTP) and oxidative stress. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(14), 5215–5220. doi:10.1073/pnas.0501282102 PMID:15784737

Kinumi, T., Kimata, J., Taira, T., Ariga, H., & Niki, E. (2004). Cysteine-106 of DJ-1 is the most sensitive cysteine residue to hydrogen peroxide-mediated oxidation in vivo in human umbilical vein endothelial cells. *Biochemical and Biophysical Research Communications*, *317*(3), 722–728. doi:10.1016/j. bbrc.2004.03.110 PMID:15081400

Kitada, T., Asakawa, S., Hattori, N., Matsumine, H., Yamamura, Y., Minoshima, S., ... Shimizu, N. (1998). Mutations in the *Parkin* gene cause autosomal recessive juvenile Parkinsonism. *Nature*, *392*(6676), 605–608. doi:10.1038/33416 PMID:9560156

Kumar, A., & Singh, A. (2015). A review on mitochondrial restorative mechanism of antioxidants in Alzheimer's disease and other neurological conditions. *Frontiers in Pharmacology*, 6. PMID:26441662

Kwiatkowski, T. J., Bosco, D. A., Leclerc, A. L., Tamrazian, E., Vanderburg, C. R., Russ, C., & Valdmanis, P. (2009). Mutations in the FUS/TLS gene on chromosome 16 cause familial amyotrophic lateral sclerosis. *Science*, *323*(5918), 1205–1208. doi:10.1126cience.1166066 PMID:19251627

Larsson, N. G., Wang, J., Wilhelmsson, H., Oldfors, A., Rustin, P., Lewandoski, M., & Clayton, D. A. (1998). Mitochondrial transcription factor A is necessary for mtDNA maintenance and embryogenesis in mice. *Nature Genetics*, *18*(3), 231–236. doi:10.1038/ng0398-231 PMID:9500544

Lee, S. J., Kim, S. J., Kim, I. K., Ko, J., Jeong, C. S., Kim, G. H., & Cha, S. S. (2003). Crystal structures of human DJ-1 and Escherichia coli Hsp31, which share an evolutionarily conserved domain. *The Journal of Biological Chemistry*, 278(45), 44552–44559. doi:10.1074/jbc.M304517200 PMID:12939276

Lee, V. M., Giasson, B. I., & Trojanowski, J. Q. (2004). More than just two peas in a pod: Common amyloidogenic properties of tau and α -synuclein in neurodegenerative diseases. *Trends in Neurosciences*, 27(3), 129–134. doi:10.1016/j.tins.2004.01.007 PMID:15036877

Lesage, S., & Brice, A. (2009). Parkinson's disease: From monogenic forms to genetic susceptibility factors. *Human Molecular Genetics*, *18*(1), 48–59. doi:10.1093/hmg/ddp012 PMID:19297401

Lev, N., Ickowicz, D., Melamed, E., & Offen, D. (2008). Oxidative insults induce DJ-1 upregulation and redistribution: Implications for neuroprotection. *Neurotoxicology*, *29*(3), 397–405. doi:10.1016/j. neuro.2008.01.007 PMID:18377993

Li, Y., Tomiyama, H., Sato, K., Hatano, Y., Yoshino, H., Atsumi, M., & Toda, T. (2005). Clinicogenetic study of PINK1 mutations in autosomal recessive early-onset Parkinsonism. *Neurology*, *64*(11), 1955–1957. doi:10.1212/01.WNL.0000164009.36740.4E PMID:15955953

Lin, M. T., & Beal, M. F. (2006). Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature*, 443(7113), 787–795. doi:10.1038/nature05292 PMID:17051205

Lockrow, J. P., Fortress, A. M., & Granholm, A. C. E. (2012). Age-related neurodegeneration and memory loss in Down syndrome. *Current Gerontology and Geriatrics Research*. PMID:22545043

Lu, B. (2009). Mitochondrial dynamics and neurodegeneration. *Current Neurology and Neuroscience Reports*, 9(3), 212–219. doi:10.100711910-009-0032-7 PMID:19348710

Lustbader, J. W., Cirilli, M., Lin, C., Xu, H. W., Takuma, K., Wang, N., & Trinchese, F. (2004). ABAD directly links AB to mitochondrial toxicity in Alzheimer's disease. *Science*, *304*(5669), 448–452. doi:10.1126cience.1091230 PMID:15087549

Malkus, K. A., Tsika, E., & Ischiropoulos, H. (2009). Oxidative modifications, mitochondrial dysfunction, and impaired protein degradation in Parkinson's disease: How neurons are lost in the Bermuda triangle. *Molecular Neurodegeneration*, *4*(1), 24. doi:10.1186/1750-1326-4-24 PMID:19500376

Manczak, M., Jung, Y., Park, B. S., Partovi, D., & Reddy, P. H. (2005). Time course of mitochondrial gene expressions in mice brains: Implications for mitochondrial dysfunction, oxidative damage, and cytochromecin aging. *Journal of Neurochemistry*, *92*(3), 494–504. doi:10.1111/j.1471-4159.2004.02884.x PMID:15659220

Manczak, M., Park, B. S., Jung, Y., & Reddy, P. H. (2004). Differential expression of oxidative phosphorylation genes in patients with Alzheimer's disease. *Neuromolecular Medicine*, *5*(2), 147–162. doi:10.1385/NMM:5:2:147 PMID:15075441

Martin, L. J., Pan, Y., Price, A. C., Sterling, W., Copeland, N. G., Jenkins, N. A., ... Lee, M. K. (2006). Parkinson's disease α -synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death. *The Journal of Neuroscience*, 26(1), 41–50. doi:10.1523/JNEUROSCI.4308-05.2006 PMID:16399671

Maurer, I., Zierz, S., & Möller, H. J. (2000). A selective defect of cytochrome c oxidase is present in brain of Alzheimer disease patients. *Neurobiology of Aging*, 21(3), 455–462. doi:10.1016/S0197-4580(00)00112-3 PMID:10858595

Mills, R. D., Sim, C. H., Mok, S. S., Mulhern, T. D., Culvenor, J. G., & Cheng, H. C. (2008). Biochemical aspects of the neuroprotective mechanism of PTEN induced kinase 1 (PINK1). *Journal of Neurochemistry*, *105*(1), 18–33. doi:10.1111/j.1471-4159.2008.05249.x PMID:18221368

Monte, De Ia., S. M., Lu, B. X., Sohn, Y. K., Etienne, D., Kraft, J., Ganju, N., & Wands, J. R. (2000). Aberrant expression of nitric oxide synthase III in Alzheimer's disease: relevance to cerebral vasculopathy and neurodegeneration. *Neurobiology of Aging*, *21*(2), 309-319.

Morán, M., Moreno-Lastres, D., Marín-Buera, L., Arenas, J., Martín, M. A., & Ugalde, C. (2012). Mitochondrial respiratory chain dysfunction: Implications in neurodegeneration. *Free Radical Biology & Medicine*, 53(3), 595–609. doi:10.1016/j.freeradbiomed.2012.05.009 PMID:22595027

Mounsey, R. B., & Teismann, P. (2011). Mitochondrial dysfunction in Parkinson's disease: Pathogenesis and neuroprotection. *Parkinson's Disease*. PMID:21234411

Mourelatos, Z., Adler, H., Hirano, A., Donnenfeld, H., Gonatas, J. O., & Gonatas, N. K. (1990). Fragmentation of the Golgi apparatus of motor neurons in amyotrophic lateral sclerosis revealed by organellespecific antibodies. *Proceedings of the National Academy of Sciences of the United States of America*, 87(11), 4393–4395. doi:10.1073/pnas.87.11.4393 PMID:2349244

Musatov, A., & Robinson, N. C. (2012). Susceptibility of mitochondrial electron-transport complexes to oxidative damage. Focus on cytochrome c oxidase. *Free Radical Research*, *46*(11), 1313–1326. doi: 10.3109/10715762.2012.717273 PMID:22856385

Nakano, Y., Hirayama, K., & Terao, K. (1987). Hepatic ultrastructural changes and liver dysfunction in amyotrophic lateral sclerosis. *Archives of Neurology*, *44*(1), 103–106. doi:10.1001/archneur.1987.00520130079022 PMID:3800708

Oliveira, J. (2010). Nature and cause of mitochondrial dysfunction in Huntington's disease: Focusing on huntingtin and the striatum. *Journal of Neurochemistry*, *114*(1), 1–12. PMID:20403078

Orr, A. L., Li, S., Wang, C. E., Li, H., Wang, J., Rong, J., & Li, X. J. (2008). N-terminal mutant huntingtin associates with mitochondria and impairs mitochondrial trafficking. *The Journal of Neuroscience*, 28(11), 2783–2792. doi:10.1523/JNEUROSCI.0106-08.2008 PMID:18337408

Pagliarini, D. J., & Rutter, J. (2013). Hallmarks of a new era in mitochondrial biochemistry. *Genes & Development*, 27(24), 2615–2627. doi:10.1101/gad.229724.113 PMID:24352419

Paisan-Ruiz, C., Lang, A. E., Kawarai, T., Sato, C., Salehi-Rad, S., Fisman, G. K., & Rogaeva, E. (2005). LRRK2 gene in Parkinson disease Mutation analysis and case control association study. *Neurology*, *65*(5), 696–700. doi:10.1212/01.WNL.0000167552.79769.b3 PMID:16157901

Paradies, G., Paradies, V., de Benedictis, V., Ruggiero, F. M., & Petrosillo, G. (2014). Functional role of cardiolipin in mitochondrial bioenergetics. *Biochimica et Biophysica Acta*, *1837*(4), 408–417. doi:10.1016/j.bbabio.2013.10.006 PMID:24183692

Park, J., Kim, S. Y., Cha, G. H., Lee, S. B., Kim, S., & Chung, J. (2005). Drosophila DJ-1 mutants show oxidative stress-sensitive locomotive dysfunction. *Gene*, *361*, 133–139. doi:10.1016/j.gene.2005.06.040 PMID:16203113

Paterna, J. C., Leng, A., Weber, E., Feldon, J., & Büeler, H. (2007). DJ-1 and Parkin modulate dopaminedependent behavior and inhibit MPTP-induced nigral dopamine neuron loss in mice. *Molecular Therapy*, *15*(4), 698–704. doi:10.1038j.mt.6300067 PMID:17299411

Petit, A., Kawarai, T., Paitel, E., Sanjo, N., Maj, M., Scheid, M., ... Wang, L. (2005). Wild-type PINK1 prevents basal and induced neuronal apoptosis, a protective effect abrogated by Parkinson disease-related mutations. *The Journal of Biological Chemistry*, *280*(40), 34025–34032. doi:10.1074/jbc.M505143200 PMID:16079129

Polymeropoulos, M. H., Lavedan, C., Leroy, E., Ide, S. E., Dehejia, A., Dutra, A., ... Stenroos, E. S. (1997). Mutation in the α -synuclein gene identified in families with Parkinson's disease. *Science*, 276(5321), 2045–2047. doi:10.1126cience.276.5321.2045 PMID:9197268

Rakhit, R., Cunningham, P., Furtos-Matei, A., Dahan, S., Qi, X. F., Crow, J. P., ... Chakrabartty, A. (2002). Oxidation-induced misfolding and aggregation of superoxide dismutase and its implications for amyotrophic lateral sclerosis. *The Journal of Biological Chemistry*, 277(49), 47551–47556. doi:10.1074/jbc.M207356200 PMID:12356748

Reddy, P. H. (2007). Mitochondrial dysfunction in aging and Alzheimer's disease: Strategies to protect neurons. *Antioxidants & Redox Signalling*, 9(10), 1647–1658. doi:10.1089/ars.2007.1754 PMID:17696767

Reddy, P. H. (2008). Mitochondrial medicine for aging and neurodegenerative diseases. *Neuromolecular Medicine*, *10*(4), 291–315. doi:10.100712017-008-8044-z PMID:18566920

Reddy, P. H., & Beal, M. F. (2005). Are mitochondria critical in the pathogenesis of Alzheimer's disease? *Brain Research. Brain Research Reviews*, 49(3), 618–632. doi:10.1016/j.brainresrev.2005.03.004 PMID:16269322

Reddy, P. H., & Beal, M. F. (2008). Amyloid beta, mitochondrial dysfunction and synaptic damage: Implications for cognitive decline in aging and Alzheimer's disease. *Trends in Molecular Medicine*, *14*(2), 45–53. doi:10.1016/j.molmed.2007.12.002 PMID:18218341

Redeker, V., Lachkar, S., Siavoshian, S., Charbaut, E., Rossier, J., Sobel, A., & Curmi, P. A. (2000). Probing the Native Structure of Stathmin and Its Interaction Domains with Tubulin Combined Use of Limited Proteolysis, Size Exclusion Chromatography, and Mass Spectrometry. *The Journal of Biological Chemistry*, 275(10), 6841–6849. doi:10.1074/jbc.275.10.6841 PMID:10702243

Rendón, W. O., Martínez-Alonso, E., Tomás, M., Martínez-Martínez, N., & Martínez-Menárguez, J. A. (2013). Golgi fragmentation is Rab and SNARE dependent in cellular models of Parkinson's disease. *Histochemistry and Cell Biology*, *139*(5), 671–684. doi:10.100700418-012-1059-4 PMID:23212845

Rothstein, J. D. (2009). Current hypotheses for the underlying biology of amyotrophic lateral sclerosis. *Annals of Neurology*, *65*(1). PMID:19191304

Sakurai, A., Okamoto, K., Fujita, Y., Nakazato, Y., Wakabayashi, K., Takahashi, H., & Gonatas, N. K. (2000). Fragmentation of the Golgi apparatus of the ballooned neurons in patients with corticobasal degeneration and Creutzfeldt-Jakob disease. *Acta Neuropathologica*, *100*(3), 270–274. doi:10.1007004010000182 PMID:10965796

Sakurai, A., Okamoto, K., Yaguchi, M., Fujita, Y., Mizuno, Y., Nakazato, Y., & Gonatas, N. K. (2002). Pathology of the inferior olivary nucleus in patients with multiple system atrophy. *Acta Neuropathologica*, *103*(6), 550–554. doi:10.100700401-001-0500-x PMID:12012086

Sas, K., Robotka, H., Toldi, J., & Vécsei, L. (2007). Mitochondria, metabolic disturbances, oxidative stress and the kynurenine system, with focus on neurodegenerative disorders. *Journal of the Neurological Sciences*, 257(1), 221–239. doi:10.1016/j.jns.2007.01.033 PMID:17462670

Schapira, A. H. V. (2008). Mitochondrial dysfunction in neurodegenerative diseases. *Neurochemical Research*, *33*(12), 2502–2509. doi:10.100711064-008-9855-x PMID:18998208

Schapira, A. H. V., Cooper, J. M., Dexter, D., Clark, J. B., Jenner, P., & Marsden, C. D. (1990). Mitochondrial complex I deficiency in Parkinson's disease. *Journal of Neurochemistry*, *54*(3), 823–827. doi:10.1111/j.1471-4159.1990.tb02325.x PMID:2154550

Scheuner, D., Eckman, C., Jensen, M., Song, X., Citron, M., Suzuki, N., ... Larson, E. (1996). Secreted amyloid β -protein similar to that in the senile plaques of Alzheimer's disease is increased *in vivo* by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nature Medicine*, 2(8), 864–870. doi:10.1038/nm0896-864 PMID:8705854

Shibata, N. (2001). Transgenic mouse model for familial amyotrophic lateral sclerosis with superoxide dismutase 1 mutation. *Neuropathology*, 21(1), 82–92. doi:10.1046/j.1440-1789.2001.00361.x PMID:11304046

Silhavy, T. J., Kahne, D., & Walker, S. (2010). The bacterial cell envelope. *Cold Spring Harbor Perspectives in Biology*, *2*(5), a000414. doi:10.1101/cshperspect.a000414 PMID:20452953

Smith, W. W., Pei, Z., Jiang, H., Dawson, V. L., Dawson, T. M., & Ross, C. A. (2006). Kinase activity of mutant LRRK2 mediates neuronal toxicity. *Nature Neuroscience*, *9*(10), 1231–1233. doi:10.1038/nn1776 PMID:16980962

Spillantini, M. G., Schmidt, M. L., Lee, V. M. Y., Trojanowski, J. Q., Jakes, R., & Goedert, M. (1997). [alpha]-Synuclein in Lewy bodies. *Nature*, *388*(6645), 839–840. doi:10.1038/42166 PMID:9278044

Stamer, K., Vogel, R., Thies, E., Mandelkow, E., & Mandelkow, E. M. (2002). Tau blocks traffic of organelles, neurofilaments, and APP vesicles in neurons and enhances oxidative stress. *The Journal of Cell Biology*, *156*(6), 1051–1063. doi:10.1083/jcb.200108057 PMID:11901170

Strey, C. W., Spellman, D., Stieber, A., Gonatas, J. O., Wang, X., Lambris, J. D., & Gonatas, N. K. (2004). Dysregulation of stathmin, a microtubule-destabilizing protein, and up-regulation of Hsp25, Hsp27, and the antioxidant peroxiredoxin 6 in a mouse model of familial amyotrophic lateral sclerosis. *American Journal of Pathology*, *165*(5), 1701–1718. doi:10.1016/S0002-9440(10)63426-8 PMID:15509539

Su, B., Wang, X., Zheng, L., Perry, G., Smith, M. A., & Zhu, X. (2010). Abnormal mitochondrial dynamics and neurodegenerative diseases. *Biochimica et Biophysica Acta (BBA)-. Molecular Basis of Disease*, *1802*(1), 135–142. doi:10.1016/j.bbadis.2009.09.013 PMID:19799998

Sundaramoorthy, V., Walker, A. K., Tan, V., Fifita, J. A., Mccann, E. P., Williams, K. L., ... Atkin, J. D. (2015). Defects in optineurin-and myosin VI-mediated cellular trafficking in amyotrophic lateral sclerosis. *Human Molecular Genetics*, *24*(13), 3830–3846. doi:10.1093/hmg/ddv126 PMID:25859013

Swerdlow, R. H., & Khan, S. M. (2004). A "mitochondrial cascade hypothesis" for sporadic Alzheimer's disease. *Medical Hypotheses*, *63*(1), 8–20. doi:10.1016/j.mehy.2003.12.045 PMID:15193340

Tabrizi, S. J., Cleeter, M. W. J., Xuereb, J., Taanman, J. W., Cooper, J. M., & Schapira, A. H. V. (1999). Biochemical abnormalities and excitotoxicity in Huntington's disease brain. *Annals of Neurology*, *45*(1), 25–32. doi: PMID:9894873

Takamine, K., Okamoto, K., Fujita, Y., Sakurai, A., Takatama, M., & Gonatas, N. K. (2000). The involvement of the neuronal Golgi apparatus and trans-Golgi network in the human olivary hypertrophy. *Journal of the Neurological Sciences*, *182*(1), 45–50. doi:10.1016/S0022-510X(00)00447-0 PMID:11102638

Tan, W., Pasinelli, P., & Trotti, D. (2014). Role of mitochondria in mutant SOD1 linked amyotrophic lateral sclerosis. *Biochimica et Biophysica Acta* (*BBA*)-. *Molecular Basis of Disease*, *1842*(8), 1295–1301. doi:10.1016/j.bbadis.2014.02.009 PMID:24568860

Turner, B. J., & Atkin, J. D. (2006). ER stress and UPR in familial amyotrophic lateral sclerosis. *Current Molecular Medicine*, *6*(1), 79–86. doi:10.2174/156652406775574550 PMID:16472115

Valla, J., Schneider, L., Niedzielko, T., Coon, K. D., Caselli, R., Sabbagh, M. N., ... Reiman, E. M. (2006). Impaired platelet mitochondrial activity in Alzheimer's disease and mild cognitive impairment. *Mitochondrion*, *6*(6), 323–330. doi:10.1016/j.mito.2006.10.004 PMID:17123871

Vance, C., Rogelj, B., Hortobágyi, T., De Vos, K. J., Nishimura, A. L., Sreedharan, J., & Ganesalingam, J. (2009). Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science*, *323*(5918), 1208–1211. doi:10.1126cience.1165942 PMID:19251628

Vonsattel, J. P. G., & DiFiglia, M. (1998). Huntington disease. *Journal of Neuropathology and Experi*mental Neurology, 57(5), 369. doi:10.1097/00005072-199805000-00001 PMID:9596408

Vyas, S., Zaganjor, E., & Haigis, M. C. (2016). Mitochondria and cancer. *Cell*, *166*(3), 555–566. doi:10.1016/j.cell.2016.07.002 PMID:27471965

Waak, J., Weber, S. S., Görner, K., Schall, C., Ichijo, H., Stehle, T., & Kahle, P. J. (2009). Oxidizable residues mediating protein stability and cytoprotective interaction of DJ-1 with apoptosis signal-regulating kinase 1. *The Journal of Biological Chemistry*, 284(21), 14245–14257. doi:10.1074/jbc.M806902200 PMID:19293155

Wang, T., & Hay, J. C. (2015). Alpha-synuclein toxicity in the early secretory pathway: How it drives neurodegeneration in Parkinsons disease. *Frontiers in Neuroscience*, 9. PMID:26617485

Wang, W. X., Rajeev, B. W., Stromberg, A. J., Ren, N., Tang, G., Huang, Q., ... Nelson, P. T. (2008). The expression of microRNA miR-107 decreases early in Alzheimer's disease and may accelerate disease progression through regulation of β -site amyloid precursor protein-cleaving enzyme 1. *The Journal of Neuroscience*, 28(5), 1213–1223. doi:10.1523/JNEUROSCI.5065-07.2008 PMID:18234899

Waris, G., & Ahsan, H. (2006). Reactive oxygen species: Role in the development of cancer and various chronic conditions. *Journal of Carcinogenesis*. 5. doi:10.1186/1477-3163-5-14

Wiedemann, F. R., Manfredi, G., Mawrin, C., Beal, M. F., & Schon, E. A. (2002). Mitochondrial DNA and respiratory chain function in spinal cords of ALS patients. *Journal of Neurochemistry*, *80*(4), 616–625. doi:10.1046/j.0022-3042.2001.00731.x PMID:11841569

Winklhofer, K. F. (2007). The *Parkin* protein as a therapeutic target in Parkinson's disease. *Expert Opinion on Therapeutic Targets*, *11*(12), 1543–1552. doi:10.1517/14728222.11.12.1543 PMID:18020977

Wood-Kaczmar, A., Gandhi, S., Yao, Z., Abramov, A. S., Miljan, E. A., Keen, G., ... Downward, J. (2008). PINK1 is necessary for long term survival and mitochondrial function in human dopaminergic neurons. *PLoS One*, *3*(6), 2455. doi:10.1371/journal.pone.0002455 PMID:18560593

Yin, H., & Zhu, M. (2012). Free radical oxidation of cardiolipin: Chemical mechanisms, detection and implication in apoptosis, mitochondrial dysfunction and human diseases. *Free Radical Research*, *46*(8), 959–974. doi:10.3109/10715762.2012.676642 PMID:22468920

Yokota, T., Sugawara, K., Ito, K., Takahashi, R., Ariga, H., & Mizusawa, H. (2003). Down regulation of DJ-1 enhances cell death by oxidative stress, ER stress, and proteasome inhibition. *Biochemical and Biophysical Research Communications*, *312*(4), 1342–1348. doi:10.1016/j.bbrc.2003.11.056 PMID:14652021

Zarei, S., Carr, K., Reiley, L., Diaz, K., Guerra, O., Altamirano, P. F., ... Chinea, A. (2015). A comprehensive review of amyotrophic lateral sclerosis. *Surgical Neurology International*, 6. PMID:26629397

Zhang, B., Tu, P. H., Abtahian, F., Trojanowski, J. Q., & Lee, V. M. Y. (1997). Neurofilaments and orthograde transport are reduced in ventral root axons of transgenic mice that express human SOD1 with a G93A mutation. *The Journal of Cell Biology*, *139*(5), 1307–1315. doi:10.1083/jcb.139.5.1307 PMID:9382875

Zhu, X., Perry, G., Moreira, P. I., Aliev, G., Cash, A. D., Hirai, K., & Smith, M. A. (2006). Mitochondrial abnormalities and oxidative imbalance in Alzheimer disease. *Journal of Alzheimer's Disease*, 9(2), 147–153. doi:10.3233/JAD-2006-9207 PMID:16873962

Chapter 6 Neurodegenerative Disorders Progression: From Synaptic Dysfunction to Transmission Failure

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ABSTRACT

Neurodegenerative disorders (NDs) are a diverse group of disorders characterized by selective and progressive loss of neural systems that cause dysfunction of the central nervous system (CNS). The examples of NDs include Alzheimer's disease (AD), amyotrophic lateral sclerosis (ALS), Parkinson's disease (PD), and Huntington's disease (HD). The aggregated proteins block or disrupt the normal proteosomal turnover, autophagy, and become abnormally modified with time, generating toxicity via pathways thereby resulting in neurodegeneration and neuron death. The chapter highlights the understanding in the areas of AD, PD, HD as illustrative of major research so as to define the key factors and events in the improvement of NDs. It defines the physiological functioning of neural transmission (presynaptic, postsynaptic activity) at neural signaling pathway, then the dynamics of neuronal dysfunctioning and its molecular mechanism. Further, it also discusses the progression from synaptic dysfunction to transmission failure followed by NDs.

DOI: 10.4018/978-1-5225-5282-6.ch006

INTRODUCTION

Neurological disorders affect millions of people worldwide which can vary from the nerve disease that causes Tourette's into the serious CNS diseases, impacting either the brain or spinal cord, leading to the neurological or psychiatric deficits. It was observed that up to 1 billion people i.e., almost one in six of the world's population, suffers from either one or the other forms of neurological ailments such as AD, PD, stroke, brain injury, epilepsy, multiple sclerosis, neuroinfections and migraine (*Nearly 1 in 6 of* world's population suffer from neurological disorders – UN report, 2007). Also, it was being estimated in United Nations report issued, that around 6.8 million people worldwide dye of these maladies each year. Hence, understanding the neuronal dysfunctioning and resolving its pathological complexity becomes all the way a necessity now. The common aspects of the mentioned disease pathogenesis can be simply summarized with regards to the downstream implications of uncontrollable protein oligomerization and aggregations from post mitotic cells. It was being reported that the polyglutamine protein aggregates in the neurons causes the cells to undergo a stress reaction (Lim & Yue, 2015). Also, a study from the Gladstone Institutes showed for the very first time that mitochondrial damages (brain's cellular power plants) can diminish the energy levels and causes neural dysfunctions in a model of disease (Institutes, 2015). The normal autophagy, proteosomal turnover are disrupted by the aggregated proteins, which modifies with time abnormally thereby, causing toxicity through various mechanisms thereby, leading to neurodegeneration and cellular death (Sweeney et al., 2017). The postulation is coherent with the key genetic similarity between these diseases - e.g., the familial forms are generally caused by autosomal dominant mutation that favours aggregation (in case of PrP, tau, SOD1 and asyn) or formation of disease aggregation prone proteins (in case of CAG and APP repeat sequences) (G. F. Hall, 2011).

The synergistic interaction amongst proteins [synuclein and tau; amyloid precursor protein (APP)/ amyloid β (A β) and prion protein (PrP); tau and PrP; α -synuclein and PrP] occurs both at interneuronal and cellular level that lead to interneuronal lesion and eventually, pathogenesis of disease (Jellinger, 2012). Amongst all the NDs, AD is the most frequent and clinically known dementia in elderly population. It was recorded that almost 43% of elderly population that are above 85 years are suffering from AD and another CNS associated disorder PD; affects around 1-3% of population over 60 in USA (Qiu, Kivipelto, & von Strauss, 2009). Hence, owing to the above stated facts, the prevention, financial and societal effect of diseases, determination of causes, and exploration of effective treatment has been a foremost emphasis of clinical and basic research globally. Consequently, cellular mitochondria are known to play a key role in age related NDs as they are the important regulators of cell death, a critical characteristic of neurodegeneration (Jellinger, 2010). However, genetic mutations in mitochondrial DNA and oxidative stress cause ageing, attributing massively in initiating the progression of NDs (Lagouge & Larsson, 2013). This chapter focuses and highlights the basic understanding of neuronal irregularities that leads to NDs and its further progression, as illustrative of major research so as to define the key factors and events in the improvement of NDs.

BACKGROUND

NDs affect the CNS causing the dysfunction of nervous system. These incurable and debilitating conditions are indicated by loss in activity of neurons and are linked with degeneration of affected structures of nervous system (Ghavami et al., 2014). An integral subset of NDs includes dementia associated with aging (Wood, Winslow, & Strasser, 2015). Neurodegeneration is continuous loss of structural and functional properties of neurons corresponds to some pathological conditions which including neurons death (Gorman, 2008). In manner, NDs signify a huge group of neurological disorders with varied clinical and pathological expressions disturbing particular neuronal subsets in detailed functional anatomic systems that progress in a persistent manner. Different NDs like AD, PD, HD, and AML are common in therapeutic research studies. The NDs are mostly characterized through the factors like genetic risk factors, certain age ranges, courses of progression, clinical symptoms, dysfunction and death of specific subsets of neurons, particular biochemical abnormalities, and presence of intracellular and extracellular protein (Kovacs, 2016). In the advance study, the beginning of NDs is instigated by the aggregation of proteins called proteinopathies

DYNAMICS OF NEURONAL DYSFUNCTIONING

With the increase in counts of NDs cases universally, the research and findings has improved over the years but clinical trials have generated unsatisfactory results. While, gradual loss in neuron is the characteristic of NDs, some neurodegeneration may indicate neurological dysfunction instead of neuronal loss (Nixon & Yang, 2012). The abnormal proteins activate atypical activation of neurons, cause modifications in receptors of neurotransmitters and signalling cascades thereby, leading to neural network disintegration, synaptic insufficiencies and finally, breakdown of neurological functions. It was revealed in various *in vivo* models; that many pathogenic pathways can be reversed or prevented by taking off the abnormal proteins or pharmacologically regulate activities of neurons (Palop, Chin, & Mucke, 2006). Also, the increased plasticity of neurons benefits the remaining neuronal circuits to counteract for broken or lost circuits and improve the neuronal function and performance of networks (Greenwood & Parasuraman, 2010).

As the gradual neurological decline in NDs is directly related to the loss of neurons but, this association is also debatable. Further, it is doubtful that the variations in neuronal number credits for reversible and rapid fluctuations in neurological functions (Andres, Ducray, Schlattner, Wallimann, & Widmer, 2008). These variations possibly indicate the multifaceted fluctuations in molecules, modifications in synapse, neuronal circuits, neuronal networks and interactions between different networks (Figure 1). Many research studies have shown that the transgenic mouse models express abnormal proteins related with PD, AD, HD, ALS acquired disease linked neurological impairments. Further, the removal of abnormal proteins in them can inverse the neurological deficits without causing the changes in neuronal number. Hence, indicating towards the fact that the neurological destruction is also linked with neuronal dysfunction instead of loss in neurons. The surviving neurons left at various phases of impairment, comprise of functional neuronal state in the affected area and compensate to the alternative neural network for the lost ones, so if a probable target for therapeutic intrusion can be directed towards exploring the strategies to overcome vascular insufficiencies, co morbidities and genetic aspects may prove to be a more targeted approach.

However, even in the absence of disease, the neural system is depicted by enormous "degeneracy"the capacity of structurally dissimilar elements to carry out the same function or produce the same result (Price & Friston, 2002). The nervous-system degeneracy can be helpful to elucidate reason for neuron in substantia nigra dies and NDs progresses quickly. The intricacy of compensatory mechanisms is exemplified by many alterations that happen in cortical-basal-ganglia-thalamocortical network at various

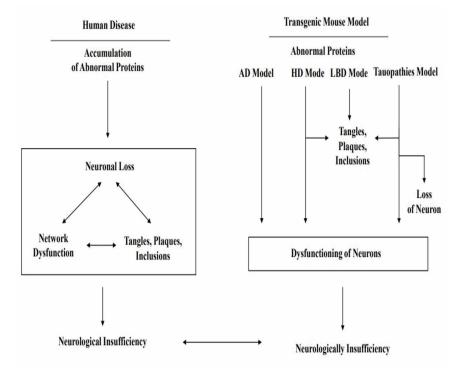


Figure 1. Schematic diagram presenting the neurodegenerative models (Alzheimer's, Huntington's, Lewy body and transporters model)

levels of PD and linked animal models. The gradual breakdown of the compensatory mechanisms may lead to the pronounced daily alterations in efficacy of drug L-DOPA (L-3,4 dihydroxyphenylalanine) which is frequently noticed in advanced PD (Rinne, Sonninen, & Siirtola, 1970). The processes that can disrupt or promote plasticity of neurons can be an effective therapeutic target.

NEURODEGENERATIVE DISORDERS MODELS

In NDs, the aggregation of proteins, neurological shortfalls and network dysfunction are linked with pathological characteristics (tangles, inclusion bodies and plaques) and loss of neurons (Takalo, Salminen, Soininen, Hiltunen, & Haapasalo, 2013). Transgenic mouse models are used to check specific hypothesis concerning the association between the features (Figure 1). The study underlined below supports model HD and Lewy body disease (LBD) model which includes dementia with Lewy bodies (DLB) and PD displaying that neurological insufficiency and neuronal dysfunction produced by congregation of abnormal proteins can be independent of tangles, plaques, loss of neurons and inclusions. It also elucidates that decreasing the production, increasing the removal or neutralizing the activity of the abnormal protein assembly efficiently reverses the neurological insufficiency (Palop et al., 2006).

Oxidative injury precedes deposition of $A\beta$ in transgenic APP mice (Praticò, Uryu, Leight, Trojanoswki, & Lee, 2001), with increase of enzymes pertaining to apoptosis and mitochondrial metabolism process occurring sooner and colocalizing with the neurons undertaking cognitive impairment (Reddy et al.,

2004). In addition, dysfunction of mitochondria and oxidative injury probably promote the pathogenesis of AD. In the neurons of fetal guinea pig, the treatment with hydrogen peroxide upregulated intracellular levels of A β (Ohyagi et al., 2000). Also, the treatment of astrocytes with mitochondrial carbonyl cyanide m-chlorophenylhydrazone (CCCP) promote cells to mimic the processing of amyloidogenic APP and accumulation of intracellular A β which is seen in the astrocytes suffering from Down syndrome (Busciglio et al., 2002). In a transgenic mouse which was APP mutant, deficiency of the anti-oxidant mitochondrial enzyme MnSOD significantly enhanced A β plaque (F. Li et al., 2004). In another study, a transgenic APP deficient mouse, inhibitors of energy metabolic procedure (kainic acid, insulin, 3-nitropropionic acid and 2-deoxyglucose) increased the activity of β -secretase levels and A β amounts. The knockout mice also displays hyperphosphorylation of tau, behavioural and motor deficiency, and neural degeneration (Liou, Sun, Ryo, & Zhou, 2003).

FACTORS CAUSING NEURONAL DYSFUNCTION

Oxidative Stress and Mitochondrial Dysfunction in Neurodegenerative Disorders

The evidences propose that mitochondria play a key role in age-related NDs and regulate the cell death, a characteristic aspect of neurodegeneration (Martin, 2010). Now, a mutation in the DNA of mitochondria and oxidative stress contributes towards the process of ageing, an utmost risk for NDs. There has been evidences that dysfunctioning of mitochondria occurs early and proceeds casually in the pathogenesis of NDs along with disease-specific proteins which interacts with the mitochondria (Table 1).

Mitochondria and Alzheimer's Disease

AD is symbolized by gradual cognitive degeneration and clinically by the existence of extracellular senile plaque (mostly composed of $A\beta$) and intracellular neurofibrillary tangles (NFTs, composed of hyperphosphorylated tau) (Murphy & LeVine, 2010). Approximately, 5-10% of the Alzheimer's cases are hereditary, happening at premature onset and autosomal-dominant way however, there are 3 proteins associated with AD specifically: amyloid precursor protein (APP that is consecutively broken by β - and γ -secretases to form A β), presentiins 1, and presentiins 2 (PS 1 and PS 2) (components of γ -secretase complex (O'Brien & Wong, 2011). There are considerable literature supporting a function for mitochondrial dysfunction and cognitive injury in the development of AD (Mancuso et al., 2009). Many pathways linking AD pathology and oxidative stress have recently been discovered (Bennett, Grant, & Aldred, 2009). Pathways that alter tau processing or amyloid precursor protein (APP) might be activated by stress. In a study, PIN1 (prolyl isomerase) was discovered to be very susceptible to cognitive impairment (Sultana et al., 2006). PIN1 helps in catalysing the conformational changes in the protein that affects both tau and processing of APP. Pin1 knockout intensifies amyloidogenic processing of APP and intracellular A β levels in mice (S. L. Ma, Pastorino, Zhou, & Lu, 2012). There are evidences which show that mtDNA (mitochondrial DNA) might be associated with dysfunctioning of mitochondria observed in AD (Mancuso, Orsucci, Siciliano, & Murri, 2008). When mtDNA of patient is moved to cell lines which are mtDNA deficient, the cybrid produces the respiratory enzyme deficiency observed from the mind along with some other cells in AD, indicating that the defect is conceded at least in part by abnormalities of

Type of Disease	Genetic Causes	Function	References
Alzheimer's disease	APP	Forms A β , main component of senile plaques.	(Murphy & LeVine, 2010)
	PS1, PS2	Component of γ -secretase, that cleaves APP to form A β	(Ya-Ping Tang & Elliot S. Gershon, 2003)
Parkinson's disease	SNCA	The principal constituent of Lewy body	(Stefanis, 2012)
	Parkin	Ubiquitin E3 ligase	(Dawson & Dawson, 2010)
	DJ-1	Protection of cell from oxidant-induced cell death.	(Lev, Roncevic, Ickowicz, Melamed, & Offen, 2006)
	PINK1	Protein kinase localized in mitochondria and protects the cell against death. Mutation in <i>PINK1</i> leads to building up of improper folded proteins in mitochondria of human cells ad fly.	(De Castro et al., 2012; Pickrell & Youle, 2015)
	LRRK2	A protein kinase. Mutation in <i>LRRK2</i> can cause cell death and PD by impairing the pathways responsible for protein degradation. <i>LRRK2</i> is essential for regulation of autophagy-lysosomal pathway.	(JQ. Li, Tan, & Yu, 2014; Tong et al., 2012)
	HTRA2	Serine protein kinase in the intermembrane space of mitochondria. HTRA2 helps in degradation of proteins within mitochondria. Helps in degrading the inhibitor of apoptosis proteins and stimulate cell death if released to cytosol.	(Strauss et al., 2005)
Amyotrophic lateral sclerosis	SOD1	Forms hydrogen peroxide from superoxide. Mutations in the enzyme lead to toxic functional gain.	(Bunton-Stasyshyn, Saccon, Fratta, & Fisher, 2015)
Huntington's disease	HTT	The protein interacts with the proteins of the brain. Thus, mutant protein is disruptive for the nerve cells and leads to expanded polyglutamine repeats.	(Zheng & Diamond, 2012)

Table 1. Specific proteins that involve with mitochondria in major neurodegenerative disorders.

mtDNA (Swerdlow et al., 1997). Nonetheless, recognizing AD particular mutations of mtDNA was a struggle. mtDNA sequencing from 128 controls and 145 AD patients did not uncover any considerable link with mutations of mtDNA and haplogroup of mitochondria (Elson et al., 2006). There was no link observed in the obtained mutations of mtDNA while examination of coding region for cytochrome c oxidase subunit I (CO1) (Lin, Simon, Ahn, Kim, & Beal, 2002). Nevertheless, in the similar manner, the promoters seemed more susceptible to injury than the coding region of genes 15, control region of mtDNA which displayed an enhanced mutation in AD (Coskun, Beal, & Wallace, 2004). The cortical areas suffering from AD had approximately 63% rise in heteroplasmic mtDNA control-region mutations, and in individuals there was a 130% increment in mutations (Coskun et al., 2004).

Mitochondria and Parkinson's Disease

PD is marked clinically by bradykinesia, tremor and progressive rigidity and biochemically by occurrence of Lewy body-loss of pigmented neurons in the substantia nigra and the presence of Lewy bodiescharacteristic inclusions in cytoplasm which immunostain for ubiquitin and α -synuclein (Lin & Beal, 2006). Mitochondria was first associated in PD as MPTP (1-methyl4-phenyl-1,2,3,6-tetrahydropyridine), whose metabolite MPP⁺ obstructs the complex I of the mitochondrial ETC (electron-transport chain), lead to parkinsonism in drug-designer abusers. The model is refined in lab animals, wherein rotenone's (Betarbet et al., 2000) (MPTP or inhibitor of Complex-I) chronic infusion-outcomes clinically at a parkinsonian phenotype and pathologically in degeneration of nigral with inclusions in cytoplasm for ubiquitin and α synuclein (Fornai et al., 2005).

The toxicity mechanism in the inhibition of complex-I models possibly includes stress (Sherer et al., 2003). Oxidative stress and complex-I inhibition have been presented to be pertinent to PD when glutathione deficiency and complex-I depletion were present in substantia nigra of patients suffering from pre-symptomatic PD and idiopathic PD (Schapira et al., 1990). To date, polymorphisms or mutations in mtDNA and 9 nuclear genes have been recognized as causing the risk of PD. These genes are: parkin, phosphatase and tensin homologue (*PTEN*)-induced kinase 1 (*PINK1*), α -synuclein, leucine-rich-repeat kinase 2 (LRRK2), DJ-1, ubiquitin carboxy-terminal hydrolase L1, the nuclear receptor HTRA2, NURR1 and tau. Of the nuclear genes: parkin, α -synuclein, DJ-1, HTRA2, PINK1 and LRRK2 directly or indirectly implicate mitochondria (Deas, Wood, & Plun-Favreau, 2011). In a number of cases, inherited mutations in mtDNA resulted in parkinsonism, generally as one characteristic of a larger syndrome. It was discovered that the Leber's atrophy G11778A mutation was associated with L-DOPA-responsive parkinsonism, variably co-occurring with ophthalmoplegia, ataxia, dementia and dystonia, and (D. Simon et al., 1999). Notably, this mutation is at a subunit of complex I. Mutations in the nuclear-encoded mtDNA polymerase-y (POLG) receptor impair mtDNA replication and result in multiple mtDNA deletions, typically causing chronic progressive external ophthalmoplegia and myopathy. POLG mutations also co-segregate with parkinsonism, myopathy and ophthalmoplegia (Luoma et al., 2004). There's less evidence for mtDNA participation in non-syndromic PD. Nigral neurons from PD patients contain increased levels of clonally expanded mtDNA deletions compared with those from controls, although elevated levels can also be seen in ageing (Bender et al., 2006; Kraytsberg et al., 2006) and discovered no distinction between PD and control subjects in inherited or acquired complex-I or tRNA point mutations (D. K. Simon et al., 2004; Vives-Bauza et al., 2002). Interestingly, however, several groups have discovered that particular continent-specific clusters of polymorphisms, termed mtDNA haplogroups, might decrease the probability of developing PD. Among Europeans, the haplogroup cluster UJKT is associated with a decreased risk for PD compared with haplogroup H (Pyle et al., 2005). The haplogroups underrepresented in PD patients are overrepresented in healthy centenarians (Tanaka, 2002). Protective mtDNA lineages seem to have arisen from areas requiring cold-adaptation, including uncoupling of mitochondria to increase heat generation at the expense of ATP production. It's been suggested that this partial uncoupling increases longevity and decreases risk of neurodegeneration by decreasing free-radical generation (Wallace, 2005). Mutations in α -synuclein are associated with autosomal dominant familial PD. α -Synuclein is a significant component of Lewy bodies, along with the primary effect of α -synuclein mutations is likely to be an increased formation of fibrillar or oligomeric aggregates (Recchia et al., 2004). Nevertheless, there seem to be close interrelationships between protein accumulation or degradation, oxidative stress and mitochondrial dysfunction. In transgenic mice, overexpression of-synuclein impairs mitochondrial function, increases oxidative stress and enhances nigral pathology induced by MPTP (Song, Shults, Sisk, Rockenstein, & Masliah, 2004). Furthermore, at a latest research study of mice overexpressing alpha A53T mutant α -synuclein mitochondria were immune stained for α -synuclein, raising the possibility that mutant α -synuclein may damage mitochondria directly (Martin et al., 2006). Whereas overexpression of α -synuclein increases sensitivity to MPTP, α -synuclein-devoid-mice are more immune to MPTP41 along with other mitochondrial toxins like malonate and 3-nitropropionic acid (Klivenyi et al., 2006). Therefore, α -synuclein seems to mediate the toxicity effects of MPTP. Mutations in parkin are associated with autosomal recessive juvenile PD. Parkin encodes ubiquitin E3 ligase, and also the primary abnormality, therefore, is in the ubiquitin proteasome system (UPS).

Mitochondria and Amyotrophic Lateral Sclerosis

ALS is marked clinically by spasticity, progressive weakness and atrophy of muscular tissue, indicating the degeneration of both lower and upper motor neurons in the brainstem, spinal cord and cortex. About 90% of cases are sporadic ALS (SALS) and 10% are familial ALS (FALS). Approximately, 20% of FALS cases are triggered by Cu/Zn-superoxide dismutase (SOD1) mutations (Rezania et al., 2003). In both SALC and FALC, biopsy and post mortem samples from the nerves, muscles and spinal cord present anomalies in structure of mitochondria, localization and number. Nonetheless, it's hard to analyse from glimpses of already symptomatic individuals if mitochondria contribute to progression of disease or are harmless bystanders. So, a study on the involvement of mitochondria in ALS has focussed on expression of mutant SOD1 in animal and cellular models of ALS. Overexpression of G93A SOD1 mutation induces impaired mitochondrial energy metabolic process in the spinal cord and brain at the onset of disease (Mattiazzi et al., 2002). Nonetheless, before the onset of disease, the calcium loading capacity is decreased in the mitochondria from spinal cord and brain, but, not in the liver of mice overexpressing G85R or G93A mutant SOD1 (Damiano et al., 2006). Back in G93A mice, there's an explosive upsurge in degeneration of vacuolar mitochondria thereby, leading to death of motor neuron, which suggests that abnormalities in mitochondrial trigger onset of the disease (Kong & Xu, 1998). Interestingly, SOD1 is concentrated within vacuolated mitochondria (Jaarsma et al., 2001). SOD1 has conventionally been believed to be a cytoplasmic protein, although localization of a portion of mobile SOD1 to the mitochondrial intermembrane space, matrix and outer membrane has now been confirmed (Higgins, Jung, & Xu, 2003; Vijayvergiya, Beal, Buck, & Manfredi, 2005) and only the affected cells localize SOD1 (Liu et al., 2004). Interaction between mitochondria and SOD1 suggests different mechanisms by which cell survival and mitochondrial function might be negatively influenced. The targeting of mutant SOD1 to mitochondria triggered apoptosis and release of cytochrome c, while targeting to nucleus or endoplasmic reticulum (ER) did not trigger cell death (Takeuchi, Kobayashi, Ishigaki, Doyu, & Sobue, 2002). It was suggested that mutant SOD1 collects and accumulates from the outer membrane of mitochondria and interrupts the importation of protein, thus, leading to dysfunctioning of mitochondria (Liu et al., 2004). Mutant SOD1 is suggested to encourage production of ROS, and cause oxidative damage to proteins and lipids of mitochondria, together with impaired synthesis of ATP and respiration in mice expressing mutant human SOD1 (Mattiazzi et al., 2002).

Mitochondria and Huntington's Disease

HD is characterized by psychiatric disturbances, dementia and chorea and biochemically by damage of long projection neurons, and anticipated to cause the development of cytosine, adenine and guanine (CAG) trinucleotide recurrence in the huntingtin (*HTT*) gene, that leads to polyglutamine stretch expansion from the consequent protein (Finkbeiner, 2011). The standard number of repeats of CAG repeats is less than 36 and greater than 40, which leads to occurrence of disease. Many evidences establish the association of dysfunctioning of mitochondrial in HD. NMR (Nuclear magnetic resonance spectroscopy) shows enhanced lactate from basal ganglia and cortex (Jenkins, Koroshetz, Beal, & Rosen, 1993). Biochemical researches reveal reduced performances of complexes II and III of the ETC (electron- transfer chain)

in the human HD brain (Gu et al., 1996). In striatal cells of mutant HTT-knock-in mouse embryos, the production of ATP and respiration in mitochondria were considerably damaged (Milakovic & Johnson, 2005). The dysfunctioning of mitochondria noticed previously is important pathogenically, as malonatemitochondrial toxins and 3-nitropropionic acid selectively restrains succinate dehydrogenase and complex II- stimulates a clinical and behavioural phenotype that looks like HD closely (Brouillet et al., 1995). Also, by adding 82 glutamines to 171 amino acids of *HTT*, restoration of complex-II activity, obstruction of cell death and dysfunctioning of mitochondria was observed in striatal neurons (Benchoua et al., 2006). Many mechanisms are known by which the mutation can lead to dysfunctioning of mitochondria. Firstly, there can be a direct interaction between *HTT* and mitochondria. In a research (Panov et al., 2002), mitochondria from lymphoblast of HD patients and mitochondria from brain of YAC transgenic mice expressing *HTT* with 72 repeats was discovered to have a membrane potential and had an ability to depolarize in lower calcium loads in comparison to mitochondria of control.

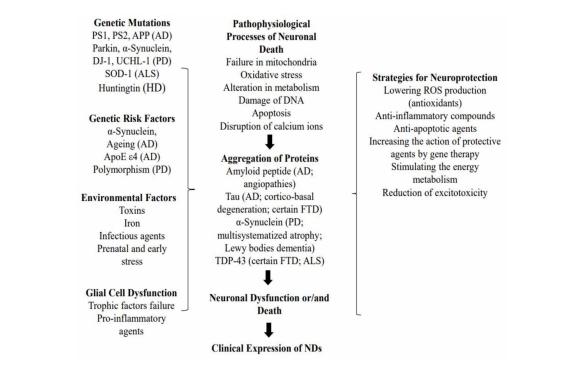
Other Causes for Neuronal Dysfunction

The factors causing the NDs still remain unidentified till now. For AD and possibly PD, the significant risk factor is ageing due to the increased prevalence of the disease with age. In case of AD the situation is apparent, however in the other case such an assessment is open to question since for a sizable proportion of patients the starting place of PD is quite premature, and disguised with a long prodromal stage of the disease. Certain recognizable kinds of the diseases find a genetic component but monogenic forms are extremely infrequent and even exceptional (Corti, Fournier, & Brice, 2009). It is the case in early-onset of AD where the amyloid hypothesis is supported by mutations of genes causing the disease, such as mutations of gene encoding amyloid protein precursor (chromosome 21) or presenilins (PS1 and PS2 found on chromosomes 1 and 14 (respectively) in rare autosomal dominant kinds (<5%) (Ya-Ping Tang & Elliot S Gershon, 2003). A comparable situation can be found, for instance, at autosomal dominant forms of PD, with the gene encoding LRRK2 (mutation PARK8) and α - synuclein (mutation PARK1 and PARK4) among 15 varied known mutations (Figure 2). In some families, where ALS and FTD co-segregate, mutations on a single gene associated to chromosome 19 can cause both clinical phenotypes, that affirms the hypothesis of a common mechanism, as discovered by new data on accumulation of proteins (Sleegers, Cruts, & Van Broeckhoven, 2010). While, it is contemplated that environmental factors are also involved in the development of a certain NDs (Cicchetti, Drouin-Ouellet, & Gross, 2009). For example, compounds like heavy metals, pesticides, fungicides, chemicals derived from viruses and addictive drugs may perhaps play a part in causing PD (Moisan et al., 2011). Nevertheless, epidemiologic research has to be further developed since present conclusions are usually not adequately apparent, therefore suggesting that only exposures to these environmental factors even when prolonged and intensive aren't usually sufficient to cause the diseases (Dick, 2006). Interestingly, in relation to environmental factors that could influence the occurrence of diseases, a brand new direction has recently been taken focusing on anxiety. Intensive chronic stress in humans was for instance associated with decreased hippocampal and right orbito-frontal volumes, (Gianaros et al., 2007) which might affect cognitive and emotional functions.

Abbreviations: PS1, Presenilin1; PS2, Presenilin2; APP, Amyloid precursor protein; UCHL-1, Ubiquitin carboxyl-terminal hydrolase isozyme L1; SOD-1, Superoxide dismutase 1; Apo E ε 4, Apolipoprotein E ε 4; FTD, Frontotemporal dementia; ROS, reactive oxygen species; TDP-43, TAR DNA-binding protein 43; NDs, Neurodegenerative disorders.

Neurodegenerative Disorders Progression

Figure 2. Summary of the processes that can be associated in neurodegenerative disorders, comprising of genetic factors and mutations that interfere with environmental factors to cause disease



PROTEIN AGGREGATION AND INTERNEURONAL LESION

NDs like AD, PD, HD, ALS and prion diseases are increasingly being realized to have common cellular and molecular mechanisms including protein aggregation and inclusion body formation (Lansbury & Lashuel, 2006). The aggregates usually consist of fibres containing misfolded protein with a β -sheet conformation, termed amyloid. The more than likely explanation is that inclusions along with other protein aggregates represent an end phase of a cascade of steps, and that earlier phases in the cascade can be more directly connected to pathogenesis than the inclusions themselves. For diseases, genetic variants help in describing the pathogenesis of the more common sporadic forms and developing mouse along with other models. There's now increased understanding of the pathways involved with protein aggregation, and a few latest hints have emerged as to the molecular mechanisms of toxicity. These are leading to strategies toward rational therapeutics.

Genetic and Neuropathological Features in the Pathogenesis of Neurodegenerative Disorders

The main focus of preliminary research into the pathogenesis of NDs is on the processes and its mechanisms accountable for protein aggregation as well as the description of cytotoxic changes that accompany and result from the aggregation (G. Hall, 2011). As a result, events which are associated to aggregation and downstream implications of aggregation like loss of protein turnover mechanism in neurons are among

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the best-characterized features of NDs. This work has created a broad consensus that aggregation leads to the breakdown of normal protein turnover mechanisms and the subsequent growth of abnormal toxicity routes of protein disposal are main pathogenic events of the NDs that affect the human CNS. Common toxicity elements which are downstream of protein aggregation in NDs consist of:

- Association of aggregation which harms the protein turnover mechanisms (De Baets et al., 2011),
- Dysfunctioning of mitochondria or/and maldistribution resulting in changes associated with apoptosis because of oxidative stress generation, low ATP and abnormal Ca²⁺ fluxes and (Cui, Kong, & Zhang, 2012)
- Sequestration of normal proteins because of aggregation thereby, leading to damage in normal functioning of sequestered proteins (Yang & Hu, 2016).

Alzheimer's Disease and Tauopathies

AD is a late-onset dementing illness, with progressive memory loss, task performance, speech, and recognition of people and objects. There's degeneration of neurons (particularly in the basal forebrain and hippocampus), however at least as necessary for pathogenesis could be synaptic pathology and modified neuronal connections (Hardy & Selkoe, 2002; Selkoe, 2002). AD involves two types of protein aggregates. Extracellular aggregates known as neuritic plaques have as their major constituent the A^β peptide that can be derived from proteolytic processing of the APP. The A β -containing aggregates have β -sheet structure along with Congo red (an azo dye) and reactivity characteristic of amyloid (Serpell & Smith, 2000). There are also intracellular aggregates of the microtubule-associated protein tau, called neurofibrillary tangles. The pathogenesis of AD has been greatly clarified by the identification of genetic mutations accountable for rare forms of the disease. These mutations have been in APP itself and also in the presenilins, that are involved with the cleavage of APP (Citron, 2002; Esler & Wolfe, 2001). Additionally, tauopathies like frontotemporal dementia with parkinsonism may be brought on by mutations in the tau protein (Goedert, 2004; Ingram & Spillantini, 2002). The characteristic example of a neuropathogenesis pattern which suggests the lesion spread in NDs (outside of prion diseases) is stipulated by AD. It is evident after the studies of Heiko and Eva Braak (Braak & Braak, 1991) that neurofibrillary degenerative changes of AD progress in a distinctive order which comprehends the development of symptoms (Armstrong, Cairns, & Lantos, 2001; Su, Deng, & Cotman, 1997). The initial alterations happen in particular limbic regions which are associated with, spatial localization, olfaction and formation and consolidation of episodic memory (entorhinal, pyriform and transentorhinal cortices), functions which are usually compromised in the earliest preclinical and clinical phases of AD.

It is followed by increasing involvement of paralimbic and limbic centers which includes the insula, anterior cingulate cortex, hippocampus and adjacent allocortical regions of the medial temporal lobe (e.g. subiculum). These neuropathological changes match the progression of AD symptoms with alterations in short term memory and emotional processing becoming obvious by the time AD can be recognized as such in the clinic, along with the beginning of cognitive changes. The most significantly affected limbic centers are intensely interrelated with one another functionally and synaptically as will be essential for propagation of lesion by transsynaptic toxicity transfer (Su et al., 1997).

The developing association of synaptically interconnected brain regions as observed in AD is reflected in non-AD pathophysiological conditions like Pick's disease (PiD), frontotemporal dementia (FTD), corticobasal degeneration (CBD) and progressive supranuclear palsy (PSP) implicates some areas of brain (insula, frontal/prefrontal cortex temporal cortex). Likewise, PD and LBD share a joint set of susceptible loci (olfactory bulb, dopaminergic neuron, neocortical loci) nevertheless, differ immensely in extent of involvement of other parts of brain and initial nidus of vulnerability (Aguzzi, Sigurdson, & Heikenwaelder, 2008; Clinton, 2007; Cookson, 2009). Additional characteristic of NDs as a group is the appearance of each disorder in both familial and sporadic forms, with intronic or exonic mutations in a particular aggregation-susceptible protein synthesises a dominant allele which is able to replicate all the attributes of the sporadic disease with high penetrance (Spillantini, Bird, & Ghetti, 1998). The interesting characteristics of this pattern are a) the extent of similarity between familial and sporadic disease forms, b) higher tendency of sporadic as compared to familial, disease forms to present the asymmetric development, particularly in non-AD conditions (Dickson, 2010; Lee, Goedert, & Trojanowski, 2001).

Linking Aggregation to Lesion Spreading-Prion Protein

The prion protein (PrP) is an example of protein which is prone to aggregation that spreads its misfolded state of protein at protein, cellular and organismal level. NDs brought on by prions may be sporadic or could be obtained either by genetic mutations or environmental transmission (Pruisner, 2001). Mutations in the prion gene cause the disease thereby, leading to changes in the prion protein. The pathology of the disease includes the formation of amyloid plaques that appears comparable to AD which can be labelled by prion antibody. Prion disease is caused by changes in protein conformation and the research has shown that it is brought out by construction of abnormal protein. The prion disease mechanism is lit from the finding of prion like protein conformation changes in yeast (Lindquist, Krobitsch, Li, & Sondheimer, 2001; Scheibel, Bloom, & Lindquist, 2004). In all cases, disease occurs by the presence of abnormally folded prion proteins. The aggregation of prion protein might take place extracellularly and intracellularly (J. Ma & Lindquist, 2002; J. Ma, Wollmann, & Lindquist, 2002). The misfolding of PrP facilitates a range of neurological degenerative diseases like transmissible spongiform encephalopathy in humans. The "Prion hypothesis" states that the individual subunit of PrP is misprocessed and misfolded in a way that makes it form a neurotoxic conformation (PrP^{sc}), and promotes the transmission of this conformation to other prion proteins with normal conformation (PrP^c). The controversial and peculiar history of prion biology offers us the example of how the misprocessed protein aggregation-prone protein can lead to interneuronal propagation with neurotoxic characteristics and thereby influence the spreading of neurofibrillary lesions to postsynaptic, adjacent and presynaptic neurons. The relevance of misprocessing mechanism of PrP to development of tauopathies, NDs and synucleinopathies is highlighted by various findings that massive differences in the misprocessing of PrP and PrP^{sc} structure appear to cause neuropathological and clinical manifestations of different prion diseases (Caughey & Baron, 2006; Collinge & Clarke, 2007; Parchi et al., 1996). Additionally, various studies of the physiological functions of PrP^C propose its involvement in function of cell adhesion, actin-rich subcortical cytoskeleton and its interaction with microtubules, signal transduction pathways (Cushman, Johnson, King, Gitler, & Shorter, 2010). PrP^c plays an integral role in synaptic plasticity. The similarities in the localization, misprocessing and cellular functions of asyn, tau, APP/Abeta and PrP recognize likely points of interactions between these proteins and synergy in the process of misprocessing.

PROGRESSION FROM SYNAPTIC DYSFUNCTION TO TRANSMISSION FAILURE

Neural plasticity is extremely adaptive during the disease and health. It allows the animals to interact efficiently with the environment and to manage better with neurological damages. A chief constituent of neural plasticity is located in synapse, which are actively weakened and strengthened by complicated methods to form the dynamic network and circuit which stores memory and generates thought process. Particularly, the abnormal proteins that are supposed to cause NDs hamper the function or integrity of postsynaptic specializations and presynaptic terminals (Mahley, Weisgraber, & Huang, 2006; Muchowski & Wacker, 2005b; Walsh & Selkoe, 2004). Many mechanisms can be included, such as excitotoxicity (Hynd, Scott, & Dodd, 2004; Mark, Ashford, Goodman, & Mattson, 1995), inflammation (Wyss-Coray & Mucke, 2002), cognitive stress (Beal, 2005) along with other processes (Handley, Naji, Dunnett, & Rosser, 2006; Mattson, 2004; Muchowski & Wacker, 2005b). In AD, synaptic loss surpasses the neuronal loss, and reduction of synaptic proteins and synapses relates better with cognitive degeneration than the massive quantity of tangles or plaques (Honer, 2003; Levine, Cepeda, Hickey, Fleming, & Chesselet, 2004; Van Dellen, Grote, & Hannan, 2005). Chronic variations in neurotransmission and synaptic plasticity can impact expression of gene and activity-dependent signalling, thereby, causing the degeneration of neural network and, in the end, the collapse of functioning of neurons. Conversely, in mouse models, environmental stimulation, upsurges the synaptic plasticity and can both decrease and delay pathological alterations (Lazarov et al., 2005; Van Dellen et al., 2005). The lipid provider (APOE) regulates sensitivity to typical bronchial ailments and neural plasticity. Different injuries produce an efficient neural plasticity reactions with the protective APOE3 and APOE2 isoforms (not the APOE4) which intensifies the menace and accelerates the onset of PD AD, and different neurological conditions (Mahley et al., 2006).

FUTURE RESEARCH DIRECTIONS

There is a constant research to discover mechanism by which disease-associated proteins aggregate, misfold and produce toxicity (Moreno-Gonzalez & Soto, 2011). Presently, various efforts are focussed on eliminating the abnormal protein assembly themselves (Muchowski & Wacker, 2005a; Selkoe & Schenk, 2003). By increasing the removal, nullifying the actions of the assembly and decreasing the synthesis helps in improving the survival of neurons. If the functional decline in NDs is produced by the neuronal loss, it might take many years for the detection of benefits of disease-modifying intrusions-a discouraging possibility that depresses developing better drugs for AD (Tasakis & Tsolaki, 2015). The continued progress in our ability to examine amyloid-forming protein and the association with some proteins provide assurance that novel treatments will be recognized for different disease states (Kaiser et al., 2012). The therapeutic options are explored which includes the targeting of the interaction between a chaperone-protein at various points from the route, thereby, encouraging the clearance of proteins and network rebalancing (Saibil, 2013). Nevertheless, the recognition and *in vivo* validation of new therapeutics is obstructed by the deficiency of the drivers of diseases and biomarkers for examining the therapeutic responses in model organisms. Nonetheless, the acceleration in research and collaboration amongst the community of drug discovery (regulatory agencies, pharmaceutical businesses, academia, advocacy groups, foundations, patients, contract research organizations, clinicians) is a constructive change which may help hasten the identification of novel therapeutic modalities.

CONCLUSION

In the research of neuroscience, the arena of NDs is one of the most studied with respect to both medical and concerned social issues. The current studies have significantly increased our knowledge of the role of mitochondria in the pathogenesis of NDs. The interaction of mitochondria with a particular protein which is related to the disease unties the prospects for discovery of therapeutic targets. Whereas, in PD, ALS and AD, genetic aspects of disease relate to only small fraction of cases and the significance of interaction of mitochondria with these proteins should be established for sporadic cases. The advances in this area have led to the essential information of the diseases, assessment of pathological processes and estimation of therapeutic approaches to deal with them. These strategies include therapies which are based on better understanding of the physiopathological mechanisms and efficient neuroprotection. These approaches to NDs are perhaps extremely global to be effective in short term. Nonetheless, advances in the understanding of etiopathological factors of the disease at genetic and molecular levels will cause new developments. The information of NDs has advanced rapidly in the previous years, and the area holds an immense promise for expanding our knowledge and understanding and subsequent treatment of the destructive taupathies.

REFERENCES

Aguzzi, A., Sigurdson, C., & Heikenwaelder, M. (2008). Molecular mechanisms of prion pathogenesis. *Annual Review of Pathology: Mechanisms of Disease*, *3*(1), 11–40. doi:10.1146/annurev.pathmechdis.3.121806.154326 PMID:18233951

Andres, R. H., Ducray, A. D., Schlattner, U., Wallimann, T., & Widmer, H. R. (2008). Functions and effects of creatine in the central nervous system. *Brain Research Bulletin*, *76*(4), 329–343. doi:10.1016/j. brainresbull.2008.02.035 PMID:18502307

Armstrong, R. A., Cairns, N. J., & Lantos, P. L. (2001). What does the study of the spatial patterns of pathological lesions tell us about the pathogenesis of neurodegenerative disorders? *Neuropathology*, *21*(1), 1–12. doi:10.1046/j.1440-1789.2001.00373.x PMID:11304036

Beal, M. F. (2005). Mitochondria take center stage in aging and neurodegeneration. *Annals of Neurology*, *58*(4), 495–505. doi:10.1002/ana.20624 PMID:16178023

Benchoua, A., Trioulier, Y., Zala, D., Gaillard, M.-C., Lefort, N., Dufour, N., ... Hantraye, P. (2006). Involvement of mitochondrial complex II defects in neuronal death produced by N-terminus fragment of mutated huntingtin. *Molecular Biology of the Cell*, *17*(4), 1652–1663. doi:10.1091/mbc.E05-07-0607 PMID:16452635

Bender, A., Krishnan, K. J., Morris, C. M., Taylor, G. A., Reeve, A. K., Perry, R. H., ... Klopstock, T. (2006). High levels of mitochondrial DNA deletions in substantia nigra neurons in aging and Parkinson disease. *Nature Genetics*, *38*(5), 515–517. doi:10.1038/ng1769 PMID:16604074

Bennett, S., Grant, M. M., & Aldred, S. (2009). Oxidative stress in vascular dementia and Alzheimer's disease: A common pathology. *Journal of Alzheimer's Disease*, *17*(2), 245–257. doi:10.3233/JAD-2009-1041 PMID:19221412

Betarbet, R., Sherer, T. B., MacKenzie, G., Garcia-Osuna, M., Panov, A. V., & Greenamyre, J. T. (2000). Chronic systemic pesticide exposure reproduces features of Parkinson's disease. *Nature Neuroscience*, *3*(12), 1301–1306. doi:10.1038/81834 PMID:11100151

Braak, H., & Braak, E. (1991). Neuropathological stageing of Alzheimer-related changes. *Acta Neuropathologica*, 82(4), 239–259. doi:10.1007/BF00308809 PMID:1759558

Brouillet, E., Hantraye, P., Ferrante, R. J., Dolan, R., Leroy-Willig, A., Kowall, N. W., & Beal, M. F. (1995). Chronic mitochondrial energy impairment produces selective striatal degeneration and abnormal choreiform movements in primates. *Proceedings of the National Academy of Sciences of the United States of America*, 92(15), 7105–7109. doi:10.1073/pnas.92.15.7105 PMID:7624378

Bunton-Stasyshyn, R. K., Saccon, R. A., Fratta, P., & Fisher, E. M. (2015). SOD1 Function and Its Implications for Amyotrophic Lateral Sclerosis Pathology: New and Renascent Themes. *The Neuroscientist*, *21*(5), 519–529. doi:10.1177/1073858414561795 PMID:25492944

Busciglio, J., Pelsman, A., Wong, C., Pigino, G., Yuan, M., Mori, H., & Yankner, B. A. (2002). Altered metabolism of the amyloid β precursor protein is associated with mitochondrial dysfunction in Down's syndrome. *Neuron*, *33*(5), 677–688. doi:10.1016/S0896-6273(02)00604-9 PMID:11879646

Caughey, B., & Baron, G. S. (2006). Prions and their partners in crime. *Nature*, 443(7113), 803–810. doi:10.1038/nature05294 PMID:17051207

Cicchetti, F., Drouin-Ouellet, J., & Gross, R. E. (2009). Environmental toxins and Parkinson's disease: What have we learned from pesticide-induced animal models? *Trends in Pharmacological Sciences*, *30*(9), 475–483. doi:10.1016/j.tips.2009.06.005 PMID:19729209

Citron, M. (2002). Alzheimer's disease: Treatments in discovery and development. *Nature Neuroscience*, *5*(11s), 1055–1057. doi:10.1038/nn940 PMID:12403985

Clinton, L. K., Blurton-Jones, M., Myczek, K., Trojanowski, J. Q., & LaFerla, F. M. (2007). Synergistic Interactions Among Abeta, Tau, and Alpha-synuclein: Acceleration of Neuropathology and Cognitive Decline. *The Journal of Neuroscience*, *30*(21), 7281–7289. doi:10.1523/JNEUROSCI.0490-10.2010 PMID:20505094

Collinge, J., & Clarke, A. R. (2007). A general model of prion strains and their pathogenicity. *Science*, *318*(5852), 930–936. doi:10.1126cience.1138718 PMID:17991853

Cookson, M. R. (2009). α-Synuclein and neuronal cell death. *Molecular Neurodegeneration*, 4(1), 9. doi:10.1186/1750-1326-4-9 PMID:19193223

Corti, O., Fournier, M., & Brice, A. (2009). Neurodegeneration in Parkinson's disease: Genetics enlightens physiopathology. In Birth, Life and Death of Dopaminergic Neurons in the Substantia Nigra (pp. 215–221). Springer. doi:10.1007/978-3-211-92660-4_17

Coskun, P. E., Beal, M. F., & Wallace, D. C. (2004). Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proceedings of the National Academy of Sciences of the United States of America*, *101*(29), 10726–10731. doi:10.1073/ pnas.0403649101 PMID:15247418

Cui, H., Kong, Y., & Zhang, H. (2012). Oxidative Stress, Mitochondrial Dysfunction, and Aging. *Journal of Signal Transduction*, 646354. doi:10.1155/2012/646354 PMID:21977319

Cushman, M., Johnson, B. S., King, O. D., Gitler, A. D., & Shorter, J. (2010). Prion-like disorders: Blurring the divide between transmissibility and infectivity. *Journal of Cell Science*, *123*(8), 1191–1201. doi:10.1242/jcs.051672 PMID:20356930

Damiano, M., Starkov, A. A., Petri, S., Kipiani, K., Kiaei, M., Mattiazzi, M., ... Manfredi, G. (2006). Neural mitochondrial Ca2+ capacity impairment precedes the onset of motor symptoms in G93A Cu/ Zn-superoxide dismutase mutant mice. *Journal of Neurochemistry*, *96*(5), 1349–1361. doi:10.1111/j.1471-4159.2006.03619.x PMID:16478527

Dawson, T. M., & Dawson, V. L. (2010). The role of parkin in familial and sporadic Parkinson's disease. *Journal of Movement Disorders*, 25(S1), S32–S39. doi:10.1002/mds.22798 PMID:20187240

De Baets, G., Reumers, J., Blanco, J. D., Dopazo, J., Schymkowitz, J., & Rousseau, F. (2011). An evolutionary trade-off between protein turnover rate and protein aggregation favors a higher aggregation propensity in fast degrading proteins. *PLoS Computational Biology*, 7(6), e1002090. doi:10.1371/journal. pcbi.1002090 PMID:21731483

De Castro, I. P., Costa, A., Lam, D., Tufi, R., Fedele, V., Moisoi, N., ... Martins, L. M. (2012). Genetic analysis of mitochondrial protein misfolding in Drosophila melanogaster. *Cell Death and Differentiation*, *19*(8), 1308–1316. doi:10.1038/cdd.2012.5 PMID:22301916

Deas, E., Wood, N. W., & Plun-Favreau, H. (2011). Mitophagy and Parkinson's disease: The PINK1– parkin link. *Biochimica et Biophysica Acta (BBA)- Molecular Cell Research*, *1813*(4), 623–633.

Dick, F. D. (2006). Parkinson's disease and pesticide exposures. *British Medical Bulletin*, 79(1), 219–231. doi:10.1093/bmb/ldl018 PMID:17242039

Dickson, D. W. (2010). Neuropathology of non-Alzheimer degenerative disorders. *International Journal of Clinical and Experimental Pathology*, *3*(1), 1. PMID:19918325

Elson, J. L., Herrnstadt, C., Preston, G., Thal, L., Morris, C. M., Edwardson, J., ... Howell, N. (2006). Does the mitochondrial genome play a role in the etiology of Alzheimer's disease? *Human Genetics*, *119*(3), 241–254. doi:10.100700439-005-0123-8 PMID:16408223

Esler, W. P., & Wolfe, M. S. (2001). A portrait of Alzheimer secretases--new features and familiar faces. *Science*, *293*(5534), 1449–1454. doi:10.1126cience.1064638 PMID:11520976

Finkbeiner, S. (2011). Huntington's Disease. *Cold Spring Harbor Perspectives in Biology*, *3*(6), a007476. doi:10.1101/cshperspect.a007476 PMID:21441583

Fornai, F., Schlüter, O. M., Lenzi, P., Gesi, M., Ruffoli, R., Ferrucci, M., ... Battaglia, G. (2005). Parkinson-like syndrome induced by continuous MPTP infusion: Convergent roles of the ubiquitinproteasome system and α-synuclein. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(9), 3413–3418. doi:10.1073/pnas.0409713102 PMID:15716361 Ghavami, S., Shojaei, S., Yeganeh, B., Ande, S. R., Jangamreddy, J. R., Mehrpour, M., ... Kashani, H. H. (2014). Autophagy and apoptosis dysfunction in neurodegenerative disorders. *Progress in Neurobiology*, *112*, 24–49. doi:10.1016/j.pneurobio.2013.10.004 PMID:24211851

Gianaros, P. J., Jennings, J. R., Sheu, L. K., Greer, P. J., Kuller, L. H., & Matthews, K. A. (2007). Prospective reports of chronic life stress predict decreased grey matter volume in the hippocampus. *NeuroImage*, *35*(2), 795–803. doi:10.1016/j.neuroimage.2006.10.045 PMID:17275340

Goedert, M. (2004). Tau protein and neurodegeneration. *Seminars in Cell & Developmental Biology*, 15(1), 45–49. doi:10.1016/j.semcdb.2003.12.015 PMID:15036206

Gorman, A. M. (2008). Neuronal cell death in neurodegenerative diseases: Recurring themes around protein handling. *Journal of Cellular and Molecular Medicine*, *12*(6a), 2263–2280. doi:10.1111/j.1582-4934.2008.00402.x PMID:18624755

Greenwood, P. M., & Parasuraman, R. (2010). Neuronal and cognitive plasticity: A neurocognitive framework for ameliorating cognitive aging. *Frontiers in Aging Neuroscience*, 2. PMID:21151819

Gu, M., Gash, M., Mann, V., Javoy-Agid, F., Cooper, J., & Schapira, A. (1996). Mitochondrial defect in Huntington's disease caudate nucleus. *Annals of Neurology*, *39*(3), 385–389. doi:10.1002/ana.410390317 PMID:8602759

Hall, G. (2011). What is the Link Between Protein Aggregation and Interneuronal Lesion Propagation in Neurodegenerative Disease? In *Neurodegenerative Diseases-Processes, Prevention, Protection and Monitoring*. InTech.

Handley, O. J., Naji, J. J., Dunnett, S. B., & Rosser, A. E. (2006). Pharmaceutical, cellular and genetic therapies for Huntington's disease. *Clinical Science*, *110*(1), 73–88. doi:10.1042/CS20050148 PMID:16336206

Hardy, J., & Selkoe, D. J. (2002). The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science*, 297(5580), 353–356. doi:10.1126cience.1072994 PMID:12130773

Higgins, C. M., Jung, C., & Xu, Z. (2003). ALS-associated mutant SOD1 G93A causes mitochondrial vacuolation by expansion of the intermembrane space and by involvement of SOD1 aggregation and peroxisomes. *BMC Neuroscience*, *4*(1), 16. doi:10.1186/1471-2202-4-16 PMID:12864925

Honer, W. G. (2003). Pathology of presynaptic proteins in Alzheimer's disease: More than simple loss of terminals. *Neurobiology of Aging*, *24*(8), 1047–1062. doi:10.1016/j.neurobiolaging.2003.04.005 PMID:14643376

Hynd, M. R., Scott, H. L., & Dodd, P. R. (2004). Glutamate-mediated excitotoxicity and neurodegeneration in Alzheimer's disease. *Neurochemistry International*, 45(5), 583–595. doi:10.1016/j.neuint.2004.03.007 PMID:15234100

Ingram, E. M., & Spillantini, M. G. (2002). Tau gene mutations: Dissecting the pathogenesis of FTDP-17. *Trends in Molecular Medicine*, 8(12), 555–562. doi:10.1016/S1471-4914(02)02440-1 PMID:12470988

Institutes, G. (2015). Loss of cellular energy leads to neuronal dysfunction in neurodegenerative disease model: Scientists developed new tests to accurately measure brain's energy supply, which is essential for understanding how impairments in the system cause neurodegeneration. Retrieved from www.sci-encedaily.com/releases/2015/09/150914215617.htm

Jaarsma, D., Rognoni, F., van Duijn, W., Verspaget, H. W., Haasdijk, E. D., & Holstege, J. C. (2001). CuZn superoxide dismutase (SOD1) accumulates in vacuolated mitochondria in transgenic mice expressing amyotrophic lateral sclerosis-linked SOD1 mutations. *Acta Neuropathologica*, *102*(4), 293–305. PMID:11603803

Jellinger, K. A. (2010). Basic mechanisms of neurodegeneration: A critical update. *Journal of Cellular and Molecular Medicine*, *14*(3), 457–487. PMID:20070435

Jellinger, K. A. (2012). Interaction between pathogenic proteins in neurodegenerative disorders. *Journal of Cellular and Molecular Medicine*, *16*(6), 1166–1183. doi:10.1111/j.1582-4934.2011.01507.x PMID:22176890

Jenkins, B. G., Koroshetz, W. J., Beal, M. F., & Rosen, B. R. (1993). Evidence for impairment of energy metabolism in vivo in Huntington's disease using localized 1H NMR spectroscopy. *Neurology*, *43*(12), 2689–2695. doi:10.1212/WNL.43.12.2689 PMID:8255479

Kaiser, D. M., Acharya, M., Leighton, P. L., Wang, H., Daude, N., Wohlgemuth, S., ... Allison, W. T. (2012). Amyloid beta precursor protein and prion protein have a conserved interaction affecting cell adhesion and CNS development. *PLoS One*, *7*(12), e51305. doi:10.1371/journal.pone.0051305 PMID:23236467

Klivenyi, P., Siwek, D., Gardian, G., Yang, L., Starkov, A., Cleren, C., ... Beal, M. F. (2006). Mice lacking alpha-synuclein are resistant to mitochondrial toxins. *Neurobiology of Disease*, 21(3), 541–548. doi:10.1016/j.nbd.2005.08.018 PMID:16298531

Kong, J., & Xu, Z. (1998). Massive mitochondrial degeneration in motor neurons triggers the onset of amyotrophic lateral sclerosis in mice expressing a mutant SOD1. *The Journal of Neuroscience*, *18*(9), 3241–3250. PMID:9547233

Kovacs, G. G. (2016). Molecular pathological classification of neurodegenerative diseases: Turning towards precision medicine. *International Journal of Molecular Sciences*, *17*(2), 189. doi:10.3390/ ijms17020189 PMID:26848654

Kraytsberg, Y., Kudryavtseva, E., McKee, A. C., Geula, C., Kowall, N. W., & Khrapko, K. (2006). Mitochondrial DNA deletions are abundant and cause functional impairment in aged human substantia nigra neurons. *Nature Genetics*, *38*(5), 518–520. doi:10.1038/ng1778 PMID:16604072

Lagouge, M., & Larsson, N. G. (2013). The role of mitochondrial DNA mutations and free radicals in disease and ageing. *Journal of Internal Medicine*, 273(6), 529–543. doi:10.1111/joim.12055 PMID:23432181

Lansbury, P. T., & Lashuel, H. A. (2006). A century-old debate on protein aggregation and neurodegeneration enters the clinic. *Nature*, 443(7113), 774–779. doi:10.1038/nature05290 PMID:17051203

Lazarov, O., Robinson, J., Tang, Y.-P., Hairston, I. S., Korade-Mirnics, Z., Lee, V. M.-Y., ... Sisodia, S. S. (2005). Environmental enrichment reduces A β levels and amyloid deposition in transgenic mice. *Cell*, *120*(5), 701–713. doi:10.1016/j.cell.2005.01.015 PMID:15766532

Lee, V. M., Goedert, M., & Trojanowski, J. Q. (2001). Neurodegenerative tauopathies. *Annual Review of Neuroscience*, 24(1), 1121–1159. doi:10.1146/annurev.neuro.24.1.1121 PMID:11520930

Lev, N., Roncevic, D., Ickowicz, D., Melamed, E., & Offen, D. (2006). Role of DJ-1 in Parkinson's disease. *Journal of Molecular Neuroscience*, 29(3), 215–225. doi:10.1385/JMN:29:3:215 PMID:17085780

Levine, M. S., Cepeda, C., Hickey, M. A., Fleming, S. M., & Chesselet, M.-F. (2004). Genetic mouse models of Huntington's and Parkinson's diseases: Illuminating but imperfect. *Trends in Neurosciences*, 27(11), 691–697. doi:10.1016/j.tins.2004.08.008 PMID:15474170

Li, F., Calingasan, N. Y., Yu, F., Mauck, W. M., Toidze, M., Almeida, C. G., ... Lin, M. T. (2004). Increased plaque burden in brains of APP mutant MnSOD heterozygous knockout mice. *Journal of Neurochemistry*, 89(5), 1308–1312. doi:10.1111/j.1471-4159.2004.02455.x PMID:15147524

Li, J.-Q., Tan, L., & Yu, J.-T. (2014). The role of the LRRK2 gene in Parkinsonism. *Molecular Neurodegeneration*, 9(1), 47. doi:10.1186/1750-1326-9-47 PMID:25391693

Lim, J., & Yue, Z. (2015). Neuronal aggregates: Formation, clearance and spreading. *Developmental Cell*, *32*(4), 491–501. doi:10.1016/j.devcel.2015.02.002 PMID:25710535

Lin, M. T., & Beal, M. F. (2006). Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature*, 443(7113), 787–795. doi:10.1038/nature05292 PMID:17051205

Lin, M. T., Simon, D. K., Ahn, C. H., Kim, L. M., & Beal, M. F. (2002). High aggregate burden of somatic mtDNA point mutations in aging and Alzheimer's disease brain. *Human Molecular Genetics*, *11*(2), 133–145. doi:10.1093/hmg/11.2.133 PMID:11809722

Lindquist, S., Krobitsch, S., Li, L., & Sondheimer, N. (2001). Investigating protein conformation–based inheritance and disease in yeast. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, *356*(1406), 169–176. doi:10.1098/rstb.2000.0762 PMID:11260797

Liou, Y.-C., Sun, A., Ryo, A., Zhou, X. Z., Yu, Z.-X., Huang, H.-K., ... Lu, K. P. (2003). Role of the prolyl isomerase Pin1 in protecting against age-dependent neurodegeneration. *Nature*, *424*(6948), 556–561. doi:10.1038/nature01832 PMID:12891359

Liu, J., Lillo, C., Jonsson, P. A., Velde, C. V., Ward, C. M., Miller, T. M., ... Andersen, P. M. (2004). Toxicity of familial ALS-linked SOD1 mutants from selective recruitment to spinal mitochondria. *Neuron*, *43*(1), 5–17. doi:10.1016/j.neuron.2004.06.016 PMID:15233913

Luoma, P., Melberg, A., Rinne, J. O., Kaukonen, J. A., Nupponen, N. N., Chalmers, R. M., ... Majamaa, K. (2004). Parkinsonism, premature menopause, and mitochondrial DNA polymerase γ mutations: Clinical and molecular genetic study. *Lancet*, *364*(9437), 875–882. doi:10.1016/S0140-6736(04)16983-3 PMID:15351195

Ma, J., & Lindquist, S. (2002). Conversion of PrP to a self-perpetuating PrPSc-like conformation in the cytosol. *Science*, *298*(5599), 1785–1788. doi:10.1126cience.1073619 PMID:12386336

Ma, J., Wollmann, R., & Lindquist, S. (2002). Neurotoxicity and neurodegeneration when PrP accumulates in the cytosol. *Science*, 298(5599), 1781–1785. doi:10.1126cience.1073725 PMID:12386337

Ma, S. L., Pastorino, L., Zhou, X. Z., & Lu, K. P. (2012). Prolyl Isomerase Pin1 Promotes Amyloid Precursor Protein (APP) Turnover by Inhibiting Glycogen Synthase Kinase-3β (GSK3β) Activity novel mechanism for pin1 to protect against Alzheimer disease. *The Journal of Biological Chemistry*, 287(10), 6969–6973. doi:10.1074/jbc.C111.298596 PMID:22184106

Mahley, R. W., Weisgraber, K. H., & Huang, Y. (2006). Apolipoprotein E4: A causative factor and therapeutic target in neuropathology, including Alzheimer's disease. *Proceedings of the National Academy of Sciences of the United States of America*, *103*(15), 5644–5651. doi:10.1073/pnas.0600549103 PMID:16567625

Mancuso, M., Calsolaro, V., Orsucci, D., Carlesi, C., Choub, A., Piazza, S., & Siciliano, G. (2009). Mitochondria, Cognitive Impairment, and Alzheimer's Disease. *International Journal of Alzheimer's Disease*, *951548*. doi:10.4061/2009/951548 PMID:20798880

Mancuso, M., Orsucci, D., Siciliano, G., & Murri, L. (2008). Mitochondria, mitochondrial DNA and Alzheimer's disease. What comes first? *Current Alzheimer Research*, 5(5), 457–468. doi:10.2174/156720508785908946 PMID:18855587

Mark, R. J., Ashford, J. W., Goodman, Y., & Mattson, M. P. (1995). Anticonvulsants attenuate amyloid β-peptide neurotoxicity, Ca 2+ deregulation, and cytoskeletal pathology. *Neurobiology of Aging*, *16*(2), 187–198. doi:10.1016/0197-4580(94)00150-2 PMID:7777136

Martin, L. J. (2010). Mitochondrial and cell death mechanisms in neurodegenerative diseases. *Pharmaceuticals*, *3*(4), 839–915. doi:10.3390/ph3040839 PMID:21258649

Martin, L. J., Pan, Y., Price, A. C., Sterling, W., Copeland, N. G., Jenkins, N. A., ... Lee, M. K. (2006). Parkinson's disease α -synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death. *The Journal of Neuroscience*, 26(1), 41–50. doi:10.1523/JNEUROSCI.4308-05.2006 PMID:16399671

Mattiazzi, M., D'Aurelio, M., Gajewski, C. D., Martushova, K., Kiaei, M., Beal, M. F., & Manfredi, G. (2002). Mutated human SOD1 causes dysfunction of oxidative phosphorylation in mitochondria of transgenic mice. *The Journal of Biological Chemistry*, 277(33), 29626–29633. doi:10.1074/jbc. M203065200 PMID:12050154

Mattson, M. P. (2004). Pathways towards and away from Alzheimer's disease. *Nature*, 430(7000), 631–639. doi:10.1038/nature02621 PMID:15295589

Milakovic, T., & Johnson, G. V. (2005). Mitochondrial respiration and ATP production are significantly impaired in striatal cells expressing mutant huntingtin. *The Journal of Biological Chemistry*, *280*(35), 30773–30782. doi:10.1074/jbc.M504749200 PMID:15983033

Moisan, F., Spinosi, J., Dupupet, J. L., Delabre, L., Mazurie, J. L., Goldberg, M., ... Elbaz, A. (2011). The relation between type of farming and prevalence of Parkinson's disease among agricultural workers in five French districts. *Movement Disorders*, *26*(2), 271–279. doi:10.1002/mds.23370 PMID:21412834

Moreno-Gonzalez, I., & Soto, C. (2011). Misfolded protein aggregates: Mechanisms, structures and potential for disease transmission. *Seminars in Cell & Developmental Biology*, 22(5), 482–487. doi:10.1016/j. semcdb.2011.04.002 PMID:21571086 Muchowski, P. J., & Wacker, J. L. (2005a). Modulation of neurodegeneration by molecular chaperones. *Nature Reviews. Neuroscience*, *6*(1), 11–22. doi:10.1038/nrn1587 PMID:15611723

Murphy, M. P., & LeVine, H. (2010). Alzheimer's Disease and the β-Amyloid Peptide. *Journal of Alzheimer's disease. JAD*, *19*(1), 311. doi:10.3233/JAD-2010-1221 PMID:20061647

Nearly 1 in 6 of world's population suffer from neurological disorders – UN report. (2007). Retrieved from http://www.un.org/apps/news/story.asp?cr=neurological&newsid=21689#.WhBcS2iCzIU

Nixon, R. A., & Yang, D.-S. (2012). Autophagy and neuronal cell death in neurological disorders. *Cold Spring Harbor Perspectives in Biology*, 4(10), a008839. doi:10.1101/cshperspect.a008839 PMID:22983160

O'Brien, R. J., & Wong, P. C. (2011). Amyloid Precursor Protein Processing and Alzheimer's Disease. *Annual Review of Neuroscience*, *34*(1), 185–204. doi:10.1146/annurev-neuro-061010-113613 PMID:21456963

Ohyagi, Y., Yamada, T., Nishioka, K., Clarke, N. J., Tomlinson, A. J., Naylor, S., ... Younkin, S. G. (2000). Selective increase in cellular A β 42 is related to apoptosis but not necrosis. *Neuroreport*, *11*(1), 167–171. doi:10.1097/00001756-200001170-00033 PMID:10683851

Palop, J. J., Chin, J., & Mucke, L. (2006). A network dysfunction perspective on neurodegenerative diseases. *Nature*, 443(7113), 768–773. doi:10.1038/nature05289 PMID:17051202

Panov, A. V., Gutekunst, C.-A., Leavitt, B. R., Hayden, M. R., Burke, J. R., Strittmatter, W. J., & Greenamyre, J. T. (2002). Early mitochondrial calcium defects in Huntington's disease are a direct effect of polyglutamines. *Nature Neuroscience*, *5*(8), 731–736. doi:10.1038/nn884 PMID:12089530

Parchi, P., Castellani, R., Capellari, S., Ghetti, B., Young, K., Chen, S. G., ... Trojanowski, J. Q. (1996). Molecular basis of phenotypic variability in sporadc creudeldt-jakob disease. *Annals of Neurology*, *39*(6), 767–778. doi:10.1002/ana.410390613 PMID:8651649

Pickrell, A. M., & Youle, R. J. (2015). The Roles of PINK1, Parkin and Mitochondrial Fidelity in Parkinson's Disease. *Neuron*, 85(2), 257–273. doi:10.1016/j.neuron.2014.12.007 PMID:25611507

Praticò, D., Uryu, K., Leight, S., Trojanoswki, J. Q., & Lee, V. M.-Y. (2001). Increased lipid peroxidation precedes amyloid plaque formation in an animal model of Alzheimer amyloidosis. *The Journal of Neuroscience*, 21(12), 4183–4187. PMID:11404403

Price, C. J., & Friston, K. J. (2002). Degeneracy and cognitive anatomy. *Trends in Cognitive Sciences*, *6*(10), 416–421. doi:10.1016/S1364-6613(02)01976-9 PMID:12413574

Pruisner, S. (2001). Shattuck lecture-neurodegenerative diseases and prions. *The New England Journal of Medicine*, 344(20), 1516–1526. doi:10.1056/NEJM200105173442006 PMID:11357156

Pyle, A., Foltynie, T., Tiangyou, W., Lambert, C., Keers, S. M., Allcock, L. M., ... Barker, R. (2005). Mitochondrial DNA haplogroup cluster UKJT reduces the risk of PD. *Annals of Neurology*, *57*(4), 564–567. doi:10.1002/ana.20417 PMID:15786469

Qiu, C., Kivipelto, M., & von Strauss, E. (2009). Epidemiology of Alzheimer's disease: Occurrence, determinants, and strategies toward intervention. *Dialogues in Clinical Neuroscience*, *11*(2), 111. PMID:19585947

Recchia, A., Debetto, P., Negro, A., Guidolin, D., Skaper, S. D., & Giusti, P. (2004). α-Synuclein and Parkinson's disease. *The FASEB Journal*, *18*(6), 617–626. doi:10.1096/fj.03-0338rev PMID:15054084

Reddy, P. H., McWeeney, S., Park, B. S., Manczak, M., Gutala, R. V., Partovi, D., ... Mori, M. (2004). Gene expression profiles of transcripts in amyloid precursor protein transgenic mice: Up-regulation of mitochondrial metabolism and apoptotic genes is an early cellular change in Alzheimer's disease. *Human Molecular Genetics*, *13*(12), 1225–1240. doi:10.1093/hmg/ddh140 PMID:15115763

Rezania, K., Yan, J., Dellefave, L., Deng, H. X., Siddique, N., & Pascuzzi, T. (2003). A rare Cu/Zn superoxide dismutase mutation causing familial amyotrophic lateral sclerosis with variable age of onset, incomplete penetrance and a sensory neuropathy. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, 4(3), 162–166. doi:10.1080/aml.4.3.162.166 PMID:13129803

Rinne, U., Sonninen, V., & Siirtola, T. (1970). L-dopa treatment in Parkinson's disease. *European Neurology*, 4(6), 348–369. doi:10.1159/000113990 PMID:4932969

Saibil, H. (2013). Chaperone machines for protein folding, unfolding and disaggregation. *Nature Reviews*. *Molecular Cell Biology*, *14*(10), 630–642. doi:10.1038/nrm3658 PMID:24026055

Schapira, A., Cooper, J., Dexter, D., Clark, J., Jenner, P., & Marsden, C. (1990). Mitochondrial complex I deficiency in Parkinson's disease. *Journal of Neurochemistry*, *54*(3), 823–827. doi:10.1111/j.1471-4159.1990. tb02325.x PMID:2154550

Scheibel, T., Bloom, J., & Lindquist, S. L. (2004). The elongation of yeast prion fibers involves separable steps of association and conversion. *Proceedings of the National Academy of Sciences of the United States of America*, *101*(8), 2287–2292. doi:10.1073/pnas.0308754101 PMID:14983002

Selkoe, D. J. (2002). Alzheimer's disease is a synaptic failure. *Science*, 298(5594), 789–791. doi:10.1126cience.1074069 PMID:12399581

Selkoe, D. J., & Schenk, D. (2003). Alzheimer's disease: Molecular understanding predicts amyloid-based therapeutics. *Annual Review of Pharmacology and Toxicology*, *43*(1), 545–584. doi:10.1146/annurev. pharmtox.43.100901.140248 PMID:12415125

Serpell, L. C., & Smith, J. M. (2000). Direct visualisation of the β-sheet structure of synthetic Alzheimer's amyloid. *Journal of Molecular Biology*, 299(1), 225–231. doi:10.1006/jmbi.2000.3650 PMID:10860734

Sherer, T. B., Betarbet, R., Testa, C. M., Seo, B. B., Richardson, J. R., Kim, J. H., ... Greenamyre, J. T. (2003). Mechanism of toxicity in rotenone models of Parkinson's disease. *The Journal of Neuroscience*, 23(34), 10756–10764. PMID:14645467

Simon, D., Pulst, S., Sutton, J., Browne, S., Beal, M., & Johns, D. (1999). Familial multisystem degeneration with parkinsonism associated with the 11778 mitochondrial DNA mutation. *Neurology*, *53*(8), 1787–1787. doi:10.1212/WNL.53.8.1787 PMID:10563629

Simon, D. K., Lin, M. T., Zheng, L., Liu, G.-J., Ahn, C. H., Kim, L. M., ... Johns, D. R. (2004). Somatic mitochondrial DNA mutations in cortex and substantia nigra in aging and Parkinson's disease. *Neurobiology of Aging*, 25(1), 71–81. doi:10.1016/S0197-4580(03)00037-X PMID:14675733

Sleegers, K., Cruts, M., & Van Broeckhoven, C. (2010). Molecular pathways of frontotemporal lobar degeneration. *Annual Review of Neuroscience*, *33*(1), 71–88. doi:10.1146/annurev-neuro-060909-153144 PMID:20415586

Song, D. D., Shults, C. W., Sisk, A., Rockenstein, E., & Masliah, E. (2004). Enhanced substantia nigra mitochondrial pathology in human α -synuclein transgenic mice after treatment with MPTP. *Experimental Neurology*, *186*(2), 158–172. doi:10.1016/S0014-4886(03)00342-X PMID:15026254

Spillantini, M. G., Bird, T. D., & Ghetti, B. (1998). Frontotemporal dementia and Parkinsonism linked to chromosome 17: A new group of tauopathies. *Brain Pathology (Zurich, Switzerland)*, 8(2), 387–402. doi:10.1111/j.1750-3639.1998.tb00162.x PMID:9546295

Stefanis, L. (2012). α-Synuclein in Parkinson's Disease. *Cold Spring Harbor Perspectives in Medicine*, 2(2), a009399. doi:10.1101/cshperspect.a009399 PMID:22355802

Strauss, K. M., Martins, L. M., Plun-Favreau, H., Marx, F. P., Kautzmann, S., Berg, D., ... Kruger, R. (2005). Loss of function mutations in the gene encoding Omi/HtrA2 in Parkinson's disease. *Human Molecular Genetics*, *14*(15), 2099–2111. doi:10.1093/hmg/ddi215 PMID:15961413

Su, J. H., Deng, G., & Cotman, C. W. (1997). Transneuronal degeneration in the spread of Alzheimer's disease pathology: Immunohistochemical evidence for the transmission of tau hyperphosphorylation. *Neurobiology of Disease*, *4*(5), 365–375. doi:10.1006/nbdi.1997.0164 PMID:9440125

Sultana, R., Boyd-Kimball, D., Poon, H. F., Cai, J., Pierce, W. M., Klein, J. B., ... Butterfield, D. A. (2006). Oxidative modification and down-regulation of Pin1 in Alzheimer's disease hippocampus: A redox proteomics analysis. *Neurobiology of Aging*, *27*(7), 918–925. doi:10.1016/j.neurobiolaging.2005.05.005 PMID:15950321

Sweeney, P., Park, H., Baumann, M., Dunlop, J., Frydman, J., Kopito, R., ... Hodgson, R. (2017). Protein misfolding in neurodegenerative diseases: Implications and strategies. *Translational Neurodegeneration*, *6*(1), 6. doi:10.118640035-017-0077-5 PMID:28293421

Swerdlow, R., Parks, J., Cassarino, D., Maguire, D., Maguire, R., Bennett, J., ... Parker, W. (1997). Cybrids in Alzheimer's disease: A cellular model of the disease? *Neurology*, 49(4), 918–925. doi:10.1212/WNL.49.4.918 PMID:9339668

Takalo, M., Salminen, A., Soininen, H., Hiltunen, M., & Haapasalo, A. (2013). Protein aggregation and degradation mechanisms in neurodegenerative diseases. *American Journal of Neurodegenerative Disease*, 2(1), 1. PMID:23516262

Takeuchi, H., Kobayashi, Y., Ishigaki, S., Doyu, M., & Sobue, G. (2002). Mitochondrial localization of mutant superoxide dismutase 1 triggers caspase-dependent cell death in a cellular model of familial amyotrophic lateral sclerosis. *The Journal of Biological Chemistry*, 277(52), 50966–50972. doi:10.1074/jbc.M209356200 PMID:12393885

Tanaka, M. (2002). Mitochondrial genotypes and cytochrome b variants associated with longevity or Parkinson's disease. *Journal of Neurology*, 249. PMID:12375058

Tang, Y.-P., & Gershon, E. S. (2003). Genetic studies in Alzheimer's disease. *Dialogues in Clinical Neuroscience*, 5(1), 17–26. PMID:22033785

Tasakis, R. N. S., & Tsolaki, M. (2015). Mitochondria; pathogenesis and dysfunction in Alzheimer's disease. *Hellenic Journal of Nuclear Medicine*, 18, 10.

Tong, Y., Giaime, E., Yamaguchi, H., Ichimura, T., Liu, Y., Si, H., ... Shen, J. (2012). Loss of leucinerich repeat kinase 2 causes age-dependent bi-phasic alterations of the autophagy pathway. *Molecular Neurodegeneration*, 7(1), 2. doi:10.1186/1750-1326-7-2 PMID:22230652

Van Dellen, A., Grote, H. E., & Hannan, A. J. (2005). Gene–environment interactions, neuronal dysfunction and pathological plasticity in Huntington's disease. *Clinical and Experimental Pharmacology & Physiology*, *32*(12), 1007–1019. doi:10.1111/j.1440-1681.2005.04313.x PMID:16445565

Vijayvergiya, C., Beal, M. F., Buck, J., & Manfredi, G. (2005). Mutant superoxide dismutase 1 forms aggregates in the brain mitochondrial matrix of amyotrophic lateral sclerosis mice. *The Journal of Neuroscience*, *25*(10), 2463–2470. doi:10.1523/JNEUROSCI.4385-04.2005 PMID:15758154

Vives-Bauza, C., Andreu, A. L., Manfredi, G., Beal, M. F., Janetzky, B., Gruenewald, T. H., & Lin, M. T. (2002). Sequence analysis of the entire mitochondrial genome in Parkinson's disease. *Biochemical and Biophysical Research Communications*, 290(5), 1593–1601. doi:10.1006/bbrc.2002.6388 PMID:11820805

Wallace, D. C. (2005). A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: A dawn for evolutionary medicine. *Annual Review of Genetics*, *39*(1), 359–407. doi:10.1146/ annurev.genet.39.110304.095751 PMID:16285865

Walsh, D. M., & Selkoe, D. J. (2004). Deciphering the molecular basis of memory failure in Alzheimer's disease. *Neuron*, 44(1), 181–193. doi:10.1016/j.neuron.2004.09.010 PMID:15450169

Wood, L. B., Winslow, A. R., & Strasser, S. D. (2015). Systems biology of neurodegenerative diseases. *Integrative Biology*, 7(7), 758–775. doi:10.1039/C5IB00031A PMID:26065845

Wyss-Coray, T., & Mucke, L. (2002). Inflammation in neurodegenerative disease—a double-edged sword. *Neuron*, *35*(3), 419–432. doi:10.1016/S0896-6273(02)00794-8 PMID:12165466

Yang, H., & Hu, H. Y. (2016). Sequestration of cellular interacting partners by protein aggregates: Implication in a loss-of-function pathology. *The FEBS Journal*, 283(20), 3705–3717. doi:10.1111/ febs.13722 PMID:27016044

Zheng, Z., & Diamond, M. I. (2012). Huntington disease and the huntingtin protein. *Progress in Molecular Biology and Translational Science*, *107*, 189–214. doi:10.1016/B978-0-12-385883-2.00010-2 PMID:22482451

Chapter 7 Stroke: A Potential Risk Factor of Neurodegenerative Disorders

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ABSTRACT

The brain relies on a specialized endothelial system, the blood brain barrier (BBB), which is capable of regulating the transfer of substances from the blood to the neurons. Stroke is the most frequent cause of disability in adulthood. Lesions of vascular origin also include asymptomatic small infarcts, microbleeds, dilated perivascular spaces, and atrophy. Vascular cognitive impairment (VCI) is the second most prevalent cause of dementia. Several mechanisms are implied, including strategic infarct dementia, post-stroke dementia, cerebral amyloid angiopathy, and subcortical vascular dementia. As there is no disease modifying therapies currently available, treatment of comorbidities and adequate control of the vascular risk factors remain the standard strategies to reduce the vascular contributions to neurodegeneration. This chapter represents the basic concepts of pathophysiology of cerebrovascular diseases, and describes the subtypes of VCI, as well as treatment and primary prevention strategies.

DOI: 10.4018/978-1-5225-5282-6.ch007

INTRODUCTION

There is no single mechanism that satisfactorily explains the complex relation between stroke and neurodegenerative disorders, predominantly those related to cognition impairment. Rather, this relation may be explained by multiple pathophysiological mechanisms, such as microstructural defects, leading to blood brain barrier and neurovascular unit dysfunction; by vascular mechanisms, when traditional vascular risk factors lead to failure on the capacity of cerebral blood flow regulation; and by genetic predisposition, exemplified by CADASIL, apolipoprotein ε polymorphisms and variations of homocysteine's metabolism. Furthermore, these are not necessarily exclusive, and a combination of these factors may occur in individual patients (Iadecola, 2013; Schneider *et al*, 2003).

In patients with cerebrovascular disease, cognitive decline, affecting mainly executive domains and language functions, may occur. A variety of cerebrovascular conditions will result in distinct profiles of cognitive impairment, by either lowering the threshold for neurodegenerative diseases or unmasking an already present, often mild or subclinical, dementia.

Vascular cognitive impairment (VCI) is a condition in which vascular lesions can lead or contribute to impaired cognitive function, and ranges from mild cognitive disorder to dementia (Bowler and Hachinsky, 1996; Gorelick *et al.*, 2011). It is the second most prevalent cause of cognitive impairment, and remains vastly underdiagnosed (Godefroy *et al.*, 2013).

There are several clinical subtypes of VCI, such as poststroke dementia, with a short-term incidence after stroke as high as 40% (Pendlebury and Rothwell, 2009); strategic infarct dementia, caused by interruptions of strategic networks, leading to impaired executive functions; subcortical vascular dementia, by far the most common subtype of VCI; microinfarcts dementia, caused by small lesions detected only by high resolution brain imaging or biopsy; microbleeds related VCI, which could be caused either by hypertensive vasculopathy or cerebral amyloid angiopathy; hypoperfusion dementia, as a consequence of hemodynamic failure in border zone (watershed) regions; and mixed disease, when vascular pathology and Alzheimer Disease (AD) overlap.

In this context, VCI appears as a continuum of different clinical entities, which makes it the perfect prototype to understand the fascinating connection between cerebrovascular disease and neuronal degeneration (Iadecola, 2013; Schneider *et al*, 2003).

This chapter review the concepts of blood brain barrier, neurovascular unit and brain autoregulation; define stroke and its subtypes (ischemic, hemorrhagic and cerebral venous thrombosis), with their main risk factors, clinical features and morbidity; explain the impact of cerebrovascular diseases on cognition and neurodegeneration; and describe the subtypes of VCI, as well as treatment and primary prevention strategies.

BACKGROUND

The attempt to divide normal ageing from pathological senility began in the late 19th century, which was a critical period for developing the concept of dementia as we know it today. At that time, the definition of dementia was similar to the one used nowadays, although much more inclusive: a general compromise of intelligence, with prominent memory loss and alterations of personality. Studies by French and German psychiatrists were crucial to separate psychotic syndromes, such as schizophrenia, from other forms of cognitive impairment. This interest in understanding pathological aging has encouraged a great number

Stroke

of histological studies, which helped divide organic and psychiatric diseases into different nosological entities (Caixeta *et al.*, 2014).

In 1907, Dr. Alois Alzheimer first described the case of August Deter, a 51 year-old female patient showing language comprehension and memory disturbances. The pathological study of her brain revealed neurofibrillary tangles and amyloid senile plaques, establishing the typical histopathological findings of Alzheimer's disease. After that, the amyloid hypothesis became the most accepted explanation for senile dementia (Ramirez-Bermudez, 2012).

The vascular contributions to dementia were first described simultaneously to Alzheimer's work. Another neuropathologist, Dr. Gaetano Perusini, also studied the brain of August Deter, and described that "*the large cerebral vessels, the arterial circle of Willis and the Sylvian arteries showed no significant signs of arteriosclerosis*" (Ramirez-Bermudez, 2012). Despite that, until the 1990's, there was a general belief that cerebrovascular disease only caused dementia in the presence of many cortical infarcts – the "multi-infarct" approach. In 1974, Dr. Vladimir Hachinski stated that most cases of senile dementia were attributable to Alzheimer-like degeneration and not to atherosclerosis - for the Canadian doctor, cerebral infarcts accounted for only a minority of cases of mental deterioration in the elderly, often associated with hypertension or extracerebral vascular disease (Hachinski V.D. *et al.*, 1974).

In the past three decades epidemiological studies showed that dementia dramatically increases with cerebrovascular diseases and vascular risk factors and the vascular hypothesis for neurodegenerative process began to be better understood (O'Brien *et al.*, 2015).

Until the 21st century, VCI and neurodegenerative dementia (Alzheimer's disease) where thought to be two distinct nosological entities. However, in the past two decades, the vascular contributions to the pathology of every types of dementia have been increasingly recognized (O'Brien *et al.*, 2015). Nowadays we know that mixed dementia, with the co-existence of amyloid and vascular pathological processes, is the rule, and not the exception (Schneider, J.A. *et al.*, 2007).

THE BLOOD BRAIN BARRIER AND THE NEUROVASCULAR UNIT

Composed mainly by vascular endothelial cells, the blood brain barrier (BBB) also interacts with different vascular, immune and neural cells (Daneman and Prat, 2015). Different from the vasculature from other tissues, BBB allows the brain to stringently regulate the transfer of ions, molecules and cells between the blood and brain tissue itself, through tight junctions, enzymes and other transport systems (Ramirez-Bermudez, 2012). In order to maintain brain homeostasis, the endothelial cells deliver oxygen and nutrients, thus guaranteeing proper neuronal function, and filter substances from the brain back to the bloodstream, protecting the neural tissue from toxins and pathogens (Daneman and Prat, 2015).

BBB is involved in the pathology and progression of different neurological diseases. There is a great number of proinflammatory cytokines and other proteins related to specific diseases, which might mediate BBB disruption. Such dysfunction can be seen in traumatic brain injuries, viral encephalitis, Alzheimer's disease (AD), brain expanding lesions, stroke, epilepsy, multiple sclerosis and idiopathic Parkinson's disease (Persidsky *et al.*, 2006).

Together, endothelial cells, pericytes, basement membrane, neurons and glial cells form the neurovascular unit (Sá-Pereira *et al.*, 2012). Similar to the BBB, the neurovascular unit has many regulating and protective mechanisms, a process also called "neurovascular coupling", which refers to the capacity of the brain vasculature to regulate tissue perfusion, responding to changes in neuronal activity, systemic perfusion pressure and local changes to the chemical environment (Sá-Pereira *et al.*, 2012). Through vasodilation and vasoconstriction, the neurovascular unit maintains oxygen and nutrients in appropriate levels (Muoio *et al.*, 2014).

Autoregulation of cerebral blood flow is the main component of brain vasculature reactivity responsible for keeping cerebral blood flow constant. Regulation of cerebral blood flow predominantly occurs in parenchymal and leptomeningeal arterioles, where the precapillary sphincters dynamically change their resistance to blood flow through either contraction or dilatation (Muoio *et al.*, 2014).

Chemical, neurogenic and myogenic mechanisms are implied in cerebral autoregulation. Byproducts of metabolism, such as H^+ , K^+ , O_2 and adenosine, are vasoactive substances that cause arterial dilatation. Metabolic parenchymal substances, monoamine neurotransmitters, secreted by dorsal raphe nucleus, locus coeruleus and oral pontine reticular nucleus, also play a role regulating vessel tone (Hilz *et al.*, 2000). In addition, sympathetic noradrenergic fibers, which modulate mainly proximal large vessels tone, exert significant vasoconstrictor effects on these arteries. Parasympathetic and vasodilating impulses are of lesser influence (Hilz *et al.*, 2000). Finally, the smooth muscles of cerebral vessels constrict in response to elevated pressure and dilate in response to decreased pressure. If these mechanisms are compromised, significant brain injury occurs (Cipolla, 2009).

Cerebrovascular injury to the brain happens when the vasculature becomes diseased, contributing to cognitive impairment and neurodegeneration. This type of cerebrovascular injury may be easily detected by magnetic resonance imaging (MRI) (Smith *et al.*, 2017). Most cerebral infarcts are actually clinically unrecognized. As silent cerebral infarcts occur over time, cognitive impairment worsens and the risk for dementia increases. White matter lesions of vascular origin, microbleeds, dilated perivascular spaces and atrophy represents the broad neuroimaging spectrum of cerebrovascular disease found on brain MRI (Figure 1) (Smith *et al.*, 2017).

Cerebral microinfarcts are thought to represent the pathological link between cerebrovascular disease and dementia and are clinically associated with stroke and cognitive decline. There are probably multiple causes for small, deep cerebral infarcts, such as cerebral small vessel disease, amyloid angiopathy, microemboli and hypoperfusion mechanisms (Van Veluw *et al.*, 2017).

Apart from small vessel disease, large vessel intracranial and extracranial atherosclerosis also co-occurs with cerebral microinfarcts (Van Veluw *et al.*, 2017). In a sub analysis of the SAMMPRIS trial, 49.5% of subjects with stroke due to intracranial stenosis also showed patterns of small vessel disease in brain imaging (Kwon *et al.*, 2016). These patients scored significantly less in the Montreal Cognitive Assessment (MoCA) examination, confirming the association between intracranial stenosis, microinfarcts and cognitive decline (Kwon *et al.*, 2016). As for extracranial atherosclerosis, in 2017 a prospective study demonstrated an incidence of 13.8% of cerebral microinfarcts after carotid endarterectomy. Previous abnormal neurological exam and the necessity of shunting during surgery were the main risk factors for small cerebral infarctions (Gwon *et al.*, 2017).

In patients with small vessel disease and microinfarcts, cognitive decline occurs independently from Alzheimer's pathology. Executive domains and language functions are most commonly affected (Van Veluw *et al.*, 2017) and these dysfunctions are thought to be secondary to disruption of structural brain connections. It is estimated that a 0.2mm³ lesion may cause pathological alterations at least 12-fold greater than the volume of the lesion core (Summers *et al.*, 2017). Perilesional alterations include astrogliosis, reduced expression of aquaporin-4 and demyelination. Small-vessel disease also leads to dysfunction of the glymphatic system, responsible for the clearance of beta-amyloid peptide aggregates (Van Veluw *et al.*, 2017).

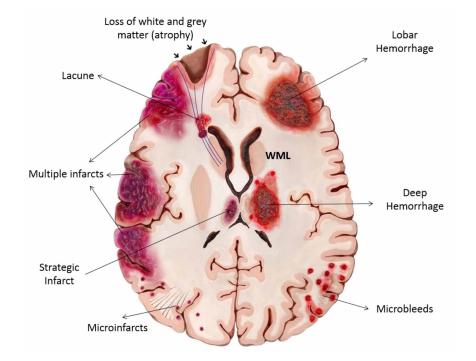


Figure 1. Figure 1. Lesions of vascular origin on brain pathology, WML = White matter lesions.

STROKE

Stroke is the second most common cause of death and the most frequent cause of disability in adulthood worldwide (Obviagele *et al.*, 2013). According to the World Stroke Organization, 1 out of 6 people will suffer a stroke in their lifetime. Stroke diagnosis is essentially clinical, based on the sudden onset of a focal neurologic deficit, leading to motor and/or sensory disabilities. It is primarily classified as ischemic stroke (IS), which represents 87% of all cases, and intracranial hemorrhage (ICH), completing the remaining 13% (Roger *et al.*, 2011).

Ischemic Stroke

Further categorization of subtypes of ischemic stroke (IS) is important to estimate the odds of permanent disability, as well as mortality and recurrence rates, as determining the cause of stroke directly influences on the choices for management and secondary prevention. According to the TOAST classification, IS can be divided into large-vessel atherosclerosis, cardioembolism, small-vessel occlusion, other etiologies and stroke of undetermined etiology (Adams *et al.*, 1993).

The prevalence of risk factors in patients with IS varies according to subtype and the population studied. Hypertension, smoking habit, history of diabetes mellitus, waist-to-hip ratio, alcohol intake, cardiac causes and psychosocial issues are risk factors for stroke consistent in all studied populations (O'Donnell, M.J. et al., 2016).

For the acute phase treatment, intravenous administration of recombinant tissue-type plasminogen activator (rtPA) is associated with significantly less disability for patients treated within a treatment window of 4.5 hours since the last time they were known to be well (asymptomatic) (Hacke *et al.*, 2008). Among patients with occlusions of intracranial large vessels, endovascular mechanical thrombectomy results in higher rates of reperfusion than intravenous t-PA alone (Church, E.W. *et al.*, 2017). Unfortunately, reported intravenous rT-PA treatment rates are still low, ranging from 3.4 to 9.1% for patients with IS (Scholten *et al.*, 2013).

In the Framingham Heart Study (Romero and Wolf, 2013), among survivors of IS, follow-up at six months after stroke showed that 26% were dependent for activities of daily living (ADL), 50% had reduced mobility or hemiparesis, 19% had aphasia and 35% exhibited signs of depression (Ma V.Y. *et al.*, 2014).

Hemorrhagic Stroke

Spontaneous ICH, the second most common subtype of stroke, is defined as non-traumatic bleeding into the brain tissue, which may extend into the ventricles and subarachnoid space (de Oliveira Manoel *et al.*, 2016). It affects more than 5 million people worldwide annually (Krishnamurthi *et al.*, 2013) and its most important modifiable risk factor is chronic arterial hypertension (Ariesen *et al.*, 2003). ICH survivors are more frequently left with severe disability, when compared to IS patients. In some series, less than 40% of patients regained functional independence (Qureshi *et al.*, 2009) and death rates at 1 month after ICH may be as high as 40% (van Asch *et al.*, 2010).

Cerebral Venous Thrombosis

Cerebral venous thrombosis (CVT) is not as common as ischemic stroke or intracerebral hemorrhage, representing about 0.5% to 1% of all strokes (Stam, 2005). It affects mainly adults in their third and fourth decades, with an estimated incidence of 4 cases per million of the population (Algahtani and Aldarmahi, 2014).

Clinically, CVT may present as isolated headache, symptoms of elevated intracranial pressure, seizures, focal lobar syndrome or decreased level of consciousness. The main risk factor is an underlying hypercoagulable state, either genetic (hereditary thrombophilias) or acquired (oral contraceptives, pregnancy and puerperium, smoking habits, infection - especially mastoiditis -, underlying malignancy, and head injury - skull fractures extending to a dural venous sinus) (Algahtani and Aldarmahi, 2014).

Treatment is based on temporary anticoagulation and long-term prognosis is usually favorable. Nevertheless, about 15% of patients remain dependent or die, particularly if there is a diagnostic delay (Ferro *et al.*, 2009).

GENETIC SUSCEPTIBILITY TO CEREBROVASCULAR DISEASE

Cerebral Autosomal Dominant Arteriopathy With Subcortical Infarcts and Leukoencephalopathy

Cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy (CADA-SIL) is an autosomal dominant, inherited, small artery disease which causes a combination of cognitive

impairment, migraine with aura, lacunar ischemic infarction and mood alterations. The pathology results from *NOTCH3* gene mutations, an exon located on chromosome 19p13.2 – p13.1 (Bersano *et al.*, 2017).

CADASIL has an estimated prevalence of 10.7 per 100.000 individuals and is now considered the most common cause of inherited stroke and vascular dementia in adults. Ischemic strokes usually occurs between 30 and 66 years of age (medium age of 49 to 57 years) (Majersik, 2017; Moreton *et al.*, 2014).

The *NOTCH3* gene encodes for a 2321-amino-acid-long single pass transmembrane receptor protein (Joutel *et al.*, 1996) and is mostly expressed in the vascular smooth muscle cells of small arteries and pericytes of the brain (Joutel *et al.*, 2010). This receptor is composed by a large extracellular domain containing 34 tandem epidermal growth factor like repeats, as well as transmembrane and intracellular domains. THE EGF-like domains are formed by 6 cysteine residues, which form 3 disulfide bridges that stabilize the EGF-like domain structure (Smith *et al.*, 2008). There are more than 150 documented *NOTCH3* gene dominant mutations that can cause CADASIL and most of them are missense mutations. These mutations result is alteration of the number of cysteine residues in one of the 34 EGF-like domains, which causes tottering disulfide bridge formation in the mutant extracellular domain, with increased multimerization properties. These alterations may impair arterial differentiation, maturation of muscle cells and also blood pressure regulation. Nonetheless, the way this correlates with neurodegeneration and dementia remains uncertain (Smith *et al.*, 2008).

As for clinical features, migraine is found in 70% of patients, typically with aura (Eikermann-Haerter *et al.*, 2011). Stroke or transitory ischemic attack (TIA) happens in 61%, commonly in the absence of typical cerebrovascular risk factors (Singhal *et al.*, 2004). Cognitive impairment is observed in 48%, with prominent reduction in executive function and processing speed (Chabriat et al., 2016), and psychiatric disorders are seen in 47% (Eikermann-Haerter *et al.*, 2011).

Diagnosis of CADASIL is based on clinical history and complementary diagnostic tests, such as MRI, skin biopsy (with accumulation of granular osmophilic material – GOM) and genetic tests of *NOTCH3* gene exons, the latter being considered as the gold-standard diagnostic tool (Peters *et al.*, 2005; Joutel *et al.*, 1997).

No specific treatment has been established yet, so treatment is focused on symptoms' control and management of vascular risk factors (Di Donato et al., 2017).

Homocysteine

Serum elevation of homocysteine is an independent risk factor for either cerebrovascular, coronary and peripheral arterial occlusive disease or peripheral venous thrombosis (Fridman, 1999).

Homocysteine is an amino acid formed by the demethylation of methionine. In healthy individuals, it is catalyzed to cystathionine by cystathionine beta-synthase, one of the pyridoxal phosphate-dependent enzymes. Following that, homocysteine is remethylated to methionine by methionine synthase, a cyanocobalamin-dependent enzyme. Therefore, nutritional conditions such as vitamin B12, vitamin B6 or folate deficiencies may contribute to the increasing of plasma homocysteine levels (Ganguly and Alam, 2015).

Elevation of plasmatic homocysteine causes atherosclerosis by several mechanisms: not only it promotes direct damage to the endothelium, but also stimulates proliferation of smooth muscle cells, enhances peroxidation of low-density lipoprotein and increases platelet aggregation, affecting the co-agulation system (Ganguly and Alam, 2015).

As a result of the disruption of the BBB, after a stroke for example, the brain is exposed to constituents of plasma, including homocysteine (Sharma *et al.*, 2015). This molecule is also a direct neurotoxic agent, acting through the overstimulation of N-methyl-D-aspartate (NMDA) receptors. The excessive stimulation of NMDA receptors allows high levels of calcium to enter the nerve cells, a pathological process known as excitotoxicity. This contributes to neuronal damage and neurodegeneration (Ganguly and Alam, 2015; Sharma *et al.*, 2015).

VASCULAR COGNITIVE IMPAIRMENT

The ways that vascular brain injury contributes to cognitive impairment depend on the patient and the nature of the vascular insult itself (Figure 2). Location of lesions within networks with an important role in cognition, and also size and number of lesions are important determinants of the type and severity of cognitive impairment (Alexander *et al.*, 1986).

The co-occurrence of vascular and neurodegenerative pathologies, such as AD (amyloidopathy and tauopathy) and Lewy body disease (synucleinopathy) occurs rather frequently, especially with increasing age (Schneider *et al.*, 2007; Arvanitakis *et al.*, 2011; Soontornniyomkij *et al.*, 2010). Sufficient evidence suggests that neurodegenerative lesions and ischemic infarcts combine to increase the risk of cognitive decline and dementia (Schineider *et al.*, 2004; White, 2009; Troncoso *et al.*, 2008; Toledo *et al.*, 2013).

Cognitive decline and behavioral changes may reveal vascular cognitive impairment even in the absence of acute symptoms like stroke; in fact, this is possibly by far the most frequent situation (Godefroy, 2013). These "silent infarcts" constitute a major cognitive risk factor. Thus, diagnosis of VCI may be very difficult and needs to be evaluated in two different settings: the post-stroke outpatient and non-stroke patients evaluated for cognitive complaints (Godefroy *et al.*, 2013; Moorhouse and Rockwood, 2013).

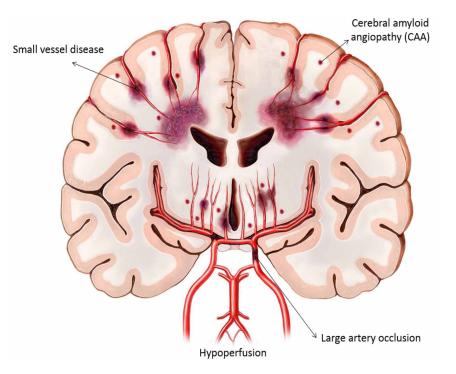


Figure 2. Mechanisms underlying vascular cognitive impairment.

Epidemiology

Previous meta-analysis and systematic reviews have shown that the prevalence of major post-stroke cognitive impairment ranges from 7% to 67.3%, depending on the study setting (hospital or populationbased studies), type of stroke (ischemic or hemorrhagic), frequency of pre-stroke dementia, frequency of recurrent stroke and the post-stroke interval (Middleton *et al.*, 2014). Compared with normal controls, previous stroke doubles the risk of subsequent dementia over 10 years (Ivan *et al.*, 2004). History of diabetes and high systolic blood pressure are associated with a greater risk of dementia in late life, especially among those with the apolipoprotein £4 (APOE £4) allele. The relationship between APOE £4 and diabetes as an increased risk factor for dementia seems to rely on an increased risk of cerebral amyloid angiopathy formation (Peila *et al.*, 2002; Elias *et al.*, 2005).

Clinical Subtypes of Vascular Cognitive Impairment

Poststroke Dementia

Cognitive dysfunction is among the most common and severe consequences of stroke, considered by patients and caregivers one of the top 10 priorities in post-stroke long-term care (Pantoni, 2017; Pollock *et al.*, 2014).

Short-term incidence of post-stroke dementia may be as high as 40% (Pendlebury and Rothwell, 2009), while mild cognitive impairment (MCI) might be observed in as much as 80% of patients (Popovic *et al.*, 2007). Long-term incidence of dementia starting from 3 months after stroke has been estimated to be 3% to 6% per year, with slightly lower rates reported for patients with TIA and minor stroke (Pendlebury and Rothwell, 2009).

Risk factors for cognitive impairment at the time of stroke include older age, lower education levels, diabetes mellitus, atrial fibrillation, smoking, female sex and pre-stroke cognitive decline (Mandzia *et al.*, 2016; Allan *et al.*, 2011; Pendlebury, 2009; Yang *et al.*, 2015). As for those related to stroke itself, the most important predictors are left hemisphere stroke, stroke severity, early seizures, hypoxic ischemic episodes and hypotension. Brain imaging predictors of post-stroke dementia include silent brain injury, white matter hyperintensities and medial temporal lobe atrophy (Yang *et al.*, 2015; Mandzia *et al.*, 2016).

Liu and colleagues, in a recent study with 72 persons with cognitive impairment after stroke/TIA, found that those with positive amyloid- β deposition after the index event, as detected by positron emission tomography (PET) imaging with C-labeled Pittsburgh Compound-B (C-PIB), experienced a more severe and rapid cognitive decline on cognition evaluation by the Mini Mental State Examination (MMSE) and the Montreal Cognitive Assessment (MoCA) during a 3-year follow-up as compared with those with negative amyloid PET scans (Mandzia *et al.*, 2016).

Data on the cognitive profile of post-stroke dementia and post-stroke MCI is somewhat limited. The most pronounced and early abnormalities are deficits in executive function and attention, but other cognitive domains (orientation, memory and language) are also affected, particularly in those with more severe cognitive impairment (Stephens *et al.*, 2004). Therefore, recent secondary stroke prevention trials are including cognitive endpoints to their study protocols, such as the Cambrigde Assessment Mental Disorder in the Elderly, section B (CAMCOG-R), a standardized test (maximum score of 107), encompassing memory, orientation, language, comprehension, praxis, executive function and calculation (Diener *et al.*, 2008; Pearce *et al.*, 2014).

Strategic Infarct Dementia

Dementia can also occur after a single strategic infarct. The left angular gyrus, the inferomesial temporal gyrus and the medial frontal lobe are considered strategic regions perfused by large arteries. When it comes to executive functions, the frontal-subcortical loops are considered strategic networks located in the prefrontal cortex, the dorsomedial and the anterior thalamic nuclei, the head of the caudate, the anterior limb of the internal capsule and the capsular genu (Cummings *et al.*, 1993; Duering M *et al.*, 2011).

Subcortical Vascular Dementia

By far, the most common cause of vascular cognitive impairment is subcortical vascular dementia (SVD), which typically manifests itself with white matter lesions, termed white matter hyperintensities (WMH) on MRI and lacunes, mainly confined to subcortical white and grey matter (Vermeer *et al.*, 2002).

In the general population, prevalence rates for WMH rise from 50% around 45 years of age to 95% for people in their eighties (Wen *et al.*, 2009; de Leeuw *et al.*, 2001). Small brain infarcts are also common, and like WMH, they have been shown to be associated with cognitive deficits and dementia (Vermeer *et al.*, 2007; Debette *et al.*, 2010).

In addition to post-stroke dementia and the strategic location infarct dementia caused by small vessel disease, subcortical arteriosclerotic encephalopathy and lacunar state are included in the subcortical vascular dementia (Dozono *et al.*, 1991).

Lacunar state is a striking phenotype of SVD, characterized by multiple lacunar infarcts in thalamus, the basal ganglia, and white matter. Clinical features comprise abrupt onset hemiparesis, lack of volition, dysarthria, akinetic mutism, pseudo-bulbar palsy and affect, urinary incontinence and small-stepped gait (Dozono *et al.*, 1991).

The consequences of SVD may extend into the cortex, manifesting both as microscopic vascular lesions and cortical atrophy and now cortical changes are considered a clinically relevant component of SVD (Duering *et al.*, 2015; Peres *et al.*, 2016; Gouw *et al.*, 2011; Smith *et al.*, 2012).

Subcortical arteriosclerotic encephalopathy, the so-called Binswanger's syndrome, is characterized by demyelination of deep white matter, attributed to stenosis of deep penetrating arteries (Roman, 1987; Bennett *et al.*, 1990). The typical clinical triad is comprised of a slowly progressive dementia, urinary incontinence and gait apraxia. Binswanger's disease may easily be misdiagnosed as normal pressure hydrocephalus because of similar clinical findings (Erkinjuntti *et al.*, 2000).

Clinical criteria for SVD include executive dysfunction, cognitive slowing, extrapyramidal signs, depression and gait disturbances (Erkinjuntti *et al.*, 2000).

Multi-Infarct Dementia

The presence of multiple infarcts has been recognized as a cause of dementia (Hachinski *et al.*, 1974), and both the number and the volume of the lesions are related with impaired cognitive performance and higher risk of dementia (White *et al.*, 2005; Sonnen *et al.*, 2007; Vemuri *et al.*, 2015). There are no clear criteria for what should be considered a threshold of overall volume of brain lesions required for the occurrence of the VCI or vascular dementia. Other than that, countless factors need to be taken into account to this definition. First and foremost, some brain regions (pre-frontal cortex, for example) are more eloquent to cognitive functions than others; second, many patients have associated comorbidities

such as AD; and finally, there are individual variations in the ability to compensate for both vascular and neurodegenerative pathologies (Zieren *et al.*, 2013; Cordonnier *et al.*, 2010).

Silent Vascular Brain Injury

The prevalence of silent brain injuries is up to 5 times greater than that of symptomatic stroke and, as shown in the Rotterdam study, they are common in elderly people (Vermeer et al, 2002). It increases progressively with age and is even higher in subjects with risk factors for vascular disease (Vermeer *et al.*, 2007); most silent infarcts are related to SVD. The Rotterdam scan study concluded that the presence of silent brain injury doubled the risk of dementia, and also led to worse results in the performance of cognitive testing and a steeper decline in the cognition (Vermeer *et al.*, 2003). Non-thalamic infarcts were related to decreased psychomotor speed, whereas thalamic infarcts were related with worsening of memory performance. Recent studies have shown similar results (Debette *et al.*, 2010).

Microinfarcts: Imperceptible Lesions

Cerebral microinfarcts are ischemic injuries almost exclusively detected through brain biopsy, typically smaller than 1 mm (Sonnen *et al.*, 2007; Vemuri *et al.*, 2015). Usually attributable to SVD, they could also be related to vasoconstriction, microemboli and cerebral hypoperfusion. The presence of 1 or 2 microinfarcts in a cerebral biopsy specimen corresponds with the presence of hundreds of microinfarcts throughout the brain (Cordonnier *et al.*, 2010). However, this type of injury is also common in patients with AD, as wells as otherwise healthy people. Additionally, some cohorts have shown that the presence of microinfarcts increases the risk of dementia (Arvanitakis *et al.*, 2011). The cognitive profile of patients with microinfarcts is not well established, but it may be associated with disturbances in semantic and episodic memory, and perceptual speed (Arvanitakis *et al.*, 2011).

Microbleeds and Cortical Superficial Siderosis

Cerebral microbleeds (MBs) are round hypointensities, smaller than 10 mm in diameter, apparent on gradient echo T2*-weighted scan MRI (Cordonnier *et al.*, 2010). MBs are usually considered in the presence of SVD caused by either cerebral amyloid angiopathy (CAA) or hypertensive vasculopathy (Yates *et al.*, 2014). Strong evidence suggests that MBs discontinue structural connectivity and, consequently, network function (Reijmer *et al.*, 2015). Similar to strategic infarcts, the location of MBs is of the utmost importance, as lobar and deep MBs are associated with hypertensive vasculopathy and only lobar MBs are related with CAA (Yates *et al.*, 2014).

Cortical superficial siderosis (cSS) is linear deposits of blood products within the leptomeninges, subarachnoid space and superficial cortical layers. This condition is closely connected to CAA (Linn *et al.*, 2010). The prevalence of cSS in patients evaluated in memory clinics ranges from 2% to 6%, whereas in non-demented elderly people it is estimated to be around 0.5%. (Zonneveld *et al.*, 2014; Wotlenweber *et al.*, 2014; Na *et al.*, 2015). There is some association between cSS and the apolipoprotein £4 allele, confirming the relation between cSS and CAA (Keage *et al.*, 2009). Despite that, the process by which cSS affects cognition still needs to be better understood.

Cerebral Amyloid Angiopathy

Cerebral amyloid angiopathy (CAA) is a consequence of the accumulation of β -amyloid in the media and adventitia layers of small and mid-sized arteries of the brain. It is an important cause of lobar hemorrhage and dementia in the elderly (Charidimou *et al.*, 2012).

The incidence of CAA is highly age-dependent; very rarely presenting before 50 years of age, whereas in a study with 784 autopsies, 12% of people over 85 years old exhibited pathological features of CAA (Greenberg *et al.*, 1997). There is a significant association with AD, and in these patients the frequency of CAA may reach 98% (Keage *et al.*, 2009).

Although asymptomatic in most cases, CAA might result in dementia, ICH and transient neurologic spells. ICH is the most recognized result of CAA, and this diagnosis should always be suspected in patients above 55 years with consecutive cortical bleedings (Passero *et al.*, 1995). It is known that CAA contributes to cognitive impairment and dementia - moderate to severe CAA was related to lower performances in some specific cognitive domains, mainly perceptual speed and episodic memory (Boyle *et al.*, 2015).

Although definitive diagnosis can only be made with full postmortem examination, according to the Boston Criteria (Knudsen *et al.*, 2001), CAA has typical neuroradiological findings that lead to *in vivo* diagnosis. Gradient echo T2*-weighted brain MRI allows the detection of cerebral MB, typically found in the cortical-subcortical transition over the posterior regions of the brain (Pendlebury *et al.*, 2009). White matter changes, convexity subarachnoid hemorrhage, cSS and silent acute ischemic lesions also represent the spectrum of hemorrhagic and ischemic manifestations of CAA seen on brain MRI (Pendlebury *et al.*, 2009).

Currently, there are no specific disease-modifying agents available for treating CAA. In subjects with previous ICH, the adequate management of hypertension is a wise strategy to avoid a new episode of bleeding, as patients with ICH have an annual recurrence risk of up to 10%, and anticoagulant therapy appears to increase this risk by 7 to 10 fold (Izumihara *et al.*, 1999).

Hypoperfusion Dementia

Reduction of cerebral perfusion can lead to transient or permanent ischemia, and consequently to cognitive decline. The RECON trial (Randomized Evaluation of Carotid Occlusion and Neurocognition) showed that hemodynamic failure was associated with cognitive impairment (Marshall *et al.*, 2012). Histopathologically, global hypoperfusion induces border zone infarcts, hippocampal sclerosis and cortical laminar necrosis contributing to cognitive impairment (Jellinger KA *et al.*, 2013).

Mixed Dementia

Many patients with dementia have a high prevalence of mixed disease with vascular and AD pathology contributions (Schneider *et al.*, 2007). A longitudinal cohort study (Rush Memory and Aging Project) found that the most cases of dementia in community-dwelling older persons were of the mixed type, due to a combination of vascular and AD-type pathology (Schneider Ja *et al.*, 2004). Subjects with multiple pathologies had a 3-times higher chance of developing dementia in their life. In most cases, it is difficult to say how these multiples pathologies contribute individually to cognitive decline, but it is assumed that vascular brain injuries lower the threshold of AD-pathology required to induce dementia. On the other

hand, AD-like pathology increases the chance of dementia after stroke (Toledo *et al.*, 2013; Mathews FE *et al.*, 2009). The relationship between AD-like and vascular pathology remains quite complex considering its additive and/or multiplying effect (Launer *et al.*, 2011; Chui *et al.*, 2015).

Atrophy of the hippocampus and amnesia are consistent with early AD, while the cognitive profile of vascular cognitive impairment is widely variable and depends on size and region of vascular lesions. Mostly, the cognitive profile of mixed dementia is dominated by that found in isolated AD.¹⁴⁹ Emerging amyloid PET scans findings could help to differentiate between pure dementia from mixed dementia (Heiss *et al.*, 2016).

TREATMENT

The general principles for management of VCI, MCI and dementia are basically the same and encompass the treatment of comorbidities, adequate control of the risk factors, treatment of psychological and the behavioral symptoms, providing support to the patients themselves and their caregivers, with the goal of maximizing independence for ADLs. Trials with acetylcholinesterase inhibitors showed at best only modest effects on global functioning, and discreet improvements in cognition. Despite the small effects on cognition, it is reasonable to consider using donepezil in patients with vascular dementia (Pase *et al.,* 2017; O'Brien and Thomas, 2015).

PREVENTION

Prevention includes control of the lifestyle risk factors (such as lower educational level, Western diet, sedentary lifestyle, smoking habit and obesity) and can be separated in primary prevention with early identification and reduction of risk factors for atherosclerosis, such as arterial hypertension, type 2 diabetes mellitus and dyslipidemia; and secondary stroke prevention (Pase *et al.*, 2017).

FUTURE RESEARCH DIRECTIONS

Currently, the most effective strategy to eliminate the vascular contributions to neurodegeneration is the aggressive control of vascular risk factors. Primary prevention politics, early diagnosis and adequate long-term care of stroke victims are important ways to prevent and/or reduce the morbidity associated with neurodegenerative disorders (Pase *et al.*, 2017).

In 2016, the Framingham Heart Study demonstrated a progressive decrease in the incidence of dementia (Pase *et al.*, 2017). This could be explained by increasing scholarity, better cardiovascular health and better management of stroke risk factors. Therefore, there is great hope that the vascular contributions to cognitive impairment can be prevented and treated. For therapeutic development, the most success has been achieved with symptomatic treatments. The scientific community is still hoping for a disease modifying therapy, which should probably aim to improve neurovascular unit function or enhance cerebral resilience to injury (Smith *et al.*, 2017).

CONCLUSION

There is a close interaction between stroke and neurodegenerative disorders, predominantly AD. From the disruption of the vascular unit to neurological progressive deterioration, there is a broad spectrum of pathologies, including white matter brain abnormalities, transient ischemic attacks, symptomatic stroke and vascular cognitive impairment, that contribute to varying degrees. However, further studies are still needed to uncover the link between cerebrovascular disease and neurodegeneration in all its aspects.

REFERENCES

Adams, H. P. Jr, Bendixen, B. H., Kappelle, L. J., Biller, J., Love, B. B., Gordon, D. L., & Marsh, E. E. III. (1993). Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. TOAST. Trial of Org 10172 in Acute Stroke Treatment. *Stroke*, *24*(1), 35–41. doi:10.1161/01. STR.24.1.35 PMID:7678184

Alexander, G. E., DeLong, M. R., & Strick, P. L. (1986). Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annual Review of Neuroscience*, *9*(1), 357–381. doi:10.1146/ annurev.ne.09.030186.002041 PMID:3085570

Algahtani, H. A., & Aldarmahi, A. A. (2014). Cerebral venous sinus thrombosis. *Neurosciences (Riyadh)*, *19*(1), 11–16. PMID:24419443

Allan, L. M., Rowan, E. N., Firbank, M. J., Thomas, A. J., Parry, S. W., Polvikoski, T. M., ... Kalaria, R. N. (2011). Long term incidence of dementia, predictors of mortality and pathological diagnosis in older stroke survivors. *Brain*, *134*(12), 3716–3727. doi:10.1093/brain/awr273 PMID:22171356

Ariesen, M. J., Claus, S. P., Rinkel, G. J., & Algra, A. (2003). Risk factors for intracerebral hemorrhage in the general population: A systematic review. *Stroke*, *34*(8), 2060–2065. doi:10.1161/01. STR.0000080678.09344.8D PMID:12843354

Arvanitakis, Z., Leurgans, S. E., Barnes, L. L., Bennett, D. A., & Schneider, J. A. (2014). Microinfarct pathology, dementia, and cognitive systems. *Stroke*, 42(3), 722–727. doi:10.1161/STROKEAHA.110.595082 PMID:21212395

Bennett, D. A., Wilson, R. S., Gilley, D. W., & Fox, J. H. (1990). Clinical diagnosis of Binswanger's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, *53*(11), 961–965. doi:10.1136/jnnp.53.11.961 PMID:2283526

Bersano, A., Bedini, G., Oskam, J., Mariotti, C., Taroni, F., Baratta, S., & Parati, E. A. (2017). CA-DASIL: Treatment and Management Options. *Current Treatment Options in Neurology*, *19*(9), 31. doi:10.100711940-017-0468-z PMID:28741120

Bowler, J., & Hachinsky, V. (1996). History of the concept of vascular dementia: two opposing views on current definitions and criteria for vascular dementia. *Vascular dementia: current concepts*, 1-24.

Boyle, P. A., Yu, L., Nag, S., Leurgans, S., Wilson, R. S., Bennett, D. A., & Schneider, J. A. (2015). Cerebral amyloid angiopathy and cognitive outcomes in community-based older persons. *Neurology*, *85*(22), 1930–1936. doi:10.1212/WNL.00000000002175 PMID:26537052

Caixeta, L., Costa, J. N., Vilela, A. C., & Nóbrega, M. D. (2014). The development of the dementia concept in 19th century. *Arquivos de Neuro-Psiquiatria*, 72(7), 564–567. doi:10.1590/0004-282X20140069 PMID:25054992

Chabriat, H., Hervé, D., Duering, M., Godin, O., Jouvent, E., Opherk, C., ... Dichgans, M. (2016). Predictors of clinical worsening in cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy: Prospective cohort study. *Stroke*, *47*(1), 4–11. doi:10.1161/STROKEAHA.115.010696 PMID:26578659

Charidimou, A., Gang, Q., & Werring, D. J. (2012). Sporadic cerebral amyloid angiopathy revisited: Recent insights into pathophysiology and clinical spectrum. *Journal of Neurology, Neurosurgery, and Psychiatry*, 83(2), 124–137. doi:10.1136/jnnp-2011-301308 PMID:22056963

Chui, H. C., & Ramirez-Gomez, L. (2015). Clinical and imaging features of mixed Alzheimer and vascular pathologies. *Alzheimer's Research & Therapy*, 7(1), 21. doi:10.118613195-015-0104-7 PMID:25722748

Church, E. W., Gundersen, A., Glantz, M. J., & Simon, S. D. (2017). Number needed to treat for stroke thrombectomy based on a systematic review and meta-analysis. *Clinical Neurology and Neurosurgery*, *156*, 83–88. doi:10.1016/j.clineuro.2017.03.005 PMID:28359980

Cipolla, M. J. (2009). The Cerebral Circulation. Morgan & Claypool Life Sciences.

Cordonnier, C., Leys, D., Dumont, F., Deramecourt, V., Bordet, R., Pasquier, F., & Hénon, H. (2010). What are the causes of pre-existing dementia in patients with intracerebral haemorrhages? *Brain*, *133*(11), 3281–3289. doi:10.1093/brain/awq246 PMID:20852266

Cummings, J. L. (1993). Frontal-subcortical circuits and human behavior. *Archives of Neurology*, *50*(8), 873–880. doi:10.1001/archneur.1993.00540080076020 PMID:8352676

Daneman, R., & Prat, A. (2015). The blood-brain barrier. *Cold Spring Harbor Perspectives in Biology*, 7(1), a020412. doi:10.1101/cshperspect.a020412 PMID:25561720

De Leeuw, F. E., de Groot, J. C., Achten, E., Oudkerk, M., Ramos, L. M., Heijboer, R., ... Breteler, M. M. (2001). Prevalence of cerebral white matter lesions in elderly people: A population based magnetic resonance imaging study. The Rotterdam Scan Study. *Journal of Neurology, Neurosurgery, and Psychiatry*, *70*(1), 9–14. doi:10.1136/jnnp.70.1.9 PMID:11118240

de Oliveira Manoel, A. L., Goffi, A., Zampieri, F. G., Turkel-Parrella, D., Duggal, A., Marotta, T. R., ... Abrahamson, S. (2016). The critical care management of spontaneous intracranial hemorrhage: A contemporary review. *Critical Care (London, England)*, *20*(1), 272. doi:10.118613054-016-1432-0 PMID:27640182

Debette, S., Beiser, A., DeCarli, C., Au, R., Himali, J. J., Kelly-Hayes, M., ... Seshadri, S. (2010). Association of MRI markers of vascular brain injury with incident stroke, mild cognitive impairment, dementia, and mortality: The Framingham Offspring Study. *Stroke*, *41*(4), 600–606. doi:10.1161/STROKEAHA.109.570044 PMID:20167919

Debette, S., & Markus, H. S. (2010). The clinical importance of white matter hyperintensities on brain magnetic resonance imaging: Systematic review and meta-analysis. *BMJ (Clinical Research Ed.)*, 341(jul26 1), c3666. doi:10.1136/bmj.c3666 PMID:20660506

Di Donato, I., Bianchi, S., De Stefano, N., Dichgans, M., Dotti, M. T., Duering, M., ... Federico, A. (2017). Cerebral Autosomal Dominant Arteriopathy with Subcortical Infarcts and Leukoencephalopathy (CADASIL) as a model of small vessel disease: Update on clinical, diagnostic, and management aspects. *BMC Medicine*, *15*(1), 41. doi:10.118612916-017-0778-8 PMID:28231783

Diener, H. C., Sacco, R. L., Yusuf, S., Cotton, D., Ounpuu, S., Lawton, W. A., ... Yoon, B. W. (2008). Prevention Regimen for Effectively Avoiding Second Strokes (PRoFESS) study group. Effects of aspirin plus extended-release dipyridamole versus clopidogrel and telmisartan on disability and cognitive function after recurrent stroke in patients with ischaemic stroke in the Prevention Regimen for Effectively Avoiding Second Strokes (PRoFESS) trial: A double-blind, active and placebo-controlled study. *Lancet Neurology*, *7*(10), 875–884. doi:10.1016/S1474-4422(08)70198-4 PMID:18757238

Dozono, K., Ishii, N., Nishihara, Y., & Horie, A. (1991). An autopsy study of the incidence of lacunes in relation to age, hypertension, and arteriosclerosis. *Stroke*, *22*(8), 993–996. doi:10.1161/01.STR.22.8.993 PMID:1866767

Duering, M., Righart, R., Wollenweber, F. A., Zietemann, V., Gesierich, B., & Dichgans, M. (2015). Acute infarcts cause focal thinning in remote cortex via degeneration of connecting fiber tracts. *Neurology*, *84*(4), 1685–1692. doi:10.1212/WNL.000000000001502 PMID:25809303

Duering, M., Zieren, N., Hervé, D., Jouvent, E., Reyes, S., Peters, N., ... Dichgans, M. (2011). Strategic role of frontal white matter tracts in vascular cognitive impairment: A voxel-based lesion-symptom mapping study in CADASIL. *Brain*, *134*(8), 2366–2375. doi:10.1093/brain/awr169 PMID:21764819

Eikermann-Haerter, K., Yuzawa, I., Dilekoz, E., Joutel, A., Moskowitz, M. A., & Ayata, C. (2011). Cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy syndrome mutations increase susceptibility to spreading depression. *Annals of Neurology*, *69*(2), 413–418. doi:10.1002/ana.22281 PMID:21387384

Elias, M. F., Elias, P. K., Sullivan, L. M., Wolf, P. A., & D'Agostino, R. B. (2005). Obesity, diabetes and cognitive deficit: The Framingham Heart Study. *Neurobiology of Aging*, *26*(1), 11–16. doi:10.1016/j. neurobiolaging.2005.08.019 PMID:16223549

Elkins, J. S., Longstreth, W. T. Jr, Manolio, T. A., Newman, A. B., Bhadelia, R. A., & Johnston, S. C. (2006). Education and the cognitive decline associated with MRI-defined brain infarct. *Neurology*, 67(3), 435–440. doi:10.1212/01.wnl.0000228246.89109.98 PMID:16894104

Erkinjuntti, T., Inzitari, D., Pantoni, L., Wallin, A., Scheltens, P., Rockwood, K., ... Desmond, D. W. (2000). Research criteria for subcortical vascular dementia in clinical trials. *Journal of Neural Transmission (Vienna, Austria)*, *59*, 23–30. PMID:10961414

Ferro, J. M., Canhão, P., Stam, J., Bousser, M. G., Barinagarrementeria, F., Massaro, A., ... Kasner, S. E. (2009). Delay in the diagnosis of cerebral vein and dural sinus thrombosis: Influence on outcome. *Stroke*, *40*(9), 3133–3138. doi:10.1161/STROKEAHA.109.553891 PMID:19608994

Fridman, O. (1999). Hyperhomocysteinemia: atherothrombosis and neurotoxicity. *Acta physiologica, pharmacologica et therapeutica latinoamericana: órgano de la Asociación Latinoamericana de Ciencias Fisiológicas y [de] la Asociación Latinoamericana de Farmacología, 49*(1), 21-30.

Ganguly, P., Alam, S.F. (2015) Role of homocysteine in the development of cardiovascular disease. *Nutrition Journal*, *10*.

Godefroy, O., Leclercq, C., & Roussel, M. (2013). Vascular cognitive impairment in the stroke unit and after the acute stage. In O. Godefroy (Ed.), *Behavioral and cognitive neurology of stroke* (2nd ed.; pp. 22–31). Cambridge University press. doi:10.1017/CBO9781139058988.004

Gorelick, P. B., Scuteri, A., Black, S. E., Decarli, C., Greenberg, S. M., Iadecola, C., ... Seshadri, S. (2011). Vascular contributions to cognitive impairment and dementia: A statement for healthcare professionals from the American Heart Association/ American Stroke Association. *Stroke*, *42*(9), 2672–2713. doi:10.1161/STR.0b013e3182299496 PMID:21778438

Gouw, A. A., Seewann, A., van der Flier, W. M., Barkhof, F., Rozemuller, A. M., Scheltens, P., & Geurts, J. J. (2011). Heterogeneity of small vessel disease: A systematic review of MRI and histopathology correlations. *Journal of Neurology, Neurosurgery, and Psychiatry*, 82(2), 126–135. doi:10.1136/jnnp.2009.204685 PMID:20935330

Greenberg, S. M., & Vonsattel, J. P. (1997). Diagnosis of cerebral amyloid angiopathy. Sensitivity and specificity of cortical biopsy. *Stroke*, 28(7), 1418–1422. doi:10.1161/01.STR.28.7.1418 PMID:9227694

Gwon, J. G., Kwon, T. W., Cho, Y. P., Kang, D. W., Han, Y., & Noh, M. (2017). Analysis of Risk Factors for Cerebral Microinfarcts after Carotid Endarterectomy and the Relevance of Delayed Cerebral Infarction. *Journal of Clinical Neurology (Seoul, Korea)*, *13*(1), 32–37. doi:10.3988/jcn.2017.13.1.32 PMID:27730766

Hachinski, V. C., Lassen, N. A., & Marshall, J. (1974). Multi-infarct dementia. A cause of mental deterioration in the elderly. *Lancet*, 2(7874), 207–210. doi:10.1016/S0140-6736(74)91496-2 PMID:4135618

Hacke, W., Kaste, M., Bluhmki, E., Brozman, M., Dávalos, A., Guidetti, D., ... Toni, D. (2008). Thrombolysis with alteplase 3 to 4.5 hours after acute ischemic stroke. *The New England Journal of Medicine*, *359*(13), 1317–1329. doi:10.1056/NEJMoa0804656 PMID:18815396

Heiss, W. D., Rosenberg, G. A., Thiel, A., Berlot, R., & de Reuck, J. (2016). Neuroimaging in vascular cognitive impairment: A state-of-the-art review. *BMC Medicine*, *14*(1), 174. doi:10.118612916-016-0725-0 PMID:27806705

Hilz, M. J., Stemper, B., Heckmann, J. G., & Neundörfer, B. (2000). Mechanisms of cerebral autoregulation, assessment and interpretation by means of transcranial doppler sonography. *Fortschritte der Neurologie-Psychiatrie*, 68(9), 398–412. doi:10.1055-2000-11798 PMID:11037638 Iadecola, C. (2013). The pathobiology of vascular dementia. *Neuron*, *80*(4), 844–866. doi:10.1016/j. neuron.2013.10.008 PMID:24267647

Ivan, C. S., Seshadri, S., Beiser, A., Au, R., Kase, C. S., Kelly-Hayes, M., & Wolf, P. A. (2004). Dementia after stroke: The Framingham Study. *Stroke*, *35*(6), 1264–1268. doi:10.1161/01.STR.0000127810.92616.78 PMID:15118167

Izumihara, A., Ishihara, T., Iwamoto, N., Yamashita, K., & Ito, H. (1999). Postoperative outcome of 37 patients with lobar intracerebral hemorrhage related to cerebral amyloid angiopathy. *Stroke*, *30*(1), 29–33. doi:10.1161/01.STR.30.1.29 PMID:9880384

Jellinger, K. A. (2013). Pathology and pathogenesis of vascular cognitive impairment - a critical update. *Frontiers in Aging Neuroscience*, *10*(5), 17. PMID:23596414

Joutel, A., Corpechot, C., Ducros, A., Vahedi, K., Chabriat, H., Mouton, P., ... Tournier-Lasserve, E. (1996). Notch3 mutations in CADASIL, a hereditary adult-onset condition causing stroke and dementia. *Nature*, *383*(6602), 707–710. doi:10.1038/383707a0 PMID:8878478

Joutel, A., Monet-Leprêtre, M., Gosele, C., Baron-Menguy, C., Hammes, A., Schmidt, S., ... Hubner, N. (2010). Cerebrovascular dysfunction and microcirculation rarefaction precede white matter lesions in a mouse genetic model of cerebral ischemic small vessel disease. *The Journal of Clinical Investigation*, *120*(2), 433–445. doi:10.1172/JCI39733 PMID:20071773

Joutel, A., Vahedi, K., Corpechot, C., Troesch, A., Chabriat, H., Vayssière, C., ... Tournier-Lasserve, E. (1997). Strong clustering and stereotyped nature of Notch3 mutations in CADASIL patients. *Lancet*, *350*(9090), 1511–1515. doi:10.1016/S0140-6736(97)08083-5 PMID:9388399

Keage, H.A., Carare, R.O., Friedland, R.P., Ince, P.G., Love, S., Nicoll, J.A., ... Brayne, C. (2009). Population studies of sporadic cerebral amyloid angiopathy and dementia: a systematic review. *BMC Neurology*, *13*.

Knudsen, K. A., Rosand, J., Karluk, D., & Greenberg, S. M. (2001). Clinical diagnosis of cerebral amyloid angiopathy: Validation of the Boston criteria. *Neurology*, *56*(4), 537–539. doi:10.1212/WNL.56.4.537 PMID:11222803

Krishnamurthi, R. V., Feigin, V. L., Forouzanfar, M. H., Mensah, G. A., Connor, M., Bennett, D. A., ... Murray, C. (2013). Global and regional burden of first-ever ischaemic and haemorrhagic stroke during 1990-2010: Findings from the Global Burden of Disease Study 2010. *The Lancet. Global Health*, *1*(5), e259–e281. doi:10.1016/S2214-109X(13)70089-5 PMID:25104492

Kwon, H. M., Lynn, M. J., Turan, T. N., Derdeyn, C. P., Fiorella, D., Lane, B. F., ... Chimowitz, M. I. (2016). Frequency, Risk Factors, and Outcome of Coexistent Small Vessel Disease and Intracranial Arterial Stenosis: Results From the Stenting and Aggressive Medical Management for Preventing Recurrent Stroke in Intracranial Stenosis (SAMMPRIS) Trial. *JAMA Neurology*, *73*(1), 36–42. doi:10.1001/jamaneurol.2015.3145 PMID:26618534

Launer, L. J., Hughes, T. M., & White, L. R. (2011). Microinfarcts, brain atrophy, and cognitive function: The Honolulu Asia Aging Study Autopsy Study. *Annals of Neurology*, *70*(5), 774–780. doi:10.1002/ana.22520 PMID:22162060

Linn, J., Halpin, A., Demaerel, P., Ruhland, J., Giese, A. D., Dichgans, M., ... Greenberg, S. M. (2010). Prevalence of superficial siderosis in patients with cerebral amyloid angiopathy. *Neurology*, 74(17), 1346–1350. doi:10.1212/WNL.0b013e3181dad605 PMID:20421578

Ma, V. Y., Chan, L., & Carruthers, K. J. (2014). Incidence, prevalence, costs, and impact on disability of common conditions requiring rehabilitation in the United States: Stroke, spinal cord injury, traumatic brain injury, multiple sclerosis, osteoarthritis, rheumatoid arthritis, limb loss, and back pain. *Archives of Physical Medicine and Rehabilitation*, *95*(5), 986–995. doi:10.1016/j.apmr.2013.10.032 PMID:24462839

Majersik, J. J. (2017). Inherites and Uncommon Causes of Stroke. *Continuum*, 23(1), 211–237. PMID:28157751

Mandzia, J. L., Smith, E. E., Horton, M., Hanly, P., Barber, P. A., Godzwon, C., ... Coutts, S. B. (2016). Imaging and baseline predictors of cognitive performance in minor ischemic stroke and patients with transient ischemic attack at 90 days. *Stroke*, *47*(3), 726–731. PMID:26846862

Marshall, R. S., Festa, J. R., Cheung, Y. K., Chen, R., Pavol, M. A., Derdeyn, C. P., ... Lazar, R. M. (2012). Cerebral hemodynamics and cognitive impairment: Baseline data from the RECON trial. *Neurology*, 78(4), 250–255. doi:10.1212/WNL.0b013e31824365d3 PMID:22238418

Matthews, F. E., Brayne, C., Lowe, J., McKeith, I., Wharton, S. B., & Ince, P. (2009). Epidemiological pathology of dementia: Attributable-risks at death in the Medical Research Council Cognitive Function and Ageing Study. *PLoS Medicine*, *6*(11), e1000180. doi:10.1371/journal.pmed.1000180 PMID:19901977

Middleton, L. E., Lam, B., Fahmi, H., Black, S. E., McIlroy, W. E., Stuss, D. T., ... Turner, G. R. (2014). Frequency of domain-specific cognitive impairment in sub-acute and chronic stroke. *NeuroRehabilitation*, *34*(2), 305–312. PMID:24401826

Moorhouse, P., & Rockwood, K. (2013). Vascular cognitive impairment in the memory clinic. In O. Godefroy (Ed.), *Behavioral and cognitive neurology of stroke* (2nd ed.; pp. 9–21). Cambridge University Press. doi:10.1017/CBO9781139058988.003

Moreton, F. C., Razvi, S. S., Davidson, R., & Muir, K. W. (2014). Changing clinical patterns and increasing prevalence in CADASIL. *Acta Neurologica Scandinavica*, *130*(3), 197–203. doi:10.1111/ane.12266 PMID:24840674

Muoio, V., Persson, P. B., & Sendeski, M. M. (2014). The neurovascular unit - concept review. *Acta Physiologica (Oxford, England)*, *210*(4), 790–798. doi:10.1111/apha.12250 PMID:24629161

Na, H. K., Park, J. H., Kim, J. H., Kim, H. J., Kim, S. T., Werring, D. J., ... Na, D. L. (2015). Cortical superficial siderosis: A marker of vascular amyloid in patients with cognitive impairment. *Neurology*, *84*(8), 849–855. doi:10.1212/WNL.00000000001288 PMID:25632096

O'Brien, J. T., & Thomas, A. (2015). Vascular dementia. *Lancet*, *386*(10004), 1698–1706. doi:10.1016/S0140-6736(15)00463-8 PMID:26595643

O'Donnell, M. J., Chin, S. L., Rangarajan, S., Xavier, D., Liu, L., Zhang, H., ... Yusuf, S. (2016). Global and regional effects of potentially modifiable risk factors associated with acute stroke in 32 countries (INTERSTROKE), a case-control study. *Lancet*, *388*(10046), 761–775. doi:10.1016/S0140-6736(16)30506-2 PMID:27431356

Ovbiagele, B., Goldstein, L. B., Higashida, R. T., Howard, V. J., Johnston, S. C., Khavjou, O. A., ... Trogdon, J. G. (2013). Forecasting the future of stroke in the United States: A policy statement from the American Heart Association and American Stroke Association. *Stroke*, *44*(8), 2361–2375. doi:10.1161/ STR.0b013e31829734f2 PMID:23697546

Pantoni, L. (2017). Have Stroke Neurologists Entered the Arena of Stroke-Related Cognitive Dysfunctions? Not Yet, but They Should! *Stroke*, *48*(6), 1441–1442. doi:10.1161/STROKEAHA.117.016869 PMID:28487330

Pase, M. P., Satizabal, C. L., & Seshadri, S. (2017). Role of Improved Vascular Health in the Declining Incidence of Dementia. *Stroke*, 48(7), 2013–2020. doi:10.1161/STROKEAHA.117.013369 PMID:28596460

Passero, S., Burgalassi, L., D'Andrea, P., & Battistini, N. (1995). Recurrence of bleeding in patients with primary intracerebral hemorrhage. *Stroke*, *26*(7), 1189–1192. doi:10.1161/01.STR.26.7.1189 PMID:7604411

Pearce, L. A., McClure, L. A., Anderson, D. C., Jacova, C., Sharma, M., Hart, R. G., & Benavente, O. R. (n.d.). SPS3 Investigators. Effects of long-term blood pressure lowering and dual antiplatelet treatment on cognitive function in patients with recent lacunar stroke: A secondary analysis from the SPS3 randomised trial. *Lancet Neurology*, *13*(7), 1177–1185. PMID:25453457

Peila, R., Rodriguez, B. L., & Launer, L. J. (2002). Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: The Honolulu-Asia Aging Study. *Diabetes*, *51*(4), 1256–1262. doi:10.2337/ diabetes.51.4.1256 PMID:11916953

Pendlebury, S. T. (2009). Stroke-related dementia: Rates, risk factors and implications for future research. *Maturitas*, *64*(3), 165–171. doi:10.1016/j.maturitas.2009.09.010 PMID:19818568

Pendlebury, S. T., & Rothwell, P. M. (2009). Prevalence, incidence, and factors associated with pre-stroke and post-stroke dementia: A systematic review and meta-analysis. *Lancet Neurology*, 8(11), 1006–1018. doi:10.1016/S1474-4422(09)70236-4 PMID:19782001

Peres, R., De Guio, F., Chabriat, H., & Jouvent, E. (2016). Alterations of the cerebral cortex in sporadic small vessel disease: A systematic review of in vivo MRI data. *Journal of Cerebral Blood Flow and Metabolism*, *36*(4), 681–695. doi:10.1177/0271678X15625352 PMID:26787108

Persidsky, Y., Ramirez, S. H., Haorah, J., & Kanmogne, G. D. (2006). Blood-brain barrier: Structural components and function under physiologic and pathologic conditions. *Journal of Neuroimmune Pharmacology*, *1*(3), 223–236. doi:10.100711481-006-9025-3 PMID:18040800

Peters, N., Opherk, C., Bergmann, T., Castro, M., Herzog, J., & Dichgans, M. (2005). Spectrum of mutations in biopsy-proven CADASIL: Implications for diagnostic strategies. *Archives of Neurology*, 62(7), 1091–1094. doi:10.1001/archneur.62.7.1091 PMID:16009764

Pollock, A., St George, B., Fenton, M., & Firkins, L. (2014). Top 10 research priorities relating to life after stroke--consensus from stroke survivors, caregivers, and health professionals. *International Journal of Stroke*, *9*(3), 313–320. doi:10.1111/j.1747-4949.2012.00942.x PMID:23227818

Popović, I. M., Serić, V., & Demarin, V. (2007). Mild cognitive impairment in symptomatic and asymptomatic cerebrovascular disease. *Journal of the Neurological Sciences*, 257(1-2), 185–193. doi:10.1016/j. jns.2007.01.029 PMID:17328916

Qureshi, A. I., Mendelow, A. D., & Hanley, D. F. (2009). Intracerebral haemorrhage. *Lancet*, *373*(9675), 1632–1644. doi:10.1016/S0140-6736(09)60371-8 PMID:19427958

Ramirez-Bermudez, J. (2012). Alzheimer's disease: Critical notes on the history of a medical concept. *Archives of Medical Research*, *43*(8), 595–599. doi:10.1016/j.arcmed.2012.11.008 PMID:23178566

Reijmer, Y. D., Fotiadis, P., Martinez-Ramirez, S., Salat, D. H., Schultz, A., Shoamanesh, A., ... Greenberg, S. M. (2015). Structural network alterations and neurological dysfunction in cerebral amyloid angiopathy. *Brain*, *138*(1), 179–188. doi:10.1093/brain/awu316 PMID:25367025

Roger, V. L., Go, A. S., Lloyd-Jones, D. M., Adams, R. J., Berry, J. D., Brown, T. M., ... Wylie-Rosett, J. (2011). Heart disease and stroke statistics—2011 update: A report from the american heart association. *Circulation*, 23(4), e18–e209. doi:10.1161/CIR.0b013e3182009701 PMID:21160056

Roman, G. C. (1987). Senile dementia of the Binswanger type. A vascular form of dementia in the elderly. *Journal of the American Medical Association*, 258(13), 1782–1788. doi:10.1001/jama.1987.03400130096040 PMID:3625988

Romero, J. R., & Wolf, P. A. (2013). Epidemiology of Stroke: Legacy of the Framingham Heart Study. *Global Heart*, 8(1), 67–75. doi:10.1016/j.gheart.2012.12.007 PMID:23527318

Sá-Pereira, I., Brites, D., & Brito, M. A. (2012). Neurovascular unit: A focus on pericytes. *Molecular Neurobiology*, *45*(2), 327–347. doi:10.100712035-012-8244-2 PMID:22371274

Schneider, J. A., Arvanitakis, Z., Bang, W., & Bennett, D. A. (2007). Mixed brain pathologies account for most dementia cases in community-dwelling older persons. *Neurology*, *69*(24), 2197–2204. doi:10.1212/01.wnl.0000271090.28148.24 PMID:17568013

Schneider, J. A., Wilson, R. S., Bienias, J. L., Evans, D. A., & Bennett, D. A. (2004). Cerebral infarctions and the likelihood of dementia from Alzheimer disease pathology. *Neurology*, *62*(7), 148–155. doi:10.1212/01.WNL.0000118211.78503.F5 PMID:15079015

Scholten, N., Pfaff, H., Lehmann, H. C., Fink, G. R., & Karbach, U. (2013). Thrombolysis for acute stroke--a nationwide analysis of regional medical care. *Fortschritte der Neurologie-Psychiatrie*, *81*(10), 579–585. PMID:24081518

Sharma, M., Tiwari, M., & Tiwari, R. K. (2015). Hyperhomocysteinemia: Impact on Neurodegenerative Diseases. *Basic & Clinical Pharmacology & Toxicology*, *117*(5), 287–296. doi:10.1111/bcpt.12424 PMID:26036286

Siddiqui, F. M., Dandapat, S., Banerjee, C., Zuurbier, S. M., Johnson, M., Stam, J., & Coutinho, J. M. (2015). Mechanical thrombectomy in cerebral venous thrombosis: Systematic review of 185 cases. *Stroke*, *46*(5), 1263–1268. doi:10.1161/STROKEAHA.114.007465 PMID:25899238

Singhal, S., Bevan, S., Barrick, T., Rich, P., & Markus, H. S. (2004). The influence of genetic and cardiovascular risk factors on the CADASIL phenotype. *Brain*, *127*(9), 2031–2038. doi:10.1093/brain/awh223 PMID:15229130

Smith, E. E., Cieslak, A., Barber, P., Chen, J., Chen, Y. W., Donnini, I., ... Hachinski, V. (2017). Therapeutic Strategies and Drug Development for Vascular Cognitive Impairment. *Journal of the American Heart Association*, *6*(5), e005568. doi:10.1161/JAHA.117.005568 PMID:28476873

Smith, E. E., Schneider, J. A., Wardlaw, J. M., & Greenberg, S. M. (2012). Cerebral microinfarcts: The invisible lesions. *Lancet Neurology*, *11*(13), 272–282. doi:10.1016/S1474-4422(11)70307-6PMID:22341035

Smith, R. A., Curtain, R., Ovcaric, M., Tajouri, L., Macmillan, J., & Griffiths, L. (2008). Investigation of the NOTCH3 and TNFSF7 genes on C19p13 as candidates for migraine. *The Open Neurology Journal*, 2(1), 1–7. doi:10.2174/1874205X00802010001 PMID:19018300

Sonnen, J. A., Larson, E. B., Crane, P. K., Haneuse, S., Li, G., Schellenberg, G. D., ... Montine, T. J. (2007). Pathological correlates of dementia in a longitudinal, population-based sample of aging. *Annals of Neurology*, *62*(4), 406–413. doi:10.1002/ana.21208 PMID:17879383

Soontornniyomkij, V., Lynch, M. D., Mermash, S., Pomakian, J., Badkoobehi, H., Clare, R., & Vinters, H. V. (2010). Cerebral microinfarcts associated with severe cerebral beta-amyloid angiopathy. *Brain Pathology (Zurich, Switzerland)*, 20(2), 459–467. doi:10.1111/j.1750-3639.2009.00322.x PMID:19725828

Stam, J. (2005). Thrombosis of the cerebral veins and sinuses. *The New England Journal of Medicine*, 352(17), 1791–1798. doi:10.1056/NEJMra042354 PMID:15858188

Stephens, S., Kenny, R. A., Rowan, E., Allan, L., Kalaria, R. N., Bradbury, M., & Ballard, C. G. (2004). Neuropsychological characteristics of mild vascular cognitive impairment and dementia after stroke. *International Journal of Geriatric Psychiatry*, *19*(11), 1053–1057. doi:10.1002/gps.1209 PMID:15481073

Summers, P. M., Hartmann, D. A., Hui, E. S., Nie, X., Deardorff, R. L., McKinnon, E. T., ... Jensen, J. H. Sh. (2017). Functional deficits induced by cortical microinfarcts. *Journal of Cerebral Blood Flow* and *Metabolism*, *37*(11), 3599–3614. doi:10.1177/0271678X16685573 PMID:28090802

Tatemichi, T. K., Desmond, D. W., & Prohovnik, I. (1995). Strategic infarcts in vascular dementia. A clinical and brain imaging experience. *Arzneimittel-Forschung*, *45*(3A), 371–385. PMID:7763329

Toledo, J. B., Arnold, S. E., Raible, K., Brettschneider, J., Xie, S. X., Grossman, M., ... Trojanowski, J. Q. (2013). Contribution of cerebrovascular disease in autopsy confirmed neurodegenerative disease cases in the National Alzheimer's Coordinating centre. *Brain*, *136*(Pt 9), 2697–2706. doi:10.1093/brain/awt188 PMID:23842566

Troncoso, J. C., Zonderman, A. B., Resnick, S. M., Crain, B., Pletnikova, O., & O'Brien, R. J. (2008). Effect of infarcts on dementia in the Baltimore longitudinal study of aging. *Annals of Neurology*, *64*(2), 168–176. doi:10.1002/ana.21413 PMID:18496870

Van Asch, C. J., Luitse, M. J., Rinkel, G. J., van der Tweel, I., Algra, A., & Klijn, C. J. (2010). Incidence, case fatality, and functional outcome of intracerebral haemorrhage over time, according to age, sex, and ethnic origin: A systematic review and meta-analysis. *Lancet Neurology*, *9*(2), 167–176. doi:10.1016/S1474-4422(09)70340-0 PMID:20056489

Van Veluw, S. J., Shih, A. Y., Smith, E. E., Chen, C., Schneider, J. A., Wardlaw, J. M., ... Biessels, G. J. (2017). Detection, risk factors, and functional consequences of cerebral microinfarcts. *Lancet Neurology*, *16*(9), 730–740. doi:10.1016/S1474-4422(17)30196-5 PMID:28716371

Vemuri, P., Lesnick, T. G., Przybelski, S. A., Knopman, D. S., Preboske, G. M., Kantarci, K., ... Jack, C. R. Jr. (2015). Vascular and amyloid pathologies are independent predictors of cognitive decline in normal elderly. *Brain*, *138*(3), 761–771. doi:10.1093/brain/awu393 PMID:25595145

Vermeer, S. E., Koudstaal, P. J., Oudkerk, M., Hofman, A., & Breteler, M. M. (2002). Prevalence and risk factors of silent brain infarcts in the population-based Rotterdam Scan Study. *Stroke*, *33*(1), 21–25. doi:10.1161/hs0102.101629 PMID:11779883

Vermeer, S. E., Longstreth, W. T. Jr, & Koudstaal, P. J. (2007). Silent brain infarcts: A systematic review. *Lancet Neurology*, *6*(7), 611–619. doi:10.1016/S1474-4422(07)70170-9 PMID:17582361

Vermeer, S. E., Prins, N. D., den Heijer, T., Hofman, A., Koudstaal, P. J., & Breteler, M. M. (2003). Silent brain infarcts and the risk of dementia and cognitive decline. *The New England Journal of Medicine*, *348*(13), 1215–1222. doi:10.1056/NEJMoa022066 PMID:12660385

Wen, W., Sachdev, P. S., Li, J. J., Chen, X., & Anstey, K. J. (2009). White matter hyperintensities in the forties: Their prevalence and topography in an epidemiological sample aged 44-48. *Human Brain Mapping*, *30*(9), 1155–1167. doi:10.1002/hbm.20586 PMID:18465744

Westover, M. B., Bianchi, M. T., Yang, C., Schneider, J. A., & Greenberg, S. M. (2013). Estimating cerebral microinfarct burden from autopsy samples. *Neurology*, *80*(15), 1365–1369. doi:10.1212/ WNL.0b013e31828c2f52 PMID:23486880

Wollenweber, F. A., Buerger, K., Mueller, C., Ertl-Wagner, B., Malik, R., Dichgans, M., ... Opherk, C. (2014). Prevalence of cortical superficial siderosis in patients with cognitive impairment. *Journal of Neurology*, 261(2), 277–282. doi:10.100700415-013-7181-y PMID:24221645

Yang, J., Wong, A., Wang, Z., Liu, W., Au, L., Xiong, Y., ... Mok, V. C. (2015). Risk factors for incident dementia after stroke and transient ischemic attack. *Alzheimer's & Dementia*, *11*(1), 16–23. doi:10.1016/j. jalz.2014.01.003 PMID:24603162

Yates, P. A., Villemagne, V. L., Ellis, K. A., Desmond, P. M., Masters, C. L., & Rowe, C. C. (2014). Cerebral microbleeds: A review of clinical, genetic, and neuroimaging associations. *Frontiers in Neurology*, *6*(4), 205. PMID:24432010

Zieren, N., Duering, M., Peters, N., Reyes, S., Jouvent, E., Hervé, D., ... Dichgans, M. (2013). Education modifies the relation of vascular pathology to cognitive function: Cognitive reserve in cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy. *Neurobiology of Aging*, *34*(4), 400–407. doi:10.1016/j.neurobiolaging.2012.04.019 PMID:22626524

Zonneveld, H. I., Goos, J. D., Wattjes, M. P., Prins, N. D., Scheltens, P., van der Flier, W. M., ... Barkhof, F. (2014). Prevalence of cortical superficial siderosis in a memory clinic population. *Neurology*, *82*(8), 698–704. doi:10.1212/WNL.00000000000150 PMID:24477113

Chapter 8 Zinc and Neurodegenerative Disorders

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ABSTRACT

Zinc (Zn) is an essential trace element that is abundantly present in humans. Despite its importance in normal brain functions, alterations in zinc homeostasis cause various neurological pathologies such as dementia, Parkinson's disease, Prion's disease, etc. A growing body of evidence has shown that zinc might play a dual role: in which both zinc depletion and excess zinc cause severe damage and hence neurotoxicity develops. Homeostatic controls are put in place to avoid the accumulation of excess zinc or its deficiency. This cellular zinc homeostasis results from the actions of a coordinated regulation effected by different proteins involved in the uptake, excretion, and intracellular storage or trafficking of zinc. Further investigation has also shown the role of endogenous carnosine (beta-alanyl-L-histidine) in binding excess zinc. Hence, it has the ability to prevent neurotoxicity. Also, the role of a zinc-rich diet cannot be overemphasized. The authors of the chapter, however, provide an insight into the link between zinc homeostasis and neurodegenerative disorders (NDs).

DOI: 10.4018/978-1-5225-5282-6.ch008

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INTRODUCTION

One of the most important mineral identified in the human system is zinc, a crucial trace element in the metabolic activities, cell division, immune system and as a co-factor of over 300 well-known enzymes (Hambidge, 2000) in the control of different cellular activities and signaling pathways expedient for both neuro and systemic operation (Takeda, 2000). This divalent metal constitutes a cofactor of a number of enzymes like carbonic anhydrase, alcohol dehydrogenase, alkaline phosphatase, phospholipase, carboxypeptidase, zinc/copper superoxide dismutase (SOD) and other allosteric proteins (Mc-Call et al., 2000). This is the reason why zinc is referred to as antioxidative element in human beings because these enzymes which have zinc as cofactor are involved in combating oxidative stress (Feng et al., 2013). Zinc is used to make the hormone thyrotropin-releasing hormone (TRH) that signals the thyroid to make thyroid hormones. It converts the protein we eat into amino acids, including tyrosine which powers the thyroid hormone production. Finally, it is involved in the making of triiodothyronine (T3) the active form that is used in the muscles (Soh et al., 2012). Furthermore, in the formation of bone, zinc is used by enzymes in the production of collagen and alkaline phosphatase (ALP), which are important for bone formation (Hyun-Ju et al., 2010). It is also used to make calcitonin, a hormone that inhibits the breakdown of bone. Zinc is also a critical component of deoxyribonucleic acid (DNA) and ribonucleic acid (RNA) polymerases and has a fundamental role for nucleic acid metabolism and gene activation and repression. 30% of the zinc found in a cell is found in the nucleus (Ali et al., 2012). This makes sense as it is very involved with DNA and the replication of cells and proteins needed by the body. At the transcription level during protein expression, zinc fingers structure enables transcription factors to anchor to DNA helix. Cyclins and cyclin-dependent kinases are directly influenced by this divalent metal and in essence regulate cell cycle. (Chesters and Petrie, 1999). The activities of many growth factors are zinc dependent, hence cell proliferation is regulated by the concentration of zinc ions (Hamza et al., 2012). Zinc functions as a neurotransmitter (Tóth, 2011); in addition, it modulates the function of glutamate and other neurotransmitter receptors. Recent studies have indicated zinc to play important signaling roles in different human biological activities (Hirano et al., 2008). Zinc deficiency in neonates is known to cause dwarfism, the retardation of mental and physical development, immune dysfunction, furthermore, learning incapacities and in grown-up certain neurodegenerative issue, for example, sadness, schizophrenia, Alzheimer's diseases (AD), Parkinson's diseases (PD), maturing, or amyotrophic horizontal sclerosis (AHS) (Prasad, 2009).

The mechanism of action of zinc is that, it modulates certain receptors at the post synaptic cleft. During the neuronal activity, when zinc with glutamate is released from synaptic vesicles into synaptic cleft, zinc interactions with postsynaptic receptors may occur (Morris and Levenson, 2012). The well-known process of zinc inhibition of *N*-Methyl-D-aspartic acid receptors (NMDAr) in synapses is one of such interaction. Excessive influx of zinc into neurons has been found to result in neurotoxicity and damage to post synaptic neurons. Zinc is additionally proposed as a hazard factor for melancholy, AD, maturing and other neurodegenerative issue (Izumi, 2006).

Then again, a developing assemblage of proof proposes that a lack as opposed to an overabundance of zinc prompts an expanded hazard for the improvement of neurological issue (Szewczyk, 2013). Without a doubt, zinc insufficiency has been appeared to influence neurogenesis and increment neuronal apoptosis, which can prompt learning and memory deficiencies. There is need for homeostatic controls to be put in place to avoid the accumulation of excess zinc or its deficiency. Disturbances of zinc homeostasis are considered as important factors in neurodegenerative brain disorders (Konoha *et al.*, 2006). The involve-

ment of zinc in the pathology of neurodegenerative has been reported hundreds of times. It is, however, still a matter of debate whether the disease progression can be influenced by modifying zinc in the diet or regulating amount of zinc in the body (Foster *et al.*, 2012). Zinc activities could be as a supporter of the sickness in one section, and as a defender in another. Therefore, zinc may assume a part like that of Janus, who is the antiquated Roman divine force of entryways and who has 2 unique countenances, in the cerebrum: both zinc exhaustion and abundance zinc make serious harm neurons (Dai and Masahiro, 2013). This chapter aims at discussing the current findings relating to neuronal zinc metabolism and the way in which zinc can modulate normal brain activity. To be discussed also is the contribution of zinc to the formation, aggregation, and mechanism of zinc induced toxicity, the contribution of zinc to the pathogenesis of NDs. Possible treatment will also be looked into for zinc induced neurotoxicity.

BACKGROUND

NDs such as AD, PD, Huntington's disease (HD), AHS and Prion disease (PrD) etc. are the major neurological diseases being reported lately (Vijay *et al.*, 2016 a). Unfortunately, the pathogenesis of these chronic NDs is not fully understood, and current treatments do not stop or slow down progression of these pathological conditions. Increased accumulation of heavy metals, such as iron (Fe), zinc and manganese (Mg) in the brain patients emphasizes their possible role in NDs (Dexter et al., 1991). Knowledge about zinc has rapidly evolved over the years with the last two decades have brought, interesting new insights about the role of zinc in molecular and cellular processes as well as health and disease (Bernadeta, 2013) stated in Figure 1.

Actual fields of research in neurobiology are not only aimed at understanding the different aspects of central nervous system but also at developing strategies useful to preserve brain compensatory capacity and to prevent the onset of NDs. Consistent with this trend much attention has been addressed to zinc metabolism (Mocchegiani *et al.*, 2005).

ZINC AND THE CENTRAL NERVOUS SYSTEM

In the central nervous system (CNS), zinc occurs in two forms: the first being tightly bound to proteins and, secondly, the free, cytoplasmic, or extracellular form found in presynaptic vesicles (Bernadeta, 2013). In the brain, zinc is present in its free ionic form (Zn^{2+}) within synaptic vesicles, mostly at the glutamatergic terminals (Frederickson *et al.*, 2000; Paoletti *et al.*, 2009; Sensi *et al.*, 2011). In this role, zinc is highly concentrated in the synaptic vesicles of a specific contingent of neurons called "zinc containing" neurons, which co-release zinc with the neurotransmitter glutamate upon excitation. The majorities of these "gluzincergic" neurons (Frederickson and Bush, 2001) have their cell bodies located in either the cerebral cortex or the limbic structures of the forebrain (Slomianka *et al.*, 1990), and so an extensive network uniting limbic and cerebrocortical functions is created (Koh, 2005). These neurons are discovered solely in the fore mind, where in well evolved creatures they have advanced into an intricate and expand associational system that interconnects a large portion of the cerebral cortices. There is increasing evidence that zinc not only act as neurotransmitter to mediate intercellular communication, but also act as intracellular signaling molecule much like calcium (Hirano *et al.*, 2008).

Zinc and Neurodegenerative Disorders

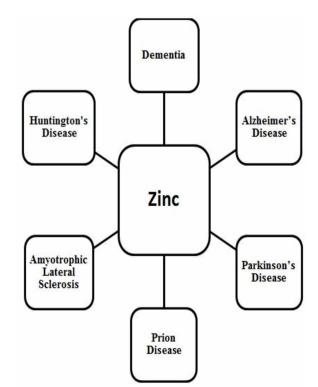


Figure 1. The relationship between zinc and neurodegenerative disorders

Recent studies have suggested that secreted Zn^{2+} plays crucial roles in information processing, synaptic plasticity, learning, and memory (Vijay *et al.*, 2016 b). Indeed, Zn^{2+} has been shown to be essential in the hippocampus for the induction of long-term potentiation, a form of synaptic information storage that has become a well-known paradigm for the mechanisms underlying memory formation (Tamano and Takeda, 2011).

Highest concentrations of zinc in the brain are found in the hippocampus, amygdala, cerebral cortex, thalamus, and olfactory cortex (Frederickson *et al.*, 2000). Because the hippocampus is the region of the brain which plays a critical role in memory, learning and neurogenesis, the impact of zinc deficiency or zinc supplementation on these processes will be critical (Muhammad and Sandrine, 2012). Indeed, it was found that a zinc deficient diet, decreases the number of progenitor cells and immature neurons in the dentate gyrus (DG) in rodents and that reversal to a normal diet containing zinc restored a number of these cells (Gao *et al.*, 2009; Suh *et al.*, 2009). In human, the dietary zinc is mostly absorbed in duodenum, ileum and jejunum by active transport. Zinc is transported from blood trough blood brain barrier system, mostly in the form of complexes with amino acids, especially L-histidine and cysteine (Costello *et al.*, 2011; Pavlica and Gebhardt, 2010).

ZINC HOMEOSTASIS AND NEUROTOXICITY

Intracellular zinc is tightly controlled because zinc is essential but potentially toxic. Adequate zinc level is critical for CNS development and the differentiation of nervous stem cells in mammals (Levenson and Morris, 2011; Xu *et al.*, 2011). The exact controls of zinc homeostasis are basic for focal sensory system and for the entire creature. As perturbations of zinc either above or below physiological concentrations are detrimental to cell survival, neurons must maintain tight homeostatic control on their zinc content.

Disturbances of zinc homeostasis are considered as important factors in neurodegenerative brain disorders. Adequate zinc intake is crucial for proper cognitive functions, especially in children and elderly human (Brewer, 2010; Lovell et al., 2006; Levenson and Morris, 2011; Pavlica and Gebhardt, 2010; Toren et al., 1996; Yasuda et al., 2011). Zinc lopsidedness can come about from inadequate dietary admission, as well as from disabled movement of zinc transport proteins and zinc subordinate direction of metabolic pathways. Under normal CNS physiology, homeostatic controls are put in place to avoid the accumulation of excess zinc or its deficiency (Szewczyk, 2013). This cellular zinc homeostasis results from the actions of a coordinated regulation effected by different proteins involved in the uptake, excretion and intracellular storage/trafficking of zinc. Several different groups of proteins are involved in managing cellular levels of zinc. The first group consists of are membranous zinc transporters (ZnTs) mediating the zinc efflux from cells or influx into cellular compartments or organelles (Huang and Tepaamorndech, 2013). The second group is members of the zip family (zinc-regulated and iron regulated transporter proteins) that promote zinc transport from the extracellular space or from intracellular vesicles to the cytoplasm (Cousins et al., 2006). The changes in zinc levels can be caused by the altered expression of zinc transporters, especially ZnT1, 3, 4, 6 (Zinc transporters) and metallothionein proteins. The extracellular concentration of zinc in hippocampus and cerebral cortex depends on the activity of zinc transporter-ZnT3 (Math and Gordon, 1997). The abnormally increased activity of ZnT3 may additionally elevate the extracellular zinc and in that way it induces the formation of β -amyloid plagues. β -amyloid plaques are the protein fragment which is also a biomarker to identify people at risk for AD (Christian, 2011). This process is thought to be the first step in zinc homeostasis disruption in course of that pathological state. The evidently, unbalanced homeostasis of this biometal affects cell function. Free or chelatable zinc which is not associated with proteins or amino acids ligands appears to participate in the neurotoxic accumulation of zinc in neurons. Damage to the CNS induced by seizure, trauma, or ischemia can all result in the accumulation of zinc from vesicular and non-vesicular zinc pools (Deborah and Cathy, 2012). Under these conditions, excessive free zinc is released into the synaptic cleft and in return, zinc modulates a number of postsynaptic neuronal receptors, with excess zinc leading to death of neurons (Inoue et al., 2015). There is evidence that the influx and accumulation of excess zinc causes excitotoxicity, generates oxidative stress, and impairs neuronal energy production (Morris and Levenson, 2012). N-methyl-D-aspartate receptors (NMDA), for example, have a high-affinity zinc-binding site that can bind synaptically released zinc at nanomolar concentrations. Zinc is thus responsible for fine-tuning the activity of these important glutamate receptors. Additionally, there is now evidence of mutations in NMDA receptors with altered zinc affinities that may have implications for neurodevelopment leading to a variety of developmental disorders including childhood epilepsy and cognitive deficits (Serraz et al., 2016).

Other toxic effects that zinc can potentially induce include the disruption of normal tubulin assembly (Kress *et al.*, 1981), and the over activation of calcium-mediated enzymes (Csermely *et al.*, 1988). Furthermore, zinc reacts with the thiol and imidazole moieties of many proteins (Chvapil *et al.*, 1972) thus, can disrupt structure and function by binding proteins.

ZINC TOXICITY AND NEURODEGENERATIVE DISORDERS

Neurodegenerative disorder (ND) is an umbrella term for a range of conditions which primarily affect the neurons in the human brain. Neurons are the building block of the nervous system which includes the brain and spinal cord. NDs are incurable and debilitating conditions that result in progressive degeneration and / or death of nerve cells. This causes problems with movement called ataxia, or mental functioning called dementias (Choi *et al.*, 1988; Perry *et al.*, 1997).

In physiological concentrations, zinc exhibits neuroprotective activity, but at high concentrations, zinc can be neurotoxic (Cote *et al.*, 2005; Plum *et al.*, 2010). Also, effects of malnutrition may be particularly relevant for NDs as the brain is the organ with the highest zinc levels (in mean: approx. 150 μ mol/l) separated in delicately balanced pools (Weiss et al., 2000). Zinc dyshomeostasis have also been reported in such diseases as depression and attention deficit hyperactivity disorder (ADHD), Senile dementia which can be classified into AD and vascular dementia (VD), PD as well as in brain ischemia and traumatic brain injury (Bernadeta, 2013).

Zinc and Senile Dementia

Senile dementia is characterized by a decrease in cognitive abilities. This may include the person's ability to concentrate, to recall information, and to properly judge a situation. Senility is a deterioration of body and mind associated with advanced aging. Senile dementia is mostly divided into AD and vascular-sort dementia (VSD) (Mizuno and Kawahara, 2013). VD is a degenerative cerebrovascular disease, and its peril factors join age, the male sex, diabetes, and hypertension. The most generally perceived sort of VSD is caused by a movement of little strokes or ischemia (Lee *et al.*, 2000).

In ischemic conditions, a considerable amount of zinc (up to 300 μ M) is associated with glutamate in synaptic clefts due to membrane depolarization. Zinc causes the apoptotic downfall of basic refined cortical neurons. Also, chelatable zinc evidently moves from presynaptic terminals into postsynaptic neuronal cell bodies. A development in intracellular Zn²⁺ levels, zinc translocation, occurs in vulnerable neurons in the districts of the hippocampus going before the start of conceded neuronal going after transient overall ischemia (Koh et al., 1996). This zinc translocation has been represented to enhance the nearness of infarcts. Thus, Zn translocation has been recognized as the primary event in the pathway of Zn-induced neuronal death. (Sensi *et al.*, 2003) a process that might be involved in the pathogenesis of VSD (Plum *et al.*, 2010; Weiss *et al.*, 1993). Understanding the molecular mechanisms of Zn-induced neuronal death is of great importance for the treatment of VSD.

Zinc and Alzheimer's Disease

Another chronic ND is AD and it is the most common cause of dementia. It is estimated that AD represents 60– 80% of all dementia cases (Daviglus *et al.*, 2010). AD is a progressive ND that symptomatically leads to intellectual decline including memory loss, language breakdown, difficulties in learning and orientation, and changes in behavior with neurofibrillary tangles (NFTs), the aggregates of hyperphosphorylated tau protein which are most commonly known as the primary marker (Goedert *et al.*, 2006). It is described as amyloid accumulation in the cerebral neuropil and vasculature. These amyloid deposits contain primarily fragments and full-length (40 or 42 residue) forms of the amyloid β -protein (A β) structured into fibrillar assemblies (Louise, 2000).

The neurotic highlights of AD are the collection of β -amyloid (A β) and the total of A β is recommended as the reason for neurodegeneration saw in AD (Small and Cappai, 2006). A β is the result of proteolytic cleavage from the amyloid antecedent protein (AAP) by the catalyst known as β -secretase or β -site AAP severing chemical 1(BACE-1) (Masters *et al.*, 1985). Although the key part of A β in the pathogenesis of AD is unequivocally settled now, the instrument by which A β prompts lethality or the causes and factors related with the hazard or movement of AD is still inadequately caught on (Paul and Harry, 2010).

One of the several hypotheses proposed for the pathophysiology of AD is the trace elements hypothesis, with zinc taking the center stage. Zinc was first described as a possible factor leading to dementia by Burnet (1981) and, since then, the knowledge base regarding the role of zinc in the pathogenesis and therapy of AD has evolved rapidly. Several path ways for the involvement of zinc in AAP processing or A β aggregation has been suggested. As stated earlier, AD is characterized by the presence of extracellular amyloid plaques and intracellular neurofibrillary tangles within the afflicted brain, which cause neuronal loss in the neocortex, hippocampus, and basal forebrain, leading to progressive cognitive and behavioral decline (Selkoe, 2001). It was found that AAP synthesis is regulated by zinc-containing transcription factors nuclear factor-kappa B (NF- κ B) and specificity protein 1 (sp1) (Grilli *et al.*, 1996). Zinc is also involved in processing of AAP protein (Lee *et al.*, 2009).

Zinc and Prion Disease

PrD represents a group of conditions that affect the nervous system in humans and animals. In people, these conditions impair brain function, causing changes in memory, personality, and behavior; a decline in intellectual function (dementia); and abnormal movements, particularly difficulty with coordinating movements (ataxia) (Hernández *et al.*,1999). The signs and symptoms of PrD typically begin in adulthood and worsen with time, leading to death within a few months to several years. The prion protein (*PRNP*) gene provides instructions for making a protein called cellular prion protein (PrP^C), conformational change of PrP^C into pathological prion protein PrP (scrapie) leads to onset of PrD (Laura *et al.*, 2007). Although the precise function of this protein is unknown, researchers have proposed roles in several important processes. These include the transport of copper into cells, protection of brain cells (neurons) from injury (neuroprotection), and communication between neurons and recently it was discovered that

it facilitates zinc uptake (Frederickson *et al.*, 2006). A valuable evidence of PrP^{c} -zinc interaction is that not only does PrP^{c} facilitate zinc uptake, zinc also potently stimulates the endocytosis of PrP^{c} . PrP^{c} via α -amino-3-hydroxy-5-methyl-4-isoxazolepropionate (AMPA) receptors directly influences the zinc content of neuronal cells (Spevacek *et al.*, 2013). Hence changes in zinc levels will definitely affect the protein. PrP^{c} has been identified as a neuronal zinc sensor, modulating zinc binding, and promoting zinc uptake via α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors, with a subsequent downstream effect on the activity of intracellular protein tyrosine phosphatases (Hajo and Lothar, 2009). Research has shown that zinc concentration decreases in PrD (Wong *et al.*, 2001). Mutation in PrP^{c} prevents the inter-domain interactions, and disrupts the interaction between PrP^{c} and the AMPA receptor, providing a possible explanation for the reduction in zinc in PrD -affected brains.

Zinc and Parkinson's Disease

PD is a severe, progressive motor disorder caused by changes in the central nervous system, tightly linked to degeneration of dopaminergic neurons of basal ganglia known as substantia nigra (Pietro *et al.*, 2012). PD is characterized by extensive loss of dopaminergic neurons in the substantia nigra. Recently, it has been found that zinc ions directly bind to the peptide fragments from the PD gene Park9 (Remelli *et al.*, 2013). Expulsion of Zn from parkinson protein 2 (Park2) causes almost total unfurling of the protein and loss of its movement. In any case, vague and opposing outcomes exist in the writing with respect to circling Zn levels in PD patients. For instance, a few examinations detailed a lessening of circling Zn in patients with PD contrasted and wellbeing controls, and different investigations found no critical distinction or even expanded Zn levels in PD patients (Remelli *et al.*, 2013).

Actually, altered metal homeostasis has been reported in PD. While zinc is required for normal functioning of the brain, its excessive exposure may lead to PD. Clinical studies with PD patients suggested significantly decreased zinc status in patient blood serum compared with healthy elders control group. Another meaningful observation concerned protein deposits connected with metal ions disturbances in patient brain compartments, which showed some similarities to those observed in AD (Brewer *et al.*, 2010). Additionally, lowered zinc concentration may have influence on pathophysiology of PD by increased inflammation processes, generation of reactive oxygen species and affected neurotropic factors levels, resulted in the proceeding neuronal degeneration.

ZINC AND AGING

Aging is an inevitable process associated with progressive pathological features such as: oxidative stress, altered cell metabolism, damaged nucleic acid, or deposition of abnormal forms of proteins (Bernadeta, 2013). In the brain, aging is characterized by neuronal loss, cognitive impairment, and susceptibility to neurological disorders (Mocchegiani *et al.*, 2005). Recent progress in studies involving age related processes provide evidence that changes occurring in the brain during aging are related to zinc homeostasis and that zinc deficiency is a common cause of morbidity among the elderly (Mocchegiani *et al.*, 2005). Because aging changes dietary habits as a result of physiological, psychological, and social reasons, (McClain *et al.*, 2002) the elderly may be at particular risk of a zinc deficiency and influencing malnutrition. The UK national diet and nutrition survey reported that the mean average daily intake was 8.8 mg for men and 6.9 mg for women aged 65 years or older (Smithers *et al.*, 1998) while the intake for men

and women in the age range 19–64 was 10.2 and 7.4 mg/day respectively (Henderson *et al.*, 2003). Data from the second and third national health and nutrition examination survey (NHANES II, NHANES III) study reported that serum Zn levels peak between the age of 18–25 years, slowly decrease in adulthood and drop off after the age of 65–70 years (Hotz *et al.*, 2003). Serum Zn levels of the oldest-old (age range 90–107) were consistently found to be lower than the levels of adult (20–65 years) and elderly control groups (65–89 years) (Maret and Sandstead, 2006; Ravaglia *et al.*, 2000). When adequacy of zinc intake was defined as a total intake of more than 77% of the 1989 recommended daily allowance (RDA) (12 mg for non-pregnant and non-lactating women and 15 mg for males > 11 years) only 55.6% of the total sample had an adequate intake with the elderly at particular risk of malnutrition.

In aging, zinc deficiency is usually the result of an inadequate zinc dietary intake. It has been reported that only 40% of elderly people have a sufficient intake of zinc (Andriollo-Sanchez et al., 2005; Mocchegiani et al., 2008). Studies comparing old and young mice fed with low dietary zinc indicated that zinc is an important nutritional factor for a proper inflammatory/immune response (Kelly et al., 1996). Accordingly, zinc has anti-inflammatory properties and a low zinc status is associated with increased susceptibility to infection plus intracellular zinc has been found to play a key role in signaling in immune cells (Haase and Rink, 2009; Hasan et al., 2012). On the other hand, aging is characterized by the progressive dysregulation of immune responses. Therefore, zinc has been suggested as a good factor in providing the remodeling of some age- associated changes and also as leading to healthy ageing through the reduction of inflammation (Kahmann et al., 2008). The other mechanism linking age, zinc and inflammation is associated with Metallothionein (MT). It was found that ageing is associated with a higher MT expression and consequently, low availability of intra-cellular zinc for normal immune responses. On the other hand, the supplementation of zinc in aging improves immune function and leads to decreased mortality from infections (Mocchegiani et al., 2010). In another study, Mocchegiani et al. (2011) showed evidence that zinc deficiency and an altered immune response is more evident in people with a polymorphism in interleukin 6 (IL-6) and metal-response element binding transcription factor-1 (Metallothionein 1A, MT1A) and that these individuals will benefit more from zinc supplementation.

REMEDY FOR ZINC-INDUCED NEUROTOXICITY

Since zinc deficiency is prevalent in patients with NDs, the appropriate preventive measures should be considered. Sufficient endogenous buffering must normally be present to prevent release of zinc in concentrations which can cause neurotoxicity (Michelle *et al.*, 2000). Hence, zinc homeostasis is necessary in the prevention and treatment of zinc neurotoxicity. The body has its own way of preventing neurotoxicity of zinc one of such is production of carnosine. Carnosine, a dipeptide has been discovered as one of the endogenous protection against zinc neurotoxicity particularly in cases of dementia (Mizuno and Kawahara, 2013). It is abundantly present in muscles of fishes, birds and mammals (Severin and Boldyrev, 1991). Carnosine play important role in body pH Balances (Abe, 2000), serve as endogenous suppressor of oxidative damages, plays protective role in zinc induced neurodegeneration in after ischemia. Carnosine has been discovered to decrease in aged animals, therefore dietary supplementation is required. The molecular mechanism of carnosine for the prevention of zinc induced neuronal death involves binding of zinc and inhibitory effect is generated (Baran, 2000). Physiologically relevant concentrations of carnosine provide a protective effect against toxicity induced by zinc. The chelating ability of the histidine residue of this dipeptide (Brown and Antholine, 1979) may contribute to its attenuation of zinc

mediated toxicity. Another means of preventing zinc neurotoxicity is modification of zinc diet in order to lower the risk of related diseases. Due to the lack of mechanisms which allow storing this biometal for a long time in the body, it is essential that zinc homeostasis be ensured by the body. It is evident that zinc supplementation may be useful in aging to prevent age-related NDs (Andriollo-Sanchez *et al.*, 2005; Mocchegiani *et al.*, 2008). If zinc supplementation is performed with caution, it may be beneficial for cognitive functions in several age related NDs. Another remedy for treatment of zinc neurotoxicity is treatment of cells *in vitro* and in animals with a zinc chelator, named N,N,N',N'-tetrakis (2-pyridylmethyl) ethylenediamine (TPEN), it attenuated neurological deficit, reduced the rate of neuronal apoptosis and the cerebral infarct area, increased superoxide dismutase activity and reduced plasma concentrations of malondialdehyde and IL-6 (Dilina *et al.*, 2017). An alternative approach to minimizing the consequences of zinc dyshomeostasis could be to promote mechanisms which enhance reuptake.

FUTURE RESEARCH DIRECTIONS

The maintenance of discrete subcellular pools of zinc is critical for the functional and structural

Integrity of neuronal and other body cells that is dependent on zinc (Cuajungco and Lees, 1997). Zinc metabolism in the brain: relevance to human NDs. One of the least understood but very important areas in zinc-related NDs is the relationship of these disorders to physiological and pathological changes in altered zinc homeostasis (Bernadeta, 2013). Why, for instance, does zinc deficiency primarily increase diseases like PrD, depression etc.? While the presence of free zinc in neurons and the role of excess zinc in neuronal damage and death have been known for almost three decades, the mechanisms responsible for zinc-mediated neurotoxicity are still being explored and debated.

Another area that is very much underdeveloped is the relationship between zinc and ND, in particular, the fluxes of zinc that either participate in or result from biological and pathological processes (Frederickson, 1989). There is relatively little knowledge of the differences in content and distribution of pools of zinc in different cells, tissues and organs at different stages of development, in different metabolic states and in local or systemic disease, nor do we understand how these fluxes of zinc interact with the pathways causing cell death. Much has yet to be learned about how cells handle and use zinc, concentrate it in organelles and insert it in zinc MT (Truong-Tran *et al.*, 2000). Zinc in relation to several tissues should be appraised. After exocytosis, where and how is the secretory zinc reacquired? It is only when the cellular biology of zinc is better understood will the full implications of zinc-related toxicity become apparent. Recently, the zinc-homeostasis regulating proteins such as transporters and MT have been gaining more prominence in related literature indicating they may be very important players in the pathophysiology of NDs. Therefore, more studies are needed to fully understand the influence of peripheral zinc deficiency or an overdose on these proteins (Szewczyk, 2013).

CONCLUSION

The disruption of zinc homeostasis, namely zinc depletion and excess zinc, cause severe damage to neurons and linked with various NDs. Whereas, zinc plays an essential role in the number of processes crucial for proper cells and organism function. During human growth, zinc can influence development and proper function of nervous system and neuronal plasticity. The adequate zinc levels in specific brain

compartments seem to be critical for the proper brain functioning because even slight disturbances in zinc homeostasis may lead to or participate in the development of several disorders. However, the beams of search light of researchers are needed to explain the exact mechanisms linking zinc and its various underlying processes involved in diseases. First, zinc deficiency is prevalent in patients with psychiatric and NDs. Conversely, even if the beneficial effects of zinc supplementation were reported either in treatment or in the prevention of ND, zinc supplement users should be overly cautious and avoid overdosing. Understanding the role of endogenous compounds with putative neuroprotective actions, such as carnosine, may be helpful in the development of clinical approaches for the treatment of neuropathy that involves metals.

REFERENCES

Andriollo-Sanchez, M., Hininger-Favier, I., Meunier, N., Venneria, E., O'Connor, J.M., Maiani, G., Coudray, C., & Roussel, A.M. (2005). Age-related oxidative stress and antioxidant parameters in middle-aged and older European subjects: the ZENITH study. *European Journal of Clinical Nutrition*, *59*(Suppl. 2), S 58–S62.

Ann, R. S., Eric, G. B. E., Jillian, L. M., Heidi, C. M., Jeffrey, G. P., & Glenn, L. M. (2013). Zinc drives a tertiary fold in the prion protein with familial disease mutation sites at the interface. *Structure (London, England)*, *21*(2), 236–246. doi:10.1016/j.str.2012.12.002 PMID:23290724

Baran, E. J. (2000). Metal complexes of carnosine. Biochemistry, 65, 789-797. PMID:10951097

Bernadeta, S. (2013). Zinc homeostasis and neurodegenerative disorders. *Frontiers in Aging Neuroscience*, *5*, 33. PMID:23882214

Brewer, G. J., Kanzer, S. H., Zimmerman, E. A., Molho, E. S., Celmins, D. F., Heckman, S. M., & Dick, R. (2010). Subclinical zinc deficiency in Alzheimer's disease and parkinson's disease. *American Journal of Alzheimer's Disease and Other Dementias*, 2(7), 572–575. doi:10.1177/1533317510382283 PMID:20841345

Brown, C. E., & Antholine, W. E. (1979). Chelation chemistry of carnosine. Evidence that mixed complexes may occur in vivo. *Journal of Physical Chemistry*, 83(26), 3314–3319. doi:10.1021/j100489a002

Burnet, F. M. (1981). A possible role of zinc in the pathology of dementia. *Lancet*, *1*(8213), 186–188. doi:10.1016/S0140-6736(81)90062-3 PMID:6162062

Choi, D. W., Yokoyama, M., & Koh, J. (1988). Zinc neurotoxicity in cortical cell culture. *Neuroscience*, 24(1), 67–79. doi:10.1016/0306-4522(88)90312-0 PMID:3368058

Christian, H. (2011). Identifying and validating biomarkers for Alzheimer's disease. *Trends in Biotechnology*, 29(1), 26–32. doi:10.1016/j.tibtech.2010.09.007 PMID:20971518

Chvapil, M., Ryan, J. N., & Zukoski, C. F. (1972). The effect of zinc and other metals on the stability of lysosomes. *Proceedings of the Society for Experimental Biology and Medicine*, *140*(2), 642–646. doi:10.3181/00379727-140-36521 PMID:5037603

Costello, L. C., Fenselau, C. C., & Franklin, R. B. (2011). Evidence for operation of the direct zinc ligand exchange mechanism for trafficking, transport, and reactivity of zinc in mammalian cells. *Journal of Inorganic Biochemistry*, *105*(5), 589–599. doi:10.1016/j.jinorgbio.2011.02.002 PMID:21440525

Cote, A., Chiasson, M., Peralta, M. R. III, Lafortune, K., Pellegrini, L., & Tóth, K. (2005). Cell typespecific action of seizure-induced intracellular zinc accumulation in the rat hippocampus. *The Journal of Physiology*, *566*(3), 821–837. doi:10.1113/jphysiol.2005.089458 PMID:15919712

Cousins, R. J., Liuzzi, J. P., & Lichten, L. A. (2006). Mammalian zinc transport, trafficking, and signals. *The Journal of Biological Chemistry*, 281(34), 24085–24089. doi:10.1074/jbc.R600011200 PMID:16793761

Csermely, P., Szamel, M., Resch, K., & Somogyi, J. (1988). Zinc can increase the activity of protein kinase C and contributes to its binding to plasma membranes in T-lymphocytes. *The Journal of Biological Chemistry*, 263(14), 6487–6490. PMID:3258866

Cuajungco, M. P., & Lees, G. J. (1997). Zinc metabolism in the brain: Relevance to human neurodegenerative disorders. *Neurobiology of Disease*, 4(3-4), 137–169. doi:10.1006/nbdi.1997.0163 PMID:9361293

Dai, M., & Masahiro, K. (2013). The Molecular Mechanisms of zinc neurotoxicity and the pathogenesis of vascular type senile dementia. *International Journal of Molecular Sciences*, *14*(11), 22067–22081. PMID:24213606

Daviglus, M. L., Bell, C. C., Berrettini, W., Bowen, P. E., Connolly, E. S. Jr, & Cox, N. J. (2010). national institutes of health state-of-the-science conference statement: Preventing Alzheimer disease and cognitive decline. *Annals of Internal Medicine*, *153*(3), 176–181. doi:10.7326/0003-4819-153-3-201008030-00260 PMID:20547888

Deborah, R. M., & Cathy, W. L. (2012). Ion channels and zinc: Mechanisms of neurotoxicity and neurodegeneration. *Journal of Toxicology*, 2012, 785647. PMID:22645609

Dexter, D. T., Carayon, A., Javoy-Agid, F., Agid, Y., Wells, F. R., Daniel, S. E., ... Marsden, C. D. (1991). Alterations in the levels of iron, ferritin and other trace metals in parkinson's disease and other neurodegenerative diseases affecting the basal ganglia. *Brain*, *114*(4), 1953–1975. doi:10.1093/brain/114.4.1953 PMID:1832073

Feng, B., Ruiz, M. A., & Chakrabarti, S. (2013). Oxidative-stress-induced epigenetic changes in chronic diabetic complications- Review. *Canadian Journal of Physiology and Pharmacology*, *91*(3), 213–220. doi:10.1139/cjpp-2012-0251 PMID:23537434

Frederickson, C. J. (1989). Neurobiology of zinc and zinc-containing neurons. *International Review of Neurobiology*, *31*, 145–238. doi:10.1016/S0074-7742(08)60279-2 PMID:2689380

Frederickson, C. J., & Bush, A. I.Dilina do. (2001). Synaptically released zinc: Physiological functions and pathological effects. *Biometals*, *14*(3/4), 353–366. doi:10.1023/A:1012934207456 PMID:11831465

Frederickson, C. J., Giblin, L. J., Balaji, R. V., Masalha, R., Frederickson, C. J., Zeng, Y., ... Kreze, A. (2006). Synaptic release of zinc from brain slices: Factors governing release, imaging, and accurate calculation of concentration. *Journal of Neuroscience Methods*, *154*(1-2), 19–29. doi:10.1016/j.jneumeth.2005.11.014 PMID:16460810

Frederickson, C. J., Suh, S. W., Silva, D., Frederickson, C. J., & Thompson, R. B. (2000). Importance of zinc in the central nervous system: The zinc- containing neuron. *The Journal of Nutrition*, *130*(5), 1471S–1483S. doi:10.1093/jn/130.5.1471S PMID:10801962

Gao, H. L., Zheng, W., Xin, N., Chi, Z.-H., Wang, Z.-Y., Chen, J., & Wang, Z.-Y. (2009). Zinc deficiency reduces neurogenesis accompanied by neuronal apoptosis through caspase-dependent and-independent signaling pathways. *Neurotoxicity Research*, *16*(4), 416–425. doi:10.100712640-009-9072-7 PMID:19548052

Goedert, M., Klug, A., & Crowther, R. A. (2006). Tau protein, the paired helical filament and Alzheimer disease. *Journal of Alzheimer's Disease*, 9(s3), 195–207. doi:10.3233/JAD-2006-9S323 PMID:16914859

Grilli, M., Goffi, F., Memo, M., & Spano, P. F. (1996). Interleukin-1beta and glutamate activate the NF kappa B/ Rel binding site from the regulatory region of the amyloid precursor protein gene in primary neuronal cultures. *The Journal of Biological Chemistry*, 271(25), 15002–15007. doi:10.1074/ jbc.271.25.15002 PMID:8663145

Haase, H., & Rink, L. (2009). Functional significance of zinc- related signaling pathways in immune cells. *Annual Review of Nutrition*, 29(1), 133–152. doi:10.1146/annurev-nutr-080508-141119 PMID:19400701

Hajo, H., & Lothar, R. (2009). The immune system and the impact of zinc during aging. *Immunity & Ageing*, *6*(1), 9. doi:10.1186/1742-4933-6-9 PMID:19523191

Hambidge, M. (2000). Human zinc deficiency. *The Journal of Nutrition*, *130*(5), 1344S–1349S. doi:10.1093/jn/130.5.1344S PMID:10801941

Hasan, R., Rink, L., & Haase, H. (2012). Zinc signals in neutrophil granulocytes are required for the formation of neutrophil extra- cellular traps. *Innate Immunity*, *19*(3), 253–264. doi:10.1177/1753425912458815 PMID:23008348

Henderson, L., Gregory, J., Irving, K., & Swan, G. (2003). *The national diet & nutrition survey: adults aged 19 to 64 years*. London: Stationery Office.

Hernández, A. S., García, T. A., Torres, R. E., Pacheco, C. R., & Vázquez, R. J. J. (1999). Prions: Definition and diseases. *Anales de Medicina Interna (Madrid, Spain)*, *16*, 647–653. PMID:10686720

Hirano, T., Murakami, M., Fukada, T., Nishida, K., Yamasaki, S., & Suzuki, T. (2008). Roles of zinc and zinc signaling in immunity: Zinc as an intracellular signaling molecule. *Advances in Immunology*, *97*, 149–176. doi:10.1016/S0065-2776(08)00003-5 PMID:18501770

Hotz, C., Peerson, J. M., & Brown, K. H. (2003). Suggested lower cutoffs of serum zinc concentrations for assessing zinc status: Reanalysis of the second national health and nutrition examination survey data (1976–1980). *The American Journal of Clinical Nutrition*, 78(4), 756–764. doi:10.1093/ajcn/78.4.756 PMID:14522734

Huang, L., & Tepaamorndech, S. (2013). The SLC30 family of zinc transporters are view of current understanding of their biological and pathophysiological roles. *Molecular Aspects of Medicine*, *34*(2-3), 548–560. doi:10.1016/j.mam.2012.05.008 PMID:23506888

Hyun-Ju, S., Young-Eun, C., Taewan, K., Hong-In, S., & In-Sook, K. (2010). Zinc may increase bone formation through stimulating cell proliferation, alkaline phosphatase activity and collagen synthesis in osteoblastic MC3T3-E1 cells. *Nutrition Research and Practice*, *4*(5), 356–361. doi:10.4162/nrp.2010.4.5.356 PMID:21103080

Inoue, K., O'Bryant, Z., & Xiong, Z. G. (2015). Zinc-permeable ion channels: Effects on intracellular zinc dynamics and potential physiological/pathophysiological significance. *Current Medicinal Chemistry*, 22(10), 1248–1257. doi:10.2174/0929867322666150209153750 PMID:25666796

Jéssica, B. B., Juliana, S. S., & Ana, R. S. O. (2017). Zinc and oxidative stress: Current Mechanisms. *Antioxidant*, *6*(4), 24. doi:10.3390/antiox6020024

Kahmann, L., Uciechowski, P., Warmuth, S., Plümäkers, B., Gressner, A. M., Malavolta, M., ... Rink, L. (2008). Zinc supplementation in the elderly reduces spontaneous inflammatory cytokine release and restores T-cell functions. *Rejuvenation Research*, *11*(1), 227–237. doi:10.1089/rej.2007.0613 PMID:18279033

Kelly, E. J., Quaife, C. J., Froelick, G. J., & Palmiter, R. D. (1996). Metallothione in I and II protect against zinc deficiency and zinc toxicity in mice. *The Journal of Nutrition*, *126*(7), 1782–1790. PMID:8683339

Koh, J. Y. (2005). Endogenous zinc in neurological diseases. *Journal of Clinical Neurology (Seoul, Korea)*, 1(2), 121–133. doi:10.3988/jcn.2005.1.2.121 PMID:20396459

Koh, J. Y., Suh, S. W., Gwag, B. J., He, Y. Y., Hsu, C. Y., & Choi, D. W. (1996). The role of zinc in selective neuronal death after transient global cerebral ischemia. *Science*, 272(5264), 1013–1016. doi:10.1126cience.272.5264.1013 PMID:8638123

Kress, Y., Gaskin, F., Brosnan, C. F., & Levine, S. (1981). Effects of zinc on the cytoskeletal proteins in the central nervous system of the rat. *Brain Research*, 220(1), 139–149. doi:10.1016/0006-8993(81)90217-1 PMID:6974032

Laura, W., Heather, M. C., & David, A. H. (2007). The cellular prion protein (PrPC): Its physiological function and role in disease. *Biochimica et Biophysica Acta*, *1772*(6), 629–644. doi:10.1016/j.bba-dis.2007.02.011 PMID:17451912

Lee, J., Kim, C. H., Kim, D. G., & Ahn, Y. S. (2009). Zinc inhibits amyloid beta production from Alzheimer's amyloid precursor protein in SH-SY5Y Cells. *The Korean Journal of Physiology & Pharmacology; Official Journal of the Korean Physiological Society and the Korean Society of Pharmacology, 13*(3), 195–200. doi:10.4196/kjpp.2009.13.3.195 PMID:19885037

Lee, J. M., Grabb, M. C., Zipfel, G. J., & Choi, D. W. (2000). Brain tissue responses to ischemia. *The Journal of Clinical Investigation*, *106*(6), 723–731. doi:10.1172/JCI11003 PMID:10995780

Levenson, C. W., & Morris, D. (2011). Zinc and neurogenesis: Making new neurons from development to adulthood. *Advances in Nutrition*, 2(2), 96–100. doi:10.3945/an.110.000174 PMID:22332038

Louise, C. S. (2000). Alzheimer's amyloid fibrils: Structure and assembly. *Biochimica et Biophysica Acta*, *1502*(1), 16–30. doi:10.1016/S0925-4439(00)00029-6 PMID:10899428

Lovell, M. A., Smith, J. L., & Markesbery, W. R. (2006). Elevated zinc transporter-6 in mild cognitive impairment, Alzheimer disease, and pick disease. *Journal of Neuropathology and Experimental Neurology*, *65*(5), 489–498. doi:10.1097/01.jnen.0000229237.98124.91 PMID:16772872

Martin, L., Nikolaus-von, S., & Harald, W. (2012). Zinc diet and Alzheimer's disease: A systematic review. *Nutritional Neuroscience*, *15*(5), 1–12. PMID:22305647

Masters, C. L., Simms, G., Weinman, N. A., Multhaup, G., McDonald, B. L., & Beyreuther, K. (1985). Amyloid plaque core protein in Alzheimer disease and Down syndrome. *Proceedings of the National Academy of Sciences of the United States of America*, 82(12), 4245–4249. doi:10.1073/pnas.82.12.4245 PMID:3159021

Math, P. C., & Gordon, J. L. (1997). Zinc Metabolism in the brain: Relevance to human neurodegenerative disorders. *Neurobiology of Disease*, 4(3-4), 137–169. doi:10.1006/nbdi.1997.0163 PubMed

McCall, K. A., Huang, C., & Fierke, C. A. (2000, May 01). Function and mechanism of zinc metalloenzymes. *The Journal of Nutrition*, *130*(5), 1437S–1446S. doi:10.1093/jn/130.5.1437S PubMed

McClain, C. J., McClain, M., Barve, S., & Boosalis, M. G. (2002). Trace metals and the elderly. *Clinics in Geriatric Medicine*, *18*(4), 801–818. doi:10.1016/S0749-0690(02)00040-X PMID:12608504

Michelle, S. H., Laura, J. B., & Paul, Q. T. (2000). Endogenous mechanisms of neuroprotection: Role of zinc, copper, and carnosine. *Brain Research*, 852(1), 56–61. doi:10.1016/S0006-8993(99)02215-5 PMID:10661495

Mocchegiani, E., Bertoni-Freddari, C., Marcellini, F., & Malavolta, M. (2005). Brain, aging and neurodegeneration:role of zinc ion availability. *Progress in Neurobiology*, *75*(6), 367–390. doi:10.1016/j. pneurobio.2005.04.005 PMID:15927345

Mocchegiani, E., Burkle, A., & Fulop, T. (2008). Zinc and ageing (ZINCAGEProject). *Experimental Gerontology*, *43*(5), 361–362. doi:10.1016/j.exger.2008.03.009 PMID:18417310

Morris, D. R., & Levenson, C. W. (2012). Ion channels and zinc: Mechanisms of neurotoxicity and neurodegeneration. *Journal of Toxicology*, 2012, 785647. doi:10.1155/2012/785647 PMID:22645609

Muhammad, S. A. Z., & Sandrine, T. (2012). Nutrition, adult hippocampal neurogenesis and mental health. *British Medical Bulletin*, *103*(1), 89–114. doi:10.1093/bmb/lds021 PMID:22833570

Paoletti, P., Vergnano, A. M., Barbour, B., & Casado, M. (2009). Zinc at glutamatergic synapses. *Neuroscience*, *158*(1), 126–136. doi:10.1016/j.neuroscience.2008.01.061 PMID:18353558

Paul, M. M., & Harry, L. (2010). Alzheimer's disease and the β-amyloid peptide. *Journal of Alzheimer's Disease*, *19*(1), 311–323. doi:10.3233/JAD-2010-1221 PMID:20061647

Pavlica, S., & Gebhardt, R. (2000). Comparison of uptake and neuroprotective potential of seven zincsalts. *Neurochemistry International*, *56*(1), 84–93. doi:10.1016/j.neuint.2009.09.005 PMID:19782114 Perry, D. K., Smyth, M. J., Stennicke, H. R., Salvesen, G. S., Duriez, P., Poirier, G. G., & Hannun, Y. A. (1997). Zinc is a potent inhibitor of the apoptotic protease, caspase-3. A novel target for zinc in the inhibition of apoptosis. *The Journal of Biological Chemistry*, 272(30), 18530–18533. doi:10.1074/jbc.272.30.18530 PMID:9228015

Pietro, M., Britne, S., & Juan, C. C. (2012). Motor control abnormalities in Parkinson's disease. *Cold Spring Harbor Perspectives in Medicine*, 2, a009282. PMID:22675667

Plum, L. M., Rink, L., & Haase, H. (2010). The essential toxin: Impact of zinc on human health. *International Journal of Environmental Research and Public Health*, 7(4), 1342–1365. doi:10.3390/ ijerph7041342 PMID:20617034

Plum, L. M., Rink, L., & Haase, H. (2010). The essential toxin: Impact of zinc on human health. *International Journal of Environmental Research and Public Health*, 7(12), 1342–1365. doi:10.3390/ ijerph7041342 PMID:20617034

Prasad, A. S. (2009). Impact of the discovery of human zinc deficiency on health. *Journal of the American College of Nutrition*, 28(3), 257–265. doi:10.1080/07315724.2009.10719780 PMID:20150599

Ravaglia, G., Forti, P., Maioli, F., Nesi, B., Pratelli, L., Savarino, L., ... Cavalli, G. (2000). Blood micronutrient and thyroid hormone concentrations in the oldest-old. *The Journal of Clinical Endocrinology and Metabolism*, 85(6), 2260–2265. doi:10.1210/jcem.85.6.6627 PMID:10852460

Remelli, M., Peana, M., Medici, S., Delogu, L. G., & Zoroddu, M. A. (2013). Interaction of divalent cations with peptide fragments from Parkinson's disease genes. *Dalton Transactions (Cambridge, England)*, 42(17), 5964–5974. doi:10.1039/C2DT32222F PMID:23202360

Selkoe, D. J. (2001). Alzheimer's disease: Genes, proteins, and therapy. *Physiological Reviews*, 81(2), 741–766. doi:10.1152/physrev.2001.81.2.741 PMID:11274343

Sensi, S. L., Ton-That, D., Sullivan, P. G., Jonas, E. A., Gee, K. R., Kaczmarek, L. K., & Weiss, J. H. (2003). Modulation of mitochondrial function by endogenous Zn²⁺ pools. *Proceedings of the National Academy of Sciences of the United States of America*, *100*(10), 6157–6162. doi:10.1073/pnas.1031598100 PMID:12724524

Serraz, B., Grand, T., & Paoletti, P. (2016). Altered zinc sensitivity of NMDA receptors harboring clinically-relevant mutations. *Neuropharmacology*, *109*, 196–204. doi:10.1016/j.neuropharm.2016.06.008 PMID:27288002

Severin, S. E., & Boldyrev, A. A. (1991). Effect of carnosine, a specific component of striated muscle, on muscles and other tissues. *Biomedical Science*, *2*, 91–94. PMID:1912249

Slomianka, L., Danscher, G., & Frederickson, C. J. (1990). Labeling of the neurons of origin of zinccontaining pathways by intraperitoneal injections of sodium selenite. *Neuroscience*, *38*(3), 843–854. doi:10.1016/0306-4522(90)90076-G PMID:2176723

Small, D. H., & Cappai, R. (2006). Alois Alzheimer and Alzheimer's disease: A centennial perspective. *Journal of Neurochemistry*, *99*(3), 708–710. doi:10.1111/j.1471-4159.2006.04212.x PMID:17076655

Smithers, G., Finch, S., Doyle, W., Lowe, C., Bates, C. J., Prentice, A., & Clarke, P. C. (1998). The national diet and nutrition survey: People aged 65 years and over. *Nutrition & Food Science*, *98*(3), 133–134. doi:10.1108/00346659810209791

Suh, S. W., Jensen, K. B., Jensen, M. S., Silva, D. S., Kesslak, P. J., Danscher, G., & Frederickson, C. J. (2000). Histochemically- reactive zinc in amyloid plaques, angiopathy and degenerating neurons of Alzheimer's disease brains. *Brain Research*, *852*(2), 274–278. doi:10.1016/S0006-8993(99)02096-X PMID:10678753

Szewczyk, B. (2013). Zinc homeostasis and neurodegenerative disorders. *Frontiers in Aging Neuroscience*, 19.

Takeda, A. (2000). Movement of zinc and its functional significance in the brain. *Brain Research. Brain Research Reviews*, *34*(3), 137–148. doi:10.1016/S0165-0173(00)00044-8 PMID:11113504

Tamano, H., & Takeda, A. (2011). Dynamic action of neurometals at the synapse. *Metallomics*, *3*(7), 656–661. doi:10.1039/c1mt00008j PMID:21409223

Toren, P., Eldar, S., Sela, B. A., Wolmer, L., Weitz, R., Inbar, D., ... Laor, N. (1996). Zinc deficiency in attention-deficit hyperactivity disorder. *Biological Psychiatry*, 40(12), 1308–1310. doi:10.1016/S0006-3223(96)00310-1 PMID:8959299

Tóth, K. (2011). Zinc in neurotransmission. *Annual Review of Nutrition*, 21(1), 139–153. doi:10.1146/ annurev-nutr-072610-145218 PMID:21548772

Truong-Tran, A. Q., Ho, L. H., Chai, F., & Zalewski, P. D. (2000). Zinc and health: Current Status and future directions. *The Journal of Nutrition*, 130, 1459S–1466S. doi:10.1093/jn/130.5.1459S PMID:10801960

Vijay, K., Ashok, K., Sandeep, K.S., Sanjeev, K.T., Dinesh K, Ragni, S., Seema, D. (2016b). Zinc deficiency and its effect on the brain: An update. *International Journal of Genetics and Gene Therapy*. doi: 10.16966/2471-4968.105

Vijay, K., Neha, S., Tara, K., Asimul, I., Faizan, A., & Imtaiyaz, H. (2016a). Protein aggregation and neurodegenerative diseases: From theory to therapy. *European Journal of Medicinal Chemistry*, *124*, 1105–1120. doi:10.1016/j.ejmech.2016.07.054 PMID:27486076

Weiss, J. H., Hartley, D. M., Koh, J. Y., & Choi, D. W. (1993). AMPA receptor activation potentiates zinc neurotoxicity. *Neuron*, *10*(1), 43–49. doi:10.1016/0896-6273(93)90240-R PMID:7678965

Weiss, J. H., Sensi, S. L., & Koh, J. Y. (2000). Zn²⁺: A novel ionic mediator of neural injury in brain disease. *Trends in Pharmacological Sciences*, *21*(10), 395–401. doi:10.1016/S0165-6147(00)01541-8 PMID:11050320

Wong, B. S., Brown, D. R., Pan, T., Whiteman, M., Liu, T., Bu, X., ... Sy, M. S. (2001). Oxidative impairment in scrapie-infected mice is associated with brain metals perturbations and altered antioxidant activities. *Journal of Neurochemistry*, *79*(3), 689–698. doi:10.1046/j.1471-4159.2001.00625.x PMID:11701772

Wong, B. S., Chen, S. G., Colucci, M., Xie, Z., Pan, T., Liu, T., ... Brown, D. R. (2001). Aberrant metal binding by prion protein in human prion disease. *Journal of Neurochemistry*, 78(6), 1400–1408. doi:10.1046/j.1471-4159.2001.00522.x PMID:11579148

Xu, H., Gao, H.-L., Zheng, W., Xin, N., Chi, Z.-H., Bai, S.-L., & Wang, Z.-Y. (2011). Lactational zinc deficiency-induced hippocampal neuronal apoptosis by a BDNF-independent TrkB signaling pathway. *Hippocampus*, *21*(5), 495–501. doi:10.1002/hipo.20767 PMID:20101602

Yasuda, H., Yoshida, K., Yasuda, Y., & Tsutsui, T. (2011). Infantile zinc deficiency: Association with autism spectrum disorders. *Scientific Reports*, 1(1), 129. doi:10.1038rep00129 PMID:22355646

Section 2 Imperious Neurodegenerative Disorders

Chapter 9 Neurodegenerative Disorders: An Introduction

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ABSTRACT

Neurodegenerative diseases (NDs) are characterized by specific dysfunction and damage of neurons related to pathologically changed proteins that deposit in the patient brain but also in peripheral organs. These proteins can be used for therapy or used as biomarkers. Except for a plethora of alterations revealed for dissimilar neurodegeneration-related proteins, amyloid- β , prion protein, TAR DNA-binding protein 43 (TDP-43, transactive response DNA binding protein 43 kDa), tau and α -synuclein, or fused in sarcoma protein (FUS), molecular classification of NDs depend on the full morphological assessment of protein deposits, their spreading in the brain, and their correspondence to clinical signs with specific genetic modifications. The current chapter represents the etiology of neurodegeneration, classification of NDs, concentrating on the maximum applicable biochemical and anatomical characteristics and most imperative NDs.

INTRODUCTION

NDs are usually characterized as disorders with particular loss of neurons and distinctive association of utilitarian frameworks characterizing clinical symptoms (Kovacs et al., 2010). Full biochemical, and molecular pathological inspections have extended this definition. Many investigations have proven that proteins with changed physicochemical properties were aggregated in the human brain in NDs. Deposition of proteins has controlled to the meaning of the conformational ailments (Carrell and Lomas, 1997). Thus, the physical compliance of a physiological protein adjustment, which outcomes in a changed capacity or harmful intra or additional cell deposition. Changes in the encoding genes are identified with innate types of infection. Every one of these investigations have prompted recategorization of many issues, and opened new ways for remedial methodologies. The focal part of proteins has been converted into biomarker explore and furthermore into the advancement of novel restorative procedures. To be sure, immunization against α -synuclein, amyloid- β (A β), or tau has been investigated, specifically

DOI: 10.4018/978-1-5225-5282-6.ch009

that these proteins appear to proliferate cell-to-cell and might be available to antibodies (Kovacs et al., 2010; Kovacs and Budka, 2010). Infection changing remedial methodologies may require decreasing the production, keeping the conglomeration or potentially upgrading the leeway of the neurotic types of proteins. These perspectives likewise stress the significance of protein-based grouping of NDs and its interpretation into in vivo biomarkers equipped for distinguishing illnesses as right on time as could be allowed. Protein-based biomarkers would be required for the stratification of patients for against protein treatments, specifically since a significant number of the NDs demonstrate covering clinical highlights and furthermore consolidated affidavit of proteins (Kovacs and Budka, 2010). Many scatters are related to the degeneration of neurons, including immunological clutters; besides, a few quality changes prompt the brokenness of the encoded proteins (Kovacs et al., 2010). Be that as it may, not these procedures connect with minutely perceivable protein testimonies, in any event not with the right now connected systems. For instance, in inherited spastic paraplegia, the neuropathological examination, without learning of the clinical manifestations, can propose the condition yet there are no particular protein incorporations that enable the onlooker to connect the pathology to a particular gene transformation (Kovacs et al., 2010; Kovacs and Budka, 2010). Using specific antibodies against ND-related proteins leads to a specific of description of novel neuropathological phenotypes and the development of trusted diagnostic standards (Rahimi and Kovacs, 2014). There are a reasonable number of possible mixtures of proteinopathies, also related to in the link of varied pathologies (Rahimi and Kovacs, 2014; Kovacs, 2016). The objectives of the chapter are to focus on causes of NDs, factors which are implemented in the pathophysiology of different NDs, classification of NDs and the outline of the some important NDs.

BACKGROUND

Neurodegeneration is a big term for a range of conditions which mainly affect cells (neurons) in the brain. These cells degenerate, which result in the development of dementia and/or movement disorders. Alzheimer's disease (AD) and Parkinson's disease (PD) are the two most public neurodegenerative disorders (Tutar and Tutar, 2010). At present much knowledge is being accumulated concerning the disease mechanisms, but the causative factors of these conditions are still largely unknown. Development of NDs has not been illustrated for a long time. To date, an assortment of components have been proposed for clarifying protein misfolding and protein accumulation, in any case we can't comprehend the system plainly at the molecular and cell premise. Useful proteins must pass a quality control process as far as collapsing to perform catalysis, cell transport, signal transmission and direction. In any case, an assortment of basic and ecological variables impact this procedure adversely (Tutar and Tutar, 2010). The rate of sickness movement (i.e., the length of a given neuropathological arrange) and clinical introduction additionally fluctuate starting with one patient then onto the next. Youthful beginning PD patients, for instance, regularly have a more incessant family history of PD and a more factor survival rate in respect to those without the familial history (Tutar and Tutar, 2010). A few investigations of the A β protein totals, which cause AD, likewise demonstrate that the presence of particular shapes in Aβ totals, 40 deposit Aβ (Aβ40) and 42 buildup Aβ fibril structures, and distinguish the unmistakable strain-particular qualities (characterized as "strainness") of the types of AD by the diverse adaptation of the totals (Tutar and Tutar, 2010).

There is however restricted learning about the components that decide singular variety. People that convey a similar transformation in a similar illness causing quality may show a scope of various clinical

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indications. For instance, evaluating 6-year change in verbal memory, preparing pace, and official capacity in AD distinguished impacts of MS4A4E, CLU, and NME8 in whites and of ZCWPW1 and CDS33 variations in African Americans (Przedborski et al., 2003). For MS4A4E and CLU, this affiliation was just noteworthy in people bearing no less than one APOE ϵ 4 allele (AD hazard quality) (Przedborski et al., 2003). Aside from way of life and condition, these individual contrasts are caused by the one of a kind hereditary foundation of every individual. The hereditary foundation, subsequently, could be characterized as the hereditary cosmetics of all alleles that associate with the malady identified with the "sickness causing" transformation.

CAUSES OF NEURODEGENERATION

The causes of NDs are principally unidentified, and when they have been recognized, the factors by which they start the disease remain unclear (Przedborski et al., 2003). One of the most important reasons related to the etiology of NDs depends on the basis of hereditary and environmental inducers at the beginning of these diseases. Some NDs have a strong familial incidence, signifying a genetic origin (Przedborski et al., 2003). Furthermore, these genetic NDs, others are basically infrequent but display a small depending on patients in whom the disease is congenital. Exactly as PD, AD, and even amyotrophic lateral sclerosis (ALS), of which about 10% of all cases are clearly familial (Przedborski et al., 2003).

When the disease is actually infrequent, it seems that any genetic involvement to the neurodegenerative course is insignificant (Tanner et al., 1999). As an alternative, poisonous ecological elements may be the primary reason for introducing neurodegenerative mechanisms. Accordingly, the case for the PD-ALS difficult, which is, due to a poisonous component in *Cycas circinalis*, a plant usually consumed as a food or treatment by the Chamorros of Guam (Kurland, 1988). poisoning cases with 1-methyl-4phenyl- 1,2,3,6-tetrahydropyridine, a by-product of the production process of a meperidine compound, is known to make a hard and permanent parkinsonian condition, which is nearly matching to PD (Przedborski et al., 2001). Additionally, numerous great scale epidemiological literatures were unsuccessful to demonstrate any relative relationship between ecological influences and incidence of diseases such as PD (Tanner, 1989). All these outcomes say that infrequent cases are neither obviously genetic nor noticeably environmental, but maybe, they arise from both hereditary and ecological triggers. The opportunity that such both triggers characterize a respected idea causal of infrequent neurodegeneration needs serious attention.

Risk factors for ND can be gender, endocrine disorders, oxidative stress, viral or bacterial infection, inflammations, dietary habits, vascular diseases, depression, and cancer (Marder et al., 1998). Ethnicity and culture can also have consequences and be responsible for intuitions into the prognosis of neurode-generation (Marder et al., 1998). Especially, smoking, caffeine, and alcohol intake have been assumed to have a relation with neurodegenerative syndromes. There is less proof for a relationship among smoking and parkinsonian conditions, but single study revealed a defensive influence for multiple system atrophy (MSA) but not for progressive supranuclear palsy (PSP) (Vanacore et al., 2000). Likewise, caffeine intake has been shown to be defensive against the progress of PD (Ross and Petrovitch, 2001), although there is less reliable evidence for a relationship with AD (Tyas et al., 2001; Lindsay et al., 2002; Brown et al., 2005).

Neurodegeneration and Oxidative Stress

Mitochondrial dysfunctions, excitotoxicity, lastly apoptosis have been demonstrated as neurotic reason for neurodegenerative sicknesses, for example, PD, AD, MS, and ALS (Janus et al., 2000). Free radical generation catalyzed by redox particles have been shown to assume a basic part in overseeing redox responses bringing about receptive nitrogen species (RNS) and receptive oxygen species (ROS), principle reasons in neurodegeneration (Janus et al., 2000). ROS incorporates hydrogen peroxide (H2O2), nitric oxide (NO), receptive hydroxyl (OH•), and monoxide radicals (NO•). Impeded mitochondria and animated microglia play as pool of ROS. ROS age was believed to be because of lopsidedness amongst generation and leeway of ROS and RNS yet numerous sciences have been uncovered controlling ROS those assume a real part in managing key cell capacities (Apel and Hirt, 2004). Free radicals have been expressed for their significant impact to neuronal harm in cerebral ischemia, seizure issue, schizophrenia, PD and AD (Cadet, 1988). Neuronal biochemical piece is for the most part powerless to ROS since it includes pool of unsaturated lipids those are labile to peroxidation and oxidative adjustment. Twofold obligations of unsaturated fats are problem areas for assault by free radicals those start course or fasten response to harm neighboring unsaturated fats (Zaleska and Floyd, 1985). A few analysts viewed cerebrum as anomalous delicate to oxidative harm and many examinations decisive of the simplicity of peroxidation of brain membranes bolstered this idea. Brain contains abnormal state of unsaturated fats which are more affordable to peroxidation, that expends an over the top part (20%) of aggregate oxygen utilization for its moderately little weight (2%). Moreover, it isn't especially advanced in cell reinforcement barriers. Brain is brought down in cell reinforcement action in examination with different tissues, for instance, around 10% of liver. In addition, human cerebrum has more elevated amount of iron in specific districts and as a rule has large amounts of ascorbate. As apparent from above information, neural cells are thought to be more powerless to oxidative harm when contrasted with other body tissues (Zaleska and Floyd, 1985).

Protein Misfolding and Aggregation in Neurodegenerative Disorders

Protein misfolding prompts protein total and collection of these totals is embroiled as the principle reason of NDs. In brain, some local proteins (prion, tau, β -amyloid, α -syn, and Huntington) experience conformational changes through hereditary and natural variables (Soto and Estrada, 2008). In this way, optional structures of protein change over from α -helix/irregular coil to β -sheet. Thusly, neurotoxic misfolded protein totals are accumulated in CNS and cerebrum harm that prompt NDs (Soto and Estrada, 2008).

Factors Affecting Protein Aggregation in Neurodegenerative Disorders

Protein Structures

The principal basic factor is protein structure. Particularly, essential and optional structures of a protein are two of the most imperative variables for physical and synthetic highlights. Encoded data in amino acid grouping of a protein decides the three dimensional structure. Position and number of various specific amino acids buildups in essential structure may prompt an expansion or a diminishing in total synthesis. Number of hydrophobic amino acids in proteins is relative to propensity of accumulation (Wang, 2005). Auxiliary structures of proteins include in protein misfolding and in addition steadiness.

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Proteins frequently crease locally into stable structures that incorporate α -helix and β -sheet. For the most part, some β -sheet-rich proteins, (for example, scrapie tainted prion protein) connect with neurotic states. Amid protein collection, the auxiliary structure is changed over from α -helix to β -sheet. In this manner, protein gets strictness and wide surface region (Philo and Arakawa, 2009).

Protein Concentration

Protein concentration is a critical parameter in protein collection. High protein concentration can improve the probability of accumulation. Protein-protein collaborations and intermolecular associations (particularly cooperations among hydrophobic amino acids) may produce irregular protein structures. Some misfolded protein accumulations can be constituted neurodegenerative sicknesses over a specific concentration. In addition, proteins are refolded at low concentrations unexpectedly. For instance, lysozyme and immunoglobulin G refold itself at low protein concentration be that as it may, refolding yield diminishes with expanding protein concentration. In this manner, the ideal unconstrained protein concentration extend is acknowledged as $10-50 \mu g/ml$. (Treuheit et al., 2002).

Post-Translational Modifications

After a protein is manufactured, the posttranslational adjustments (PTM) of amino acids may expand the decent variety of proteins by extra useful accumulations (acetic acid derivation, phosphate, different proteins and so on.) and auxiliary changes (Karve and Cheema, 2011). Specifically, phosphorylation assumes a huge part in NDs. It is likewise realized that, event of AD is related with tauopathy because of conglomeration of the tau protein. In the brain, tau protein is found in neurons and it can be phosphorylated with kinase catalysts. Therefore, deviant tau totals are framed and they can be amassed in neurons, in this manner their harmful impacts are caused neuronal misfortune and synaptic adjustment (Karve and Cheema, 2011). Glycosylation is an essential PTM for protein solidness and conglomeration potential. Human prion protein has two potential N-glycosylation destinations (Asn181 and Asn197). Notwithstanding, in prion pathology, transformation of PrPc to PrPSc happens effortlessly if the PrPc is glycosylation are appeared to be required for protein collection and misfolding in neurodegenerative sicknesses. Also, alternate PTMs, for example, glycation, nitration, truncation, polyamination and so forth include in protein misfolding infections (Martin et al., 2011).

Oxidative Stress

Oxidative tension prompts protein oxidation which is a biomarker for some neurodegenerative disorders. Specifically, free radicals and ROS (receptive oxygen species) leads to protein oxidation. An assortment of oxidants can be happened in ordinary oxygen consuming digestion (Rudd et al., 2001). Additionally, absence of cell reinforcements, abundance of oxygen and lipid and metal particles can create free radicals. The oxidation of proteins greatly relies upon their amino acid synthesis. For the most part; lysine, histidine, arginine, methionine, cysteine, phenylalanine, tryptophan, threonine, glutamic acid, and proline buildups slant oxidation. A few proteins have metal restricting districts alone structure. Metal particles, for example, copper, zinc, and iron, are equipped for redox responses and electrons are exchanged from particles to oxidizing mixes. In this way, lethal free radicals are shaped and proteins can be changed

over into conglomeration structures or proteins can be amassed by conformational changes (Moulton and Yang, 2012).

Mutations

Transformations assume determinative part in protein collection and they may drastically change solvency, strength, and accumulation propensity of proteins (Lee and Yu, 2005). Thermally steady proteins may change its solidness even with a point transformation in its structure. For instance, a human lysozyme I56T and D67H mutants significantly diminish the lysozyme efficacy and subsequently the lysozyme accumulates effortlessly after warming. Promote conglomeration cause amyloid fibrils and these fibrils are kept in tissues and are related to neurodegenerative sicknesses (Lee and Yu, 2005). As of late, researchers have been proposed another protein for comprehension of ALS, AD, cystic fibrosis (CF) and frontotemporal lobar degeneration (FTLD) systems. The TDP-43 is found by every single mammalian tissue, conformational changes in this protein cause collection and loss of capacity. TDP-43 has been appeared to chelate to DNA (deoxyribonucleic acid) and mRNA (dispatcher ribonucleic acid) and partake in control of transcription and translation. TDP-43 has a glycine rich C-terminal tail and change happens from this area. Thus, TDP-43 is changed over to amassed frame which is gathered in tissues (Wilson et al., 2011).

pН

Ecological pH is to be basic for protein collection because of changes in net charge on protein. Protonation condition of ionizable destinations of protein and positive net charge are expanded in acidic conditions. Particularly, association of salt scaffolds is changed in parallel with formed new optional structures. In prion sickness, acidic pH encourages production of PrPSc. At low pH, PrPc picks up β -sheet structures and shows accumulation inclination. As per the Finl (2006), α -syn brooded at various temperature and pH esteems, and the best development conditions were resolved as pH 7.4 and 37°C (Fink, 2006). α -syn can be lost its local structures and PD is quickened in these conditions.

CLASSIFICATION OF NEURODEGENERATIVE DISORDERS

Organization of NDs is built on scientific demonstration, structural areas and cell kinds damaged, conformational changed proteins complicated in the pathogenesis (Kovacs, 2016). In many cases, there is a similarity of the medical signs in the progression of the disease. Thus, medical categorization is important when early clinical signs are assessed (Kovacs, 2016).

Clinical and Anatomical Classification

Behavioral Disorders and Changes in Brain Functions

The main anatomical sections affected are the hippocampus, limbic system, and neocortical sections. In focal cortical signs, deterioration of the frontal, temporal, or the occipital lobe can be affected. Fronto-

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temporal dementia (FTD), which is famed with deterioration of the frontal and temporal sections (Kovacs, 2016).

Movement Changes and Motor Dysfunction

The main anatomical sections affected are the basal ganglia, thalamus, brainstem nuclei, cerebellar cortex and nuclei and lower motor neurons (Kovacs, 2016).

Mixed

Behavioral and movement disorders both of these signs may be detected in certain disease early during the clinical course and during the progression (Kovacs, 2016).

Altered Proteins Associated Classification

The following proteins are mostly associated with NDs:

- The microtubule-associated protein tau (MAPT) is essential for the assembly of microtubules (Kovacs et al., 2010).
- Amyloid- β (A β), which originates from the amyloid precursor protein (APP). Additional genes with relation to the pathogenesis of AD include presentiin-1 (*PSEN1*) and *PSEN2*.
- α -syn is a 140-amino acid protein that related to a group of brain proteins (α , β , and γ -synuclein).
- Prion protein (PrP) is a 253-amino acid protein significant in the pathogenesis of prion diseases or spongiform encephalopathies.
- TDP-43 is a 414- amino acid nuclear protein. This protein is encoded by the *TARDBP* gene. Among others, also the most related genes for TDP-43 proteinopathy include progranulin and C9orf72 (Ou et al., 1995).
- FET proteins, which contain the fused in sarcoma (FUS), Ewing's sarcoma RNA-binding protein 1 (EWSR1) (Neumann et al., 2011).

Neuropathological Classification

It depends mainly on the assessment of the structural spreading of neuronal damage, and other histological properties, and the type of intracellular and extracellular protein aggregations, which are identified by immunohistochemistry and biochemistry (Kovacs, 2016). It is important to differentiate between the subcellular site of the intracellular aggregations and if they are nuclear, cytoplasmic, or neurotic, or in cellular processes. In certain cases, only morphological principles are used for classifying, but in certain other cases biochemical alterations or gene polymorphism are also included (Kovacs, 2016).

Molecular Pathological Classification

The most critical step based on this classification is to assess the site and spreading of proteins (Kovacs and Budka, 2010). Extracellular deposits mainly deposit with positive immune reaction for A β or PrP. Essentially, sickness related PrP represents a synaptic site of aggregation. Most important proteins that

aggregate intracellular are: tau, α -syn, TDP-43, FUS/FET proteins, and they are related with rare genetic diseases (Walker et al., 2013).

Current studies propose that disease related protein helps as important key role in the start and increase of aggregated proteins in NDs (Walker et al., 2013) investigation in humans showed specific mechanisms by which definite protein pathology might transmit by the central nervous system (CNS) accompanying evidence of the molecular pathways of protein proliferation in AD (tau and A β), FTLD-tau, and FTLD-TDP (Brettschneider et al., 2015). Furthermore to building a basis for improving therapeutically policies that stop the spreading of protein deposits, this all ideas have essential effects on categorization and pathological relationship (Kovacs, 2016).

SYNTHESIS OF BIOCHEMISTRY, GENETICS AND MORPHOLOGY

Biochemical considerations revealed variances among the proteins in physiological and in sickness states. These variances can prevent the solubility, and ability of fibril development, or can be toxic. Examples of these interferences between proteins, for instance generation of β -sheet, proteinase-resistant cores, and phosphorylation (Kovacs, 2016). Yet, from a large number of biochemical alterations only some have been turned into the investigative kits like Proteinase K (PK)-resistance for PrP, phosphorylation for tau, isoform alterations for tau, and by products for A β . But, the spatial and progressive spreading of the diverse pathologic protein types has not been explained in particulars (Kovacs, 2016).

Anatomical spreading and morphology of pathological changes still control the categorization NDs. Inspection of important gene alterations, besides with biological inspection, has been integrated into the classification strategy (Kovacs, 2016). Concerning the spreading of pathological protein aggregation, there are coinciding properties seen in diverse proteinopathies, like:

- Discrimination relies on extra- (Aβ, PrP) or intra-cellular (tau, α-syn, TDP-43, FUS) control of protein deposits (Kovacs, 2016);
- 2. Differentiating neuronal, mixed neuronal-glial, and glial predominant types of proteinopathies (Kovacs, 2016);
- 3. Estimation of periods is applied in an increasing number of disorders (Kovacs, 2016);
- 4. Existence of intracellular protein deposits showing primary phases expressing future steps of intracellular protein pathologies (Kovacs, 2016).

Another fact is the common incidence of associated proteinopathies, leading to complications of disease identification. So, additionally to the specific injuries of an ND structure, extra pathological variations can be showed in the similar case (Kovacs, 2016). Aggregation of deferent neurodegeneration linked proteins in conjunction with non neurodegenerative pathologies, is a communal occurrence (Rahimi and Kovacs, 2014; Kovacs et al., 2008). These outcomes might have consequences on therapy techniques directing to specific pathological protein in the brains of old patients with dementia. Accepting the thought of "decreasing" for a medical indication is mandatory for the medical and neuropathological experience. Lastly, it essential be well-known that numerous gene alterations related also with depositions of various proteins not only specific to their gene (Kovacs, 2016).

FEW IMPERIOUS NEURODEGENERATIVE DISORDERS

Alzheimer's Disease

AD is portrayed by the extracellular aggregation of A β fibrils and by the intraneuronal aggregation of irregularly phosphorylated tau protein (Duyckaerts et al., 2009). Pr-symptomatic AD can be confirmed *in vivo* depending on the incidence of lab results (Jack et al., 2014) or after death by the incidence of AD-type neuropathological changes although without manifestation of mental weakening throughout lifetime. For tau pathology only neuronal tau immunoreactivity are measured. A β -fibrils could be deposited in the parenchyma in the shape of signs and in the vessel walls similar to cerebral amyloid angiopathy (CAA). Parenchymal A β aggregations appear different morphologies, like stellate (related to astrocytes), prolix aggregations (further divided into fleecy, lake-like) or focal deposits (with or without a dense core), and further sporadic morphologies similar to cotton-wool plaques (Duyckaerts et al., 2009). Different plaque kinds are associated frequently to their structural spreading (Thal et al., 2015; Kovacs, 2016).

Amyotrophic Lateral Sclerosis

ALS is a deadly motor neuron syndrome that is, manifested by advanced damage of the upper and lower motor neurons (MNs) at the spinal or bulbar level (Rowland and Shneider, 2001). The clearest signs that appear in ALS are muscle weakness, twitching, and cramping, which in time can result in the injury of muscles (Wijesekera and Leigh, 2009). In the most progressive periods, ALS patients will develop signs of dyspnea and dysphagia. Magnetic resonance imaging (MRI) readings of the brain and spinal cord are the most valuable neuroimaging method in ALS principally to exclude disorders that like ALS (Hardiman et al., 2011). For instance, new chromosome 9p-linked FTD-ALS shows a dissimilar form of brain atrophy and neuropathological definition that can help to discriminate from typical ALS (Boxer et al., 2009). Innovative neuroimaging machineries are beneficial research means that may aid to recognize specific ALS related pathologies in a non-invasive way. Neuroimaging is frequently done to aid eliminate differential diagnosis rather than approving the diagnosis of ALS (Hardiman et al., 2011; Zarei et al., 2015).

Batten Disease

Batten disease (BD) is a very infrequent and lethal autosomal recessive neurodegenerative syndrome that begins in childhood. It is the most public form of a group of syndromes termed the neuronal ceroid lipofuscinoses (NCLs). Primary symptoms of the syndrome frequently appear around ages 2 to 10, with steady onset of vision difficulties, or seizures (Weimer et al., 2002). Early signs may be refined personality and behavior alterations, limited education or regression, repetitive speech, or stumbling. There may be decelerating head development in the infantile type, weak circulation in lower extremities, diminished body fat and muscle build, curvature of the spine, hyperventilation, teeth crushing, and constipation (Weimer et al., 2002). By the time, affected children undergo mental weakening, increasing seizures, and progressive loss of vision, speech and mechanical abilities. Together denoted to as BD, the NCLs are responsible for the most of neurodegenerative syndromes that affect children. Precisely, the incidence of this disease is around 1 per 12,500 persons (Weimer et al., 2002). The exact type of NCL is categorized by the age of symptomatic beginning and genetic alteration involved. Recently, it has been found that

alterations in ten genes lead to the growth of BD (Weimer et al., 2002). Urinalysis and blood examining can aid to distinguish aberrations that may point to BD. For instance, increased levels of (dolichol) in urine have been shown in many individuals with NCL. The incidence of vacuolated lymphocytes that include holes or cavities when shared with other results that indicate NCL, is indicative for CLN3 mutations (Weimer et al., 2002). Investigative imaging test let doctors to better imagine the appearance of the brain. MRI uses magnetic and radio waves to any one either help or produce images of the brain. The computerized tomography (CT) is additional type of imaging test that uses X-rays and computers to generate a full image of the brain's tissues and constructions. Both diagnostic imaging tests can help to show brain areas that are damaging in persons with NCL (Weimer et al., 2002).

Creutzfeldt-Jakob Disease

Creutzfeldt-Jakob disease (CJD) is an infrequent, deteriorating, always lethal brain sickness. CJD related to a group of human NDs, recently involving Gerstmann-Straussler-Scheinker disease (GSS), kuru, fatal familial insomnia (FFI), and prion protein cerebral amyloid angiopathy (PrPCAA) and variably proteasesensitive prionopathy (VPSPr) (Head et al., 2012). Neuropathologically, the disorder is manifested by many florid signs in the cerebral and cerebellar cortex and frequent mass plaques, amorphous pericellular and perivascular prion protein aggregations in the same regions. There is a high spongiform modification and perineuronal and axonal prion protein conglomerations in the caudate core and putamen, stamped astrocytosis and neuronal harm in the back thalamus and reticular and perineuronal accumulation in the mind stem (Parchi et al., 1997). First problems over nomenclature having been principally fixed, most of the researchers use a PrPr^{es} typing system in which the two main differentially N-terminally truncated 21 kDa and 19 kDa protease-resistant fragments are called type 1 and type 2 PrPr^{es} respectively (Parchi et al., 1996) using western blotting. PrPr^{es} type and glycoform ratio are measured distinctly in this nomenclature (Head et al., 2012). Glycotypes are best characterized as a proportion (% diglycosylated: % monoglycosylated: % nonglycosylated), however a shorthand has been built up in which illustrations where the diglycosylated band leads are given the postfix B and those in which the monoglycosylated band leads are given the addition A (Parchi et al., 1997; Head et al., 2012).

Corticobasal Degeneration

Corticobasal degeneration (CBD) is a pathologic syndrome (Dickson et al., 2002) resulting from unusual accumulation of hyper-phosphorylated tau isoforms with four preserved rehash arrangements (4R tau). Prior to 2013, CBD analytic standards uncovered just a single displaying phenotype, corticobasal syndrome (CBS), once thought pathognomonic for CBD. CBS is an engine appearance with atypical parkinsonism, dystonia, and myoclonus alongside higher cortical manifestations including apraxia, alien limb phenomena, cortical sensory loss, and cognitive alterations (Armstrong, 2014). It is currently realized that CBS is single one showing phenotype of CBD, revealed in updated clinical research diagnostic principles for CBD (Armstrong et al., 2012). Imaging could be valuable to inspect other origins of presenting signs, like signs of prion disease on MRI (Lee et al., 2011) in cases with CBS and quickly worsening dementia or amyloid imaging to predict causal AD. Voxel based morphometry analysis of MRI weaken forms may be prognostic of underlying pathology in CBS cases (Whitwell et al., 2010; Lee et al., 2011; Armstrong, 2014), but these results cannot yet be practical to daily clinical diagnosis.

Friedreich's Ataxia

Friedreich's ataxia (FRDA) is the furthermost shared genetic ataxia (Harding, 1984). It is the outcome of aggregation of iron in mitochondria causing additional generation of free radicals, which at that time leads to cellular loss and loss (Eder et al., 1998). The principal medical structures are advanced gait and limb ataxia, absent lower limb responses, extensor plantar responses, dysarthria, and decrease in or damage of vibration sense and proprioception (sensory modalities connected by posterior column neurons) (Delatycki et al., 2000). Cardiomyopathy, scoliosis, and foot abnormality are shared but non-important signs (Harding, 1984). The chief locations of pathology in FRDA are the dorsal root ganglia, posterior columns of the spinal cord, corticospinal tracts, and the heart. Macroscopically there is a minor part spinal cord with the posterior and lateral columns mainly exaggerated (Harding, 1984). The nervous system variations seem to be a dying back manner from the margin (Hughes et al., 1968). Declined phospholipid heights have been established in the cerebellar and occipital cortex of brains of patients with FRDA (Eder et al., 1998; Delatycki et al., 2000).

Huntington's Disease

Huntington's disease (HD) is an uncommon NDs of the CNS manifested by unwanted choreatic actions, communicative and psychiatric complaints and dementia. Occurrence in the Caucasian people is expected at 1 from 10000 to 20000. The nuclear indications and signs of HD involve motor, mental and psychiatric disorders. Further less familiar, but predominant and often bad structures of HD includes unintentional weight decline, sleep and circadian rhythm troubles and autonomic nervous system abnormalities. The average age at beginning is between 30 and 50 years, with a variety of 2 to 85 years. The mean period of the sickness is 17 to 20 years. The development of the illness clues to additional needs in regular life and lastly death. The greatest shared reason of death is pneumonia, tailed by suicide (Roos, 2010). HD is an autosomal dominantly hereditary sickness triggered by an extended CAG repeat on the short arm of chromosome 4p16.3 in the huntingtin gene (Trottier et al., 1994). This gene codes for the Huntington's protein and, on exon 1, holds the CAG tract. The worst type contains a CAG repeat, coding for a polyglutamine stretch in the protein at that position in the range 6 to 26 (Trottier et al., 1994). Certain medical demonstration will happen if the quantity of repeats goes beyond 40 (Roos, 2010). The range between 29 and 35, is unbalanced, which means that these isoforms are susceptible to alternate throughout reproduction (Roos, 2010). Repetition the gene can lead to errors and frequently leads to elongation and rarely to restriction. This incidence is mostly seen in the man type of reproduction (Trottier et al., 1994). The analysis is established on the medical indications and symptoms in a patient with a parent with established HD (Roos, 2010). Mainly, it is mandatory to take a full history from the person with symptoms followed by a full family history. When all data has been attained the analysis is not very hard, even though nonspecific medical images can be false. Also when the parent is not known or has died due to another reason at an early age, the medical picture can be hard to distinguish (Roos, 2010). It is often essential to demand old data in the form of medical archives and autopsy information. The present important standard is DNA determination, viewing a CAG-repeat of as a minimum 36 on the huntingtin gene on chromosome 4 (Roos, 2010).

Lewy Body Disease

Lewy body disease (LBD) is now known as a broad term that involves PD, PD dementia (PDD)and dementia with Lewy bodies (DLB) (McKeith et al., 2005; Lippa et al., 2007). Many dementia experts, misinterpret the correlation between LBD and DLB. LBD is defined as, a chronic advanced neuropsychiatric syndrome, which is clinically manifested by parkinson signs of pre-senile or senile, or occasionally younger onset, frequently followed by dementia at the advanced stages (Lippa et al., 2007). Progressive dementia or several kinds of psychiatric signs including distinctive visual hallucination and delusions are the main symptoms, commonly followed by parkinson indications. It is neuropathologically described by numerous Lewy bodies (LBs) and neuritis, and neuronal cell damage in the central and autonomic nervous system. DLBD is manifested clinically by advanced dementia and parkinson signs of pre-senile or senile, or occasionally of younger onset, and neuropathologically by frequent LB and neuronal cell damage in the central and autonomic nervous systems, commonly followed by several degrees of Alzheimer pathology. Certain genetic indicators for the judgment of DLB have also been improved, such as brain single photon emission computed tomography/positron emission tomography (SPECT/PET), dopamine transporter imaging [¹²³I-Fluoropropyl-2-beta-carbomethoxy-3-beta(4-iodophenyl) nortropane (FP-CIT) SPECT] (Walker et al., 2007), and meta-iodobenzylguanidine (MIBG) myocardial scintigraphy (Yoshita et al., 2001). α -syn was found to be the key component of LB. α -syn is a 140 amino acid protein encoded by the SNCA gene, found in nuclei and presynaptic parts, but its purpose has not yet been known (Kosaka, 2014). Though, the cerebral type of LBD in which plentiful LB was found in the cerebral cortex even with there being only a little in the brain stem nuclei, proposes the probability that Lewy pathology happens in the cerebral cortex and propagates downward to the brain stem. Lewy pathology may possibly also start from Auerbach's plexus of the lower esophagus or the olfactory bulb (Kosaka, 2014). Recently, the possibility that accumulation of α -syn could spread trans-cellular all over the brain in a prion-like technique has been found (Kosaka, 2014).

Parkinson's Disease

PD is a frequently restricting syndrome seen in individuals from all countries and geographical spots, with medical symptoms developing also in a broad age variety (Lees et al. 2009). Motor symptoms might involve bradykinesia (tardiness of movements with an advanced damage of adequacy or speed amid endeavored quick sporadic developments of body parts), rest tremor (cadenced oscillatory involuntary movement that begins when the influenced body part is relaxed and remained by a surface, accordingly make away from the move of gravitational powers). Notwithstanding unbending nature (a hoisted muscle tone detected through examination by aloof movement of the influenced part, including both flexor and extensor muscle gatherings), and postural and stride disability (Massano and Bhatia, 2012). While, the non-motor symptoms could include neuropsychiatric disorders, dysautonomia, sleep disorders, and sensory dysfunction (Massano and Bhatia, 2012). Brain anatomical scanning, whichever by CT or MRI should continuously be done; but the latter is chosen, since certain constructive results infrequently expose other investigative things (Sitburana and Ondo 2009). Dopamine functional scanning might be measured to approve that progressive parkinsonism is the source of signs. Positron emission tomography (PET) with fluorodopa is one of the techniques presented, but the prices and inadequate availability make it hard for using it. So, dopamine transporter (DAT) imaging with single-photon emission CT (DAT-SPECT) is a very valuable method, because it is delicate for the revealing of presynaptic dopaminergic neuron damage in the striatum (Kagi et al., 2010). Nowadays, MRI is chosen over CT, and family report is not considered as a prohibiting principle.

Posterior Cortical Atrophy

Posterior cortical atrophy (PCA) is a neurodegenerative disease that is manifested by an advanced decrease in visuospatial, visuoperceptual, literacy and praxis abilities (Lehmann et al., 2011). The advanced neurodegeneration damaging parietal, occipital and occipito-temporal cortices which cause PCA is referred to AD in the most of cases. Though, other causal etiologies counting DLB, CBD and prion syndrome have also stood recognized, and not all PCA cases have deteriorated on medical examination (Lehmann et al., 2011). Cross-sectional voxel-based morphometry (VBM) has shown extensive grey matter changes between PCA cases and normal people, with the maximum important decreases established in sections of the occipital and parietal lobes, trailed by sections in the temporal lobe (Lehmann et al., 2011). Direct assessment between PCA and typical AD by both VBM and cortical thickness methods have confirmed more right parietal and less left medial temporal and hippocampal degeneration in PCA (Migliaccio et al., 2012). It must be well-known that a quantity of readings report asymmetric atrophy patterns in PCA (right > lift), but these changes may reproduce choice preferences in the diagnosis and staffing of cases with noticeable visual impairment. Incomplete diffusion tensor imaging (DTI) records also propose PCA decreases the integrity of white matter tracts in posterior brain regions (Migliaccio et al., 2012). Records from practical imaging readings using single photon emission computed tomography (SPECT) and fluorodeoxyglucose positron emission tomography (FDG-PET) are mainly reliable with anatomical alterations in parieto-occipital parts (Gardini et al., 2011).

Progressive Supranuclear Palsy

PSP is a widespread subcortical neurofibrillary deterioration mostly originate in the globus pallidus, subthalamic nucleus, substantia nigra and cerebellar dentate nucleus were manifested as the pathological substrates of the clinic-pathologic entity of progressive supranuclear palsy (PSP), also recognized as Steele-Richardson-Olszewski syndrome (Ling, 2016). Medical subtypes of PSP-parkinsonism (PSP-P) and PSP-pure akinesia with gait freezing (PSP-PAGF) have a further benign growth with a survival period of 10 years or more. Also, both subclasses have a general tau burden fewer than those in PSP-RS and the spreading of irregular tau is comparatively limited to the brain stem (Williams et al., 2007). Clinical signs could involve primary postural instability and/or falls, eye movement irregularities, cognitive weakening, frontal behavior, non-fluent aphasia and/or apraxia of speech, limb dystonia, pyramidal and Babinski's signs, levodopa response, and dysautonomia (Ling, 2016). Conventional MRI of the brain is valuable to eliminate extensive cerebrovascular disease, leukodystrophy, regular pressure hydrocephalus, structural midbrain lesions and, infrequently, manganese poisoning which can all pretense as PSP (Ling, 2016). Laboratory investigations for syphilis and human immunodeficiency virus (HIV) serology, autoimmune disorders, paraneoplastic disease autoantibodies, antibodies for stiff-person syndrome and Niemann Pick type C may be measured (Ling, 2016). Dopamine transporter single photon emission computed tomography (SPECT) imaging displays diminished tracer uptake in the striatum which is a helpful finding to discriminate PSP from other diseases such as cerebrovascular disease and normal pressure hydrocephalus (Kagi et al., 2010). PET imaging with tau ligand is a favorable radiologic method for investigative and check-up of PSP (Kepe et al., 2013).

Pick's Disease

Pick's disease (PiD) is well-defined pathologically by the occurrence of neuronal presence bodies (Pick bodies) immune-reactive to tau and ubiquitin inside cerebral cortical neurons and inflated achromatic neurons (Pick cells) inside the cerebral cortex (Neary et al., 1998). Depending on the site of the cortical degeneration dissimilar medical patterns have stood illustrated in PiD (Neary et al., 1993). Cases with noticeable medial temporal lobe pathology, particularly deteriorating the amygdala, can have a Klüver-Bucy syndrome (Cummings and Duchen, 1983). Cases with extra damaged frontal lobe pathology usually have FLS (Dickson, 1998). As is pure from this information of this syndrome, the neuropathology of FLS is varied; only a lesser percent have classical PiD. In regions with extreme pathology, the cortex has nearly widespread damage of big pyramidal neurons with the breakdown of the parenchyma, diffuse spongiosis and dense gliosis (Dickson, 1998). The cytoarchitectural properties of the cortex turn into cover. In initial phases or zones with less severe pathology, adequate vacuolation of the neuropil is existing in the upper cortex. Swollen neurons (also denoted to as Pick cells) are existing in the central and lower cortical layers. Functional imaging readings such as PET or SPECT can show bifrontal shortages. This outline compares with the biparietal shortages representative of AD and the irregular superior frontal or para-central deficits of CBD (Dickson, 1998).

Primary Progressive Aphasia

Primary progressive aphasia (PPA) is a medical condition distinguished by the instant beginning and advanced ending of language abilities (Mesulam, 2003). While further mental signs may arise far along in the progression of disease, shortages must be totally restricted to the extent of verbal for at minimum 2 years to accomplish the measures for an identification of PPA. At examination, the most constant conclusion is that of focal deterioration, which is commonly established specially and more strongly in verbal regions, and manifested by neuronal damage, gliosis, and spongiform alterations including the superficial cortical layers (Leger and Johnson, 2007). Cortex could also include infrequent ballooned neurons, named Pick cells. Staining with silver and novel histochemical stains for both the microtubule related protein tau (MAPT or tau) and ubiquitin can expose neuronal and glial insertions in an outline specific to the particular neuropathological development approaching (Leger and Johnson, 2007). Using fMRI (functional MRI) and a grammatically complex sentence comprehension pattern, presented that patients with PPA give fewer initiation within the ventral portion of the inferior frontal cortex, a part suspected to be vital to the handing out of difficult sentences (Cooke et al., 2003). They further assume that this disturbance of a supposed big scale neural system for sentence intellectual capacity forms the source for the difficult grammatical comprehension shortages shown in these cases (Cooke et al 2006).

Spinal Muscular Atrophy

The word spinal muscular atrophy (SMA) is realistic to a varied collection of hereditary syndromes that all distress the spinal MN (Arnold et al., 2015). The diverse types of SMA are related with frequent gene alterations and major phenotypic differences. SMA is generally characterized by distinct faintness (proximal or distal) and manner of inheritance (Arnold et al., 2015). SMN lacking disturbance of other cellular mechanisms could also be significant in the pathogenesis of SMA, and damage of axonal mRNA transportation could also have a key role in SMA (Arnold and Burghes, 2013). Changed transcripts (SMN

deficiency) have been recognized, but a certain relation to the disease pathogenesis has not been established. For instance, the protein Stasimon was recognized lately as a probable disease-associated mark of altered splicing produced by SMN insufficiency (Imlach et al., 2012). Muscle biopsy and electrodiagnostic analysis were typical measures for assessment, but since molecular analysis is offered, these and other analytical examinations (MRI) are not necessary. Electrophysiological methods of electromyography (EMG), compound muscle action potential (CMAP), and motor unit number estimation (MUNE) have shown excellent association with clinical progressing, age, and functional grade (Arnold et al., 2015).

Spinocerebellar Ataxias

The most common congenital ataxias, currently named spinocerebellar ataxias (SCAs). Gradually advanced ataxia along with cerebellar deterioration is frequently hereditary in basis. The previous 5 decades have observed an improvement in our knowledge of the reasons of dominantly hereditary ataxias, now acknowledged as the spinocerebellar ataxias SCAs (Paulson, 2009). SCA is a deficit of coordination, mainly of gait. So, when a doctor examines patient with ataxia, it normally is someone with gait inequity related with limb incoordination counting difficulties with gross and fine motor control (Paulson, 2009). The detection over 10 years ago that polyQ sickness brain comprises intracellular presences of the disease protein recommended that the development enhances misfolding of the disease protein, causing in accumulation. The principal advantage of investigative hereditary analysis is that it could deliver an exact and accurate diagnosis (Paulson, 2009). SCA gene test in a case whose signs are constant with a hereditary form of ataxia, but whose family report is indefinite or lacking, can approve the medical analysis with efficacy, cost, and inevitability. In an ataxic case, gene investigations are accurate and precise, whereas brain MRI is not. In SCA gene analysis can agree an identification from between a collection of clinically comparable chromosomal situations (Paulson, 2009).

CONCLUSION

This chapter targets to condense recent thoughts of disease categorization with a concentration on the molecular pathological features for neurodegenerative situations of the old age where microscopically noticeable protein aggregations have stood defined. Misrepresentation of infection gathering with the point of creating treatments for whatever number people as could be allowed indicating comparable clinical highlights has not prompted huge achievement. Characterizing novel groups of patients with NDs for stratified helpful methodologies, in light of consistent, careful, orchestrated, and continually refreshed clinical, neuroimaging, biochemical, and hereditary characterizations with lasting neuropathology-based quality control are by all accounts a superior approach in the time of precise medication.

REFERENCES

Apel, K., & Hirt, H. (2004). Reactive oxygen species: Metabolism, oxidative stress, and signal transduction. *Annual Review of Plant Biology*, *55*(1), 373–399. doi:10.1146/annurev.arplant.55.031903.141701 PMID:15377225

Armstrong, M. J. (2014). Diagnosis and treatment of corticobasal degeneration. *Current Treatment Options in Neurology*, *16*(3), 282. doi:10.100711940-013-0282-1 PMID:24469408

Armstrong, M. J., Litvan, I., Lang, A. E., Bak, T. H., Bhatia, K. P., Borroni, B., ... Weiner, W. J. (2013). Criteria for the diagnosis of corticobasal degeneration. *Neurology*, *80*(5), 496–503. doi:10.1212/ WNL.0b013e31827f0fd1 PMID:23359374

Arnold, W. D., & Burghes, A. H. (2013). Spinal muscular atrophy: Development and implementation of potential treatments. *Annals of Neurology*, 74(3), 348–362. doi:10.1002/ana.23995 PMID:23939659

Arnold, W. D., Kassar, D., & Kissel, J. T. (2015). Spinal muscular atrophy: Diagnosis and management in a new therapeutic era. *Muscle & Nerve*, *51*(2), 157–167. doi:10.1002/mus.24497 PMID:25346245

Boxer, A. L., Mackenzie, I. R., Boeve, B. F., Baker, M., Seeley, W. W., Crook, R., ... Rademakers, R. (2011). Clinical, neuroimaging and neuropathological features of a new chromosome 9p-linked FTD-ALS family. *Journal of Neurology, Neurosurgery, and Psychiatry*, 82(2), 196–203. doi:10.1136/jnnp.2009.204081 PMID:20562461

Brettschneider, J., Del Tredici, K., & Lee, V. M. (2015). Spreading of pathology in neurodegenerative diseases: A focus on human studies. *Nature Reviews. Neuroscience*, *16*(2), 109–120. doi:10.1038/ nrn3887 PMID:25588378

Brown, R. C., Lockwood, A. H., & Sonawane, B. R. (2005). Neurodegenerative diseases: An overview of environmental risk factors. *Environmental Health Perspectives*, *113*(9), 1250–1256. doi:10.1289/ehp.7567 PMID:16140637

Cadet, J. L. (1988). Free radical mechanisms in the central nervous system: An overview. *The International Journal of Neuroscience*, 40(1-2), 13–18. doi:10.3109/00207458808985722 PMID:2840405

Carrell, R. W., & Lomas, D. A. (1997). Conformational disease. *Lancet*, 350(9071), 134–138. doi:10.1016/S0140-6736(97)02073-4 PMID:9228977

Cooke, A., DeVita, C., Gee, J., Alsop, D., Detre, J., Chen, W., & Grossman, M. (2003). Neural basis for sentence comprehension deficits in frontotemporal dementia. *Brain and Language*, 85(2), 211–221. doi:10.1016/S0093-934X(02)00562-X PMID:12735939

Cooke, A., Grossman, M., DeVita, C., Gonzalez-Atavales, J., Moore, P., Chen, W., ... Detre, J. (2006). Large-scale neural network for sentence processing. *Brain and Language*, *96*(1), 14–36. doi:10.1016/j. bandl.2005.07.072 PMID:16168473

Cummings, J. L., & Duchen, L. W. (1981). Kluver-Bucy syndrome in Pick disease: Clinical and pathologic correlations. *Neurology*, *31*(11), 1415–1422. doi:10.1212/WNL.31.11.1415 PMID:7198189

Delatycki, M. B., Williamson, R., & Forrest, S. M. (2000). Friedreich ataxia: An overview. *Journal of Medical Genetics*, *37*(1), 1–8. doi:10.1136/jmg.37.1.1 PMID:10633128

Dickson, D. W. (1998). Pick's disease: A modern approach. *Brain Pathology (Zurich, Switzerland)*, 8(2), 339–354. doi:10.1111/j.1750-3639.1998.tb00158.x PMID:9546291

Neurodegenerative Disorders

Dickson, D. W., Bergeron, C., Chin, S. S., Duyckaerts, C., Horoupian, D., Ikeda, K., ... Litvan, I. (2002). Office of Rare Diseases neuropathologic criteria for corticobasal degeneration. *Journal of Neuropathology and Experimental Neurology*, *61*(11), 935–946. doi:10.1093/jnen/61.11.935 PMID:12430710

Duyckaerts, C., Delatour, B., & Potier, M. C. (2009). Classification and basic pathology of Alzheimer disease. *Acta Neuropathologica*, *118*(1), 5–36. doi:10.100700401-009-0532-1 PMID:19381658

Eder, K., Kish, S. J., Kirchgessner, M., & Ross, B. M. (1998). Brain phospholipids and fatty acids in Friedreich's ataxia and spinocerebellar atrophy type-1. *Movement Disorders*, *13*(5), 813–819. doi:10.1002/mds.870130510 PMID:9756151

Fink, A. L. (2006). The aggregation and fibrillation of alpha-synuclein. *Accounts of Chemical Research*, *39*(9), 628–634. doi:10.1021/ar050073t PMID:16981679

Gardini, S., Concari, L., Pagliara, S., Ghetti, C., Venneri, A., & Caffarra, P. (2011). Visuo-spatial imagery impairment in posterior cortical atrophy: A cognitive and SPECT study. *Behavioural Neurology*, *24*(2), 123–132. doi:10.1155/2011/547451 PMID:21606573

Hardiman, O., van den Berg, L. H., & Kiernan, M. C. (2011). Clinical diagnosis and management of amyotrophic lateral sclerosis. *Nature Reviews. Neurology*, 7(11), 639–649. doi:10.1038/nrneurol.2011.153 PMID:21989247

Harding, A. (1984). The hereditary ataxias and related disorders. Edinburgh, UK: Churchill Livingstone.

Head, M. W., & Ironside, J. W. (2012). Review: Creutzfeldt-Jakob disease: prion protein type, disease phenotype and agent strain. *Neuropathology and Applied Neurobiology*, *38*(4), 296–310. doi:10.1111/j.1365-2990.2012.01265.x PMID:22394291

Hughes, J. T., Brownell, B., & Hewer, R. L. (1968). The peripheral sensory pathway in friedreich's ataxia. An examination by light and electron microscopy of the posterior nerve roots, posterior root ganglia, and peripheral sensory nerves in cases of friedreich's ataxia. *Brain*, *91*(4), 803–818. doi:10.1093/ brain/91.4.803 PMID:4178703

Imlach, W. L., Beck, E. S., Choi, B. J., Lotti, F., Pellizzoni, L., & McCabe, B. D. (2012). SMN is required for sensory-motor circuit function in Drosophila. *Cell*, *151*(2), 427–439. doi:10.1016/j.cell.2012.09.011 PMID:23063130

Jack, C. R. Jr, Wiste, H. J., Weigand, S. D., Rocca, W. A., Knopman, D. S., Mielke, M. M., ... Petersen, R. C. (2014). Age-specific population frequencies of cerebral beta-amyloidosis and neurodegeneration among people with normal cognitive function aged 50-89 years: A cross-sectional study. *Lancet Neurology*, *13*(10), 997–1005. doi:10.1016/S1474-4422(14)70194-2 PMID:25201514

Janus, C., Pearson, J., McLaurin, J., Mathews, P. M., Jiang, Y., Schmidt, S. D., ... Westaway, D. (2000). A beta peptide immunization reduces behavioural impairment and plaques in a model of Alzheimer's disease. *Nature*, 408(6815), 979–982. doi:10.1038/35050110 PMID:11140685

Kagi, G., Bhatia, K. P., & Tolosa, E. (2010). The role of DAT-SPECT in movement disorders. *Journal of Neurology, Neurosurgery, and Psychiatry*, *81*(1), 5–12. doi:10.1136/jnnp.2008.157370 PMID:20019219

Karve, T. M., & Cheema, A. K. (2011). Small changes huge impact: The role of protein posttranslational modifications in cellular homeostasis and disease. *Journal of Amino Acids*, 2011, 207691. doi:10.4061/2011/207691 PMID:22312457

Kepe, V., Bordelon, Y., & Boxer, A. (2013). PET imaging of neuropathology in tauopathies: Progressive supranuclear palsy. *Journal of Alzheimer's Disease*, *36*(1), 145–153. PMID:23579330

Kosaka, K. (2014). Lewy body disease and dementia with Lewy bodies. *Proceedings of the Japan Academy*. *Series B, Physical and Biological Sciences*, 90(8), 301–306. doi:10.2183/pjab.90.301 PMID:25311140

Kovacs, G. G. (2016). Molecular Pathological Classification of Neurodegenerative Diseases: Turning towards Precision Medicine. *International Journal of Molecular Sciences*, *17*(2), 189. doi:10.3390/ ijms17020189 PMID:26848654

Kovacs, G. G., Alafuzoff, I., Al-Sarraj, S., Arzberger, T., Bogdanovic, N., Capellari, S., ... Budka, H. (2008). Mixed brain pathologies in dementia: The BrainNet Europe consortium experience. *Dementia and Geriatric Cognitive Disorders*, *26*(4), 343–350. doi:10.1159/000161560 PMID:18849605

Kovacs, G. G., Botond, G., & Budka, H. (2010). Protein coding of neurodegenerative dementias: The neuropathological basis of biomarker diagnostics. *Acta Neuropathologica*, *119*(4), 389–408. doi:10.100700401-010-0658-1 PMID:20198481

Kovacs, G. G., & Budka, H. (2010). Current concepts of neuropathological diagnostics in practice: Neurodegenerative diseases. *Clinical Neuropathology*, 29(5), 271–288. doi:10.5414/NPP29271 PMID:20860890

Kurland, L. T. (1988). Amyotrophic lateral sclerosis and Parkinson's disease complex on Guam linked to an environmental neurotoxin. *Trends in Neurosciences*, *11*(2), 51–54. doi:10.1016/0166-2236(88)90163-4 PMID:2465598

Lee, C., & Yu, M. H. (2005). Protein folding and diseases. *Journal of Biochemistry and Molecular Biology*, *38*(3), 275–280. PMID:15943901

Lee, S. E., Rabinovici, G. D., Mayo, M. C., Wilson, S. M., Seeley, W. W., DeArmond, S. J., ... Miller, B. L. (2011). Clinicopathological correlations in corticobasal degeneration. *Annals of Neurology*, *70*(2), 327–340. doi:10.1002/ana.22424 PMID:21823158

Lees, A. J., Hardy, J., & Revesz, T. (2009). Parkinson's disease. *Lancet*, 373(9680), 2055–2066. doi:10.1016/S0140-6736(09)60492-X PMID:19524782

Leger, G. C., & Johnson, N. (2007). A review on primary progressive aphasia. *Neuropsychiatric Disease* and *Treatment*, *3*(6), 745–752. PMID:19300609

Lehmann, M., Crutch, S. J., Ridgway, G. R., Ridha, B. H., Barnes, J., Warrington, E. K., ... Fox, N. C. (2011). Cortical thickness and voxel-based morphometry in posterior cortical atrophy and typical Alzheimer's disease. *Neurobiology of Aging*, *32*(8), 1466–1476. doi:10.1016/j.neurobiolaging.2009.08.017 PMID:19781814

Lindsay, J., Laurin, D., & Verreault, R. (2002). Risk factors for Alzheimer's disease: A prospective analysis from the Canadian Study of Health and Aging. *American Journal of Epidemiology*, *156*(5), 445–453. doi:10.1093/aje/kwf074 PMID:12196314

Ling, H. (2016). Clinical Approach to Progressive Supranuclear Palsy. *Journal of Movement Disorders*, *9*(1), 3–13. doi:10.14802/jmd.15060 PMID:26828211

Lippa, C. F., Duda, J. E., Grossman, M., Hurtig, H. I., Aarsland, D., Boeve, B. F., ... Wszolek, Z. K. (2007). DLB and PDD boundary issues: Diagnosis, treatment, molecular pathology, and biomarkers. *Neurology*, *68*(11), 812–819. doi:10.1212/01.wnl.0000256715.13907.d3 PMID:17353469

Marder, K., Logroscino, G., Alfaro, B., Mejia, H., Halim, A., Louis, E., ... Mayeux, R. (1998). Environmental risk factors for Parkinson's disease in an urban multiethnic community. *Neurology*, *50*(1), 279–281. doi:10.1212/WNL.50.1.279 PMID:9443493

Martin, L., Latypova, X., & Terro, F. (2011). Post-translational modifications of tau protein: Implications for Alzheimer's disease. *Neurochemistry International*, *58*(4), 458–471. doi:10.1016/j.neuint.2010.12.023 PMID:21215781

Massano, J., & Bhatia, K. P. (2012). Clinical approach to Parkinson's disease: Features, diagnosis, and principles of management. *Cold Spring Harbor Perspectives in Medicine*, 2(6), a008870. doi:10.1101/ cshperspect.a008870 PMID:22675666

McKeith, I. G., Dickson, D. W., Lowe, J., Emre, M., O'Brien, J. T., Feldman, H., ... Yamada, M. (2005). Diagnosis and management of dementia with Lewy bodies: Third report of the DLB Consortium. *Neurology*, *65*(12), 1863–1872. doi:10.1212/01.wnl.0000187889.17253.b1 PMID:16237129

Migliaccio, R., Agosta, F., Toba, M. N., Samri, D., Corlier, F., de Souza, L. C., ... Bartolomeo, P. (2012). Brain networks in posterior cortical atrophy: A single case tractography study and literature review. *Cortex*, 48(10), 1298–1309. doi:10.1016/j.cortex.2011.10.002 PMID:22099855

Moulton, P. V., & Yang, W. (2012). Air pollution, oxidative stress, and Alzheimer's disease. *Journal of Environmental and Public Health*, 2012, 472751. doi:10.1155/2012/472751 PMID:22523504

Neary, D., Snowden, J. S., Gustafson, L., Passant, U., Stuss, D., Black, S., ... Benson, D. F. (1998). Frontotemporal lobar degeneration: A consensus on clinical diagnostic criteria. *Neurology*, *51*(6), 1546–1554. doi:10.1212/WNL.51.6.1546 PMID:9855500

Neary, D., Snowden, J. S., & Mann, D. M. (1993). The clinical pathological correlates of lobar atrophy. *Dementia (Basel, Switzerland)*, *4*(3-4), 154–159. PMID:8401784

Neumann, M., Bentmann, E., Dormann, D., Jawaid, A., DeJesus-Hernandez, M., Ansorge, O., ... Mackenzie, I. R. A. (2011). FET proteins TAF15 and EWS are selective markers that distinguish FTLD with FUS pathology from amyotrophic lateral sclerosis with FUS mutations. *Brain*, *134*(Pt 9), 2595–2609. doi:10.1093/brain/awr201 PMID:21856723

Ou, S. H., Wu, F., Harrich, D., & (1995). Cloning and characterization of a novel cellular protein, TDP-43, that binds to human immunodeficiency virus type 1 TAR DNA sequence motifs. *Journal of Virology*, *69*(6), 3584–3596. PMID:7745706

Parchi, P., Capellari, S., Chen, S. G., Petersen, R. B., Gambetti, P., Kopp, N., ... Kretzschmar, H. (1997). Typing prion isoforms. *Nature*, *386*(6622), 232–234. doi:10.1038/386232a0 PMID:9069279

Parchi, P., Castellani, R., Capellari, S., Ghetti, B., Young, K., Chen, S. G., ... Gambetti, P. (1996). Molecular basis of phenotypic variability in sporadic Creutzfeldt-Jakob disease. *Annals of Neurology*, *39*(6), 767–778. doi:10.1002/ana.410390613 PMID:8651649

Paulson, H. L. (2009). The spinocerebellar ataxias. *Journal of Neuro-Ophthalmology*, 29(3), 227–237. doi:10.1097/WNO0b013e3181b416de PMID:19726947

Philo, J. S., & Arakawa, T. (2009). Mechanisms of protein aggregation. *Current Pharmaceutical Biotechnology*, *10*(4), 348–351. doi:10.2174/138920109788488932 PMID:19519409

Przedborski, S., Jackson-Lewis, V., Naini, A. B., Jakowec, M., Petzinger, G., Miller, R., & Akram, M. (2001). The parkinsonian toxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP): A technical review of its utility and safety. *Journal of Neurochemistry*, *76*(5), 1265–1274. doi:10.1046/j.1471-4159.2001.00183.x PMID:11238711

Przedborski, S., Vila, M., & Jackson-Lewis, V. (2003). Neurodegeneration: What is it and where are we? *The Journal of Clinical Investigation*, *111*(1), 3–10. doi:10.1172/JCI200317522 PMID:12511579

Rahimi, J., & Kovacs, G. G. (2014). Prevalence of mixed pathologies in the aging brain. *Alzheimer's Research & Therapy*, 6(9), 82. doi:10.118613195-014-0082-1 PMID:25419243

Roos, R. A. (2010). Huntington's disease: A clinical review. *Orphanet Journal of Rare Diseases*, 5(1), 40. doi:10.1186/1750-1172-5-40 PMID:21171977

Ross, G. W., & Petrovitch, H. (2001). Current evidence for neuroprotective effects of nicotine and caffeine against Parkinson's disease. *Drugs & Aging*, *18*(11), 797–806. doi:10.2165/00002512-200118110-00001 PMID:11772120

Rowland, L. P., & Shneider, N. A. (2001). Amyotrophic lateral sclerosis. *The New England Journal of Medicine*, 344(22), 1688–1700. doi:10.1056/NEJM200105313442207 PMID:11386269

Rudd, P. M., Wormald, M. R., Wing, D. R., Prusiner, S. B., & Dwek, R. A. (2001). Prion glycoprotein: Structure, dynamics, and roles for the sugars. *Biochemistry*, 40(13), 3759–3766. doi:10.1021/bi002625f PMID:11300755

Sitburana, O., & Ondo, W. G. (2009). Brain magnetic resonance imaging (MRI) in parkinsonian disorders. *Parkinsonism & Related Disorders*, *15*(3), 165–174. doi:10.1016/j.parkreldis.2008.04.033 PMID:19059803

Soto, C., & Estrada, L. D. (2008). Protein misfolding and neurodegeneration. *Archives of Neurology*, 65(2), 184–189. doi:10.1001/archneurol.2007.56 PMID:18268186

Tanner, C. M. (1989). The role of environmental toxins in the etiology of Parkinson's disease. *Trends in Neurosciences*, *12*(2), 49–54. doi:10.1016/0166-2236(89)90135-5 PMID:2469210

Tanner, C. M., Ottman, R., & Goldman, S. M. (1999). Parkinson disease in twins: An etiologic study. *Journal of the American Medical Association*, 281(4), 341–346. doi:10.1001/jama.281.4.341 PMID:9929087

Thal, D. R., Walter, J., Saido, T. C., & Fändrich, M. (2015). Neuropathology and biochemistry of Abeta and its aggregates in Alzheimer's disease. *Acta Neuropathologica*, *129*(2), 167–182. doi:10.100700401-014-1375-y PMID:25534025

The Huntington's Disease Collaborative Research Group. (1993). A novel gene containing a trinucleotide repeat that is expanded and unstable on Huntington's disease chromosomes. *Cell*, 72(6), 971-983.

Treuheit, M. J., Kosky, A. A., & Brems, D. N. (2002). Inverse relationship of protein concentration and aggregation. *Pharmaceutical Research*, *19*(4), 511–516. doi:10.1023/A:1015108115452 PMID:12033388

Trottier, Y., Biancalana, V., & Mandel, J. L. (1994). Instability of CAG repeats in Huntington's disease: Relation to parental transmission and age of onset. *Journal of Medical Genetics*, *31*(5), 377–382. doi:10.1136/jmg.31.5.377 PMID:8064815

Tutar, L., & Tutar, Y. (2010). Heat shock proteins; an overview. *Current Pharmaceutical Biotechnology*, *11*(2), 216–222. doi:10.2174/138920110790909632 PMID:20170474

Tyas, S. L., Manfreda, J., Strain, L. A., & Montgomery, P. R. (2001). Risk factors for Alzheimer's disease: A population-based, longitudinal study in Manitoba, Canada. *International Journal of Epidemiology*, *30*(3), 590–597. doi:10.1093/ije/30.3.590 PMID:11416089

Vanacore, N., Bonifati, V., Fabbrini, G., Colosimo, C., Marconi, R., Nicholl, D., ... Meco, G. (2000). Smoking habits in multiple system atrophy and progressive supranuclear palsy. European Study Group on Atypical Parkinsonisms. *Neurology*, *54*(1), 114–119. doi:10.1212/WNL.54.1.114 PMID:10636135

Walker, L. C., Diamond, M. I., Duff, K. E., & Hyman, B. T. (2013). Mechanisms of protein seeding in neurodegenerative diseases. *JAMA Neurology*, *70*(3), 304–310. doi:10.1001/jamaneurol.2013.1453 PMID:23599928

Walker, Z., Jaros, E., Walker, R. W., Lee, L., Costa, D. C., Livingston, G., ... Katona, C. L. E. (2007). Dementia with Lewy bodies: A comparison of clinical diagnosis, FP-CIT single photon emission computed tomography imaging and autopsy. *Journal of Neurology, Neurosurgery, and Psychiatry*, 78(11), 1176–1181. doi:10.1136/jnnp.2006.110122 PMID:17353255

Wang, W. (2005). Protein aggregation and its inhibition in biopharmaceutics. *International Journal of Pharmaceutics*, 289(1-2), 1–30. doi:10.1016/j.ijpharm.2004.11.014 PMID:15652195

Weimer, J. M., Kriscenski-Perry, E., Elshatory, Y., & Pearce, D. A. (2002). The neuronal ceroid lipofuscinoses: Mutations in different proteins result in similar disease. *Neuromolecular Medicine*, 1(2), 111–124. doi:10.1385/NMM:1:2:111 PMID:12025857

Whitwell, J. L., Jack, C. R. Jr, Boeve, B. F., Parisi, J. E., Ahlskog, J. E., Drubach, D. A., ... Josephs, K. A. (2010). Imaging correlates of pathology in corticobasal syndrome. *Neurology*, *75*(21), 1879–1887. doi:10.1212/WNL.0b013e3181feb2e8 PMID:21098403

Wijesekera, L. C., & Leigh, P. N. (2009). Amyotrophic lateral sclerosis. Orphanet Journal of Rare Diseases, 4(1), 3. doi:10.1186/1750-1172-4-3 PMID:19192301

Williams, D. R., Holton, J. L., Strand, K., Revesz, T., & Lees, A. J. (2007). Pure akinesia with gait freezing: A third clinical phenotype of progressive supranuclear palsy. *Movement Disorders*, 22(15), 2235–2241. doi:10.1002/mds.21698 PMID:17712855

Wilson, A. C., Dugger, B. N., & Dickson, D. W. (2011). TDP-43 in aging and Alzheimer's disease - a review. *International Journal of Clinical and Experimental Pathology*, 4(2), 147–155. PMID:21326809

Yoshita, M., Taki, J., & Yamada, M. (2001). A clinical role for [(123)I]MIBG myocardial scintigraphy in the distinction between dementia of the Alzheimer's-type and dementia with Lewy bodies. *Journal of Neurology, Neurosurgery, and Psychiatry*, 71(5), 583–588. doi:10.1136/jnnp.71.5.583 PMID:11606666

Zaleska, M. M., & Floyd, R. A. (1985). Regional lipid peroxidation in rat brain in vitro: Possible role of endogenous iron. *Neurochemical Research*, *10*(3), 397–410. doi:10.1007/BF00964608 PMID:4000395

Zarei, S., Carr, K., Reiley, L., Diaz, K., Guerra, O., Altamirano, P. F., ... Chinea, A. (2015). A comprehensive review of amyotrophic lateral sclerosis. *Surgical Neurology International*, *6*(1), 171. doi:10.4103/2152-7806.169561 PMID:26629397

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Chapter 10 **Tau Pathology:** A Step Towards Understanding Neurodegenerative Disorders Network Complexity

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ABSTRACT

A number of neurodegenerative disorders (NDs) are usually referred as tauopathies and characterized by the disappearance or disintegration of tau protein from microtubules. Alzheimer's disease (AD), Pick's disease (PiD), Parkinson's disease (PD) are directly or indirectly associated with tauopathy. Tau is a protein which is usually associated with microtubule. Microtubules are the backbone of neurons, and tau provides a support to microtubule stability. Hyperphosphorylation of tau leads to its separation from microtubule, consequently forming neurofibrillary tangles and resulting in a condition of dementia. Therapeutic implication on tauopathy is symptomatic as there is no exact regulation mechanism known till date. This chapter helps in the comprehensive study of biomarkers and pathways involved in tauopathy to decipher the complexity of the system, resulting in candidate drug target for the management of NDs.

INTRODUCTION

NDs like AD and PD are responsible for a notable increase in the proportion of mortality and morbidity in the developed world today (L. E. Hebert, Beckett, Scherr, & Evans, 2001; Liesi E. Hebert, Scherr, Bienias, Bennett, & Evans, 2003). It is due to the result of increase in the life expectancy of individuals and the change in the population demographics like the neurodegenerative movement disorders

DOI: 10.4018/978-1-5225-5282-6.ch010

which are more common nowadays, aging of baby boomers as well as the neurodegenerative dementias (Brookmeyer, Gray, & Kawas, 1998; Samii, Nutt, & Ransom, 2004). With the aging of the population will provide an improved perceptive of such diseases and this will provide a vital part in the development of many effective therapies and to combat the shocking costs of such diseases (Ernst, Hay, Fenn, Tinklenberg, & Yesavage, 1997). Unifying pathogenesis theories in ND offers an opportunity for the development of therapeutic strategies with broad range of applications in the prevention of disease and a chance for declining morbidity and mortality due to these disorders in the elderly population (Forman, Trojanowski, & Lee, 2004). The lines of analysis showing convergence have exposed a potential single common pathogenic mechanism that underlies various diverse neurodegenerative disorders (the deposition and aggregation of misfolded proteins). Almost all the major neurodegenerative disease has been characterized pathologically by the insidious buildup of insoluble filamentous aggregates of usually soluble proteins in the central nervous system (CNS) (Irvine, El-Agnaf, Shankar, & Walsh, 2008). Since the filamentous aggregates show the tinctorial and ultrastructural characteristics of amyloid, which are ~10-nm-wide fibrils with crossed β -pleated sheet structures that stain with Congo red, thioflavin-S, etc; these diseases can together be grouped as brain amyloidosis (Rajamohamedsait& Sigurdsson, 2012).

The main question comes in mind, what is actually responsible for the remarkable phenotypic diversity found in the above mentioned diseases? Every associated brain amyloidosis is differentiated by diverse temporal and regional patterns of aggregates deposition, changing cellular hosts or extracellular locales of the aggregates, and various protein constituents of the aggregates (Guo & Lee, 2014). All of these characters, along with the innate and variable reactions of the patients to the aggregates that might vary the cascade of events which guide to a particular temporal and regional pattern of neuronal dysfunction (Morris, Clark, & Vissel, 2014). This can result in death, revealing as a particular clinical syndrome like dementia in AD or a movement disorder in PD. Therefore, looking from a pathological view point, neurodegenerative entity can be well explained by the nature and pattern of the deposition of amyloid in the brain. Unluckily, the category and pattern of the amyloidosis in the brain does not always relate fine with the experimental clinical phenotype which were observed (Allen, Robinson, Snowden, Davidson, & Mann, 2014). The variability in connections has advanced to a perplexing nosology that at times need clinicians to explain phenotypes with respect to the presumed existence of the pathological lesions like dementia along with Lewy bodies. At times there is a necessity for pathologists to illustrate lesions by means of clinical language despite of the patient's actual clinical presentation as in the case of progressive supranuclear palsy (PSP). One of the right ways to get around this turmoil is by achieving chemical analytes of biological fluids and neuroimaging biomarkers. These biological entities will permit the clinicians to differentiate between brain amyloidosis based on the character and level of the brain pathology along with the particular amyloidogenic proteins caught up in disease pathogenesis (Macedo & Cordeiro, 2017). These NDs share common mechanisms which involves accumulation of CNS misfolded proteins which provides an idea that these disorders might be linked to similar targets for the advancement of diagnostic and therapeutic agents.

In this context, AD, tauopathies (PiD, cortical basal degeneration, and PSP), and the synucleinopathies (dementia with Lewy bodies, PD, and multiple system atrophy) are discussed as models of the brain amyloidosis that are found in many ageing-related neurodegenerative disorders (Hassan, Whitwell, & Josephs, 2011; Levin, Kurz, Arzberger, Giese, & Höglinger, 2016; Tsai & Boxer, 2014). It further deals with mechanistic understanding of the deregulation of neurodegenerative pathways through various signaling cascades. In NDs, the disease spread goes behind disease-specific patterns that look like the structural design of brain connectivity networks. The point which still remains unclear is that, what

makes the disease spread along such networks. Literature survey and all results together may provide support for the hypothesis that the spread of neurodegeneration across neural networks is through a specific cascade. This chapter discusses the role of tau in various forms of dementia and its involvement in molecular pathway. Screening of key biomarkers using various network approaches, understanding of neuronal signaling is simplified in this chapter.

BACKGROUND

The current spreading out of the public interactome databases allow researchers to proceed through various computational methods used for network medicine (Barabási, Gulbahce, & Loscalzo, 2011). The aim of network medicine is to investigate the pathogenic mechanism of a specific disease and additionally to deduce the complex relations of diseases in an organized way. One of the key approaches is the investigation of the human protein-protein interaction (PPI) network to study disease causing genes through their consequent protein's products which are afterwards used to create the disease PPI network (Ideker & Sharan, 2008). Research based on PPI networks for various diseases has achieved remarkable outcomes (Kann, 2007; Navlakha & Kingsford, 2010; Nguyen & Ho, 2012; Nguyen, Liu, & Jordán, 2011; Oti, Snel, Huynen, & Brunner, 2006; Schuster-Böckler & Bateman, 2008). Various on-going and recent studies have analyzed NDs using PPI; though, they generally considered a particular disease, like AD (Goñi et al., 2008; Krauthammer, Kaufmann, Gilliam, & Rzhetsky, 2004; Wang et al., 2016). Another work inferred overlapping regulators of different NDs in various organisms (Chen & Burgoyne, 2012), the direct commonality among NDs in regard to their pathways (Limviphuvadh, Tanaka, Goto, Ueda, & Kanehisa, 2007), or the construction of NDs network which is based on PPI networks, regulatory networks and Boolean networks (Vasaikar, Padhi, Jayaram, & Gomes, 2013). The earlier works which focused upon the construction of PPI networks linked to NDs have still not quantified the topological relations between NDs. Furthermore, the indirect network associations underlying functionality linkages between NDs are still not clear.

AMYLOID BETA ASSOCIATION WITH NEURODEGENERATIVE DISORDERS

Aβ is the protein that forms pathogenic aggregates in the AD brain. The largest aggregates of Aβ are present in extracellular plaques, although data suggest that Aβ aggregates may also form within neurons (Echeverria & Cuello, 2002; Jindal & Bansal, 2016; Oddo et al., 2003). Aβ can be toxic when applied to the targeted proteins, which may accumulate and cause cellular dysfunction and neuronal death (Singh, Srivastav, Yadav, Srikrishna, & Perry, 2016). Aggregation of pathogenic proteins in association with the plasma membrane can impair the function of various transporters (ion-motive adenyl pyrophosphatase or ATPases that catalyze decomposition of ATP into ADP; glucose and glutamate transporters), ion channels, receptors (for neurotransmitters, growth factors, and cell adhesion molecules) and transduction proteins (GTP-binding proteins, adenylate cyclase, phospholipases, kinases) (Mattson, 2004). The protein aggregation that occurs in association with cytoskeleton proteins may disrupt axonal transport and synaptic signaling (Mietelska-Porowska, Wasik, Goras, Filipek, & Niewiadomska, 2014). Alterations in endoplasmic reticulum functions including abnormal stress responses and perturbed calcium regulation have been documented in protein aggregation disorders (A. Kumar & Singh, 2017; Panigrahi

& Singh, 2013). Increasing evidence suggests that intracellular aggregates of α -synuclein, huntingtin and prions impair proteasome-mediated degradation of proteins resulting the accumulation of damaged proteins in neurons (Ciechanover & Kwon, 2015). Mitochondrial dysfunction has been widely documented in patients and experimental models of AD, PD, and Huntington's disease (HD). Aggregation of A β , α -synuclein, and expanded polyglutamine proteins might have direct effects on mitochondria or may indirectly compromise mitochondrial function by inducing oxidative stress and disrupting calcium homeostasis. Finally, aggregates of polyglutamine-expanded proteins and other pathogenic proteins can form in the nucleus where they may perturb the transcription of genes (Panigrahi & Singh, 2012).

 $A\beta$ aggregation and mutations in tau result in breakage of bond between microtubules which is the skeleton of neuron, ultimately resulting in neuronal damage. The impact of these selected biomarkers can be directly traced and handled until we understand the signaling mechanism in normal and neuro-degenerative conditions.

SIGNALING MECHANISMS FOR NEURODEGENERATIVE DISORDERS

Ca²⁺ Blockers and a Combinational Approach for the Treatment of Neurodegenerative Disorders

The majority of the scientific attempt is paying attention on classification of the major reasons of the mentioned diseases and in the development of the methods to target them. Like, in case of AD, the main reason of disease is believed to be amyloid accumulation (Nilsson et al., 2010). Therefore, the much needed research is headed in the path of searching ways to avoid buildup of amyloid through blocking the fabrication or by the facilitation of its clearance from the brain. The major cause of the disease in the case of HD is the expression of mutant huntingtin protein (Irvine et al., 2008). The effort here is focused on trying to lessen the expression of mutant huntingtin in the brain which can be done with the help of antisense or ribonucleic acid interference knockdown. These approaches have been tricky to decode to the clinic in spite of outstanding scientific rationales. In case of HD clinical trials, already existing hindrance is the advancement of an antisense brain-delivery system which might be helpful in humans. RNAi methodologies might not initiate in HD clinical trials, unless it gets achieved. The clinical trials involving amyloid-binding compound tramiprosate along with tarenflurbil (Flurizan) which is a γ -secretase inhibitor have failed (Lukiw, 2012). Moreover while considering AD, the amyloid-binding monoclonal antibodies have yielded a few restricted benefits in the clinical trials.

Neuronal Ca²⁺Signaling and Aging

Various studies related to comparative analysis were done over the neurons taken from rodents which show the neuronal calcium ion signaling machinery undergoing major changes which are age-related (Toescu & Verkhratsky, 2007). A model of changes dependent on age in hippocampal calcium ion handling is newly planned (Gant, Sama, Landfield, & Thibault, 2006). The leading modulations in the aging neurons comprise of the amplified release of calcium ions from the intracellular stores via different receptors like inositol (1,4,5)-trisphosphate receptors (InsP3R) and ryanodine receptors (RyanRs) (Núñez-Santana et

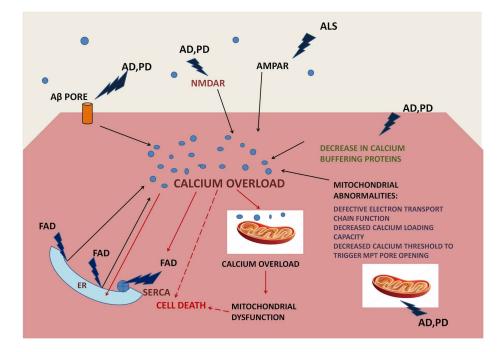


Figure 1. Role of calcium accumulation in neurodegenerative disorders through mitochondrial dysfunction (Berridge, 2010)

al., 2014). Amplified Ca^{2+} influx via L-type voltage-gated calcium channels (VGCCs), amplified slow after-hyperpolarization with the initiation of Ca^{2+} -dependent K⁺ channels, lesser input of N-methyl Daspartate receptor (NMDAR)-mediated Ca^{2+} influx and condensed cytosolic Ca^{2+} buffering capacity and moreover the induction of calcineurin and calpains (Catterall, 2011). The substantial changes in neuronal Ca^{2+} dynamics further leads to augmented susceptibility to induction of long-term depression (LTD) and an increase in the threshold frequency for initiation of long-term potentiation (LTP) in aging neurons (Foster, 2007). LTD and LTP refer to activity-dependent and constant variations in synaptic strength, which are generally considered to form a basis for formation and then storage of memories in the brain (Foster, 2007).

The pathways accountable for alterations depending on age in neuronal Ca²⁺ signaling mechanism are not clearly understood. The probable clarification is associated to defects induced with the age in the functioning of mitochondria due to cumulative oxidative damage to mitochondria. The depolarization of the mitochondria from aged neurons and lesser proficient in handling Calcium ions load (Toescu & Verkhratsky, 2007). Changes related to age in the transcription of calcium ion signaling genes were practically obtained in microarray studies (Toescu & Verkhratsky, 2007). A few of these alterations are caused directly due to aging and a few are compensatory, on the other hand, the general picture is constant with alterations due to age in neuronal calcium ion signaling at several levels. The modulations in calcium ion signaling which are detected in the NDs such as PD and AD is discussed using calcium signaling route in given Figure 2.

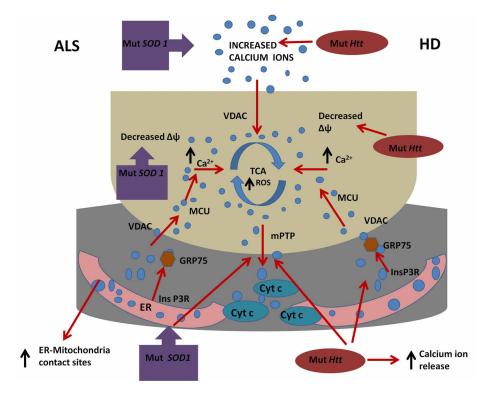


Figure 2. Calcium signaling understanding through mitochondrial route and association with cell death pathway by increase in calcium level (Rizzuto, Stefani, Raffaello, &Mammucari, 2012)

Neuronal Ca²⁺Signaling and Alzheimer's Disease

Amyloid hypothesis is the most governing representation for pathogenesis in the case AD according to which the amplified production of amyloidogenic A β 42 peptide or an enhance in A β 42:A β 40 ratio is a major reason behind neuronal and synaptic loss (Hardy & Selkoe, 2002). The tentative hold for this hypothesis is drawn from:

- 1. Buildup of amyloid plaques in the brains of patients suffering from AD, (Arispe, Rojas, & Pollard, 1993)
- 2. The Familial AD cases that are an outcome of the missense mutations of amyloid-precursor protein (Arispe et al., 1993)
- 3. The FAD cases that result from the missense mutations in presenilins forming a catalytic subunit of the amyloid precursor protein (APP)-cleaving enzyme γ -secretase (Singh et al., 2016).

The prime focus of AD drug development has been the 'amyloid-targeting' therapies. The latest clinical trial outcomes have suggested that additional targets beyond amyloid have to be considered seriously for the treatment of AD (Seabrook, Ray, Shearman, & Hutton, 2007). Amyloid body of confirmation has also recommended that neuronal Calcium ions dyshomeostasis have an essential role in AD.

One of the potential connections involving AD pathogenesis and calcium ions can be observed from A β oligomers that can form Ca²⁺permeable channels in membranes (Arispe et al., 1993). Presence of

phosphatidylserine (PtdS) on the surface of the cell ordinarily demonstrates the cells under vitality shortage conditions and additionally improves the capacity of relationship of A β with the film (G. Lee, Pollard, and Arispe, 2002). Mitochondrial disabilities which are related with the age may bring about the expanded surface PtdS levels in neurons that are influenced and arrange them up for A β -intervened pore development, Ca^{2+} attack and cell passing. The neurons with dense cytosolic ATP levels and unmistakable surface PtdS levels are predominantly vulnerable to A β poisonous quality (Simakova and Arispe, 2007). The ability of A β oligomers to frame channels which demonstrate Ca²⁺ penetrability in neuronal plasma films is steady with most recent in vivo Ca2+ imaging tests which are performed utilizing APP transgenic mice (Kuchibhotla et al., 2008). The examinations portrayed the resting levels of Ca^{2+} were extensively conspicuous in about 35% of neurites found in the quick surroundings of A β plaques. The conceivable elucidation for these results is restricted convergence of A β oligomers in the close-by areas of amyloid plaques which prompts the arrangement of Ca^{2+} porous particle diverts in the neuronal plasma film. The neurites with hoisted Ca^{2+} levels needed spines and demonstrated a unique morphology (Kuchibhotla et al., 2008). These morphological changes in the neurites may get lessened by treatment with the calcineurin (CaN) inhibitor named FK-506 (Kuchibhotla et al., 2008), which proposes that CaN has a critical part to play in obsessive reactions in order to hoist Ca^{2+} levels in the APP transgenic mouse. Moreover, straight impacts of A β on plasma layer Ca²⁺ porousness, A β oligomers even impact neuronal Ca²⁺ homeostasis by altering the activity of N-methyl-D-aspartate receptor (NMDARs) (De Felice et al., 2007; Shankar et al., 2007), alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic corrosive receptors (AMPARs) (Hsieh et al., 2006) and additionally P/Q-sort VGCCs (Nimmrich et al., 2008).

One more likely affiliation associating Ca²⁺ flagging and AD can be comprehended by means of watching different FAD transformations in presenilins impact in irregular Calcium particles flagging. The connection including presenilins and Ca^{2+} flagging was initially uncovered once it was watched that fibroblasts from FAD patients free supranormal measures of Ca^{2+} in response with InsP3 (Ito et al., 1994). Related results were acquired from tries different things with cells communicating FAD mutant presenilins (Leissring, Paul, Parker, Cotman, and LaFerla, 1999) and cortical neurons from FAD presenilin-mutant thump in mice (Stutzmann et al., 2006; Stutzmann, Caccamo, LaFerla, and Parker, 2004). Keeping in mind the end goal to illuminate these results, it is prompted that mutant presenilins impact store-worked Ca^{2+} deluge (Leissring et al., 2000; Yoo et al., 2000), help the movement and in addition the outflow of intracellular Ca²⁺discharge channels, as RyanR (S. L. Chan, Mayne, Holden, Geiger, and Mattson, 2000; Rybalchenko, Hwang, Rybalchenko, and Koulen, 2008; Yoo et al., 2000) and InsP3R (Cai et al., 2006; Cheung et al., 2008), or change the part of the sarcoplasmic and endoplasmic reticulum calcium ATPase (SERCA) ER Ca2+ pump (Green et al., 2008). Presenilins have themselves been seen to work as ER Ca^{2+} spill channels and numerous FAD changes in presentions prompt the loss of ER Ca^{2+} release work, additionally bringing about ER Ca²⁺ over-burden and supranormal Ca²⁺ discharge from the ER (Nelson et al., 2007; Tu et al., 2006). Even though they vary in the particulars of the expected mechanisms, many of the research work was accomplished with various FAD mutations in presenilins resulting in intense Ca²⁺ release from the ER via InsP3R and RyanR.

Neuronal Ca²⁺Signaling and Parkinson's Disease

PD fallout due to the selective loss of dopaminergic neurons in the substantia nigra pars compacta (SNc). The majority of the genes involved in familial PD e.g. *PINK1*, *DJ-1*, *LRRK2* and *Parkin* which encode proteins that are linked to a few aspects of mitochondrial function, pointing to mitochondria

as an essential locus in PD pathogenesis (Abou-Sleiman, Muqit, & Wood, 2006). The widespread idea of the 'dopamine hypothesis' states that dopamine (DA) which acts as a natural toxin and oxidation of cytosolic DA to 6-hydroxy-DA and other metabolites damages mitochondria and causes cell death of SNc neurons (Sulzer, 2007). Consistent with the continuous oxidative damage, affected SNc neurons accumulate large amounts of neuromelanin (NM), which is a lysosome composed of oxyradical DA derivatives of lipids and proteins. The Food and Drug Administration (FDA) approved treatment for PD is administration of levodopa (L-dopa), which is converted to DA and leads to elevation of DA levels in the cytosol and synaptic vesicles of remaining SNc neurons (Alexander, 2004). In an evident disagreement to the 'dopamine hypothesis', the management of L-dopa does not speed up disease expansion in PD patients in spite of increased levels of DA in their brain (Sulzer, 2007). Moreover, PD has lower level of penetrance and the majority of the people do not build up PD in spite of various similar levels of DA in their SNc neurons. The above mentioned observations hint towards a 'multi-hit' hypothesis of PD, which states that SNc neurons in PD provide a collective outcome of DA-related oxidative stress along with 'factor X' (Sulzer, 2007).

Midbrain dopaminergic neurons that express lifted levels of the calcium restricting protein (CaBP) calbindin are reasonably more secure in PD patients and in addition in creature models (Surmeier, 2007). Calpain actuation has additionally been inspected in sporadic PD and furthermore in creature models. α -synuclein is a principle constituent of Lewy bodies saw in the brains of sporadic PD patients and the changes in α -synuclein prompt autosomal-predominant inherited PD. The conceivable strategy for α -synuclein harmfulness is related with the development of little α -synuclein totals or protofibrils (Lashuel, Overk, Oueslati, and Masliah, 2013). Biophysicians thinks that synucleinprotofibrils deliver particle pores in manufactured lipid films (Volles et al., 2001) and empower Ca²⁺ convergence in neurons (Danzer et al., 2007; Furukawa et al., 2006). The proposed technique for synuclein-intervened Ca²⁺ flood might be similar the one proposed for A β oligomer-shaped Ca²⁺ channels. Additionally hold up for the 'Ca²⁺speculations of PD (Surmeier, 2007) was given by physiological investigations. In actuality, the majority of alternate neurons in the sensory system, the SNc dopaminergic neurons utilize CaV1.3 L-sort Ca²⁺ channels to make imprudent pacemaking development in the scope of 2 to 4 Hz (C. S. Chan et al., 2007). The steady Ca²⁺ deluge prompts an over the top metabolic load on SNc neurons and thus making them overwhelmingly vulnerable to optional abuse on mitochondrial work. The reliance of SNc neurons on L-sort Ca²⁺ channels for the control over pacemaking increments with age and this may illuminate whether why age is such a noteworthy hazard factor for growing PD (C. S. Chan et al., 2007).

Inhibition of pharmacological $Ca_v 1.3$ L-type Ca2+ channels having dihydropyridine isradipine by restoring Ca^{2+} free 'juvenile' pacemaking movement in SNc neurons. Subcutaneous release of isradipine considerably sheltered SNc neurons in animal models of PD. In support of this association to PD, a latest demonstration on epidemiological study suggested that the treatment of hypertension with Ca^{2+} channel antagonists considerably reduced the threat of rising PD (Becker, Jick, & Meier, 2008). These interpretations encouraged a restricted clinical trial for isradipine in PD patients.

Neuronal Ca²⁺Signaling and Huntington's Disease

The disorders [AD, PD and amyotrophic sidelong sclerosis (ALS)] are for the most part unpredictable with uncommon familial structures. A couple of regular point of view that shows up from the investigation of the capacity of neuronal calcium particles motioning in these sporadic issues. These disorders

are 'multi-hit' and will most likely require a blended treatment, with Ca^{2+} inhibitors included as a piece of the treatment regimen. Rather than these clutters, HD is an absolutely hereditary confusion that is caused by a solitary change in CAG rehash (polyglutamine) extension in the huntingtin (*HTT*) quality (Gusella and MacDonald, 2000). The medium prickly striatal neurons (MSNs) are most influenced in HD. Chiefly, specialists concur with mutant *Htt* exp protein securing a 'dangerous pick up of capacity' (Gusella and MacDonald, 2000; Li and Li, 2006). Crumbling of neuronal Ca^{2+} flagging is such dangerous elements of the *Htt* exp protein. Steady changes in the articulation levels of numerous Ca^{2+} flagging proteins were seen in microarray investigations of the brains from HD patients and furthermore from HD mouse models (Kuhn et al., 2007). The proof for a 'Ca²⁺ theory of HD' have been explored already (Bezprozvanny and Hayden, 2004) and are compressed and refreshed quickly here.

Neuronal Ca²⁺Signaling Association With Amyotrophic Lateral Sclerosis

Sporadic AD, PD and ALS are 'multi-hit' diseases that are activated by a few neurotic components shown to be acting. Some of these variables are normal to all said three issues, while some are 'illness specific' (Bezprozvanny and Hayden, 2004). One of the critical components which are basic to these disarranges is maturing. The 'infection particular' elements result in specificity of neuronal populaces being influenced in these disarranges – hippocampal and cortical neurons in AD, dopaminergic SNc neurons in PD and engine neurons in ALS. The critical 'infection particular' factor for AD is collection of amyloid totals; for ALS, it is incendiary harm prompted by actuated microglia and for PD, it is poisonous quality coming about because of dopamine oxidation. As these disarranges seem to be 'multi-target', combinational treatments can be just unbeaten in treating such infection, with both 'ailment particular' and in addition 'normal' pathways targeted (Alexander, 2004).

Neuronal populaces communicating lifted levels of Ca^{2+} restricting proteins are decently saved in such clutters, yet if there should be an occurrence of neuronal populaces with diminished levels of CaBP are extremely influenced. Decrease in levels of neuronal CaBPs is one of the results of the typical maturing process. Lessened capacity to cradle cytosolic Ca^{2+} is probably going to be one of the components that make maturing neurons defenseless in AD, PD and ALS (Surmeier, 2007).

Enactment of the calpain group of Ca^{2+} subordinate proteases is seen in maturing neurons and in sporadic AD, PD and ALS. The initiation of calpains is caused by raised cytosolic Ca^{2+} levels. Actuated calpains cut an assortment of substrates critical for neuronal capacity, prompting neuronal brokenness and death (Bezprozvanny and Hayden, 2004).

TAUOPATHIES

Tau pathology is not a well-known characteristic of AD, however it is additionally found in an assortment of other related NDs. Actually, PiD, corticobasal degeneration, PSP and frontal temporal dementia alongside parkinsonism identified with chromosome 17 (FTDP-17) are all defined by some specific local and in addition by cell appropriations of strangely collected tau filaments (Wszolek et al., 2006). There is presently an unmistakable affirmation demonstrating the variations from the norm in tau are sufficient to cause NDs. These abnormalities comprise of:

- 1. The occurrence of causative mutations in the gene for tau in patients with FTDP-17(Clark et al., 1998; Goedert, Crowther, & Spillantini, 1998)
- 2. The linkage of specific tau haplotypes to PSP and CBD (Houlden et al., 2001)
- 3. The deficiency of other disease specific neuropathological abnormalities in many tauopathies (Wszolek et al., 2006)
- 4. The creation of tau transgenic mice that can recapitulate the chief phenotypic hallmark so authentic human neurodegenerative tauopathies (Lee, Kenyon, & Trojanowski, 2005)

Despite the fact that tau dysfunction unaided is adequate to fortify neurodegeneration without other cerebrum amyloids however the recurrence with which tau pathology happens in a wide scope of different NDs. These NDs has prompted the theory that tau capacities in a final, normal pathway that prompts neuronal passing or brokenness however this can be actuated by some other starting occasions. In transgenic mice, communicating tau and co-expression of APP or α -synuclein brings about the accelerating of tau pathology (Morris et al., 2014). This proposes neurodegeneration intervened by tau pathology can likewise get enacted downstream of the pathways of different illness which comes about because of the collections of both A β and α synuclein amyloidogenesis. This fits into the elucidations from infection appeared by the patients with APP changes clarifying bounteous A β and tauopathology, yet patients with tau mutations demonstrate for the most part tau pathology alone (Morris et al., 2014).

Tau is basically a microtubule-linked protein that binds to microtubules and stabilizes it. At present there are more than 30 different mutations or pathogenic nucleotide substitutions in the gene intended for tau that have been exposed to be the reason behind FTDP-17 in more than 50 different disease related biomarkers (Levy et al., 2005). These mutations might lead to NDs by one or more different mechanisms:

- 1. Alterations in tau splicing that lead to abnormal patterns of tau-isoform expression (D'Souza et al., 1999).
- 2. Compromise of tau's capability to combine with and stabilize microtubules (Hasegawa, Smith, & Goedert, 1998; Hong et al., 1998).
- 3. Enhanced fibrillization of tau (Goedert, Jakes, & Crowther, 1999).

Therefore, it can be said that tau mutations and by analogy tau dysfunction in sporadic disease, can lead to pathogenesis through mechanisms that involve both losses of function i.e., decreased microtubule stabilization and toxic gain of function which is increased fibril formation (Van Dooren, Princen, De Witte, & Griffioen, 2014). Depicting this fundamental dichotomy, therapies which target both the mechanisms are presently in development.

Research on biomarkers on NDs is a hotly debated issue where much work has to be done yet. As we can gain from this examination subject, biomarkers are enabling us to grow the information on the organic and anatomical premise of NDs and to actualize demonstrative methods in clinical practice and clinical trials.

TOPOLOGICAL METHODS FOR NETWORK ANALYSIS

On the basis of the set of curated ND genes, firstly the network of disease proteins associated to the diseases of concern is recognized. In order to evaluate the topological features of the proteins, several

centralities in the network were measured; namely, degree, betweenness centrality, closeness and topological significance. The disease-networks were modeled afterward by calculating the disease linkages in terms of shortest paths among the pairs of disease proteins. Allocation of proteins and connecting proteins were identified as the major requirements to be most likely accountable for disease linkages. The functioning of these connecting proteins was studied by the Gene Ontology (GO) enrichment analysis (Bansal & Srivastava, 2018).

Modeling Interaction Network of Disease Proteins

For the investigation of the NDs from the network view point, firstly the interaction networks of diseases proteins were modeled (Hasegawa et al., 1998; A. Kumar & Bansal, 2017). The expressed proteins of disease genes are the disease proteins that are linked to particular NDs, the list of disease genes related to the top neurodiseases which were obtained from the morbid map published in the Online Mendelian Inheritance in Man (OMIM) database. Sequentially the construction of the protein interaction network related to the top neurodiseases was done and then the disease genes to disease proteins were mapped based on the mapping design of the UniProt which is a database of protein sequences and their functional information (Apweiler et al., 2004; Bansal, Singh, & Chauhan, 2017). The associations of those disease proteins were then derived by validation of the experimental interactions in the i2d database were expelled to amplify the dependability of protein interaction data. The ultimate interaction network of interest enclosed the disease proteins (nodes) and their direct interacting partners (edges) (V. Kumar, Bansal, & Chauhan, 2017). The network design was not directed and weighted as only the binary interactions were considered (Panigrahi & Singh, 2013).

Modeling Interaction Network of Neurodegenerative Disorders

On the basis of the disease proteins network, the network of NDs with meta-nodes and meta-edges were modeled that represented the diseases and the relations among them, in the same way. A meta-node was reasonable as a group of the ailment proteins related to one disease (Goñiet al., 2008). A meta-edge connecting one illness to the next was characterized as an arrangement of the ways interfacing their infection proteins (Bansal & Ramana, 2015). The meta-edges were weighted by methods for extraordinary score capacities ri to uncover the intensity of the association including two infections, similar to the two meta-hubs, called as ALS and spinal solid decay (SMA), included every one of the proteins related to ALS and SMA correspondingly (Bansal & Srivastava, 2018). Each match of proteins (one from ALS and one from SMA), were assembled by figuring their ways interfacing them together to perceive a meta-edge, called ALS-SMA alongside weight computed by ri. The meta-edges are just obvious if their score ri > 0.

ANALYZING THE NEURODEGENERATIVE DISORDERS NETWORK USING THE NETWORK MINING APPROACH

Computing Topological Properties of Protein Interaction Network

In order to study networks and their participating proteins the centrality of proteins in the network were evaluated. The practical importance of proteins may be observed by understanding their central roles in

the network. As the centrality depicts an exclusive structural characteristic, reliable possibilities of the biological traits can be attained by combinations of the above measures than depending upon a particular index (Azuaje, 2014). Various centralities changing from nearby scale were processed to transitional scale (topological hugeness upto 1 and 3 stages) and in conclusion to worldwide scale (betweenness and closeness). Different centralities have been utilized to portray the systems considered as takes after:

- 1. Firstly, the degree centrality or connectivity (*D*) of protein vertices V*i*, depicting total number of interactions *eij* the protein has with other proteins vertices V*j*. This is mostly accepted to calculate the local centrality in the network (Navlakha & Kingsford, 2010).
- 2. Secondly, the betweenness centrality (*B*) which is a measure of the positional control of proteins in the networks. The betweenness centrality of protein vertices V_i is well explained as the total number of shortest paths *p* lying between pairs of other proteins that pass through vertices V_i over the total number of shortest paths involving pairs of other proteins (Goñi et al., 2008).
- 3. Lastly, the closeness centrality (*C*) which measures whether how close a protein is to other proteins as well as the distant node (Limviphuvadh et al., 2007).

There are various online as well as standalone tools for network analysis like Cytoscape, STRING and GORILLA, BLAST2GO, etc. Network base analysis of molecular pathways will help in efficient prediction of biomarkers for diagnosis and prognosis of different forms of dementia. Moreover, network parameters and graph theory gives new insight to the available information which was earlier hidden in various data layers.

CONCLUSION

This chapter describes of molecular mechanisms occurs during neurodegeneration and helps to give overall view of pathological understanding through various modeling and network perspective. NDs have extended by the various characteristic features of the neuropathological lesions found in the brain. It is known now that these lesions cannot be considered as just biomarkers for NDs like AD and PD, but are tied essentially to their pathogenesis. For each situation of predominantly acquired neurodegeneration, the illness causing changes can be associated unswervingly to amyloid arrangement. These signs have now come into pointed concentration and neurodegenerative properties are enhanced comprehended, and they have given a well-manufactured method of reasoning to the in vitro, in vivo tests, and epidemiological investigations that have gone far towards checking this theory. These tests have established a strong framework for various potential therapeutics that is currently in preclinical and clinical advancement. Over the next few years the effectiveness of a many drugs will be experimentally tested, and there is an increasing assurance of the outcomes of these trials that will confirm current concept of the etiology of NDs.

REFERENCES

Abou-Sleiman, P. M., Muqit, M. M. K., & Wood, N. W. (2006). Expanding insights of mitochondrial dysfunction in Parkinson's disease. Nature *Reviews. Neuroscience*, *7*(3), 207–219. doi:10.1038/nrn1868 PMID:16495942

Alexander, G. E. (2004). Biology of Parkinson's disease: Pathogenesis and pathophysiology of a multisystem neurodegenerative disorder. Dialogues in Clinical Neuroscience, 6(3), 259–280. PMID:22033559

Allen, N., Robinson, A. C., Snowden, J., Davidson, Y. S., & Mann, D. M. A. (2014). Patterns of cerebral amyloid angiopathy define histopathological phenotypes in Alzheimer's disease. *Neuropathology and Applied Neurobiology*, *40*(2), 136–148. doi:10.1111/nan.12070 PMID:23808763

Apweiler, R., Bairoch, A., Wu, C. H., Barker, W. C., Boeckmann, B., & Ferro, S. (2004). UniProt: The Universal Protein knowledgebase. *Nucleic Acids Research*, *32*(Database issue), D115–D119. doi:10.1093/nar/gkh131 PMID:14681372

Arispe, N., Rojas, E., & Pollard, H. B. (1993). Alzheimer disease amyloid beta protein forms calcium channels in bilayer membranes: Blockade by tromethamine and aluminum. *Proceedings of the National Academy of Sciences of the United States of America*, 90(2), 567–571. doi:10.1073/pnas.90.2.567 PMID:8380642

Azuaje, F. J. (2014). Selecting biologically informative genes in co-expression networks with a centrality score. *Biology Direct*, *9*(1), 12. doi:10.1186/1745-6150-9-12 PMID:24947308

Bansal, A., & Ramana, J. (2015). TCGDB: A Compendium of Molecular Signatures of Thyroid Cancer and Disorders. *Journal of Cancer Science & Therapy*, 7(7). Retrieved from https://www. omicsonline.org/open-access/tcgdb-a-compendium-of-molecular-signatures-of-thyroid-cancer-and-disorders-1948-5956-1000350.php?aid=57693

Bansal, A., Singh, T. R., & Chauhan, R. S. (2017). A novel miRNA analysis framework to analyze differential biological networks. *Scientific Reports*, 7(1), 14604. doi:10.103841598-017-14973-x PMID:29097749

Bansal, A., & Srivastava, P. A. (2018). Transcriptomics to Metabolomics: A Network Perspective for Big Data. IGI Global.

Barabási, A.-L., Gulbahce, N., & Loscalzo, J. (2011). Network medicine: A network-based approach to human disease. *Nature Reviews. Genetics*, *12*(1), 56–68. doi:10.1038/nrg2918 PMID:21164525

Becker, C., Jick, S. S., & Meier, C. R. (2008). Use of antihypertensives and the risk of Parkinson disease. *Neurology*, *70*(16 Part 2), 1438–1444. doi:10.1212/01.wnl.0000303818.38960.44 PMID:18256367

Berridge, M. J. (2010). Calcium hypothesis of Alzheimer's disease. *Pflügers Archiv*, 459(3), 441–449. doi:10.100700424-009-0736-1 PMID:19795132

Brookmeyer, R., Gray, S., & Kawas, C. (1998). Projections of Alzheimer's disease in the United States and the public health impact of delaying disease onset. *American Journal of Public Health*, 88(9), 1337–1342. doi:10.2105/AJPH.88.9.1337 PMID:9736873

Catterall, W. A. (2011). Voltage-Gated Calcium Channels. *Cold Spring Harbor Perspectives in Biology*, *3*(8), a003947. doi:10.1101/cshperspect.a003947 PMID:21746798

Chen, X., & Burgoyne, R. D. (2012). Identification of common genetic modifiers of neurodegenerative diseases from an integrative analysis of diverse genetic screens in model organisms. *BMC Genomics*, *13*(1), 71. doi:10.1186/1471-2164-13-71 PMID:22333271

Ciechanover, A., & Kwon, Y. T. (2015). Degradation of misfolded proteins in neurodegenerative diseases: Therapeutic targets and strategies. *Experimental & Molecular Medicine*, 47(3), e147. doi:10.1038/ emm.2014.117 PMID:25766616

Clark, L. N., Poorkaj, P., Wszolek, Z., Geschwind, D. H., Nasreddine, Z. S., Miller, B., ... Wilhelmsen, K. C. (1998). Pathogenic implications of mutations in the tau gene in pallido-ponto-nigral degeneration and related neurodegenerative disorders linked to chromosome 17. *Proceedings of the National Academy of Sciences of the United States of America*, 95(22), 13103–13107. doi:10.1073/pnas.95.22.13103 PMID:9789048

D'Souza, I., Poorkaj, P., Hong, M., Nochlin, D., Lee, V. M.-Y., Bird, T. D., & Schellenberg, G. D. (1999). Missense and silent tau gene mutations cause frontotemporal dementia with parkinsonism-chromosome 17 type, by affecting multiple alternative RNA splicing regulatory elements. *Proceedings of the National Academy of Sciences of the United States of America*, *96*(10), 5598–5603. doi:10.1073/pnas.96.10.5598 PMID:10318930

Echeverria, V., & Cuello, A. C. (2002). Intracellular A-beta amyloid, a sign for worse things to come? *Molecular Neurobiology*, *26*(2–3), 299–316. doi:10.1385/MN:26:2-3:299 PMID:12428762

Ernst, R. L., Hay, J. W., Fenn, C., Tinklenberg, J., & Yesavage, J. A. (1997). Cognitive function and the costs of Alzheimer disease. An exploratory study. *Archives of Neurology*, *54*(6), 687–693. doi:10.1001/archneur.1997.00550180013006 PMID:9193203

Forman, M. S., Trojanowski, J. Q., & Lee, V. M.-Y. (2004). Neurodegenerative diseases: A decade of discoveries paves the way for therapeutic breakthroughs. *Nature Medicine*, *10*(10), 1055–1063. doi:10.1038/nm1113 PMID:15459709

Foster, T. C. (2007). Calcium homeostasis and modulation of synaptic plasticity in the aged brain. *Aging Cell*, 6(3), 319–325. doi:10.1111/j.1474-9726.2007.00283.x PMID:17517041

Gant, J. C., Sama, M. M., Landfield, P. W., & Thibault, O. (2006). Early and simultaneous emergence of multiple hippocampal biomarkers of aging is mediated by Ca2+-induced Ca2+ release. *The Journal of Neuroscience*, *26*(13), 3482–3490. doi:10.1523/JNEUROSCI.4171-05.2006 PMID:16571755

Goedert, M., Crowther, R. A., & Spillantini, M. G. (1998). Tau mutations cause frontotemporal dementias. *Neuron*, *21*(5), 955–958. doi:10.1016/S0896-6273(00)80615-7 PMID:9856453

Tau Pathology

Goedert, M., Jakes, R., & Crowther, R. A. (1999). Effects of frontotemporal dementia FTDP-17 mutations on heparin-induced assembly of tau filaments. *FEBS Letters*, 450(3), 306–311. doi:10.1016/ S0014-5793(99)00508-6 PMID:10359094

Goñi, J., Esteban, F. J., de Mendizábal, N. V., Sepulcre, J., Ardanza-Trevijano, S., Agirrezabal, I., & Villoslada, P. (2008). A computational analysis of protein-protein interaction networks in neurodegenerative diseases. *BMC Systems Biology*, 2(1), 52. doi:10.1186/1752-0509-2-52 PMID:18570646

Guo, J. L., & Lee, V. M. Y. (2014). Cell-to-cell transmission of pathogenic proteins in neurodegenerative diseases. *Nature Medicine*, *20*(2), 130–138. doi:10.1038/nm.3457 PMID:24504409

Hardy, J., & Selkoe, D. J. (2002). The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science*, 297(5580), 353–356. doi:10.1126cience.1072994 PMID:12130773

Hasegawa, M., Smith, M. J., & Goedert, M. (1998). Tau proteins with FTDP-17 mutations have a reduced ability to promote microtubule assembly. *FEBS Letters*, 437(3), 207–210. doi:10.1016/S0014-5793(98)01217-4 PMID:9824291

Hassan, A., Whitwell, J. L., & Josephs, K. A. (2011). The corticobasal syndrome–Alzheimer's disease conundrum. *Expert Review of Neurotherapeutics*, 11(11), 1569–1578. doi:10.1586/ern.11.153 PMID:22014136

Hebert, L. E., Beckett, L. A., Scherr, P. A., & Evans, D. A. (2001). Annual incidence of Alzheimer disease in the United States projected to the years 2000 through 2050. *Alzheimer Disease and Associated Disorders*, *15*(4), 169–173. doi:10.1097/00002093-200110000-00002 PMID:11723367

Hebert, L. E., Scherr, P. A., Bienias, J. L., Bennett, D. A., & Evans, D. A. (2003). Alzheimer disease in the US population: Prevalence estimates using the 2000 census. *Archives of Neurology*, *60*(8), 1119–1122. doi:10.1001/archneur.60.8.1119 PMID:12925369

Hong, M., Zhukareva, V., Vogelsberg-Ragaglia, V., Wszolek, Z., Reed, L., Miller, B. I., & ... (1998). Mutation-specific functional impairments in distinct tau isoforms of hereditary FTDP-17. *Science*, 282(5395), 1914–1917. doi:10.1126cience.282.5395.1914 PMID:9836646

Houlden, H., Baker, M., Morris, H. R., MacDonald, N., Pickering-Brown, S., Adamson, J., & ... (2001). Corticobasal degeneration and progressive supranuclear palsy share a common tau haplotype. *Neurology*, *56*(12), 1702–1706. doi:10.1212/WNL.56.12.1702 PMID:11425937

Ideker, T., & Sharan, R. (2008). Protein networks in disease. *Genome Research*, 18(4), 644–652. doi:10.1101/gr.071852.107 PMID:18381899

Irvine, G. B., El-Agnaf, O. M., Shankar, G. M., & Walsh, D. M. (2008). Protein Aggregation in the Brain: The Molecular Basis for Alzheimer's and Parkinson's Diseases. *Molecular Medicine (Cambridge, Mass.)*, 14(7–8), 451–464. PMID:18368143

Jindal, K., & Bansal, A. (2016). APOEɛ2 is Associated with Milder Clinical and Pathological Alzheimer's Disease. *Annals of Neurosciences*, 23(2), 112–112. doi:10.1159/000443572 PMID:27647961

Kann, M. G. (2007). Protein interactions and disease: Computational approaches to uncover the etiology of diseases. *Briefings in Bioinformatics*, 8(5), 333–346. doi:10.1093/bib/bbm031 PMID:17638813

Krauthammer, M., Kaufmann, C. A., Gilliam, T. C., & Rzhetsky, A. (2004). Molecular triangulation: Bridging linkage and molecular-network information for identifying candidate genes in Alzheimer's disease. *Proceedings of the National Academy of Sciences of the United States of America*, 101(42), 15148–15153. doi:10.1073/pnas.0404315101 PMID:15471992

Kumar, A., & Bansal, A. (2017). Integrated bioinformatics analysis of differentially expressed genes (DEGS) of Alzheimer's disease (AD) datasets from gene expression omnibus (GEO). *Alzheimer's & Dementia: The Journal of the Alzheimer's Association*, *13*(7), 953. doi:10.1016/j.jalz.2017.06.1270

Kumar, A., & Singh, T. R. (2017). A New Decision Tree to Solve the Puzzle of Alzheimer's Disease Pathogenesis Through Standard Diagnosis Scoring System. *Interdisciplinary Sciences, Computational Life Sciences*, *9*(1), 107–115. doi:10.100712539-016-0144-0 PMID:26792126

Kumar, V., Bansal, A., & Chauhan, R. S. (2017). Modular Design of Picroside-II Biosynthesis Deciphered through NGS Transcriptomes and Metabolic Intermediates Analysis in Naturally Variant Chemotypes of a Medicinal Herb, Picrorhiza kurroa. *Frontiers in Plant Science*, 8. doi:10.3389/fpls.2017.00564 PMID:28443130

Lee, V. M.-Y., Kenyon, T. K., & Trojanowski, J. Q. (2005). Transgenic animal models of tauopathies. *Biochimica et Biophysica Acta*, 1739(2–3), 251–259. doi:10.1016/j.bbadis.2004.06.014 PMID:15615643

Levin, J., Kurz, A., Arzberger, T., Giese, A., & Höglinger, G. U. (2016). The Differential Diagnosis and Treatment of Atypical Parkinsonism. *Deutsches Ärzteblatt International*, *113*(5), 61–69. PMID:26900156

Levy, S. F., LeBoeuf, A. C., Massie, M. R., Jordan, M. A., Wilson, L., & Feinstein, S. C. (2005). Threeand Four-repeat Tau Regulate the Dynamic Instability of Two Distinct Microtubule Subpopulations in Qualitatively Different Manners Implications for Neurodegeneration. *The Journal of Biological Chemistry*, 280(14), 13520–13528. doi:10.1074/jbc.M413490200 PMID:15671021

Limviphuvadh, V., Tanaka, S., Goto, S., Ueda, K., & Kanehisa, M. (2007). The commonality of protein interaction networks determined in neurodegenerative disorders (NDDs). *Bioinformatics (Oxford, England)*, 23(16), 2129–2138. doi:10.1093/bioinformatics/btm307 PMID:17553855

Lukiw, W. J. (2012). Amyloid beta (Aβ) peptide modulators and other current treatment strategies for Alzheimer's disease (AD). *Expert opinion on emerging drugs*. Retrieved from http://www.ncbi.nlm.nih. gov/pmc/articles/PMC3399957/

Macedo, B., & Cordeiro, Y. (2017). Unraveling Prion Protein Interactions with Aptamers and Other PrP-Binding Nucleic Acids. *International Journal of Molecular Sciences*, *18*(5). Retrieved from http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5454936/ PMID:28513534

Mattson, M. P. (2004). Pathways Towards and Away from Alzheimer's Disease. *Nature*, 430(7000), 631–639. doi:10.1038/nature02621 PMID:15295589

Mietelska-Porowska, A., Wasik, U., Goras, M., Filipek, A., & Niewiadomska, G. (2014). Tau Protein Modifications and Interactions: Their Role in Function and Dysfunction. *International Journal of Molecular Sciences*, *15*(3), 4671–4713. doi:10.3390/ijms15034671 PMID:24646911

Tau Pathology

Morris, G. P., Clark, I. A., & Vissel, B. (2014). Inconsistencies and Controversies Surrounding the Amyloid Hypothesis of Alzheimer's Disease. *Acta Neuropathologica Communications*, 2. Retrieved from http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4207354/ PMID:25231068

Navlakha, S., & Kingsford, C. (2010). The power of protein interaction networks for associating genes with diseases. *Bioinformatics (Oxford, England)*, *26*(8), 1057–1063. doi:10.1093/bioinformatics/btq076 PMID:20185403

Nguyen, T.-P., & Ho, T.-B. (2012). Detecting disease genes based on semi-supervised learning and protein-protein interaction networks. *Artificial Intelligence in Medicine*, *54*(1), 63–71. doi:10.1016/j. artmed.2011.09.003 PMID:22000346

Nguyen, T.-P., Liu, W., & Jordán, F. (2011). Inferring pleiotropy by network analysis: Linked diseases in the human PPI network. *BMC Systems Biology*, *5*(1), 179. doi:10.1186/1752-0509-5-179 PMID:22034985

Nilsson, P., Iwata, N., Muramatsu, S., Tjernberg, L. O., Winblad, B., & Saido, T. C. (2010). Gene therapy in Alzheimer's disease – potential for disease modification. *Journal of Cellular and Molecular Medicine*, *14*(4), 741–757. doi:10.1111/j.1582-4934.2010.01038.x PMID:20158567

Núñez-Santana, F. L., Oh, M. M., Antion, M. D., Lee, A., Hell, J. W., & Disterhoft, J. F. (2014). Surface L-type Ca2+ channel expression levels are increased in aged hippocampus. *Aging Cell*, *13*(1), 111–120. doi:10.1111/acel.12157 PMID:24033980

Oddo, S., Caccamo, A., Shepherd, J. D., Murphy, M. P., Golde, T. E., Kayed, R., ... LaFerla, F. M. (2003). Triple-transgenic model of Alzheimer's disease with plaques and tangles: Intracellular Abeta and synaptic dysfunction. *Neuron*, *39*(3), 409–421. doi:10.1016/S0896-6273(03)00434-3 PMID:12895417

Oti, M., Snel, B., Huynen, M. A., & Brunner, H. G. (2006). Predicting disease genes using protein-protein interactions. *Journal of Medical Genetics*, 43(8), 691–698. doi:10.1136/jmg.2006.041376 PMID:16611749

Panigrahi, P. P., & Singh, T. R. (2012). Computational analysis for functional and evolutionary aspects of BACE-1 and associated Alzheimer's related proteins. *International Journal of Computational Intelligence Studies*, *1*(4), 322–332. doi:10.1504/IJCISTUDIES.2012.050355

Panigrahi, P. P., & Singh, T. R. (2013). Computational studies on Alzheimer's disease associated pathways and regulatory patterns using microarray gene expression and network data: Revealed association with aging and other diseases. *Journal of Theoretical Biology*, *334*, 109–121. doi:10.1016/j.jtbi.2013.06.013 PMID:23811083

Rajamohamedsait, H. B., & Sigurdsson, E. M. (2012). Histological Staining of Amyloid and Pre-Amyloid Peptides and Proteins in Mouse Tissue. *Methods in Molecular Biology (Clifton, N.J.)*, 849, 411–424. doi:10.1007/978-1-61779-551-0_28 PMID:22528106

Rizzuto, R., Stefani, D. D., Raffaello, A., & Mammucari, C. (2012). Mitochondria as sensors and regulators of calcium signalling. *Nature Reviews. Molecular Cell Biology*, *13*(9), 566–578. doi:10.1038/nrm3412 PMID:22850819

Samii, A., Nutt, J. G., & Ransom, B. R. (2004). Parkinson's disease. *Lancet* (*London, England*), 363(9423), 1783–1793. doi:10.1016/S0140-6736(04)16305-8 PMID:15172778

Schuster-Böckler, B., & Bateman, A. (2008). Protein interactions in human genetic diseases. *Genome Biology*, 9(1), R9. doi:10.1186/gb-2008-9-1-r9 PMID:18199329

Seabrook, G. R., Ray, W. J., Shearman, M., & Hutton, M. (2007). Beyond amyloid: The next generation of Alzheimer's disease therapeutics. *Molecular Interventions*, 7(5), 261–270. doi:10.1124/mi.7.5.8 PMID:17932415

Singh, S. K., Srivastav, S., Yadav, A. K., Srikrishna, S., & Perry, G. (2016). Overview of Alzheimer's Disease and Some Therapeutic Approaches Targeting Aβ by Using Several Synthetic and Herbal Compounds. *Oxidative Medicine and Cellular Longevity*, 2016, 1–22. doi:10.1155/2016/7361613 PMID:27034741

Sulzer, D. (2007). Multiple hit hypotheses for dopamine neuron loss in Parkinson's disease. *Trends in Neurosciences*, *30*(5), 244–250. doi:10.1016/j.tins.2007.03.009 PMID:17418429

Toescu, E. C., & Verkhratsky, A. (2007). The importance of being subtle: Small changes in calcium homeostasis control cognitive decline in normal aging. *Aging Cell*, *6*(3), 267–273. doi:10.1111/j.1474-9726.2007.00296.x PMID:17517038

Tsai, R. M., & Boxer, A. L. (2014). Clinical Trials: Past, current and future for atypical parkinsonian syndromes. *Seminars in Neurology*, *34*(2), 225–234. doi:10.1055-0034-1381739 PMID:24963682

Van Dooren, T., Princen, K., De Witte, K., & Griffioen, G. (2014). Derailed Intraneuronal Signalling Drives Pathogenesis in Sporadic and Familial Alzheimer's Disease. *BioMed Research International*. Retrieved September 5, 2017, from https://www.hindawi.com/journals/bmri/2014/167024/

Vasaikar, S. V., Padhi, A. K., Jayaram, B., & Gomes, J. (2013). NeuroDNet - an open source platform for constructing and analyzing neurodegenerative disease networks. *BMC Neuroscience*, *14*(1), 3. doi:10.1186/1471-2202-14-3 PMID:23286825

Wang, L., Li, J., Zhao, H., Hu, J., Ping, Y., Li, F., ... Li, X. (2016). Identifying the crosstalk of dysfunctional pathways mediated by lncRNAs in breast cancer subtypes. *Molecular BioSystems*, *12*(3), 711–720. doi:10.1039/C5MB00700C PMID:26725846

Wszolek, Z. K., Tsuboi, Y., Ghetti, B., Pickering-Brown, S., Baba, Y., & Cheshire, W. P. (2006). Frontotemporal dementia and parkinsonism linked to chromosome 17 (FTDP-17). *Orphanet Journal of Rare Diseases*, 1(1), 30. doi:10.1186/1750-1172-1-30 PMID:16899117

Chapter 11 Amyloid Beta: The Foremost Protagonist in Alzheimer's Disease

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ABSTRACT

Alzheimer's disease (AD), exhibiting accumulation of amyloid beta (A β) peptide as a foremost protagonist, is one of the top five causes of deaths. It is a neurodegenerative disorder (ND) that causes a progressive decline in memory and cognitive abilities. It is characterized by deposition of A β plaques and neurofibrillary tangles (NFTs) in the neurons, which in turn causes a decline in the brain acetylcholine levels. A β hypothesis is the most accepted hypothesis pertaining to the pathogenesis of AD. Amyloid Precursor Protein (APP) is constitutively present in brain and it is cleaved by three proteolytic enzymes (i.e., alpha, beta, and gamma secretases). Beta and gamma secretases cleave APP to form A β . Ubiquitin Proteasome System (UPS) is involved in the clearing of A β plaques. AD also involves impairment in UPS. The novel disease-modifying approaches involve inhibition of beta and gamma secretases. A number of clinical trials are going on worldwide with moieties targeting beta and gamma secretases. This chapter deals with an overview of APP and its enzymatic cleavage leading to AD.

DOI: 10.4018/978-1-5225-5282-6.ch011

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INTRODUCTION

Dementia is considered to be an aggregation of symptoms affecting memory, cognition, social abilities on a severe magnitude. These adverse effects affecting the daily activities result from physical changes in brain. Dementia can further be of several types depending upon the underlying pathological condition and symptoms (Anand, Khurana, Chawla, Sharma, & Khurana, 2017). Dementias are progressive in nature. The sufferers may have troubles with short-term memory in the beginning which evolves into a loss of memory altogether. While there are varied symptoms of dementia, impairment in at least two of the following mentioned core mental functions should be there for the condition to be considered as dementia. These include deficit in memory, language and communication, ability to focus, capability to reason and judge and visual perception (Alzheimer's Association, 2017c). AD is the most common form of dementia. Over 47.5 million people globally were estimated to be suffering from dementia in 2016. By 2030, the figure is expected to reach as high as 75.6 million (World Health Organization, 2016). AD generally appears in mid to late adulthood. It is associated with a progressive and rather irreversible decline in memory and various other cognitive capabilities. In AD, there is neuronal destruction and deterioration of neural connections in the cerebral cortex region of the brain along with a substantial loss of brain mass (Perl, 2010). AD is lethal within 5–10 years of its onset (Dwyer et al., 2009). Mortality usually ensues due to complications of the chronic illness.

AD is characterized by the presence of two neuropathological hallmarks i.e. extracellular A β plaques and intracellular NFTs. The plaques constitute chiefly of the neurotoxic peptide A β , which forms after the sequential cleavage of a large precursor protein i.e. APP by two enzymes, namely, β -secretase (commonly known as BACE1) and γ -secretase (involving four proteins, including presenilin). However, A β is not formed if APP is first acted upon and cleaved by the enzyme α -secretase instead of β -secretase. NFTs comprise mainly of the protein tau which is a microtubule associated protein (MAP) i.e. it binds microtubules in cells to facilitate the neuronal transport system. In the development of AD, Tau uncouples from microtubules and aggregates into tangles thereby inhibiting transport and resulting in microtubule disassembly. It also depends on the phosphorylation of Tau (Anand, Patience, Sharma, & Khurana, 2017; Nisbet, Polanco, Ittner, & Götz, 2015).

The actual causes at play behind the development of AD are still not well defined. However, certain factors like anomaly in the phosphorylation of tau protein, alterations in calcium metabolism, oxidative stress, neuro-inflammation, abnormal energy metabolism and protein processing i.e. undesired A β formation and aggregation, are considered to be important factors in the pathogenesis of AD (Butterfield et al., 2002; Habibyar, Sharma, & Khurana, 2016; J Hardy & Selkoe, 2002). In the present chapter, role of A β formation and aggregation as the foremost protagonist in AD is discussed with special emphasis on APP and enzymes involved in its cleavage along with involvement of UPS in amyloid hypothesis of AD.

BACKGROUND

AD was first described by a German neuropathologist Alois Alzheimer in 1906 (Editors of Encyclopædia Britannica, 2016). AD was recognized as the most prevalent form of dementia among geriatric persons by the commencement of 21st century. It is one of the top five most common causes of mortality in population of the United States (Centers for Disease Control and Prevention, 2017). In rare cases, it may appear in people in their 40s and 50s, but otherwise it is a disease of old age. Based on clinical, population-

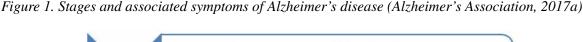
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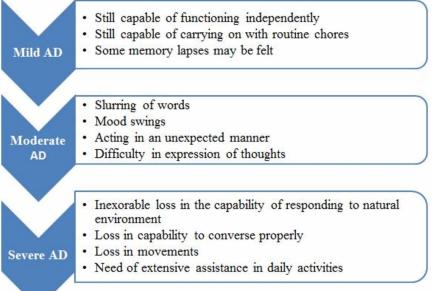
based studies, about 200,000 people under 65 years of age are suffering from AD. In contrast, around 5 million of those over 65 years of age have AD. As per speculations, a new case of AD is expected to be developed every 33 seconds, by 2050 (Alzheimer's Association, 2014).

The symptoms of AD get worse with time, although the pace at which the disease progresses is variable. Alterations in the brain related to AD commence years before any related clinical manifestations emerge. This period, spanning over a few years, is known as preclinical AD. The progression of AD occurs in three stages, namely mild AD (early-stage), moderate AD (middle-stage) and severe AD (late-stage). The symptoms associated with each stage are given in Figure 1.

There are several hypotheses which have been put up by several researchers for pathogenesis of AD out of which cholinergic hypothesis (McGleenon, Dynan, & Passmore, 1999) and A β hypothesis (Storey & Cappai, 1999) have been subjected to extensive research. A β hypothesis talks more about the actual pathological hallmarks which are visible in the patients' brains during autopsy. Cholinergic hypothesis has generated several acetylcholinesterase inhibitors as therapeutic measures for AD. However, these agents provide mere symptomatic relief (Anand, Patience, et al., 2017). Researchers are foraying more and more into the potential drugs targeting the amyloid pathway as it is considered to be a disease modifying approach.

Evidence exists suggesting involvement of both cholinergic and glutamatergic neurochemical systems in the etiology of AD. Acetylcholine (ACh) is an essential neurotransmitter responsible for cognitive functions and learning. In the brains of patients suffering from AD, it is decreased both in concentration and function. This deficit and other presynaptic cholinergic limitations, like loss of cholinergic neuronal network and reduced acetylcholinesterase activity, validate the cholinergic hypothesis of AD. Another neurochemical hypothesis for development of AD is the N-Methyl d-aspartate (NMDA) mediated glutamatergic hypothesis. Glutamate is an excitatory neurotransmitter which acts on NMDA





receptors, which are pivotal in learning and memory. However, in some circumstances, over stimulation of NMDA receptors by glutamate causes neuronal damage due to excitotoxicity (Francis, 2005). A relatively recent mitochondrial dysfunction hypothesis for pathogenesis of AD proposes that AD brain mitochondrial dysfunction leads to amyloidosis, cell cycle re-entry, and tau phosphorylation (Swerdlow, Burns, & Khan, 2014).

The appearance of $A\beta$ begins many years before the clinical manifestations of the disease appear, so it could be a reliable biomarker for AD prediction. As indicated, $A\beta$ plays a major role in the formation of both amyloid plaques and NFTs, which gradually leads to AD. $A\beta$ deposition leads to degeneration in synapses which leads to faulty interactions with different types of CNS receptors. Therefore, it disrupts neuronal homeostasis. Moreover, $A\beta$ deposition along the cerebral vessels changes their tonicity and leads to some of the cerebrovascular deficits. Furthermore, its accumulation disrupts intracellular Ca²⁺ homeostasis which ultimately reduces neuronal Ca²⁺ buffering capacity. This, in turn increases excitotoxicity outcomes. Also, $A\beta$ peptides may fold in different ways and show a prion-like pathology in the brain of AD patients (Sadigh-Eteghad et al., 2015).

An evidence base implicates $A\beta$ in the pathogenesis of AD, leading to the formulation of the amyloid cascade hypothesis, i.e., the null hypothesis (John Hardy, 2006). The post mortem observation of the brain tissue of diseased individuals has made it quite obvious why $A\beta$ is the primary suspect in disease pathogenesis. $A\beta$ is the major constituent in two of the most distinctive histopathological hallmarks, namely senile plaques and cerebral amyloid angiopathy (Glenner & Wong, 1984a, 1984b).

The first and foremost argument that $A\beta$ does not initiate AD is that deposition of $A\beta$ into senile plaques is in no way specific to AD patients and is instead a marker of normal aging, i.e. the alternate hypothesis (Davies et al., 1988). Therefore, the strong association of $A\beta$ in AD may simply mean an acceleration of senescence related deterioration. Support for this view can be found with the number of plaques in cognitively normal individuals against those seen in advanced disease (Mann, Jones, South, Snowden, & Neary, 1992; Schmitt et al., 2000). Furthermore, there exists only a weak correlation between the burden of $A\beta$ and neuronal loss or cognitive impairment (Guillozet, Weintraub, Mash, & Mesulam, 2003). The alternate hypothesis is based on the notion that $A\beta$ simply acts as a bystander or a protector rather than the causative factor of AD (H. Lee et al., 2007).

AMYLOID HYPOTHESIS OF ALZHEIMER'S DISEASE

There is an on-going debate about the A β hypothesis stating that the early, often initiating factor in development of AD is the imbalance between the production and clearance of A β 42 and other related A β peptides. There are several hypotheses that have been suggested pertaining to the pathogenesis of AD. However, A β hypothesis is becoming increasingly popular amongst researchers worldwide (Alzheimer's Association, 2014). The A β hypothesis comes with the innate advantage of involving several molecular targets for drug development. Some important molecular targets are – APP, β - secretase 2 (BACE2), A β plaques. By efficiently creating structural and/or functional alterations at any of these targets, the progression of AD could be delayed or halted altogether.

Amyloid Precursor Protein and Its Enzymatic Cleavage

APP belongs to the family of transmembrane and secreted proteins which includes the amyloid precursorlike proteins (APLP1 and APLP2) in mammals. It has several functions, which include regulation of haemostasis and mediation of neuroprotection. It is believed to stimulate neurite outgrowth and synaptogenesis. Also, it has been reported to have a role in stimulation of mitogenesis, G_0 proteins and MAP kinases for regulating cell proliferation, differentiation and survival. It seems to have a neuroprotective effect by altering cyclic guanosine monophosphate (cGMP) levels, Ca²⁺ homeostasis and activation of K⁺ channels. It has also been reported to have a protective action against excitotoxicity, hypoglycemia and brain ischemia (Storey & Cappai, 1999).

The neuroprotective activity of APP has been substantiated by *in vivo* studies on mice. In one experiment, APP-transgenic mice were mated with HIV-gp120 transgenic mice and a significant protection was reported against HIV-gp 120 induced neurotoxicity (Mucke et al., 1995).

Although APP is a transmembrane protein that is present on the surface of neurons, the majority of it undergoes degradation within the secretory pathway even before it could reach the cell surface (Caporaso, Gandy, Buxbaum, & Greengard, 1992; Citron, Teplow, & Selkoe, 1995). APP is expressed in great quantities in neurons and undergoes rapid metabolism. Several alternate pathways exist for APP proteolysis. Some of these pathways lead to formation of the A β peptide and some of them do not (M. Lee et al., 2000). There are three enzymes which are responsible for proteolytic cleavage of APP (Figure 2).

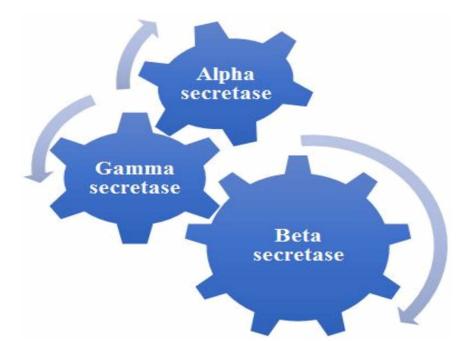


Figure 2. Enzymes involved in proteolytic cleavage of amyloid precursor protein

Alpha Secretase

Alpha secretase is synonymous with the metalloprotease activity that is believed to have an involvement in the prevention of underlying molecular pathways of AD pathogenesis. Increasing α -secretase associated proteolytic cleavage of APP is thought of as a therapeutic measure for AD (Lichtenthaler, 2012). Neurotoxic A β peptides are not formed if APP is first acted upon by α -secretase. α -secretase cleaves APP inside the A β sequence. This, in turn, prevents the formation neurotoxic A β peptides (Zhang & Saunders, 2007).

After several years of research on α -secretase, the chiefly involved enzymes have been identified as parts of a disintegrin and metalloproteinase (ADAM) family. The family comprises of three membrane proteins i.e. zinc-dependent metalloproteinases. Three members of this family i.e. ADAM 10, ADAM 17 and to some extent ADAM 9 are known to exhibit α -secretase activity. Additionally, some other membrane linked metalloproteinases may play a role in processing of APP (Postina, 2008). Several cellular signalling pathways encompassing phospholipase C (PLC), serine/threonine specific kinases (protein kinases C, MAP kinases) and phosphatidylinositol-3-kinase also contribute to expression of α -secretase. α -secretase dependent processing of APP is also known to be stimulated by direct activation of protein kinase C and activation of distinct G protein coupled receptors (Skovronsky, Moore, Milla, Doms, & Lee, 2000). Therefore, M1 muscarinic cholinergic agonists and 5-HT4/5-HT6 serotonergic agonists are being clinically evaluated for their potential in treatment of AD.

Amongst the ADAM family, ADAM 10 is the constitutive and most physiologically relevant α -secretase for shedding of APP. It partially competes with γ secretase to cleave down a C-terminal fragment generated due to β secretase (Kuhn et al., 2010). A study has reported an 81% decrease in α -secretase and 185% increase in β secretase in temporal cortex of patients of sporadic AD (Tyler, Dawbarn, Wilcock, & Allen, 2002).

 α -secretase activators show a potential to become a disease modifying therapeutic approach in the coming times.

Beta Secretase and Gamma Secretase

The most widely accepted hypothesis of AD is the aggregation of A β peptide in neurons leading to neurodegeneration. Beta secretase (BACE) is known to be the main culprit involved in the shedding of APP to generate neurotoxic A β aggregates in neurons. BACE, chiefly BACE-1, cleaves APP the product of which is further acted upon by gamma secretase to generate A β . It was a landmark discovery when BACE-1 (memapsin-2) was identified as a novel class of type 1 transmembrane aspartic protease. BACE-2 was identified soon after but it was revealed subsequently that BACE-1 is essential for majority of neuronal A β aggregation (Venugopal, Demos, Rao, Pappolla, & Sambamurti, 2008). BACE-1 causes endoproteolysis of APP to liberate A β N-terminal fragment, APPs β , and a C-terminal fragment, C99. Cleavage of APP by BACE1 is a prerequisite for γ -secretase dependent cleavage (Figure 3) (Robert Vassar, 2004). γ -secretase is multi subunit enzyme complex which plays a great role, subsequent to BACE1, in yielding A β from APP. It comprises of four principle components i.e. presenilin, nicastrin, anterior pharynx defective 1 and presenilin enhancer 2. These interact in a complex of high molecular weight to execute intramembrane proteolytic cleavage on a number of membrane-bound proteins including APP and Notch (Krishnaswamy, Verdile, Groth, Kanyenda, & Martins, 2009). Notch is a transmembrane receptor that regulates the cell-fate decisions (Barten, Meredith, Zaczek, Houston, & Albright, 2006).

Amyloid Beta

Assembly of these four subunits leads to autoproteolytic breakdown of presenilin to yield two subunits, each of which donates one aspartate to the active site of aspartyl protease. The aspartyl protease bears an initial docking site for substrate where it binds before passing between the two presenilin subunits to the internal active site. Modulators and inhibitors of γ -secretase have been in clinical assessment to evaluate therapeutic benefit in AD. The ultimate goal is to alter the function of γ secretase on APP without interfering with the processing of Notch signalling. The allosteric sites on the protease permit such selective moderation (Wolfe, 2008).

BACE 1 exists abundantly in human brain. The maximum BACE activity is seen in neural tissue while astrocytes exhibit less activity than neurons (R Vassar et al., 1999). BACE mRNA levels are low in resting glial cells. Like other members of pepsin family, BACE 1 comprises of two active site motifs and the mutation of either leads to inactivity (Bennett et al., 2000; Hussain et al., 1999). It is a type 1 membrane protease with a luminal active site which provides the optimal spatial orientation for cleavage of APP (Cole & Vassar, 2008). The gene expression of BACE1 is strictly moderated at the transcriptional level. The promoter of BACE1 comprises of a functional Sp1 response element. The gene expression of BACE gets potentiated by overexpression of transcription factor Sp1. The transcription factor Sp1 has been known to have the major involvement in modulating the BACE1 regulated cleavage of APP (Christensen et al., 2004). However, γ secretase cleavage is imprecise and yields A β isoforms of varying C terminus lengths. The longer isoforms are typically linked to AD (Robert Vassar, 2014).

BACE1 is the prime disease modifying therapeutic target for treatment of AD. Inhibition of BACE1 can lead to a substantial decrease in cerebral A β levels. Clinical development of BACE1 antagonists is being intensely researched upon. Although, BACE1 antagonism provides high hopes for prevention and treatment of AD, concerns have been rising about the mechanism based potential side effects of these agents (Yan & Vassar, 2014).

Ubiquitin-Proteasome System

Deposition of proteins is a recurring event in several NDs, including AD. Evidence is there suggesting that protein accumulation may be a consequence of an impaired UPS. Indeed, there is clear genetic and biochemical evidence of an involvement of the UPS in AD pathogenesis (Oddo, 2008) . UPS, the major intracellular protein quality control system in eukaryotes, is responsible for the clearance and degradation of A β (Hong, Huang, & Jiang, 2014). There is growing evidence showing that there exists a strong relationship between A β (Hong et al., 2014) and UPS and this relationship plays an important role in pathogenesis of AD. The disturbance in the UPS contributes to deposition of A β in brain. The impairment in the UPS in AD affects the degradation of A β and causes an abnormal aggregation of A β (Hong et al., 2014). Simultaneously, A β inhibits the proteasomal activity and subsequently causes impairment in multivesicular bodies (MVB) sorting pathway, forming an interacting relationship between A β and UPS. Mutant ubiquitin and ubiquitin-like ubiquilin-1 are related to A β accumulation (Hong et al., 2014). The proteasome is a constitutive multi-catalytic, multi-subunit protease complex which employs homopolymers of ubiquitin as a signal for targeting proteins for degradation in an ATP-dependent pathway (Baranello et al., 2015).

In mammals, the most common form is 26S proteasome comprising of a proteolytic 20S core subunit with two 19S regulatory subunits. The complex is a hollow moiety so as to provide an entrance for proteins to be takeen in and degraded (Thrower, Hoffman, Rechsteiner, & Pickart, 2000). The first event involves adenylation of ubiquitin by the ubiquitin activating enzyme E1. Then, it is transferred to

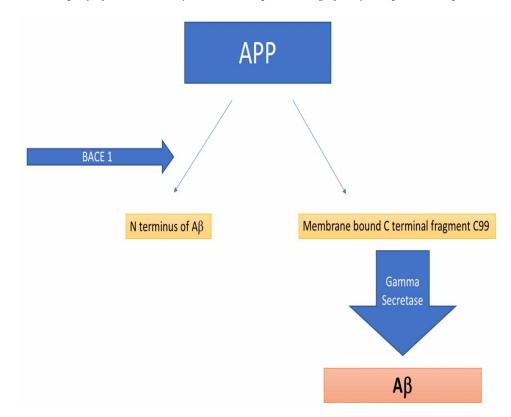


Figure 3. The interplay of BACE1 and γ secretase in processing of amyloid precursor protein to yield A β

the active site cysteine residue of E2- the ubiquitin conjugating enzyme. In the last event, a family of ubiquitin ligases – E3 is involved (Pappolla, Omar, Kim, & Robakis, 1992). It identifies specific targets and catalyses the transfer of ubiquitin from E2 to the target protein (Pappolla et al., 1992). A target protein has to be labelled by at least 4 ubiquitin molecules for it to be able to be recognized for proteolysis.

Despite the inhibitory action of A β 40 on the proteasome, it does not appear to be a substrate of the UPS. The proteasome can also moderate intracellular concentrations of PS1 and PS2, both, which indirectly influence γ secretase activity (Flood et al., 2005; Perry, Friedman, Shaw, & Chau, 1987).

AMYLOID BETA AND NEUROFIBRILLARY TANGLES: THE INTERPLAY

Apart from A β , NFTs are also invariably found in the brains of patients suffering from AD. The first experimental evidence that relates A β to tau, functionally, was described in a pair of landmark papers. Together, these studies concluded that increased tau pathology due to A β in the absence of any demonstrable effects on A β is caused by excess mutant tau. Thus, A β was reported to function upstream of tau, albeit by pathways that remained to be defined. (Gotz, Chen, van Dorpe, & Nitsch, 2001; Lewis et al., 2001).

During the past few years, ample evidence has been reported suggesting that the soluble forms of $A\beta$ and tau act in combination, independent of their accumulation into plaques and tangles, to take neurons from healthy state to the diseased state. Also, it has been reported that hallmark toxic properties of $A\beta$

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require tau (Bloom, 2014). The evidence that suggests that A β is influenced by tau, together with earlier reports that clearly demonstrated that A β clearly functions upstream of tau (Hurtado et al., 2010), creates the chances that A β triggers off a pathological feedback events with tau.

A protein that is responsible for a functional connection between A β and tau is fyn. It is a nonreceptor tyrosine kinase that causes positive regulation of NMDA receptor activity which was recently reported to be targeted to postsynaptic sites in dendrites by tau (Ittner et al., 2010) which binds fyn directly (G. Lee, Newman, Gard, Band, & Panchamoorthy, 1998). Tau is otherwise highly available in axons as compared to dendrites (Binder, Frankfurter, & Rebhun, 1985) but in response to A β , tau undergoes extensive redistribution into the somatodendritic compartment (Delacourte et al., 1990; H. Zempel, Thies, Mandelkow, & Mandelkow, 2010). Acute cytotoxicity is not the only tau-dependent manifestation of A β on cultured dissociated neurons or brain slices (Nussbaum et al., 2012; Rapoport, Dawson, Binder, Vitek, & Ferreira, 2002). A β oligomers have been reported to bring about tau-dependent microtubule disassembly (King et al., 2006), impaired long-term potentiation (Shipton et al., 2011), dendritic microtubule severing (Hans Zempel et al., 2013), inhibition of mitochondrial transport along microtubules (Vossel et al., 2010), and ectopic cell cycle re-entry of neurons (Seward et al., 2013), which eventually leads to massive neuronal death in AD.

NEW DEVELOPMENTS IN PHARMACOTHERAPEUTICS FOR THE MANAGEMENT OF ALZHEIMER'S DISEASE

The development of new therapeutic interventions for AD is under the threat of chasing the wrong pathology. Because of little concrete evidence base pertaining to a definite underlying pathological mechanism for development of AD, hypotheses based pharmacological interventions are being developed. Earlier, cholinergic hypothesis of development of AD emerged as a potential ground to build research upon. Several compounds were developed to target the acetylcholinesterase enzyme in order to elevate the levels of brain ACh. These agents are still in use but they provide only symptomatic relief. Also, a wide spectrum of side effects is associated with these agents. A few decades later, memantine came into picture as a protectant against glutamate excitotoxicity. However, this drug also could not yield effect better than mere symptomatic relief.

Several new chemical entities claiming to have potential benefits in AD have been developed by researchers all over the globe. However, a lot of them failed in the clinical trials due to different reasons mostly the severe adverse effects and lack of significant efficacy. A major factor that contributes to the increasing failures in development of new therapeutic measures for AD is that the current trial designs are quite nonspecific and blunt. Measure of cognitive abilities is a reductionist approach as the disease is too complex. Most of the pathobiological alterations do not manifest in alterations in cognitive processes and hence these two parameters cannot be correlated. The drug candidates targeting amyloid pathology are often employed at the later stages when A β related neurodegeneration has begun. This further adds to the increase in failures of new anti-AD therapeutics. This has not stopped the researchers from creating new possibilities for the treatment of AD (Anand, Patience, et al., 2017). The preclinical screening of novel entities for anti-AD effects is being carried out on a large scale worldwide. It is worth noting that no new drug (excluding nutraceutical products) has been able to obtain FDA's approval for treatment of AD in the last decade (Berk & Sabbagh, 2013). A great attrition has been found, with an

overall failure rate of 99.6% (0.4% success) in the decade spanning from 2002 to 2012 in the field of anti-AD therapeutics. The trial length and magnitude for disease modifying agents is greater than those for symptomatic agents. The success rate of an entity's progression from one phase of clinical trial to the next is low. The number of entities advancing to regulatory review is among the lowest seen in any area of therapeutics (Cummings, Morstorf, & Zhong, 2014).

Due to the limited options for the therapeutic agents, several new molecules have come in the recent years claiming to have a curative potential for AD. The existing pipeline for anti-AD drugs is quite modest in the light of the tremendous challenges associated with AD. Currently, AD is proving to be very expensive to the US economy, even more than cancer and cardiovascular diseases (Hurd, Martorell, Delavande, Mullen, & Langa, 2013).

Entities currently in clinical trials globally (as of 2017) targetting the amyloid pathological pathway of AD (Anand, Patience, et al., 2017) are given in Table 1.

FUTURE RESEARCH DIRECTIONS

Despite the exhaustive research that has happened in delineating the role of amyloid beta in AD, still certain unexplored areas remain in this amyloid hypothesis of AD. Mismatched APP cleavage by three proteolytic enzymes i.e. alpha, beta and gamma secretases, is believed to be the major reason for the production of neurotoxic A β peptides. The exact role of amyloid pathway of AD pathogenesis is still gray. There are several assumptions and hypotheses associated with it. An elaborate thrust on research areas intending to more clearly elucidate the intricate molecular mechanisms linked to amyloid pathway of AD pathogenesis needs to see the light of day. Recently unsuccessful antiamyloidogenic trials portray a strong need for an objective reassessment of the much talked about amyloid hypothesis of AD. The existing efforts with focus on A β , its deposition and accumulation, and its removal need to be supported by more intensive research in new directions. Once the exact molecular mechanisms, reaction cascades, proteolysis sequences and other associated targets are identified more clearly, effective and safe disease modifying pharmacotherapeutics can be developed.

Entity	C ompany	Description	Reference
AZD3293	AstraZeneca	Beta secretase inhibitor	Lilly, 2017
BACE inhibitor	Lilly	Beta secretase inhibitor	Lilly, 2017
CNP 520	Amgen	Beta secretase protein inhibitor	Amgen, 2017
E2609	Biogen	Beta secretase inhibitor	Biogen, 2017; Eisai, 2017
LY3314814	Lilly	Beta secretase inhibitor	Lilly, 2017
NGP 555	Neurogenetic Pharmaceuticals	Reported to cross blood brain barrier and effectively reducing the brain biomarkers A β 42 and A β 40 (in mice harbouring the human APP) by modulating the gamma secretase complex	NeuroGenetic Pharmaceuticals, 2017

Table 1. Entities currently in clinical trials for Alzheimer's disease.

Amyloid Beta

Future research directions may be oriented thereafter towards increasing the expression of alpha secretase as its action on APP does not yield a neurotoxic product (Zhang & Saunders, 2007). Beta secretase and gamma secretase inhibitors are surely to experience an elaborate and tremendous research as they are the key enzymes that are responsible for generation of A β plaques which in turn cause neurodegeneration (Venugopal et al., 2008). Also, UPS stabilizers may be developed to halt or delay the impairment in the system thereby ensuring efficient clearance of formed A β plaques (Hong et al., 2014).

CONCLUSION

AD being a growing health concern worldwide has received a lot of attention from researchers in the past few decades. Despite enormous research, only symptomatic relief could be provided to the sufferers of AD. A β hypothesis of AD is the most widely accepted hypothesis for pathogenesis of AD. It states that the beta and gamma secretase linked cleavage of APP in brain leads to the formation of neurotoxic A β plaques. Also, an impairment in UPS system further augments the intensity of AD as there is significantly reduced clearance of A β from brain. All these events contribute to the state of loss of neuronal connections in brain and eventually cause a progressive and irreversible loss in memory and cognitive abilities. From developing safer and more effective drug candidates for the existing therapeutic targets (cholinesterase, NMDA glutamate receptors) to developing moieties focusing on newer targets (A β , BACE, gamma secretase, α -secretase), the researchers have been contributing their efforts to bring a preventive and/or curative therapy for AD. With an in-depth understanding of the amyloid hypothesis of AD, an effective disease modifying therapy for AD can be developed in the coming years.

REFERENCES

Alzheimer's Association. (2014). 2014 Alzheimer's disease facts and figures. *Alzheimer's & Dementia*, *10*(2), e47–e92. doi:10.1016/j.jalz.2014.02.001 PMID:24818261

Alzheimer's Association. (2017a). *Stages of Alzheimer's and Symptoms* | *Alzheimer's Association*. Retrieved March 6, 2017, from http://www.alz.org/alzheimers_disease_stages_of_alzheimers.asp

Alzheimer's Association. (2017b). *Types of Dementia*. Retrieved April 1, 2017, from http://www.alz. org/dementia/types-of-dementia.asp

Alzheimer's Association. (2017c). *What is Dementia?* Retrieved April 1, 2017, from http://www.alz. org/what-is-dementia.asp

Amgen. (2017). Pipeline. Retrieved March 7, 2017, from http://www.amgenpipeline.com/pipeline/

Anand, A., Khurana, P., Chawla, J., Sharma, N., & Khurana, N. (2017). Emerging treatments for the behavioral and psychological symptoms of dementia. *CNS Spectrums*, 1–9. doi:10.1017/S1092852917000530 PMID:28911339

Anand, A., Patience, A. A., Sharma, N., & Khurana, N. (2017). The present and future of pharmacotherapy of Alzheimer's disease: A comprehensive review. *European Journal of Pharmacology*, *815*, 364–375. doi:10.1016/j.ejphar.2017.09.043 PMID:28978455 Baranello, R. J., Bharani, K. L., Padmaraju, V., Chopra, N., Lahiri, D. K., Greig, N. H., ... Sambamurti, K. (2015). Amyloid-beta protein clearance and degradation (ABCD) pathways and their role in Alzheimer's disease. *Current Alzheimer Research*, *12*(1), 32–46. Retrieved from http://www.ncbi.nlm. nih.gov/pubmed/25523424

Barten, D. M., Meredith, J. E. Jr, Zaczek, R., Houston, J. G., & Albright, C. F. (2006). Gamma-secretase inhibitors for Alzheimer's disease: Balancing efficacy and toxicity. *Drugs in R&D.*, 7(2), 87–97. doi:10.2165/00126839-200607020-00003 PMID:16542055

Bennett, B. D., Denis, P., Haniu, M., Teplow, D. B., Kahn, S., Louis, J.-C., ... Vassar, R. (2000). A Furinlike Convertase Mediates Propeptide Cleavage of BACE, the Alzheimer's β-Secretase. *The Journal of Biological Chemistry*, 275(48), 37712–37717. doi:10.1074/jbc.M005339200 PMID:10956649

Berk, C., & Sabbagh, M. N. (2013). Successes and failures for drugs in late-stage development for Alzheimer's disease. *Drugs & Aging*, *30*(10), 783–792. doi:10.100740266-013-0108-6 PMID:23943247

Binder, L. I., Frankfurter, A., & Rebhun, L. I. (1985). The distribution of tau in the mammalian central nervous system. *The Journal of Cell Biology*, *101*(4), 1371–1378. doi:10.1083/jcb.101.4.1371 PMID:3930508

Biogen. (2017). *Research Pipeline*. Retrieved March 7, 2017, from https://www.biogen.com/en_us/ research-pipeline/biogen-pipeline.html

Bloom, G. S. (2014). Amyloid- β and Tau. JAMA Neurology, 71(4), 505. doi:10.1001/jamaneurol.2013.5847 PMID:24493463

Butterfield, D., Castegna, A., Pocernich, C., Drake, J., Scapagnini, G., & Calabrese, V. (2002). Nutritional approaches to combat oxidative stress in Alzheimer's disease. *The Journal of Nutritional Biochemistry*, *13*(8), 444–461. doi:10.1016/S0955-2863(02)00205-X PMID:12165357

Caporaso, G. L., Gandy, S. E., Buxbaum, J. D., & Greengard, P. (1992). Chloroquine inhibits intracellular degradation but not secretion of Alzheimer beta/A4 amyloid precursor protein. *Proceedings of the National Academy of Sciences of the United States of America*, 89(6), 2252–2256. doi:10.1073/ pnas.89.6.2252 PMID:1549591

Centers for Disease Control and Prevention. (2017). *Leading Causes of Death*. Retrieved April 4, 2017, from https://www.cdc.gov/nchs/fastats/leading-causes-of-death.htm

Christensen, M. A., Zhou, W., Qing, H., Lehman, A., Philipsen, S., & Song, W. (2004). Transcriptional regulation of BACE1, the beta-amyloid precursor protein beta-secretase, by Sp1. *Molecular and Cellular Biology*, *24*(2), 865–874. doi:10.1128/MCB.24.2.865-874.2004 PMID:14701757

Citron, M., Teplow, D. B., & Selkoe, D. J. (1995). Generation of amyloid beta protein from its precursor is sequence specific. *Neuron*, *14*(3), 661–670. doi:10.1016/0896-6273(95)90323-2 PMID:7695913

Cole, S. L., & Vassar, R. (2008). The role of amyloid precursor protein processing by BACE1, the beta-secretase, in Alzheimer disease pathophysiology. *The Journal of Biological Chemistry*, 283(44), 29621–29625. doi:10.1074/jbc.R800015200 PMID:18650431

Amyloid Beta

Cummings, J. L., Morstorf, T., & Zhong, K. (2014). Alzheimer's disease drug-development pipeline: Few candidates, frequent failures. *Alzheimer's Research & Therapy*, *6*(4), 37. doi:10.1186/alzrt269 PMID:25024750

Davies, L., Wolska, B., Hilbich, C., Multhaup, G., Martins, R., Simms, G., ... Masters, C. L. (1988). A4 amyloid protein deposition and the diagnosis of Alzheimer's disease: Prevalence in aged brains determined by immunocytochemistry compared with conventional neuropathologic techniques. *Neurology*, *38*(11), 1688–1693. doi:10.1212/WNL.38.11.1688 PMID:3054625

Delacourte, A., Flament, S., Dibe, E. M., Hublau, P., Sablonnière, B., Hémon, B., ... Défossez, A. (1990). Pathological proteins Tau 64 and 69 are specifically expressed in the somatodendritic domain of the degenerating cortical neurons during Alzheimer's disease. Demonstration with a panel of antibodies against Tau proteins. *Acta Neuropathologica*, *80*(2), 111–7. Retrieved from http://www.ncbi.nlm.nih. gov/pubmed/2117840

Dwyer, B. E., Zacharski, L. R., Balestra, D. J., Lerner, A. J., Perry, G., Zhu, X., & Smith, M. A. (2009). Getting the iron out: Phlebotomy for Alzheimer's disease? *Medical Hypotheses*, 72(5), 504–509. doi:10.1016/j.mehy.2008.12.029 PMID:19195795

Editors of Encyclopædia Britannica. (2016). Alzheimer Disease. In *Encyclopaedia Britannica* (pp. 1–14). Encyclopædia Britannica, Inc. Retrieved from https://www.britannica.com/science/Alzheimer-disease

Eisai. (2017). Major R and D Pipeline. Retrieved from http://www.eisai.com/pdf/eir/erepo/epipeline.pdf

Flood, F., Murphy, S., Cowburn, R. F., Lannfelt, L., Walker, B., & Johnston, J. A. (2005). Proteasomemediated effects on amyloid precursor protein processing at the γ-secretase site. *The Biochemical Journal*, *385*(2), 545–550. doi:10.1042/BJ20041145 PMID:15473868

Francis, P. T. (2005). The interplay of neurotransmitters in Alzheimer's disease. CNS Spectrums, 10(11Suppl 18), 6–9. doi:10.1017/S1092852900014164 PMID:16273023

Glenner, G. G., & Wong, C. W. (1984a). Alzheimer's disease: Initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Biochemical and Biophysical Research Communications*, *120*(3), 885–890. doi:10.1016/S0006-291X(84)80190-4 PMID:6375662

Glenner, G. G., & Wong, C. W. (1984b). Alzheimer's disease and Down's syndrome: Sharing of a unique cerebrovascular amyloid fibril protein. *Biochemical and Biophysical Research Communications*, *122*(3), 1131–1135. doi:10.1016/0006-291X(84)91209-9 PMID:6236805

Gotz, J., Chen, F., van Dorpe, J., & Nitsch, R. M. (2001). Formation of Neurofibrillary Tangles in P301L Tau Transgenic Mice Induced by Abeta 42 Fibrils. *Science*, 293(5534), 1491–1495. doi:10.1126cience.1062097 PMID:11520988

Guillozet, A. L., Weintraub, S., Mash, D. C., & Mesulam, M. M. (2003). Neurofibrillary Tangles, Amyloid, and Memory in Aging and Mild Cognitive Impairment. *Archives of Neurology*, *60*(5), 729. doi:10.1001/archneur.60.5.729 PMID:12756137

Habibyar, A. F., Sharma, N., & Khurana, N. (2016). PASS assisted prediction and pharmacological evaluation of hesperidin against scopolamine induced amnesia in mice. *European Journal of Pharmacology*, 789, 385–394. doi:10.1016/j.ejphar.2016.07.013 PMID:27397428

Hardy, J. (2006). Alzheimer's disease: the amyloid cascade hypothesis: an update and reappraisal. *Journal of Alzheimer's Disease : JAD, 9*(3 Suppl), 151–3. Retrieved from http://www.ncbi.nlm.nih. gov/pubmed/16914853

Hardy, J., & Selkoe, D. J. (2002). The Amyloid Hypothesis of Alzheimer's Disease: Progress and Problems on the Road to Therapeutics. *Science*, 297(5580), 353–356. doi:10.1126cience.1072994 PMID:12130773

Hong, L., Huang, H.-C., & Jiang, Z.-F. (2014). Relationship between amyloid-beta and the ubiquitin– proteasome system in Alzheimer's disease. *Neurological Research*, *36*(3), 276–282. doi:10.1179/1743 132813Y.0000000288 PMID:24512022

Hurd, M. D., Martorell, P., Delavande, A., Mullen, K. J., & Langa, K. M. (2013). Monetary Costs of Dementia in the United States. *The New England Journal of Medicine*, *368*(14), 1326–1334. doi:10.1056/ NEJMsa1204629 PMID:23550670

Hurtado, D. E., Molina-Porcel, L., Iba, M., Aboagye, A. K., Paul, S. M., Trojanowski, J. Q., & Lee, V. M.-Y. (2010). Aβ Accelerates the Spatiotemporal Progression of Tau Pathology and Augments Tau Amyloidosis in an Alzheimer Mouse Model. *American Journal of Pathology*, *177*(4), 1977–1988. doi:10.2353/ajpath.2010.100346 PMID:20802182

Hussain, I., Powell, D., Howlett, D. R., Tew, D. G., Meek, T. D., Chapman, C., ... Christie, G. (1999). Identification of a Novel Aspartic Protease (Asp 2) as β -Secretase. *Molecular and Cellular Neurosciences*, *14*(6), 419–427. doi:10.1006/mcne.1999.0811 PMID:10656250

Ittner, L. M., Ke, Y. D., Delerue, F., Bi, M., Gladbach, A., van Eersel, J., ... Götz, J. (2010). Dendritic Function of Tau Mediates Amyloid-β Toxicity in Alzheimer's Disease Mouse Models. *Cell*, *142*(3), 387–397. doi:10.1016/j.cell.2010.06.036 PMID:20655099

King, M. E., Kan, H.-M., Baas, P. W., Erisir, A., Glabe, C. G., & Bloom, G. S. (2006). Tau-dependent microtubule disassembly initiated by prefibrillar β -amyloid. *The Journal of Cell Biology*, *175*(4), 541–546. doi:10.1083/jcb.200605187 PMID:17101697

Krishnaswamy, S., Verdile, G., Groth, D., Kanyenda, L., & Martins, R. N. (2009). The structure and function of Alzheimer's gamma secretase enzyme complex. *Critical Reviews in Clinical Laboratory Sciences*, *46*(5–6), 282–301. doi:10.3109/10408360903335821 PMID:19958215

Kuhn, P.-H., Wang, H., Dislich, B., Colombo, A., Zeitschel, U., Ellwart, J. W., ... Lichtenthaler, S. F. (2010). ADAM10 is the physiologically relevant, constitutive α -secretase of the amyloid precursor protein in primary neurons. *The EMBO Journal*, 29(17), 3020–3032. doi:10.1038/emboj.2010.167 PMID:20676056

Lee, G., Newman, S. T., Gard, D. L., Band, H., & Panchamoorthy, G. (1998). Tau interacts with src-family non-receptor tyrosine kinases. *Journal of Cell Science*, *111*(Pt 21), 3167–3177. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/9763511 PMID:9763511

Amyloid Beta

Lee, H., Zhu, X., Castellani, R. J., Nunomura, A., Perry, G., & Smith, M. A. (2007). Amyloid-beta in Alzheimer disease: The null versus the alternate hypotheses. *The Journal of Pharmacology and Experimental Therapeutics*, *321*(3), 823–829. doi:10.1124/jpet.106.114009 PMID:17229880

Lee, M., Kwon, Y. T., Li, M., Peng, J., Friedlander, R. M., & Tsai, L.-H. (2000). Neurotoxicity induces cleavage of p35 to p25 by calpain. *Nature*, *405*(6784), 360–364. doi:10.1038/35012636 PMID:10830966

Lewis, J., Dickson, D. W., Lin, W. L., Chisholm, L., Corral, A., Jones, G., ... McGowan, E. (2001). Enhanced Neurofibrillary Degeneration in Transgenic Mice Expressing Mutant Tau and APP. *Science*, *293*(5534), 1487–1491. doi:10.1126cience.1058189 PMID:11520987

Lichtenthaler, S. F. (2012). Alpha-secretase cleavage of the amyloid precursor protein: Proteolysis regulated by signaling pathways and protein trafficking. *Current Alzheimer Research*, 9(2), 165–177. doi:10.2174/156720512799361655 PMID:21605033

Lilly. (2017). *Clinical Development Pipeline*. Retrieved March 7, 2017, from https://www.lilly.com/pipeline/index.html

Mann, D. M., Jones, D., South, P. W., Snowden, J. S., & Neary, D. (1992). Deposition of amyloid beta protein in non-Alzheimer dementias: Evidence for a neuronal origin of parenchymal deposits of beta protein in neurodegenerative disease. *Acta Neuropathologica*, *83*(4), 415–419. doi:10.1007/BF00713534 PMID:1575018

McGleenon, B. M., Dynan, K. B., & Passmore, A. P. (1999). Acetylcholinesterase inhibitors in Alzheimer's disease. *British Journal of Clinical Pharmacology*, 48(4), 471–480. doi:10.1046/j.1365-2125.1999.00026.x PMID:10583015

Mucke, L., Abraham, C. R., Ruppe, M. D., Rockenstein, E. M., Toggas, S. M., Mallory, M., ... Masliah, E. (1995). Protection against HIV-1 gp120-induced brain damage by neuronal expression of human amyloid precursor protein. *The Journal of Experimental Medicine*, *181*(4), 1551–6. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/7699335

NeuroGenetic Pharmaceuticals. (2017). *Products and Technology*. Retrieved March 7, 2017, from http:// neurogeneticpharmaceuticals.com/

Nisbet, R. M., Polanco, J.-C., Ittner, L. M., & Götz, J. (2015). Tau aggregation and its interplay with amyloid-β. *Acta Neuropathologica*, *129*(2), 207–220. doi:10.100700401-014-1371-2 PMID:25492702

Nussbaum, J. M., Schilling, S., Cynis, H., Silva, A., Swanson, E., Wangsanut, T., ... Bloom, G. S. (2012). Prion-like behaviour and tau-dependent cytotoxicity of pyroglutamylated amyloid-β. *Nature*, 485(7400), 651–655. doi:10.1038/nature11060 PMID:22660329

Oddo, S. (2008). The ubiquitin-proteasome system in Alzheimer's disease. *Journal of Cellular and Molecular Medicine*, *12*(2), 363–373. doi:10.1111/j.1582-4934.2008.00276.x PMID:18266959

Pappolla, M. A., Omar, R. A., Kim, K. S., & Robakis, N. K. (1992). Immunohistochemical evidence of oxidative [corrected] stress in Alzheimer's disease. *American Journal of Pathology*, *140*(3), 621–628. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/1372157 PMID:1372157

Perl, D. P. (2010). Neuropathology of Alzheimer â€TM s Disease. *The Mount Sinai Journal of Medicine*, *New York*, 77(1), 32–42. doi:10.1002/msj.20157 PMID:20101720

Perry, G., Friedman, R., Shaw, G., & Chau, V. (1987). Ubiquitin is detected in neurofibrillary tangles and senile plaque neurites of Alzheimer disease brains. *Proceedings of the National Academy of Sciences of the United States of America*, 84(9), 3033–3036. doi:10.1073/pnas.84.9.3033 PMID:3033674

Postina, R. (2008). A closer look at alpha-secretase. *Current Alzheimer Research*, 5(2), 179–186. doi:10.2174/156720508783954668 PMID:18393803

Rapoport, M., Dawson, H. N., Binder, L. I., Vitek, M. P., & Ferreira, A. (2002). Tau is essential to -amyloid-induced neurotoxicity. *Proceedings of the National Academy of Sciences of the United States of America*, 99(9), 6364–6369. doi:10.1073/pnas.092136199 PMID:11959919

Sadigh-Eteghad, S., Sabermarouf, B., Majdi, A., Talebi, M., Farhoudi, M., & Mahmoudi, J. (2015). Amyloid-beta: a crucial factor in Alzheimer's disease. *Medical Principles and Practice : International Journal of the Kuwait University. Health Science Centre*, 24(1), 1–10. doi:10.1159/000369101

Schmitt, F. A., Davis, D. G., Wekstein, D. R., Smith, C. D., Ashford, J. W., & Markesbery, W. R. (2000). Preclinal AD revisited: Neuropathology of cognitively normal older adults. *Neurology*, *55*(3), 370–376. doi:10.1212/WNL.55.3.370 PMID:10932270

Seward, M. E., Swanson, E., Norambuena, A., Reimann, A., Cochran, J. N., Li, R., ... Bloom, G. S. (2013). Amyloid-signals through tau to drive ectopic neuronal cell cycle re-entry in Alzheimer's disease. *Journal of Cell Science*, *126*(5), 1278–1286. doi:10.1242/jcs.1125880 PMID:23345405

Shipton, O. A., Leitz, J. R., Dworzak, J., Acton, C. E. J., Tunbridge, E. M., Denk, F., ... Vargas-Caballero, M. (2011). Tau Protein Is Required for Amyloid -Induced Impairment of Hippocampal Long-Term Potentiation. *The Journal of Neuroscience*, *31*(5), 1688–1692. doi:10.1523/JNEUROSCI.2610-10.2011 PMID:21289177

Skovronsky, D. M., Moore, D. B., Milla, M. E., Doms, R. W., & Lee, V. M. (2000). Protein kinase C-dependent alpha-secretase competes with beta-secretase for cleavage of amyloid-beta precursor protein in the trans-golgi network. *The Journal of Biological Chemistry*, 275(4), 2568–2575. doi:10.1074/jbc.275.4.2568 PMID:10644715

Storey & Cappai. (1999). The amyloid precursor protein of Alzheimer's disease and the Abeta peptide. *Neuropathology and Applied Neurobiology*, 25(2), 81–97. 10.1046/j.1365-2990.1999.00164.x

Swerdlow, R. H., Burns, J. M., & Khan, S. M. (2014). The Alzheimer's disease mitochondrial cascade hypothesis: Progress and perspectives. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, *1842*(8), 1219–1231. doi:10.1016/j.bbadis.2013.09.010 PMID:24071439

Thrower, J. S., Hoffman, L., Rechsteiner, M., & Pickart, C. M. (2000). Recognition of the polyubiquitin proteolytic signal. *The EMBO Journal*, *19*(1), 94–102. doi:10.1093/emboj/19.1.94 PMID:10619848

Tyler, S. J., Dawbarn, D., Wilcock, G. K., & Allen, S. J. (2002). alpha- and beta-secretase: Profound changes in Alzheimer's disease. *Biochemical and Biophysical Research Communications*, 299(3), 373–376. doi:10.1016/S0006-291X(02)02635-9 PMID:12445809

Amyloid Beta

Vassar, R. (2004). BACE1: The β-Secretase Enzyme in Alzheimer's Disease. *Journal of Molecular Neuroscience*, 23(1–2), 105–114. doi:10.1385/JMN:23:1-2:105 PMID:15126696

Vassar, R. (2014). BACE1 inhibitor drugs in clinical trials for Alzheimer's disease. *Alzheimer's Research* & *Therapy*, 6(9), 89. doi:10.118613195-014-0089-7 PMID:25621019

Vassar, R., Bennett, B. D., Babu-Khan, S., Kahn, S., Mendiaz, E. A., Denis, P., ... Citron, M. (1999). Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. *Science (New York, N.Y.), 286*(5440), 735–41. Retrieved from http://www.ncbi.nlm.nih. gov/pubmed/10531052

Venugopal, C., Demos, C. M., Rao, K. S. J., Pappolla, M. A., & Sambamurti, K. (2008). Beta-secretase: Structure, function, and evolution. *CNS & Neurological Disorders - Drug Targets*, 7(3), 278–294. doi:10.2174/187152708784936626 PMID:18673212

Vossel, K. A., Zhang, K., Brodbeck, J., Daub, A. C., Sharma, P., Finkbeiner, S., ... Mucke, L. (2010). Tau Reduction Prevents A β -Induced Defects in Axonal Transport: Fig. 1. *Science*, *330*(6001), 198–198. doi:10.1126cience.1194653 PMID:20829454

Wolfe, M. S. (2008). Gamma-secretase: Structure, function, and modulation for Alzheimer's disease. *Current Topics in Medicinal Chemistry*, 8(1), 2–8. doi:10.2174/156802608783334024 PMID:18220927

World Health Organization. (2016). Dementia Fact Sheet. WHO. Retrieved from http://www.who.int/ mediacentre/factsheets/fs362/en/

Yan, R., & Vassar, R. (2014). Targeting the β secretase BACE1 for Alzheimer's disease therapy. *Lancet Neurology*, *13*(3), 319–329. doi:10.1016/S1474-4422(13)70276-X PMID:24556009

Zempel, H., Luedtke, J., Kumar, Y., Biernat, J., Dawson, H., Mandelkow, E., & Mandelkow, E.-M. (2013). Amyloid-β oligomers induce synaptic damage via Tau-dependent microtubule severing by TTLL6 and spastin. *The EMBO Journal*, *32*(22), 2920–2937. doi:10.1038/emboj.2013.207 PMID:24065130

Zempel, H., Thies, E., Mandelkow, E., & Mandelkow, E.-M. (2010). A Oligomers Cause Localized Ca2+ Elevation, Missorting of Endogenous Tau into Dendrites, Tau Phosphorylation, and Destruction of Microtubules and Spines. *The Journal of Neuroscience*, *30*(36), 11938–11950. doi:10.1523/JNEU-ROSCI.2357-10.2010 PMID:20826658

Zhang, C., & Saunders, A. J. (2007). Therapeutic targeting of the alpha-secretase pathway to treat Alzheimer's disease. *Discovery Medicine*, 7(39), 113–117. Retrieved from http://www.ncbi.nlm.nih.gov/ pubmed/18093473 PMID:18093473

Chapter 12 Parkinson's Disease: A Progressive Disorder of the Nervous System That Affects Movement

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ABSTRACT

Parkinson's disease (PD) is a neurodegenerative disorder in which a progressive loss of the dopaminergic neurons occurs. The loss of the neurons is most prominent in the substantia nigra region of the brain. The prevalence of PD is much greater among the older patients suggesting the risk of PD increases with the increase of age. The exact cause of the neurodegeneration in PD is not known. In this chapter, the authors introduce PD, demonstrate its history, pathogenesis, neurobiology, sign and symptoms, diagnosis, and pharmacotherapy.

INTRODUCTION

Parkinson's disease (PD) was first introduced by James Parkinson and arises by the progressive death of the dopamine neurons in the midbrain (Dauer & Przedborski, 2003). Neuronal cell death in the SN region in PD gives rise to the deficiency of dopamine in various region of brain including striatum, basal ganglia (BG) and subthalamic nucleus (STN) (Forno, 1996; Hornykiewicz & Kish, 1987). Increase in the age generally above 65 years increased the risk of the PD development; only a small number of individuals develop PD in early ages suggesting aging increased the risk of PD (Olanow & Tatton, 1999; Tanner, 2003). Thus older age individuals are particularly susceptible to PD (Fahn, 2003; Moghal et al,

DOI: 10.4018/978-1-5225-5282-6.ch012

Parkinson's Disease

1994). The death of the dopaminergic neurons in PD results in the development of the motor symptoms of the PD (Goldenberg et al, 2008). The exact mechanism of the neuronal cell death in PD is not known completely, however it suggested that the mutations in the number of genes linked PD (Mizuno et al, 2001; Van Den Eeden et al, 2003). The prevalence of the familial PD is approx. 5% whereas for sporadic forms it is about 95% (Farrer, 2006; Tanner, 2003). In this chapter authors demonstrate the history, pathogenesis, neurobiology, symptoms, diagnosis and pharmacotherapy of PD.

BACKGROUND

SN region of the brain is rich in the dopamine containing neurons and the neuronal cell death in the SN region increases with the increase in the age and thus increased the risk of PD (Olanow & Tatton, 1999; Tanner, 2003). The death of the dopaminergic neurons in PD results in the development of the motor symptoms of the PD (Goldenberg et al, 2008). The exact mechanism of the neuronal cell death in PD is not known completely. Further it has been reported that besides SN region the neuronal cell death also occurs in the various other regions of the brain responsible for the complexity of the pathogenesis and the variety of the symptoms observed in the patients of the PD. However there are several therapeutics options are present for the pharmacotherapeutics of the PD patients but till date there is no suitable pharmacotherapeutic agent which can stop the neuronal degeneration in the PD. Further the complexity of the symptoms often makes the diagnosis and the therapeutics of the PD patients difficult. The present chapter demonstrates the history, genetics, neuronal degeneration, symptoms and the therapeutics of PD in details

HISTORY OF PARKINSON'S DISEASE

In 1817 James Parkinsons wrote "Essay on the Shaking Palsy" and described features including flexed posture, resting tremor, and shuffling gait in his monograph (Parkinson, 1817). After Parkinson, 50 years later use the term "Parkinson's disease". He demonstrated that the PD patients did not have tremor necessarily. He suggested that PD patients performs all task, but performs the task slowly suggesting the problems lies in the execution. Charcot defines two prototypes of PD, rigid and tremorous PD (Charcot, 1872). William Gowers, suggested that males are more susceptible to PD and described the impairment in the movement of fingers in PD patients (Gowers, 1898). Richer and Meige demonstrated the clinical and morphologic details regarding the various stages of PD (Richer & Meige, 1895). Adams and colleagues first described the degeneration of neurons in the SN region (Adams et al, 1964). Brissaud described the neuronal damage in the SN region as the anatomical seat of PD (Brissaud, 1895). Involvement of midbrain in context of PD were then described (Foix & Nicolesco, 1925; Tretiakoff, 1919). Greenfield & Bosanquet, described the involvement of brainstem in PD (Greenfield & Bosanquet, 1953). Hoehn & Yahr, described the stages to described the clinical progression of the PD (Hoehn & Yahr, 1967). The development of MPTP proves beneficial as it damages SN regions, so that the potential compounds can be tested in the preclinical studies (Langston et al, 1983).

GENETIC MUTATIONS AND PARKINSON'S DISEASE

The sporadic form of the PD is more common as compared to the PD associated with the family history. However 6 genes are targeted in context to PD and the mutations in these genes is responsible for the PD cases. These genes are located on the autosomes and therefore the PD arises from the mutations in these genes can be of 2 types, autosomal recessive and autosomal dominant. Further the autosomal-dominant PD develops due to the mutations in *SNCA* and *LRRK2* genes while autosomal-recessive PD develops due to the mutations in *Parkin, PINK1, DJ-1*, and *ATP13A2* (Klein & Westenberger, 2012).

 α -synuclein is a filamentous protein present in LBs and the presence of LBs is the hallmark of PD. α -synuclein performs various functions including synaptic function and vesicular trafficking. It is reported that the overexpression of α -synuclein disrupts the mitochondrial functions, reduces tyrosine hydroxylase activity, and impairs synaptic function. It is reported that the mutation in SNCA gene results in the formation of the mutant protein which further contributes to dopamine toxicity by the oxidative stress (Campelo et al, 2017). SNCA gene encodes α -synuclein and SNCA gene mutation is responsible for the autosomal-dominant PD with the early onset (Klein & Westenberger, 2012). SNCA gene found physiologically has T instead of an A at position 53 of the protein, while the substitution of these amino acids in humans leads to the manifestation of PD. Further the A53T mutation of SNCA results in the impairments seen in the patients of the PD (Oczkowska et al., 2013). PINK1-Parkin pathway plays a key role in the pathogenesis of the neurodegenerative disorders e.g. PD (Narendra et al, 2008). PINK1 is located on the outer mitochondrial membrane upon stimulated it is translocated inside the mitochondria where it is partially degraded by presentiin-associated rhomboid like protein (PARL) resulting in the release of the degraded *PINK1* (N terminal degraded) into the cytoplasm and in the cytoplasm it is gain degraded by the ubiquitin proteasome system (UPS). Thus a very low undetectable level of PINK1 is present inside the mitochondria. Accumulation of the misfolded protein, results in the accumulation of the *PINK1* on the outer mitochondrial membrane suggesting the elimination of that mitochondria (Pickrell & Youle, 2015). PINK1 localized on the mitochondria recruits Parkin to mitochondria from the cytoplasm, followed by the activation of *Parkin* which further results in eliminates the damaged mitochondria (Youle & Narendra 2011). Mutations in PINK1 gene results in the development of the autosomal recessive form of PD (Klein & Westenberger, 2012).

Parkin was the second identified PD gene in context of PD, and the mutation in *Parkin* is responsible for early onset of the autosomal recessive form of PD (Klein & Westenberger, 2012). *Parkin* is an ubiquitin E3 ligase responsible for the degradation of the altered proteins and prevents the cellular apoptosis (Yoshii et al, 2011; Zhang et al, 2000). *Parkin* maintain the proper functioning of mitochondria and mitochondrial DNA (mtDNA) (Kuroda et al, 2006), inhibits ROS formation (Kuroda et al, 2006; Temme et al., 2009), protects mtDNA from damage from oxidative stress (Rothfuss et al, 2009, Watson et al, 2004), and regulates mitochondrial fission and fusion mechanism (Glauser et al., 2011; Narendra et al., 2008). *PINK1* phosphorylates *Parkin* at Ser65 in the N-terminal UBL domain followed by the activation of the *Parkin* (Truban et al, 2016) which then degrades the accumulated misfolded proteins and thus prevents the cell apoptosis (Yoshii et al, 2011; Zhang et al, 2001). *PINK1* phosphorylates the mitofusin 2, for the binding of the *Parkin* to the mitochondria (Pickrell & Youle, 2015). *Parkin* thus acts as a protective agent. Thus *Parkin* mediated ubiquitination serve as signal for the degradation of ubiquitinated substance by the ubiquitin proteasome system, suggesting the role of *Parkin* in the formation of inclusion and autophagic clearance of targeted proteins (Dawson & Dawson, 2010). It is reported that the mutation in the gene encodes for *Parkin* accounts for the development of the PD pathology(Miklya et al,

2014; West et al, 2007), α -synuclein aggregation mediated oxidative stress (Bendor et al. 2013; Breydo et al, 2012; Fujiwara et al, 2002; Giasson et al, 2000), structural changes in mitochondria (Botella et al, 2008) reduced complex-I activity of ETC and ROS formation (Devi et al, 2008; Parihar et al, 2008, 2009) increase mitochondrial membrane permeability responsible for the leakage of pro-apoptotic molecules (Bandopadhyay & de Belleroche, 2010) resulting in neuronal cell death.

DJ-1 is a predominantly cytosolic, homodimeric protein, ubiquitously expressed in both brain and provides the neuroprotection by protecting cells against reactive oxygen species (ROS) (Björkblom et al, 2013). *DJ-1* contains 3 cysteine residues of which only C106 is highly susceptible to oxidative damage and it is reported that the alterations in the normal functioning of this residue led to the loss of the DJ1 function completely. *DJ-1* quenches ROS (Ariga et al, 2013) inhibit p53 and stimulates SOD expression (Ariga et al, 2013). Further the activity of the DJ1 decreases with the increase in the age suggesting the risk of the PD increases with the increase in age. Further the *DJ-1* mutations are responsible for the development of autosomal recessive form of PD but found in rare cases (Klein & Westenberger, 2012).

LRRK2 belongs to the Roco family proteins (Bosgraaf & Van Haastert, 2003; Marín et al, 2008; van Egmond & van Haastert, 2010) consisting of 2 domains: Roc and COR domains. Roc domain is essential for the kinase activity (Ito et al, 2007; Xiong et al, 2010; Taymans et al, 2011) while COR domain is essential for the dimerization (Gotthardt et al, 2008). Roc domain of LRRK2 belongs to family of small G-proteins which become activated when GTP is bound and becomes inactive when GDP is bound (Vetter & Wittinghofer, 2001). LRRK2 in monomer form is inactivated, dimerization led to the activation (Berger et al, 2010; Greggio et al, 2008; James et al, 2012; Schapansky et al, 2014; Webber et al, 2011). Dimerization led to the activation of GTPase reaction (Gotthardt et al, 2008; Sen et al, 2009) required for the normal biological functioning of the Roco proteins (Webber et al, 2011). In the neurons LRRK2 colocalizes with Dynamin like protein 1 (DLP1) and its expression induced mitochondrial fragmentation and increased the level of ROS (Niu et al, 2012; West et al, 2005). LRRK2 is involved in the various cellular functions including vesicular trafficking, proteolysis, regulation of neuritic outgrowth and neuritic morphology (Li et al, 2014). LRRK2 is a positive regulator of murine microglia (Kim et al, 2012) and therefore its inhibition abolished the inflammatory mediators within the murine microglia cells (Kim et al, 2012; Moehle et al, 2012). LRRK2 is known to regulate NF-KB activity (Gardet et al, 2010) which is further responsible for the induction of TNF mediated extrinsic pathway of the cell death and the inhibition of this pathway might reduce the neuronal death in the PD (Dauer & Przedborski, 2003; Hayley et al, 2004; McCoy et al, 2006). LRRK2 gene mutations results in the onset of autosomal dominant PD in the later ages most frequently (Klein & Westenberger, 2012).

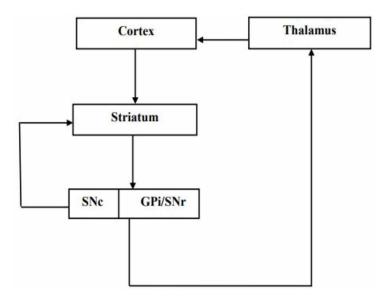
ATP13A2 gene encodes for lysosomal P5-type ATPase and it is suggested that the reduced levels of ATP13A2 or loss-of-function mutations impairs degradation mechanisms due to the inhibition of the lysosomal mediated clearance of the misfolded proteins, resulting in the accumulation of α -synuclein which is responsible for the neurotoxicity (Bento et al, 2016). Mutations of ATP13A2 is further characterized by early onset, rapid progression, dementia, pyramidal signs and supranuclear gaze palsy (Klein & Westenberger, 2012).

NEURONAL DEGENERATION IN PARKINSON'S DISEASE

SN neurons are under oxidative stress due to the metabolism of dopamine (Bender et al., 2006; Kraytsberg et al., 2006) and the metabolism of dopamine in SN generates a large number of oxidative species (Sulzer, 2007) that evokes the oxidative stress in SN. Oxidative stress promotes the misfolding, aggregation of α -synuclein protein to form protofibrils resulting in the neurotoxicity (Lotharius & Brundin, 2002) by mitochondrial dysfunction (Ved et al., 2005), membrane disruption (Conway et al., 2001), and inhibition of protein degradation by lysosomes (Martinez-Vicente et al., 2008). Protofibrils themselves raised level of the dopamine in the cytoplasm by modulating vesicular membranes (Lotharius & Brundin, 2002) and the raised levels of the dopamine further interact with the α -synuclein to exert neurotoxic effects (Caudle et al., 2008; Chen et al., 2008; Edwards, 1993; Pardo et al., 1995; Sulzer & Zecca, 2000). The neuronal cell death in the SN region that primarily gives dopaminergic innervation to the striatum results in the marked deficiency of the dopamine in the striatum responsible for the motor dysfunction in PD (Alexander, 2004). Deficiency of the dopamine then results in the decreased activity of direct pathways and increased activity of indirect pathways of BG (Figure 1 and Figure 2), resulting in difficulty in initiating movement and excessive inhibition of movements (Walia et al, 2014) Further the loss of the dopaminergic cells in the retinal cells is responsible for the visual impairments in PD patients.

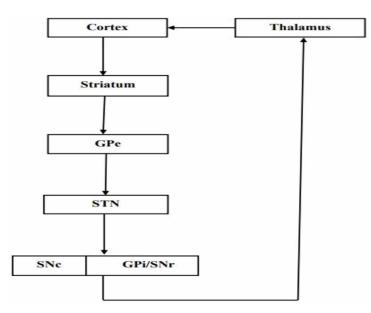
Basal forebrain consists of the cholinergic neurons (Bohnen & Albin, 2011). It is reported that the cholinergic neurons present in the pedunculopontine nucleus (PPN) and nucleus basalis of Meynert (nBM) is responsible for the maintenance of the gait (Rochester et al, 2012). It is suggested that besides SN region neuronal degeneration and accumulation of the LBs (Braak et al, 2003) also found in the PPN and nBM region (Arendt et al, 1983; Bohnen & Albin, 2011; Candy et al, 1983; Chui et al, 1986; Perry et al, 1985) of the brain resulting in the cognitive impairment (Kehagia et al, 2013) gait disturbance (Bohnen et al, 2012; Rochester et al, 2012) dopamine-resistant akinesia (Bohnen & Albin, 2011). Cholinergic

Figure 1. Direct pathway of basal ganglia. Cortex excites striatum which further receives the excitatory input from SNc region, results in the activation of striatum. Striatum sends inhibitory projections to the globus pallidus interna (GPi) and substantia nigra pars reticulate (SNr). GPi/SNr further exerts an inhibitory control on thalamus. Striatum upon excitation by SNc region, inhibits the GPi/SNr mediated inhibition of thalamus and thus facilitates movements. Therefore the damage to the neurons of direct pathway is responsible for the difficulty in initiating movement.



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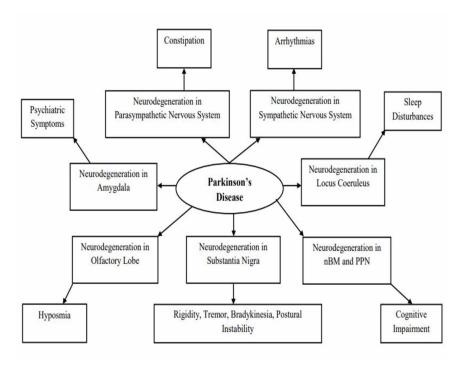
Figure 2. Indirect pathway of basal ganglia. In this pathways cortex send signal to the striatum which then send inhibitory signal to the globus pallidus externa (GPe) and reduced its activity. GPe sends projection to the subthalamic nucleus (STN) to inhibit its activity. STN further sends projections to activate GPi/SNr region resulting in the inhibition of thalamus. The reduced activity of GPe results in the increased activation of STN which is further responsible for the increased activity of GPi/SNr region and excessive inhibition of thalamus. Therefore the greater activation of this area is responsible for the inhibition of movement. In PD the activity of the indirect pathway increases resulting in the excessive inhibition of the movements.



neuronal loss in the PPN region is responsible for REM behavioral disorder in PD and in the nBM is responsible for dementia in PD (Alexander, 2004). Further the neuronal loss in these areas becomes more prominent in the presence of dementia (Candy et al, 1983; Nakano & Hirano, 1984; Rogers et al, 1985; Tagliavini et al, 1984; Whitehouse et al, 1983).

Further the destruction to the noradrenergic neuronal cells of locus coeruleus (LC) resulting in the marked decreased in the level of NA in cerebellum and frontal cortex responsible for the REM behavioral disorder in PD patients (Alexander, 2004). The destruction to the parasympathetic neurons at the preganglionic levels results in difficulty in swallowing in PD patients by the reduced esophageal motility and the destruction to the parasympathetic neurons at the preganglionic levels results in reduced GIT motility and constipation in the patients of PD (Alexander, 2004). Also the neuronal damage in the anterior olfactory nucleus and olfactory bulb is responsible for hyposmia seen in approx. 90% of PD patients (Alexander, 2004). Destruction to the sympathetic neurons at the preganglionic levels results in orthostatic hypotension in PD patients (Alexander, 2004) while at the postganglionic levels results in cardiac dysfunction (Alexander, 2004). BLA is further damaged to the glutamatergic neurons of BLA results in the visual hallucinations in PD patients (Alexander, 2004). Thus the pathogenesis of PD involves the neuronal degeneration in the different regions of the brain giving rise to the complexity and the variety of the symptoms that can be observed in the patients of PD (shown in Figure 3.)

Figure 3. Pathogenesis and symptoms of Parkinson's disease. This disease is accompanied by the neuronal damages in the different region of the brain and the symptom evoke depends on the region in which the damage occurs. Damage to the substantia nigra results in the motor dysfunction, damage to the nucleus basalis Meynert (nBM) and pedunculopontine nucleus (PPN) results in the cognitive impairment, damage in the locus coeruleus and amygdala results in the sleep disturbances and psychiatric symptoms, neuronal damage in the olfactory region is responsible for the loss of olfaction and the damage to the autonomic nervous system i.e. parasympathetic and sympathetic system is responsible for arrhythmia and constipation.



SYMPTOMS OF PARKINSON'S DISEASE

PD symptoms can be of motor and non-motor types. Motor symptoms of the PD includes resting tremor, bradykinesia, rigidity and postural instability (Clarke, 2002). Rest tremor is usually pill rolling type and is least disabling in nature as compared to the other features (DeMaagd & Philip, 2015). Besides rest tremors patients of PD often showed the presence of the various other form of tremor. Resting tremor occurs in 90%, postural tremor occurs in 88-92%, and head tremor occurs in 17% of PD patient (Thenganatt & Louis, 2012). Bradykinesia refers to the slowness of movement, a core motor feature of PD, develops initially or after the tremors (DeMaagd & Philip, 2015) and is the most disabling because of slowness imposed in the daily movements of patients they cannot perform the task quickly (Goldenberg, 2008). Rigidity is defined as the increased resistance offered during movement (Baradaran et al, 2013). Rigidity in PD is described by the term cogwheel rigidity means small jerks occurs while moving passively (DeMaagd & Philip, 2015). All the muscles in body have the opposing muscle such that if a muscle is contracted then its opposing muscles are relaxed. Rigidity occurs when this balance got disturbed. Rigidity can affect different body parts and face and therefore the patients often displayed masked expression

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(DeMaagd & Philip, 2015). Postural instability develops in the latter stages (Santamato et al, 2015) and the postural instability in PD patients increased the risk of fall due to the imbalance (DeMaagd & Philip, 2015). The imbalance is due to the change in the posture of the patients (Goldenberg, 2008). Postural instability in PD predisposed the patients to the injuries and increased the incidence of the fractures (Santamato et al, 2015). PD related postural instability might be due to the lesions in nondopaminergic systems (Santamato et al, 2015).

Non motor symptoms suggested the involvement of extra-nigral pathology in PD patients (Braak et al, 2003) and these include autonomic disturbance, olfactory dysfunctions, psychiatric problems and sleep disturbances. Autonomic disturbance presents the multiple system atrophy (MSA) in PD often responsible for the reduced levels of the dopamine, serotonin and noradrenaline (Pellicano et al, 2007). Olfactory dysfunction occurs due to the death of the neurons in the olfactory region of the brain and is responsible for anosmia and hyposmia in PD patients (Pellicano et al, 2007). Psychiatric symptoms frequently occur in the PD patients, patients often suffer from the anxiety attacks and depression (Pellicano et al, 2007). Sleep disturbance develops due to the damage in the brainstem and thalamocortical pathways in PD. Sleep disturbance is characterized by the REM behavioral disturbances further characterized by the violent activities of the patients (Pellicano et al, 2007).

DIAGNOSIS OF PARKINSON'S DISEASE

PD involves the destruction of the dopaminergic neurons and the presence of LBs in the SN region. There is exactly no definite criteria for the diagnosis of PD. However diagnosis of the clinical PD is based on the presence of the symptoms or their combination. The diagnosis should be done in a careful and systematic manner. PD can be definite, possible or probable, however if the clinical symptoms are present then possible and probable PD should be diagnosed however if the neuropathological evidences are present then definite PD should be diagnosed (Gelb et al, 1999). Inquire should be made about the family history, daily life styles, previous medical history, occupation and environmental factors, followed by the anosmia/hyposmia, REM sleep disruption behavior, constipation, dementia, visual hallucinations and psychiatric symptoms. The clinician should check the eye movement, slowness, posture, and movement of body. Thus the careful assessment of the patient history through the questioning is the first step in the diagnosis of PD to rule out the presence of the familial cases of PD or any drug induced PD symptoms. The next step is the assessment of symptoms, clinicians should check the presence of resting tremor or bradykinesia to check whether the PD is tremor dominant or rigid type. Since PD is different from the parkinsonian symptoms because PD progresses asymmetrically and generally affect one side of the body. The careful examination should be done to rule out the presence of any movement disorders, asthenia and dementia. Different types of the parkinsonism have different distinguishing features for e.g. in case of the vascular parkinsonism (or lower body parkinsonism) lower limbs and the gait is affected and resting tremor are uncommon, in case of the drug induced parkinsonism the sign are symmetrical and tremors are present. It should be noted that the essential tremor and resting tremor are two distinct phenomenon such that the essential tremor rarely occurs at the rest and is not accompanied by the PD symptoms and gait disturbances. Dementia occurs in the patients of PD in late stages of the disease. If the dementia develops before or within one year of the PD diagnosis then it is known as Parkinson's disease dementia, however the diagnosis of the dementia prior to the onset of the PD is known as dementia with LB (Massano & Bhatia, 2012). The damage to structure should be confirmed by imaging studies (Massano & Bhatia, 2012).

PHARMACOTHERAPY OF PARKINSON'S DISEASE

There are currently no standard established regarding, which should be initiate first in the treatment of PD patients. However different drugs should be used either as monotherapy or in combination for the therapeutics of PD patients. For e.g. L-dopa should be used for the treatment of the motor features, sildenafil should be used for the treatment of erectile dysfunction; cholinesterase inhibitors should be used for the treatment of constipation (Massano & Bhatia, 2012). The main aim of the pharmacotherapeutics of the PD patients is to reduce the symptoms and prevent progression of dopaminergic neuronal cell death in PD patients. The selection of the therapeutic agents should be on the basis of diagnosis and patient history to avoid the risk of adverse drug effects.

Dopaminergic Agents

L-dopa was introduced in the 1960s and its introduction revolutionized the treatment of PD (Katzenschlager & Lees, 2002). L-dopa when given cross the BBB and transported inside the brain, where it is metabolized by the enzyme aromatic amino acid decarboxylase into dopamine, which is then uptake and stored by the vesicles and released upon the neuronal depolarization (DeMaagd & Philip, 2015). L-dopa metabolized extensively in the gut and only 30% of the L-dopa reaches the systemic circulation (DeMaagd & Philip, 2015), therefore to avoid this extensive metabolism L-dopa is administered in combination carbidopa (Salat & Tolosa, 2013). The addition of the carbidopa increased the bioavailability and improved the tolerability of L-dopa. L-dopa is used as the first choice drug in the treatment of PD (Katzenschlager & Lees, 2002), however the fluctuations, on-off effect, dyskinesia, and psychiatric problems are frequently associated with the use of L-dopa (Katzenschlager & Lees, 2002; Salat & Tolosa, 2013).

Dopamine Agonists

Dopamine receptor agonists results in the activation of postsynaptic dopaminergic receptors present in the BG (Bonuccelli et al, 2009; Perez-Lloret & Rascol, 2010). Dopamine agonists are used in the symptomatic treatment of the PD and are as effective as L-dopa in the reduction of the motor symptoms of PD (Tintner & Jankovic, 2003). Dopamine agonists should be initiated at lower dose to avoid the incidence of adverse effects (Kieburtz, 2011; Hauser et al, 2010; Giladi et al, 2007; Watts et al, 2007). Instead of these facts L-dopa/carbidopa were considered more effective for controlling the motor symptoms of PD patients than the dopamine agonists (Hayes et al, 2010). Dopamine agonists should be used as monotherapy in the early PD because of their L-dopa sparing effects but used as the adjunctive to L-dopa in the advanced cases of the PD (Tan, 2003). Adverse effect includes the risk of nausea, vomiting, headache, narcolepsy, hallucination, somnolence, valvular heart disease (Borovac, 2016). Dopamine agonists are commonly divided into two groups:

Ergoline-Derivatives

Derived from the ergot and have greater risk of cancer, cardiac valve regurgitation and fibrotic changes. Ergoline derivatives include bromocriptine, pergolide, cabergoline, and lisuride. Bromocriptine shows agonism at D_2 receptors, used as monotherapy and adjunct therapy with L-dopa. Adverse effect includes nausea, vomiting, headache, confusion, hypotension, hallucinations, and delusions (Borovac, 2016). Pergolide shows agonism at D_1 and D_2 receptors and 5-HT₁ and 5-HT₂ receptors and used as the monotherapy for early PD. Adverse effect includes somnolence, sedation, and narcolepsy (Borovac, 2016). Cabergoline shows agonism at D_2 , D_3 , D_4 and 5-HT₂ receptors and antagonism at 5-HT₇ and $\alpha_2 B$ receptors. Adverse effects include nausea, vomiting, dizziness, dyspepsia, postural hypotension, peripheral edema, and narcolepsy (Borovac, 2016). Lisuride shows agonism at D_2 , D_3 , and D_4 receptor agonist. It used as an adjunct to the L-dopa therapy used in the treatment of the PD. Adverse effects include nausea, dry mouth, headache, and postural hypotension (Borovac, 2016).

Non-Ergoline-Derivatives

These dopamine agonists have better safety profile and free from the risk of the cardiac toxicity. Nonergoline derivatives include pramipexole, ropinirole, apomorphine, piribedil. Pramipexole shows agonism at D_2 and D_3 receptors and evokes beneficial effect in the early stages of PD. Adverse effects includes weight gain, constipation, and hallucinations (Borovac, 2016). Ropinirole shows agonism at D_2 , D_3 and D_4 receptors. It is used for the pharmacotherapeutics of the PD patients in the early stages of the PD. Adverse effects include nausea, vomiting, constipation, dizziness, somnolence, confusion, hallucinations, and orthostatic hypotension (Borovac, 2016). Apomorphine is a strong non-ergoline D_1 and D_2 class receptor agonist. It is used for the treatment of the L-dopa induce dyskinesia. Adverse effects includes nausea, vomiting, headache, psychiatric problems, postural instability and injection site reactions (Borovac, 2016). Piribedil shows agonism at D_2 and D_3 receptors and antagonism at α 2 receptors and produce marked vasodilatation. Adverse effect includes syncope and orthostatic hypotension (Borovac, 2016).

Catechol-O-Methyltransferase Inhibitors

Catechol-*O*-methyltransferase (COMT) inhibitors act by extending the duration of action of L-dopa and thus used in conjunction with levodopa (Waters, 2000) and increased its (Antonini et al, 2008). COMT-inhibitors including tolcapone and entacapone reduces the incidence of the wearing off associated with L-dopa/carbidopa therapy in the PD patients (Kurth et al, 1997). Adverse effect includes headache, diz-ziness, mental confusion, nausea and vomiting, dyskinesia, etc (Antonini et al, 2008).

Monoamine Oxidase Inhibitors

Monoamine oxidase (MAO) exists in 2 isoforms. MAO-A presents inside the presynaptic neurons and outside the neurons, accounts for about 20% of MAO activity in BG while MAO-B is localized in the extra-neuronal region (Sieradzan et al, 1995). MAO inhibition inhibits the metabolism of the dopamine and increased the levels of the dopamine (Sieradzan et al, 1995). MAO-A inhibitors i.e. Moclobemide has shown the symptomatic benefit in PD, safe to use and shown efficacy in the patients prescribed with L-dopa/carbidopa. However the major limitation is the incidence of cheese reaction on the consumption

of products rich in tyramine (Youdim & Bakhle, 2006). MAO-B inhibitors i.e. selegiline and rasagiline inhibit the enzyme MAO-B irreversibly, used either as the monotherapy or the adjunctive treatment to L-dopa in the therapeutics of the PD. Adverse effect includes sleeplessness, nausea, vomiting, dizziness, dry mouth, orthostatic hypotension, headache, confusion, and anxiety (Riederer & Laux, 2011).

Anticholinergic Drugs

Anticholinergic drugs are effective in the symptomatic treatments of the resting tremor seen in the patients of PD. These drugs can be used as the monotherapy or as adjunctive to L-dopa or helpful in reducing the dose of L-dopa in the advanced PD (Brocks, 1999). Anticholinergics used in the treatment of the PD are the competitive antagonists of muscarinic receptors and corrects the imbalance between the dopamine and acetylcholine as seen in the patients of PD. Adverse effects includes dryness of mouth, blurred vision urinary retention, blurred vision, tachycardia, mental confusion, constipation, and blurred vision (Brocks, 1999; Goldenberg, 2008)

CHALLENGES IN THE TREATMENT AND RESEARCH IN PARKINSON'S DISEASE

The greatest challenge is to find the reason and the mechanism of the neuronal cell death in the PD which is still unknown. Also the site of the neurodegeneration is not fixed, since multiple site of the neurodegeneration has been investigated though the stress on SN region is given primarily. The late diagnosis of the PD also promotes the progression of the disease and worsening of the symptoms in PD patients. PD is characterized by the presence of the LBs in the brainstem nuclei and the various other regions of the brain. However LBs have been noticed in the brain of the patients showing no clinical symptoms of PD and signs of dopaminergic neuronal degeneration representing a unique condition. Since PD occurs in the late phase of life which is often accompanied by the presence of the some other pathologies, further hampers the treatment of the PD. L-dopa (the main therapeutic agent reduces the motor symptoms in PD patients), itself produce dopamine and dopamine itself produces oxidative stress and exerts neurotoxicity. Further currently there is no drug available that can stop the neurodegeneration in the PD patients.

FUTURE RESEARCH DIRECTIONS

The neuronal cell death in the SN region of the midbrain might be due to the excessive release of glutamate which is known to interfere with the normal functioning of BG (Marino et al, 2003). Glutamate is the well known excitatory neurotransmitter of the CNS, produces its action by binding its receptors (Pettorruso e al, 2014). NMDA receptors have been found to be responsible for the glutamate mediated excitotoxicity and therefore the blockade of these receptors is responsible for the beneficial effect in PD. NMDA antagonists including traxoprodil and ifenprodil inhibit the glutamate mediated excitotoxicity, reduces the motor symptoms, potentiate the effect of L-dopa and reduce L-dopa induced dyskinesia associated with the L-dopa therapy. However the antiviral drug, amantadine exerts beneficial effects in the PD by its antiglutamatergic and NMDA blockade action (Johnson et al, 2009).

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Coenzyme Q10 is a unique molecule, synthesized endogenously, involved in the transfer of the electrons in the mitochondrial electron transport chain and exerts the antioxidant activity. Further CoQ10 provides protection against the ROS mediated damage and abolished oxidative stress mediated neuronal cell death (Mancuso et al, 2009). Thus this unique molecule exerts the neuroprotective effect and therefore might be a good therapeutic option used for the treatment of PD.

Another new target which can be explored in context of PD is the enzyme GSK-3, which is the target of the neurotrophic factors, i.e. BDNF. Inhibition of GSK-3 provides protection against MPTP toxicity. α -synuclein, acts a substrate and activator for GSK-3 and the activation of this enzyme results in the reduced neurogenesis and neuronal damage (Lei et al, 2011). BDNF deficiency might be responsible for the hyperactive GSK3 (Beurel et al, 2015) and therefore GSK3 inhibitors counteracts neuronal loss in PD and GSK-3 inhibitors should be explored in context of PD.

CONCLUSION

The present chapter address about the various aspects including symptoms, diagnosis, treatment and challenges related to PD. However the exact cause of the death of the dopaminergic neurons in the brain is still unknown. Therefore the future work should be focus on the elucidation of the mechanism responsible for the death of dopaminergic neurons in brain. Further there is none of therapeutic agent available today which can prevent the death of dopaminergic neurons in the brain or stop the progression of disease further. Thus future work should focus on these aspects so as to provide better therapeutic options for the treatment of PD.

REFERENCES

Adams, R. D., van Bogaert, L., & Vander Eecken, H. (1964). Striato-nigral degeneration. *Journal of Neuropathology and Experimental Neurology*, 23(4), 584–608. PMID:14219099

Alexander, G. E. (2004). Biology of Parkinson's disease: Pathogenesis and pathophysiology of a multisystem neurodegenerative disorder. *Dialogues in Clinical Neuroscience*, 6(3), 259–280. PMID:22033559

Antonini, A., Abbruzzese, G., Barone, P., Bonuccelli, U., Lopiano, L., Onofrj, M., ... Quattrone, A. (2008). COMT inhibition with tolcapone in the treatment algorithm of patients with Parkinson's disease (PD): Relevance for motor and non-motor features. *Neuropsychiatric Disease and Treatment*, *4*(1), 1–9. doi:10.2147/NDT.S2404 PMID:18728767

Arendt, T., Bigl, V., Arendt, A., & Tennstedt, A. (1983). Loss of neurons in the nucleus basalis of Meynert in Alzheimer's disease, paralysis agitans and Korsakoff's disease. *Acta Neuropathologica*, *61*(2), 101–108. doi:10.1007/BF00697388 PMID:6637393

Ariga, H., Takahashi-Niki, K., Kato, I., Maita, H., Niki, T., & Iguchi-Ariga, S. M. M. (2013). Neuroprotective Function of DJ-1 in Parkinson's Disease. *Oxidative Medicine and Cellular Longevity*, 2013, 683920. doi:10.1155/2013/683920 PMID:23766857 Bandopadhyay, R., & de Belleroche, J. (2010). Pathogenesis of Parkinson's disease: Emerging role of molecular chaperones. *Trends in Molecular Medicine*, *16*(1), 27–36. doi:10.1016/j.molmed.2009.11.004 PMID:20036196

Baradaran, N., Tan, S. N., Liu, A., Ashoori, A., Palmer, S. J., Wang, Z. J., ... McKeown, M. J. (2013). Parkinson's Disease Rigidity: Relation to Brain Connectivity and Motor Performance. *Frontiers in Neurology*, *4*, 67. doi:10.3389/fneur.2013.00067 PMID:23761780

Bender, A., Krishnan, K. J., Morris, C. M., Taylor, G. A., Reeve, A. K., Perry, R. H., ... Taylor, R. W. (2006). High levels of mitochondrial DNA deletions in substantia nigra neurons in aging and Parkinson disease. *Nature Genetics*, *38*(5), 515–517. doi:10.1038/ng1769 PMID:16604074

Bendor, J. T., Logan, T. P., & Edwards, R. H. (2013). The function of alpha-synuclein. *Neuron*, 79(6), 1044–1066. doi:10.1016/j.neuron.2013.09.004 PMID:24050397

Bento, C. F., Ashkenazi, A., Jimenez-Sanchez, M., & Rubinsztein, D. C. (2016). The Parkinson's diseaseassociated genes *ATP13A2* and *SYT11* regulate autophagy via a common pathway. *Nature Communications*, 7, 11803. doi:10.1038/ncomms11803 PMID:27278822

Berger, Z., Smith, K. A., & LaVoie, M. J. (2010). Membrane localization of LRRK2 is associated with increased formation of the highly active LRRK2 dimer and changes in its phosphorylation. *Biochemistry*, *49*(26), 5511–5523. doi:10.1021/bi100157u PMID:20515039

Beurel, E., Grieco, S. F., & Jope, R. S. (2015). Glycogen synthase kinase-3 (GSK3): Regulation, actions, and diseases. *Pharmacology & Therapeutics*, 0, 114–131. doi:10.1016/j.pharmthera.2014.11.016 PMID:25435019

Björkblom, B., Adilbayeva, A., Maple-Grødem, J., Piston, D., Ökvist, M., Xu, X. M., ... Møller, S. G. (2013). Parkinson Disease Protein DJ-1 Binds Metals and Protects against Metal-induced Cytotoxicity. *The Journal of Biological Chemistry*, 288(31), 22809–22820. doi:10.1074/jbc.M113.482091 PMID:23792957

Bohnen, N. I., & Albin, R. L. (2011). The Cholinergic System and Parkinson Disease. *Behavioural Brain Research*, 221(2), 564–573. doi:10.1016/j.bbr.2009.12.048 PMID:20060022

Bohnen, N. I., Müller, M. L. T. M., Kotagal, V., Koeppe, R. A., Kilbourn, M. R., Gilman, S., ... Frey, K. A. (2012). Heterogeneity of cholinergic denervation in Parkinson's disease without dementia. *Journal of Cerebral Blood Flow and Metabolism*, *32*(8), 1609–1617. doi:10.1038/jcbfm.2012.60 PMID:22569194

Bonuccelli, U., Del Dotto, P., & Rascol, O. (2009). Role of dopamine receptor agonists in the treatment of early Parkinson's disease. *Parkinsonism & Related Disorders*, *15*, S44–S53. doi:10.1016/S1353-8020(09)70835-1 PMID:20123557

Borovac, J. A. (2016). Side effects of a dopamine agonist therapy for Parkinson's disease: A mini-review of clinical pharmacology. *The Yale Journal of Biology and Medicine*, 89(1), 37–47. PMID:27505015

Bosgraaf, L., & Van Haastert, P. J. (2003). Roc, a Ras/GTPase domain in complex proteins. *Biochimica et Biophysica Acta (BBA)-. Molecular Cell Research*, *1643*(1), 5–10.

Parkinson's Disease

Botella, J. A., Bayersdorfer, F., & Schneuwly, S. (2008). Superoxide dismutase overexpression protects dopaminergic neurons in a Drosophila model of Parkinson's disease. *Neurobiology of Disease*, *30*(1), 65–73. doi:10.1016/j.nbd.2007.11.013 PMID:18243716

Braak, H., Del Tredici, K., Rüb, U., de Vos, R. A., Steur, E. N. J., & Braak, E. (2003). Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiology of Aging*, *24*(2), 197–211. doi:10.1016/S0197-4580(02)00065-9 PMID:12498954

Breydo, L., Wu, J. W., & Uversky, V. N. (2012). α-Synuclein misfolding and Parkinson's disease. *Biochimica et Biophysica Acta (BBA)-. Molecular Basis of Disease*, 1822(2), 261–285. doi:10.1016/j. bbadis.2011.10.002

Brissaud, É. (1895). Leçons sur les maladies nerveuses: Salpêtrière, 1893-1894 (Vol. 1). G. Masson.

Brocks, D. R. (1999). Anticholinergic drugs used in Parkinson's disease: An overlooked class of drugs from a pharmacokinetic perspective. *Journal of Pharmacy & Pharmaceutical Sciences*, 2(2), 39–46. PMID:10952768

Campêlo, C. L., Cagni, F. C., de Siqueira Figueredo, D., Oliveira Jr, L. G., Silva-Neto, A. B., Macêdo, P. T., ... de Oliveira Godeiro Jr, C. (2017). Variants in SNCA Gene Are Associated with Parkinson's Disease Risk and Cognitive Symptoms in a Brazilian Sample. *Frontiers in Aging Neuroscience*, 9. PMID:28676755

Candy, J., Perry, R. H., Perry, E. K., Irving, D., Blessed, G., Fairbairn, A. F., & Tomlinson, B. E. (1983). Pathological changes in the nucleus of Meynert in Alzheimer's and Parkinson's diseases. *Journal of the Neurological Sciences*, *59*(2), 277–289. doi:10.1016/0022-510X(83)90045-X PMID:6854353

Caudle, W. M., Colebrooke, R. E., Emson, P. C., & Miller, G. W. (2008). Altered vesicular dopamine storage in Parkinson's disease: A premature demise. *Trends in Neurosciences*, *31*(6), 303–308. doi:10.1016/j. tins.2008.02.010 PMID:18471904

Charcot, J. M. (1872). Cinquième Leçon. De la paralysie agitante. *Oeuvres completes, Recueillies et publiées par Bourneville. Bureaux du Progrès Médical, Paris, France, 1*, 155–189.

Chen, L., Ding, Y., Cagniard, B., Van Laar, A. D., Mortimer, A., Chi, W., ... Zhuang, X. (2008). Unregulated cytosolic dopamine causes neurodegeneration associated with oxidative stress in mice. *The Journal of Neuroscience*, 28(2), 425–433. doi:10.1523/JNEUROSCI.3602-07.2008 PMID:18184785

Chui, H. C., Mortimer, J. A., Slager, U., Zarow, C., Bondareff, W., & Webster, D. D. (1986). Pathologic correlates of dementia in Parkinson's disease. *Archives of Neurology*, *43*(10), 991–995. doi:10.1001/ archneur.1986.00520100013007 PMID:3753274

Clarke, C. E. (2002). Medical management of Parkinson's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 72(suppl 1), i22–i27. doi:10.1136/jnnp.72.suppl_1.i22 PMID:11870200

Conway, K. A., Rochet, J. C., Bieganski, R. M., & Lansbury, P. T. (2001). Kinetic stabilization of the α -synuclein protofibril by a dopamine- α -synuclein adduct. *Science*, 294(5545), 1346–1349. doi:10.1126cience.1063522 PMID:11701929

Dauer, W., & Przedborski, S. (2003). Parkinson's Disease, Mechanisms and Models. *Neuron*, 39(6), 889–909. doi:10.1016/S0896-6273(03)00568-3 PMID:12971891

Dawson, T. M., & Dawson, V. L. (2010). The Role of Parkin in Familial and Sporadic Parkinson's Disease. *Movement Disorders: Official Journal of the Movement Disorder Society*, 25(1), S32-S39.

DeMaagd, G., & Philip, A. (2015). Parkinson's Disease and Its Management: Part 1: Disease Entity, Risk Factors, Pathophysiology, Clinical Presentation, and Diagnosis. *P&T*, 40(8), 504–532. PMID:26236139

Devi, L., Raghavendran, V., Prabhu, B. M., Avadhani, N. G., & Anandatheerthavarada, H. K. (2008). Mitochondrial import and accumulation of α -synuclein impair complex I in human dopaminergic neuronal cultures and Parkinson disease brain. *The Journal of Biological Chemistry*, 283(14), 9089–9100. doi:10.1074/jbc.M710012200 PMID:18245082

Edwards, R. H. (1993). Neural degeneration and the transport of neurotransmitters. *Annals of Neurology*, *34*(5), 638–645. doi:10.1002/ana.410340504 PMID:7902065

Fahn, S. (2003). Description of Parkinson's disease as a clinical syndrome. *Annals of the New York Academy of Sciences*, 991(1), 1–14. doi:10.1111/j.1749-6632.2003.tb07458.x PMID:12846969

Farrer, M. J. (2006). Genetics of Parkinson disease: Paradigm shifts and future prospects. *Nature Reviews*. *Genetics*, 7(4), 306–318. doi:10.1038/nrg1831 PMID:16543934

Foix, C., & Nicolesco, J. (1925). Les noyaux gris centraux et la region mesencephalo-sous-optique: suivi d'un appendice sur l'anatomie pathologique de la maladie de Parkinson. Masson.

Forno, L. S. (1996). Neuropathology of Parkinson's disease. *Journal of Neuropathology and Experimental Neurology*, *55*(3), 259–272. doi:10.1097/00005072-199603000-00001 PMID:8786384

Fujiwara, H., Hasegawa, M., Dohmae, N., Kawashima, A., Masliah, E., Goldberg, M. S., ... Iwatsubo, T. (2002). [alpha]-Synuclein is phosphorylated in synucleinopathy lesions. *Nature Cell Biology*, *4*(2), 160–164. doi:10.1038/ncb748 PMID:11813001

Gardet, A., Benita, Y., Li, C., Sands, B. E., Ballester, I., Stevens, C., ... Podolsky, D. K. (2010). LRRK2 is involved in the IFN-γ response and host response to pathogens. *Journal of Immunology (Baltimore, Md.: 1950)*, *185*(9), 5577–5585. doi:10.4049/jimmunol.1000548 PMID:20921534

Gelb, D. J., Oliver, E., & Gilman, S. (1999). Diagnostic criteria for Parkinson disease. *Archives of Neurology*, *56*(1), 33–39. doi:10.1001/archneur.56.1.33 PMID:9923759

Giasson, B. I., Duda, J. E., Murray, I. V., Chen, Q., Souza, J. M., Hurtig, H. I., ... Lee, V. M. Y. (2000). Oxidative damage linked to neurodegeneration by selective α -synuclein nitration in synucleinopathy lesions. *Science*, 290(5493), 985–989. doi:10.1126cience.290.5493.985 PMID:11062131

Giladi, N., Boroojerdi, B., Korczyn, A. D., Burn, D. J., Clarke, C. E., & Schapira, A. H. (2007). Rotigotine transdermal patch in early Parkinson's disease: A randomized, double-blind, controlled study versus placebo and ropinirole. *Movement Disorders*, 22(16), 2398–2404. doi:10.1002/mds.21741 PMID:17935234

Parkinson's Disease

Glauser, L., Sonnay, S., Stafa, K., & Moore, D. J. (2011). Parkin promotes the ubiquitination and degradation of the mitochondrial fusion factor mitofusin 1. *Journal of Neurochemistry*, *118*(4), 636–645. doi:10.1111/j.1471-4159.2011.07318.x PMID:21615408

Goldenberg, M. M. (2008). Medical Management of Parkinson's Disease. *P&T*, 33(10), 590–606. PMID:19750042

Gotthardt, K., Weyand, M., Kortholt, A., Van Haastert, P. J., & Wittinghofer, A. (2008). Structure of the Roc–COR domain tandem of C. tepidum, a prokaryotic homologue of the human LRRK2 Parkinson kinase. *The EMBO Journal*, *27*(16), 2239–2249. doi:10.1038/emboj.2008.150 PMID:18650931

Gowers, W. R. (1898). A manual of diseases of the nervous system (Vol. 2). P. Blakiston, Son & Company.

Greenfield, J. G., & Bosanquet, F. D. (1953). The brain-stem lesions in Parkinsonism. *Journal of Neurology, Neurosurgery, and Psychiatry*, *16*(4), 213–226. doi:10.1136/jnnp.16.4.213 PMID:13109537

Greggio, E., Zambrano, I., Kaganovich, A., Beilina, A., Taymans, J. M., Daniëls, V., ... Thomas, K. J. (2008). The Parkinson disease-associated leucine-rich repeat kinase 2 (LRRK2) is a dimer that undergoes intramolecular autophosphorylation. *The Journal of Biological Chemistry*, 283(24), 16906–16914. doi:10.1074/jbc.M708718200 PMID:18397888

Hauser, R. A., Schapira, A. H., Rascol, O., Barone, P., Mizuno, Y., Salin, L., ... Poewe, W. (2010). Randomized, double-blind, multicenter evaluation of pramipexole extended release once daily in early Parkinson's disease. *Movement Disorders*, *25*(15), 2542–2549. doi:10.1002/mds.23317 PMID:20669317

Hayes, M. W., Fung, V. S., Kimber, T. E., & O'Sullivan, J. D. (2010). Current concepts in the management of Parkinson disease. *The Medical Journal of Australia*, *192*(3), 144–149. PMID:20121682

Hayley, S., Crocker, S. J., Smith, P. D., Shree, T., Jackson-Lewis, V., Przedborski, S., ... Park, D. S. (2004). Regulation of dopaminergic loss by Fas in a 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine model of Parkinson's disease. *The Journal of Neuroscience*, *24*(8), 2045–2053. doi:10.1523/JNEURO-SCI.4564-03.2004 PMID:14985447

Hoehn, M. M., & Yahr, M. D. (1967). Parkinsonism onset, progression, and mortality. *Neurology*, *17*(5), 427–427. doi:10.1212/WNL.17.5.427 PMID:6067254

Hornykiewicz, O., & Kish, S. J. (1987). Biochemical pathophysiology of Parkinson's disease. *Advances in Neurology*, *45*, 19–34. PMID:2881444

Ito, G., Okai, T., Fujino, G. O., Takeda, K., Ichijo, H., Katada, T., & Iwatsubo, T. (2007). GTP binding is essential to the protein kinase activity of LRRK2, a causative gene product for familial Parkinson's disease. *Biochemistry*, *46*(5), 1380–1388. doi:10.1021/bi061960m PMID:17260967

James, N. G., Digman, M. A., Gratton, E., Barylko, B., Ding, X., Albanesi, J. P., ... Jameson, D. M. (2012). Number and brightness analysis of LRRK2 oligomerization in live cells. *Biophysical Journal*, *102*(11), L41–L43. doi:10.1016/j.bpj.2012.04.046 PMID:22713584

Johnson, K. A., Conn, P. J., & Niswender, C. M. (2009). Glutamate receptors as therapeutic targets for Parkinson's disease. *CNS & Neurological Disorders - Drug Targets*, 8(6), 475–491. doi:10.2174/187152709789824606 PMID:19702565 Katzenschlager, R., & Lees, A. J. (2002). Treatment of Parkinson's disease: Levodopa as the first choice. *Journal of Neurology*, 249. PMID:12375059

Kehagia, A. A., Barker, R. A., & Robbins, T. W. (2013). Cognitive impairment in Parkinson's disease: The dual syndrome hypothesis. *Neurodegenerative Diseases*, *11*(2), 79–92. doi:10.1159/000341998 PMID:23038420

Kieburtz, K. (2011). Twice-daily, low-dose pramipexole in early Parkinson's disease: A randomized, placebo-controlled trial. *Movement Disorders*, 26(1), 37–44. doi:10.1002/mds.23396 PMID:20925067

Kim, B., Yang, M. S., Choi, D., Kim, J. H., Kim, H. S., Seol, W., ... Joe, E. H. (2012). Impaired inflammatory responses in murine Lrrk2-knockdown brain microglia. *PLoS One*, *7*(4), e34693. doi:10.1371/ journal.pone.0034693 PMID:22496842

Klein, C., & Westenberger, A. (2012). Genetics of Parkinson's Disease. *Cold Spring Harbor Perspectives in Medicine*, 2(1), a008888. doi:10.1101/cshperspect.a008888 PMID:22315721

Kraytsberg, Y., Kudryavtseva, E., McKee, A. C., Geula, C., Kowall, N. W., & Khrapko, K. (2006). Mitochondrial DNA deletions are abundant and cause functional impairment in aged human substantia nigra neurons. *Nature Genetics*, *38*(5), 518–520. doi:10.1038/ng1778 PMID:16604072

Kuroda, Y., Mitsui, T., Kunishige, M., Shono, M., Akaike, M., Azuma, H., & Matsumoto, T. (2006). Parkin enhances mitochondrial biogenesis in proliferating cells. *Human Molecular Genetics*, *15*(6), 883–895. doi:10.1093/hmg/ddl006 PMID:16449237

Kurth, M. C., Adler, C. H., Hilaire, M. S., Singer, C., Waters, C., LeWitt, P., ... Yoo, K. (1997). Tolcapone Improves Motor Function and Reduces Levodopa Requirement in Patients with Parkinson's Disease Experiencing Motor Fluctuations A Multicenter, Double-Blind, Randomized, Placebo-Controlled Trial. *Neurology*, *48*(1), 81–87. doi:10.1212/WNL.48.1.81 PMID:9008498

Langston, J. W., Ballard, P., Tetrud, J. W., & Irwin, I. (1983). Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. *Science*, *219*(4587), 979–980. doi:10.1126cience.6823561 PMID:6823561

Lei, P., Ayton, S., Bush, A. I., & Adlard, P. A. (2011). GSK-3 in Neurodegenerative Diseases. *International Journal of Alzheimer's Disease*, 2011, 189246. doi:10.4061/2011/189246 PMID:21629738

Li, J.-Q., Tan, L., & Yu, J.-T. (2014). The role of the LRRK2 gene in Parkinsonism. *Molecular Neuro*degeneration, 9(1), 47. doi:10.1186/1750-1326-9-47 PMID:25391693

Lotharius, J., & Brundin, P. (2002). Pathogenesis of parkinson's disease: Dopamine, vesicles and [alpha]synuclein. *Nature Reviews. Neuroscience*, *3*(12), 932–942. doi:10.1038/nrn983 PMID:12461550

Mancuso, M., Orsucci, D., Calsolaro, V., Choub, A., & Siciliano, G. (2009). Coenzyme Q10 and Neurological Diseases. *Pharmaceuticals*, 2(3), 134–149. doi:10.3390/ph203134 PMID:27713230

Marín, I., van Egmond, W. N., & van Haastert, P. J. (2008). The Roco protein family: A functional perspective. *The FASEB Journal*, 22(9), 3103–3110. doi:10.1096/fj.08-111310 PMID:18523161

Parkinson's Disease

Marino, M. J., Valenti, O., & Conn, P. J. (2003). Glutamate receptors and Parkinson's disease. *Drugs & Aging*, 20(5), 377–397. doi:10.2165/00002512-200320050-00006 PMID:12696997

Martinez-Vicente, M., Talloczy, Z., Kaushik, S., Massey, A. C., Mazzulli, J., Mosharov, E. V., ... Dauer, W. (2008). Dopamine-modified α-synuclein blocks chaperone-mediated autophagy. *The Journal of Clinical Investigation*, *118*(2), 777. PMID:18172548

Massano, J., & Bhatia, K. P. (2012). Clinical approach to Parkinson's disease: Features, diagnosis, and principles of management. *Cold Spring Harbor Perspectives in Medicine*, 2(6), a008870. doi:10.1101/ cshperspect.a008870 PMID:22675666

McCoy, M. K., Martinez, T. N., Ruhn, K. A., Szymkowski, D. E., Smith, C. G., Botterman, B. R., ... Tansey, M. G. (2006). Blocking soluble tumor necrosis factor signaling with dominant-negative tumor necrosis factor inhibitor attenuates loss of dopaminergic neurons in models of Parkinson's disease. *The Journal of Neuroscience*, *26*(37), 9365–9375. doi:10.1523/JNEUROSCI.1504-06.2006 PMID:16971520

Miklya, I., Göltl, P., Hafenscher, F., & Pencz, N. (2014). The role of parkin in Parkinson's disease. *Neuropsychopharmacologia Hungarica: a Magyar Pszichofarmakologiai Egyesulet lapja= official journal of the Hungarian Association of Psychopharmacology, 16*(2), 67-76.

Mizuno, Y., Hattori, N., Kitada, T., Matsumine, H., Mori, H., Shimura, H., ... Shimizu, N. (2000). Familial Parkinson's disease. Alpha-synuclein and parkin. *Advances in Neurology*, *86*, 13–21. PMID:11553970

Moehle, M. S., Webber, P. J., Tse, T., Sukar, N., Standaert, D. G., DeSilva, T. M., ... West, A. B. (2012). LRRK2 inhibition attenuates microglial inflammatory responses. *The Journal of Neuroscience*, *32*(5), 1602–1611. doi:10.1523/JNEUROSCI.5601-11.2012 PMID:22302802

Moghal, S., Rajput, A. H., D'Arcy, C., & Rajput, R. (1994). Prevalence of movement disorders in elderly community residents. *Neuroepidemiology*, *13*(4), 175–178. doi:10.1159/000110376 PMID:8090259

Nakano, I., & Hirano, A. (1984). Parkinson's disease: Neuron loss in the nucleus basalis without concomitant Alzheimer's disease. *Annals of Neurology*, *15*(5), 415–418. doi:10.1002/ana.410150503 PMID:6732189

Narendra, D., Tanaka, A., Suen, D. F., & Youle, R. J. (2008). Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. *The Journal of Cell Biology*, *183*(5), 795–803. doi:10.1083/ jcb.200809125 PMID:19029340

Niu, J., Yu, M., Wang, C., & Xu, Z. (2012). Leucine-rich repeat kinase 2 disturbs mitochondrial dynamics via Dynamin-like protein. *Journal of Neurochemistry*, *122*(3), 650–658. doi:10.1111/j.1471-4159.2012.07809.x PMID:22639965

Oczkowska, A., Kozubski, W., Lianeri, M., & Dorszewska, J. (2013). Mutations in PRKN and SNCA Genes Important for the Progress of Parkinson's Disease. *Current Genomics*, *14*(8), 502–517. doi:10.2 174/1389202914666131210205839 PMID:24532983

Olanow, C. W., & Tatton, W. G. (1999). Etiology and pathogenesis of Parkinson's disease. *Annual Review of Neuroscience*, 22(1), 123–144. doi:10.1146/annurev.neuro.22.1.123 PMID:10202534

Pardo, B., Mena, M. A., Casarejos, M. J., Paino, C. L., & De Yébenes, J. G. (1995). Toxic effects of L-DOPA on mesencephalic cell cultures: Protection with antioxidants. *Brain Research*, 682(1), 133–143. doi:10.1016/0006-8993(95)00341-M PMID:7552304

Parihar, M. S., Parihar, A., Fujita, M., Hashimoto, M., & Ghafourifar, P. (2008). Mitochondrial association of alpha-synuclein causes oxidative stress. *Cellular and Molecular Life Sciences*, 65(7), 1272–1284. doi:10.100700018-008-7589-1 PMID:18322646

Parihar, M. S., Parihar, A., Fujita, M., Hashimoto, M., & Ghafourifar, P. (2009). Alpha-synuclein overexpression and aggregation exacerbates impairment of mitochondrial functions by augmenting oxidative stress in human neuroblastoma cells. *The International Journal of Biochemistry & Cell Biology*, *41*(10), 2015–2024. doi:10.1016/j.biocel.2009.05.008 PMID:19460457

Parkinson, J. (1817). An Essay on the Shaking Palsy. London: Whittingham and Rowland.

Pellicano, C., Benincasa, D., Pisani, V., Buttarelli, F. R., Giovannelli, M., & Pontieri, F. E. (2007). Prodromal non-motor symptoms of Parkinson's disease. *Neuropsychiatric Disease and Treatment*, *3*(1), 145–152. doi:10.2147/nedt.2007.3.1.145 PMID:19300544

Perez-Lloret, S., & Rascol, O. (2010). Dopamine receptor agonists for the treatment of early or advanced Parkinson's disease. *CNS Drugs*, 24(11), 941–968. doi:10.2165/11537810-000000000-00000 PMID:20932066

Perry, E. K., Curtis, M., Dick, D. J., Candy, J. M., Atack, J. R., Bloxham, C. A., ... Perry, R. H. (1985). Cholinergic correlates of cognitive impairment in Parkinson's disease: Comparisons with Alzheimer's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 48(5), 413–421. doi:10.1136/jnnp.48.5.413 PMID:3998751

Pettorruso, M., De Risio, L., Martinotti, G., Di Nicola, M., Ruggeri, F., & Conte, G. (2014). ... Janiri, L. (2014). Targeting the Glutamatergic System to Treat Pathological Gambling: Current Evidence and Future Perspectives. *BioMed Research International*. PMID:25013755

Pickrell, A. M., & Youle, R. J. (2015). The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease. *Neuron*, 85(2), 257–273. doi:10.1016/j.neuron.2014.12.007 PMID:25611507

Richer, P., & Meige, H. (1895). Etude morphologique sur la maladie de Parkinson. *Nouvelle iconographie de la Salpêtriere*, *8*, 361-371.

Riederer, P., & Laux, G. (2011). MAO-inhibitors in Parkinson's Disease. *Experimental Neurobiology*, 20(1), 1–17. doi:10.5607/en.2011.20.1.1 PMID:22110357

Rochester, L., Yarnall, A. J., Baker, M. R., David, R. V., Lord, S., Galna, B., & Burn, D. J. (2012). Cholinergic dysfunction contributes to gait disturbance in early Parkinson's disease. *Brain*, *135*(9), 2779–2788. doi:10.1093/brain/aws207 PMID:22961550

Rogers, J. D., Brogan, D., & Mirra, S. S. (1985). The nucleus basalis of Meynert in neurological disease: A quantitative morphological study. *Annals of Neurology*, *17*(2), 163–170. doi:10.1002/ana.410170210 PMID:3883886

Parkinson's Disease

Rothfuss, O., Fischer, H., Hasegawa, T., Maisel, M., Leitner, P., Miesel, F., ... Patenge, N. (2009). Parkin protects mitochondrial genome integrity and supports mitochondrial DNA repair. *Human Molecular Genetics*, *18*(20), 3832–3850. doi:10.1093/hmg/ddp327 PMID:19617636

Salat, D., & Tolosa, E. (2013). Levodopa in the treatment of Parkinson's disease: Current status and new developments. *Journal of Parkinson's Disease*, *3*(3), 255–269. PMID:23948989

Santamato, A., Ranieri, M., Cinone, N., Stuppiello, L. A., Valeno, G., & De Sanctis, J. L. (2015). ... Panza, F. (2015). Postural and Balance Disorders in Patients with Parkinson's Disease: A Prospective Open-Label Feasibility Study with Two Months of Action Observation Treatment. *Parkinson's Disease*. PMID:26798551

Schapansky, J., Nardozzi, J. D., Felizia, F., & LaVoie, M. J. (2014). Membrane recruitment of endogenous LRRK2 precedes its potent regulation of autophagy. *Human Molecular Genetics*, 23(16), 4201–4214. doi:10.1093/hmg/ddu138 PMID:24682598

Sen, S., Webber, P. J., & West, A. B. (2009). Dependence of leucine-rich repeat kinase 2 (LRRK2) kinase activity on dimerization. *The Journal of Biological Chemistry*, 284(52), 36346–36356. doi:10.1074/jbc. M109.025437 PMID:19826009

Sieradzan, K., Channon, S., Ramponi, C., Stern, G. M., Lees, A. J., & Youdim, M. B. (1995). The therapeutic potential of moclobemide, a reversible selective monoamine oxidase A inhibitor in Parkinson's disease. *Journal of Clinical Psychopharmacology*, *15*(4), 51S–59S. doi:10.1097/00004714-199508001-00010 PMID:7593732

Sulzer, D. (2007). Multiple hit hypotheses for dopamine neuron loss in Parkinson's disease. *Trends in Neurosciences*, *30*(5), 244–250. doi:10.1016/j.tins.2007.03.009 PMID:17418429

Sulzer, D., & Zecca, L. (1999). Intraneuronal dopamine-quinone synthesis: A review. *Neurotoxicity Research*, *1*(3), 181–195. doi:10.1007/BF03033289 PMID:12835101

Tagliavini, F., Pilleri, G., Bouras, C., & Constantinidis, J. (1984). The basal nucleus of Meynert in idiopathic Parkinson's disease. *Acta Neurologica Scandinavica*, 70(1), 20–28. doi:10.1111/j.1600-0404.1984. tb00798.x PMID:6475484

Tan, E. K. (2003). Dopamine agonists and their role in Parkinson's disease treatment. *Expert Review of Neurotherapeutics*, *3*(6), 805–810. doi:10.1586/14737175.3.6.805 PMID:19810883

Tanner, C. M. (2003). Is the cause of Parkinson's disease environmental or hereditary? Evidence from twin studies. *Advances in Neurology*, *91*, 133. PMID:12442672

Taymans, J. M., Vancraenenbroeck, R., Ollikainen, P., Beilina, A., Lobbestael, E., De Maeyer, M., ... Cookson, M. R. (2011). LRRK2 kinase activity is dependent on LRRK2 GTP binding capacity but independent of LRRK2 GTP binding. *PLoS One*, *6*(8), e23207. doi:10.1371/journal.pone.0023207 PMID:21858031

Temme, C., Weissbach, R., Lilie, H., Wilson, C., Meinhart, A., Meyer, S., ... Wahle, E. (2009). The Drosophila melanogaster gene cg4930 encodes a high affinity inhibitor for endonuclease G. *The Journal of Biological Chemistry*, 284(13), 8337–8348. doi:10.1074/jbc.M808319200 PMID:19129189

Thenganatt, M. A., & Louis, E. D. (2012). Distinguishing essential tremor from Parkinson's disease: Bedside tests and laboratory evaluations. *Expert Review of Neurotherapeutics*, *12*(6), 687–696. doi:10.1586/ ern.12.49 PMID:22650171

Tintner, R., & Jankovic, J. (2003). Dopamine agonists in Parkinson's disease. *Expert Opinion on Investigational Drugs*, *12*(11), 1803–1820. doi:10.1517/13543784.12.11.1803 PMID:14585056

Tretiakoff, C. (1919). Contribution à l'étude de l'anatomie du locus niger de Soemmering avec quelques déductions relatives à la pathogenie des troubles du tonus musculaire et de la maladie de Parkinson. Paris: Thèse de.

Truban, D., Hou, X., Caulfield, T. R., Fiesel, F. C., & Springer, W. (2016). PINK1, Parkin, and Mitochondrial Quality Control: What can we Learn about Parkinson's Disease Pathobiology? *Journal of Parkinson's Disease*, 7(1), 13–29. doi:10.3233/JPD-160989 PMID:27911343

Van Den Eeden, S. K., Tanner, C. M., Bernstein, A. L., Fross, R. D., Leimpeter, A., Bloch, D. A., & Nelson, L. M. (2003). Incidence of Parkinson's disease: Variation by age, gender, and race/ethnicity. *American Journal of Epidemiology*, *157*(11), 1015–1022. doi:10.1093/aje/kwg068 PMID:12777365

Van Egmond, W. N., & van Haastert, P. J. (2010). Characterization of the Roco protein family in Dictyostelium discoideum. *Eukaryotic Cell*, 9(5), 751–761. doi:10.1128/EC.00366-09 PMID:20348387

Ved, R., Saha, S., Westlund, B., Perier, C., Burnam, L., Sluder, A., ... Liu, L. (2005). Similar patterns of mitochondrial vulnerability and rescue induced by genetic modification of α-synuclein, parkin, and DJ-1 in Caenorhabditis elegans. *The Journal of Biological Chemistry*, 280(52), 42655–42668. doi:10.1074/ jbc.M505910200 PMID:16239214

Vetter, I. R., & Wittinghofer, A. (2001). The guanine nucleotide-binding switch in three dimensions. *Science*, 294(5545), 1299–1304. doi:10.1126cience.1062023 PMID:11701921

Walia, V., Sharma, A., Gahlawat, M., & Dube, O. P. (2014). Dual Role of Nitric Oxide in the Pathogenesis of Parkinson's Disease. *Journal of Pharmaceutical Sciences & Pharmacology*, 1(4), 243–253. doi:10.1166/jpsp.2014.1038

Waters, C. (2000). Catechol-O-Methyltransferase (COMT) Inhibitors in Parkinson's Disease. *Journal of the American Geriatrics Society*, 48(6), 692–698. doi:10.1111/j.1532-5415.2000.tb04732.x PMID:10855610

Watson, R. E., McKim, J. M., Cockerell, G. L., & Goodman, J. I. (2004). The value of DNA methylation analysis in basic, initial toxicity assessments. *Toxicological Sciences*, 79(1), 178–188. doi:10.1093/ toxsci/kfh099 PMID:15103049

Watts, R. L., Jankovic, J., Waters, C., Rajput, A., Boroojerdi, B., & Rao, J. (2007). Randomized, blind, controlled trial of transdermal rotigotine in early Parkinson disease. *Neurology*, *68*(4), 272–276. doi:10.1212/01.wnl.0000252355.79284.22 PMID:17202432

Webber, P. J., Smith, A. D., Sen, S., Renfrow, M. B., Mobley, J. A., & West, A. B. (2011). Autophosphorylation in the leucine-rich repeat kinase 2 (LRRK2) GTPase domain modifies kinase and GTP-binding activities. *Journal of Molecular Biology*, *412*(1), 94–110. doi:10.1016/j.jmb.2011.07.033 PMID:21806997

Parkinson's Disease

West, A. B., Dawson, V. L., & Dawson, T. M. (2007). The role of Parkin in Parkinson's disease. *Neurological Disease & Therapy*, 83, 199.

West, A. B., Moore, D. J., Biskup, S., Bugayenko, A., Smith, W. W., Ross, C. A., ... Dawson, T. M. (2005). Parkinson's disease-associated mutations in leucine-rich repeat kinase 2 augment kinase activity. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(46), 16842–16847. doi:10.1073/pnas.0507360102 PMID:16269541

Whitehouse, P. J., Hedreen, J. C., White, C. L., & Price, D. L. (1983). Basal forebrain neurons in the dementia of Parkinson disease. *Annals of Neurology*, *13*(3), 243–248. doi:10.1002/ana.410130304 PMID:6847136

Xiong, Y., Coombes, C. E., Kilaru, A., Li, X., Gitler, A. D., Bowers, W. J., ... Moore, D. J. (2010). GTPase activity plays a key role in the pathobiology of LRRK2. *PLOS Genetics*, *6*(4), e1000902. doi:10.1371/journal.pgen.1000902 PMID:20386743

Yoshii, S. R., Kishi, C., Ishihara, N., & Mizushima, N. (2011). Parkin mediates proteasome-dependent protein degradation and rupture of the outer mitochondrial membrane. *The Journal of Biological Chemistry*, 286(22), 19630–19640. doi:10.1074/jbc.M110.209338 PMID:21454557

Youdim, M. B. H., & Bakhle, Y. S. (2006). Monoamine oxidase: Isoforms and inhibitors in Parkinson's disease and depressive illness. *British Journal of Pharmacology*, *147*(1), S287–S296. PMID:16402116

Youle, R. J., & Narendra, D. P. (2011). Mechanisms of mitophagy. *Nature Reviews. Molecular Cell Biology*, *12*(1), 9–14. doi:10.1038/nrm3028 PMID:21179058

Zhang, Y., Gao, J., Chung, K. K., Huang, H., Dawson, V. L., & Dawson, T. M. (2000). Parkin functions as an E2-dependent ubiquitin–protein ligase and promotes the degradation of the synaptic vesicle-associated protein, CDCrel-1. *Proceedings of the National Academy of Sciences of the United States of America*, *97*(24), 13354–13359. doi:10.1073/pnas.240347797 PMID:11078524

Chapter 13 **Alpha-Synucleinopathies:** Parkinson's Disease, Dementia With Lewy Bodies, and Multiple System Atrophy

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ABSTRACT

Alpha-synuclein is a protein that forms a major component of abnormal neuronal aggregates known as Lewy bodies. A particular group of neurodegenerative disorders (NDs) is characterized by the abnormal accumulation of α -synuclein; termed the α -synucleinopathies, this group includes Parkinson's disease (PD), dementia with Lewy bodies (DLB), and multiple system atrophy (MSA). Lysosomal storage diseases have also been linked to α -synuclein toxicity. Several therapeutic targets have been chosen among steps of metabolism of α -synuclein. Reducing α -synuclein synthesis or expression and increasing the clearance can be achieved in many ways. The development of immunotherapeutic approaches targeting α -synuclein has received considerable attention in recent years. The aim of this chapter is to present the α -synucleinopathies, as well as to present the most recent researches about treatment of synucleinopathies based on knowledge of the pathophysiology of α -synuclein pathways.

DOI: 10.4018/978-1-5225-5282-6.ch013

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INTRODUCTION

A common feature among a number of neurological disorders is the abnormal aggregation of a protein as observed with amyloid beta in Alzheimer's disease (Selkoe et al., 1990) and huntingtin protein in Huntington's disease (Vonsattel et al., 2011). In 1912, Frederick Lewy first described the cytoplasmic inclusions now known as Lewy bodies in the substantia nigra in PD Cortical Lewy bodies were first reported in association with dementia in 1961 (Okazaki et al., 1961) but they were felt to be a relatively rare finding until the 1980s, when first ubiquitin and later α -synuclein immunostains made it easier to see them (Spillantini et al., 1997) and demonstrated that Lewy bodies were a common neuropathologic finding in dementia (Gomperts, 2016). α -Synucleinopathies is a particular group of NDs characterized by the abnormal accumulation of α -synuclein (Gomperts, 2016; Kahle, 2008) (Figure 1). Lysosomal storage diseases have also been linked to α -synuclein toxicity (Wong and Krainc, 2017). There are new treatments being researched based on the pathophysiology of alpha-synuclein, and immunology. The aim of this chapter is to present the α -synucleinopathies, as well as to present the most recent researches about treatment of synucleinopathies based on knowledge of the pathophysiology of α -synuclein pathways.

BACKGROUND

Despite the early knowledge of the involvement of α -synuclein in several neurodegenerative disorders, only recently has there been, as a Holy Grail, a search for the cause and cure of these diseases through α -synuclein cellular and extracellular mechanisms and pathways. A heated discussion on this topic was

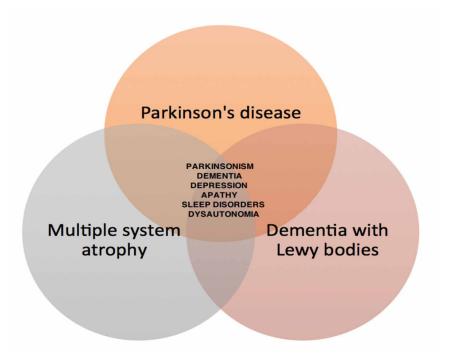


Figure 1. Main common features among α -synucleinopathies

triggered after studies suggesting a prion-like role of α -synuclein in parkinsonisms (Brundin et al., 2016). After that, studies suggested the shaping of prions from gut sharing to central nervous system. Finaly, gut microbiota changes in PD patients increased discussions and interest in this subject (Scheperjans et al., 2015).

These are not the only controversies related to synucleinopathies. Neurologists have not always contented the term synucleinopathies, a clustering of various diseases that have the Lewy bodies and alpha synuclein as the center of pathophysiology. However, this chapter will try to demonstrate after describing the differences and similarities between these diseases, new findings on the pathophysiology of α -synuclein have helped to understand the clinical differences and, therefore, generating new researches for general treatments of synucleinopathies and also specific for them.

ALPHA SYNUCLEIN

There are different strains of α -synuclein, defined as conformational variants of α -synuclein — that exhibit distinct properties such as differences in structure and toxicity and ability to seed, propagate and cross-seed tau fibrillization. α -synuclein is able to transition between multiple different conformations, including monomers, tetramers, higher-level oligomers (soluble conformations), fibrils (highly ordered insoluble conformations characterized by β -sheet conformation) and aggregates. In addition, differences in α -synuclein strains also exist between synucleinopathies, such as PD and MSA (Bendor et al., 2013; Wong and Krainc, 2017).

The presynaptic location of α -synuclein has been recognized since its original identification as a protein associated with synaptic vesicles (Maroteaux et al., 1988). The protein is relatively specific to the nervous system (Iwai et al., 1995). In addition, α -synuclein is widely expressed by many neuronal populations within both central and peripheral nervous systems, suggesting a general role in neuronal function. However, α -synuclein appears to be one of the last proteins that localizes to developing synapses, arriving after integral membrane proteins of the synaptic vesicle and the peripheral membrane synapsin proteins (Withers et al., 1997).

ALPHA SYNUCLEINOPATHIES

Parkinson's Disease

PD is a neurodegenerative disorder, characterized by movement disorders and non-motors symptoms (Salat et al., 2016), associated mainly with dopaminergic neurons death in the substantia nigra, the progressive depletion of dopaminergic nigrostriatal and mesocorticolimbic neurons (Callesen et al., 2013), abnormal deposition of α -synuclein in remaining cells and gliosis in specific areas of the nervous system (Lees et al., 2009; Salat et al., 2016). However, the changes are not restricted to these brain regions and can be found in other nuclei of the brainstem, in the cerebral cortex and even in peripheral neurons, such as those in the myenteric plexus. The presence of degenerative processes in the dopaminergic system and in different brain areas, for example the frontal lobe, can explain a series of non-motor signs and symptoms such as cognitive impairment and dementia (Lees et al., 2009). Non-dopaminergic pathways are

Alpha-Synucleinopathies

also involved, including the serotoninergic and cholinergic neurons at, in addition to the spinal cord and peripheral nervous system, correlating with the main non-motor symptoms of the disease (Poewe, 2008).

PD was for a long time characterized by its motor symptoms and signs (Parkinson, 1817). In the International Parkinson's Disease and Movement Disorders Society 2015 Criteria for PD (MDS-PD criteria), the centrality of the motor syndrome remains the core feature by which clinical PD is defined. The prerequisite to apply the MDS-PD criteria is the diagnosis of parkinsonism, which is based on three cardinal motor manifestations. Parkinsonism is defined as bradykinesia, in combination with either rest tremor, rigidity, or both. These features must be clearly demonstrable and not attributable to confounding factors. Having established that the patient has parkinsonism, the MDS-PD criteria will be applied to determine whether the patient meets criteria for PD as the cause of this parkinsonism. The diagnosis of clinically established PD requires: absence of absolute exclusion criteria (e.g. cerebellar abnormalities), at least two supportive criteria (e.g., marked improvement with dopaminergic therapy), no red flags (e.g. rapid progression of gait impairment) (Table 1) (Postuma et al., 2015).

However, over the years the non-motor manifestations became better characterized and can now be identified in most, if not all, patients (Seppi et al., 2011: Simuni et al., 2013). The non-motor manifestations of PD involve autonomic, neuropsychiatric dysfunctions, changes in sleep-wakefulness and in pain perception (Goldman and Weintraub, 2015; Poewe, 2008). The most common neuropsychiatric phenomena include dementia, visual hallucinations, apathy and depression (Aarsland et al., 2009; Lieberman, 2006).

In recent decades various sensory abnormalities have been identified as a result of more effective diagnosis and improved treatment strategies (Hughes et al., 1992). Pain is present in around 60% of PD patients and occurs two to three times more frequently in this population than in age-matched individuals

Supportive Criteria	 a 1. Response to dopaminergic therapy 2. Presence of levodopa-induced dyskinesia 3. Rest tremor of a limb 4. Positive results from at least one ancillary diagnostic test a. Olfactory loss b. Scintigraphy clearly documenting cardiac sympathetic denervation 		
Absolute Exclusion Criteria The presence of any of these features rules out PD	 e of any of 6 Absence of observable response to high-dose levedopa 		
Red Flags	 Rapid progression of gait impairment requiring regular use of wheelchair within 5 y of onset A complete absence of progression of motor symptoms or signs over 5 or more years Early bulbar dysfunction Inspiratory respiratory dysfunction Severe autonomic failure in the first 5 y of disease. Recurrent (>1/y) falls because of impaired balance within 3 y of onset. The presence of disproportionate anterocollis (dystonic in nature) or contractures of hand or feet within the first 10 y. Absence of any of the common nonmotor features of disease despite 5 y disease duration. Otherwise unexplained pyramidal tract signs Bilateral symmetric parkinsonism throughout the disease course 		

Table 1. Summary of Parkinson's disease Movement Disorders Society criteria 2015 (Postuma et al., 2015).

without PD (Beiske et al., 2009; Defazio et al., 2008; Nègre-Pagès et al., 2008). It is an early symptom and can precede motor symptoms by several years (Kim et al., 2013; Lin et al., 2013; Pont-Sunyer et al., 2015). In the case of smell, which appears to be the sense that is involved the earliest, the prevalence of dysfunction can reach 90% (Doty et al., 1988).

The initial pathological description of PD changed drastically after functional and neuropathological studies showed the extensive involvement of extranigral pathways (Rajput et al., 1991). In addition, following studies like that by Braak et al. (2003), who showed that Lewy bodies are found throughout the nervous system and affect even the peripheral nervous system, a search began for nonmotor clinical findings and pathophysiological responses in PD, particularly dysautonomias, olfactory dysfunctions and sleep disorders (Jellinger, 2009).

Hereditary Parkinson's Disease

In 1997, mutations responsible for the disease were identified in the α-synuclein gene (*SNCA*) (Polimeroloulos et al., 1997). Understanding the monogenic forms of PD provides insight more broadly into the genetic architecture of this disease, and as described later, there appears to be overlap in the genes that contain disease causing mutations and those that contain risk variants (Table 2). Mutations in three genes, *SNCA* (*PARK1*; encoding α-synuclein), *LRRK2* (*PARK8*; encoding dardarin) and *VPS35* (encoding vacuolar protein sorting 35) have been shown to cause autosomal dominant forms of PD. Mutations in six other genes, *PINK1* (*PARK6*; PTEN induced kinase-1), *DJ-1* (*PARK7*), *Parkin* (*PARK2*), *ATP13A2* (*PARK9*), *FBXO7* and *PLA2GB* have been shown to cause autosomal recessive parkinsonism. All known monogenic forms of PD combined explain only about 30% of familial and 3–5% of sporadic cases (Hernandez et al., 2016; Kumar et al., 2011).

Following the discovery of *SNCA* mutations causing a rare familial form of PD, Spillantini et al. (1997) determined that α -synuclein was a major constituent of Lewy bodies, the pathological hallmark of PD. In addition to *SNCA*, autosomal-dominant PD-causing mutations have been found in the gene encoding Leucine-rich repeat kinase 2 (*LRRK2*). Mutations in *LRRK2* are the most common known genetic cause of late-onset PD and are found in both autosomal dominant and sporadic cases (Hernandez et al., 2016). *LRRK2* encodes a large the ROCO protein family, dardarin (Beilina et al., 2014; Greggio et al., 2009; Li et al., 2009), widely expressed in brain tissue.

During recent years several susceptibility genes and numerous risk loci associated with PD have been identified. While success in this regard has been driven by largely unbiased genome wide studies, some limited success was achieved through candidate gene-based assessments in three genes: *SNCA*, *LRRK2* and *GBA* (Hernandez et al., 2016).

Lewy Bodies Dementias

DLB and dementia that arises in PD (ie, Parkinson disease dementia [PDD]) together comprise the Lewy body dementias. Despite the different temporal sequences of motor and cognitive deficits, PDD and DLB show remarkably convergent neuropathologic changes at autopsy. These changes include widespread limbic and cortical Lewy bodies and Lewy neurites composed of aggregates of α -synuclein that involve the brainstem as well as limbic and neocortical regions (referred to as Lewy body disease), loss of midbrain dopamine cells, and loss of cholinergic neurons in ventral forebrain nuclei. Neuritic

Alpha-Synucleinopathies

Locus Gene		Protein	Model	
Park1	SNCA	α-Synuclein	Autosomal Dominant	
Park2	PARK2	Parkin	Autosomal Recessive	
Park3	unknown	unknown	Autosomal Dominant	
Park4	SNCA	α-Synuclein	Autosomal Dominant	
Park5	UCHL1	Ubiquitin c terminal hydrolase Autosomal Dominant		
Park6	PINK1	Pten-induced putative kinase 1	Autosomal Recessive	
Park7	PARK7	DJ-1	Autosomal Recessive	
Park8	LRRK2	Leucine rich repeat kinase 2 (dardarin)	Autosomal Dominant	
Park9	ATP13A2	lysosomal type 5 ATPase	Autosomal Recessive	
Park10	unknown	unknown	Risk Locus	
Park11	GIGYF2	GRB interacting GYF protein 2	Autosomal Dominant	
Park12	unknown	unknown	X-Linked	
Park13	HTRA2	HTRA serine peptidase 2	Autosomal Dominant	
Park14	PLA2G6	Phospholipase A2	Autosomal Recessive	
Park15	FBXO7	F-box only protein 7	Autosomal Recessive	
Park17	VPS35	Vacuolar protein sorting 35	Autosomal Dominant	
Park18	EIF4G1	Eukaryotic translation initiation factor 4 gamma 1	Autosomal Dominant	
Park19	DNAJC6	DNAJ/HSP40 homolog subfamily C member 6	Autosomal Recessive	
Park20	SYNJ1	SYNJ1	Autosomal Recessive	
Park21	DNAJC13	DNAJ/HSP40 homolog subfamily C member 13	Autosomal Dominant	
Park22	CHCHD2	CHCHD2	Autosomal Dominant	
Park23	VPS13C	Vacuolar protein sorting 13C	Autosomal Recessive	
-	SNCA	α-Synuclein	Risk Locus	
-	LRRK2	Leucine rich repeat kinase 2	Risk Locus	
-	GBA1	Glucocerebrocidase	Risk Locus	

Table 2. Monogenic forms of hereditary Parkinson's disease (Funayama et al., 2015; Hernandez et al., 2016; Kalinderi et al., 2016; Lesage et al., 2016).

plaques that contain amyloid and neurofibrillary tangles are found in the majority of cases of DLB and are common in PD (Gomperts, 2016).

A related Lewy body disease is PD, in which dementia may ensue and thereby, shares many clinical and cognitive features with DLB (Emre et al, 2007; Goldman et al., 2014). At early stages, DLB and PDD are easy to differentiate by the predominance of dementia in DLB and of parkinsonian motor features in PD. Nevertheless, in some patients, dementia and motor signs occur in close succession, provoking debate about their nosology. For research purposes, the "one-year" rule regarding timing of dementia and parkinsonian features is used (Emre et al, 2007; Goldman et al., 2014; McKeith et al., 2005). In clinical practice, however, a diagnosis is made based on the relative prominence of the clinical features. The separation between DLB and PDD is considered by some to be artificial, since it implies that the two clinical syndromes have different pathophysiologies (Aarsland et al., 2004; Goldman et al., 2014).

DLB is the second most common form of degenerative dementia after Alzheimer's disease, with prevalence rates of up to 5% in the elderly and up to 30% of all dementia cases (McKeith et al., 2005; Zaccai et al., 2005). PDD affects about 40% to 80% (Poewe, 2008; Pigott et al., 2015) of individuals with PD during the course of the disease. It is believed that 3% to 4% of cases of dementia in the general population are related to PD, and the estimated prevalence of PDD in the general population aged over 65 years is 0.2 to 0.5% (Aarsland et al., 2007). Mild cognitive impairment in PD (MCI-PD) has been shown to be a symptom that can progress to PDD. PDD has an insidious onset and progressive evolution (Goetz et al., 2008), being a major cause of the increase in the need of care at the old people's home, higher costs in healthcare and increased mortality (de Lau et al, 2005; Poewe e Mahlknecht, 2009). When dementia becomes clinically significant, the average survival of patients drops to 5 years (Poewe e Mahlknecht, 2009). However, cognitive decline is not limited to PD advanced stages and has been identified in 20 to 35% of patients recently diagnosed with PD or in the initial stages of the disease. This high prevalence and the major impact the condition has on the patient and family members make it essential to detect and diagnose cognitive changes in PD patients as early as possible (de Lau et al, 2005; Poewe e Mahlknecht, 2009).

The characteristic features of DLB are spontaneous parkinsonism, recurrent visual hallucinations, fluctuating cognition, rapid eye movement sleep behaviour disorder (RBD), severe sensitivity to antipsychotic medications and reduction in striatal dopamine transporters on single photon emission computed tomography (SPECT) or positron emission tomography (PET) (Donaghy e McKeith, 2014). The cognitive domains affected in patients with Lewy body dementia are: attention, memory, executive function, construction and apraxia, visuospatial function and language (Goetz et al., 2008). However, Lewy body dementia is mainly a dysexecutive syndrome characterized by impaired planning, a deficit in executive function (including organization of goal-directed activities), dyspraxia, bradyphrenia, reduced problem-solving ability, learning difficulty and short-term memory loss. Most patients also present concomitantly fluctuations in attention, mood and personality, as well as hallucinations and psychoses (Emre et al., 2007; Aarsland et al., 2010). This cognitive impairment is sufficient to compromise the ability to perform activities of daily living and planning, organizational, executive, visuospatial and language abilities (Aarsland et al., 2007).

PDD and DLB are associated with dopamine loss in dopaminergic pathways to the prefrontal cortex, and, to a lesser extent, the ventral tegmental area. In PDD, the appearance of Lewy bodies, which normally occurs in the advanced stages of PD, contributes to the development of severe dementia (Aarsland et al., 2009; de la Fuente-Fernández, 2012). As PD and PDD, the main pathological lesions seen in DLB are Lewy bodies and Lewy neurites, both containing α -synuclein. The pattern of distribution of Lewy bodies pathology in DLB, PD and PDD as seen at end stage is very similar, although cortical involvement may occur earlier in DLB and brainstem involvement may be minimal (Donaghy e McKeith, 2014).

There has been shown to be an association between hypofunction in the pathways involving the dorsal caudate nucleus and the orbitofrontal cortex (including connections with the amygdala), the same circuits that have been associated with the onset of PD, when PDD is less prevalent. This may explain the finding that depression is more correlated to less severe cognitive changes (den Brok et al., 2015; Remy et al., 2005). As the disease progresses, hypofunction of the circuit involving the anterior cingulate cortex and nucleus accumbens (including connections with the amygdala) occurs in patients with apathy, followed by changes in the pathways involving the ventral caudate nucleus and orbitofrontal cortex (Monchi et al., 2007; Remy et al., 2005; Reijnders et al., 2010). In the advanced stages of PD, patients have dopamine depletion in the corpus striatum (putamen, globus pallidus and caudate nucleus) as well as the frontal

cortex, increasing the prevalence and severity of apathy as dementia and Lewy bodies' progresses (Emre et al., 2007; Reijnders et al., 2010), in agreement with the findings of the present study. Apathy, rather than depression, therefore has the strongest association with more severe cognitive dysfunction in PD (Emre et al., 2007; Reijnders et al., 2010; Seppi et al., 2011). The good effect caused by levodopa in motor PD treatment was not observed in the assessment of neuropsychiatric dysfunctions. The explanation for this paradox is that levodopa does not cause significant changes in the activity of dopaminergic projections to the frontal cortex, as they are mediated largely by D1 dopamine receptors, but stimulates mainly D2 receptors, which predominate in the corpus striatum. Levodopa therapy is therefore not expected to lead to fluctuations in non-motor disturbances in PD (de la Fuente-Fernández, 2012; Remy et al., 2005).

Multiple System Atrophy

The term multiple system atrophy (MSA) was coined in 1969 to encompass three previously distinct neurodegenerative disorders, striatonigral degeneration, olivopontocerebellar ataxia, and "Shy-Drager syndrome" (Graham and Oppenheimer, 1969).

While in recent years considerable effort has been devoted at understanding the pathogenesis of PD, less is known about MSA, which is a rapidly progressive and fatal ND characterized by parkinsonism, dysautonomia (Dickson et al., 1999; Wenning et al., 2001) and α -synuclein accumulation within oligodendroglial and neurons. This accumulation is accompanied by neuroinflammation, demyelination and neurodegeneration (Valera et al., 2016). Additionally, MSA patients can develop pyramidal and behavioral alterations such as depression and executive dysfunction that suggest frontal lobe impairment (Dujardin et al., 2003; Ubhi et al., 2011). The autonomic dysfunctions most commonly are urogenital, gastrointestinal and cardiovascular dysfunction in the form of orthostatic hypotension (Pfeiffer, 2007).

Parkinsonian features reflecting striato-nigral neurodegeneration predominate in 80% of MSA patients (MSA-P subtype), while the major motor feature in 20% of patients is cerebellar ataxia due to olivopontocerebellar atrophy (MSA-C subtype) (Gilman et al., 2008). However, epidemiological studies in North America (Gilman et al., 2008) and Japan (Yabe et al., 2006) have suggested an ethnic variation with regards to the incidence rates of MSA-P or MSA-C. The North American Study reported 60% of their patients as having MSA-P and 13% exhibiting MSA-C (Gilman et al., 2008). In contrast, the Japanese study reported a much higher percentage of patients (83.8%) exhibiting MSA-C features with only 16.2% of patients being categorized as MSA-P (Yabe et al., 2006). The underlying cause of this variability remains undetermined but may involve genetic or environmental factors, or a combination thereof (Ubhi et al., 2011).

Clinically, the rapid progression, the lack of response to levodopa (Wenning et al., 1994), and pathologically the extensive accumulation of α -synuclein within oligodendrocytes differentiates MSA from other synucleinopathies (Dickson et al., 1999b; Valera et al., 2016). However, the mechanisms through which α -synuclein accumulates within oligodendroglial cells in MSA are not completely understood. One possibility is that α -synuclein is produced by oligodendroglial cells, which in turn over-express or fail to intrinsically clear α -synuclein; the other is that α -synuclein propagates from neurons to oligodendrocytes due to neurons over-expressing and/or displaying defects in the physiological mechanisms of α -synuclein clearance (Valera et al., 2016).

STORAGE DISEASES ASSOCIATED WITH ALPHA SYNUCLEIN TOXICITY

 α -Synuclein has been implicated in several diseases, and lysosomal storage diseases have also been linked to α -synuclein toxicity.

Gaucher's Disease

Gaucher's disease (GD) is a rare, autosomal recessive lysosomal storage disorder that results from lossof-function mutations in glucocerebrosidase (GCase) (Brady et al., 1965).

Three types of GD have been described, based on the rate of clinical progression and involvement of the nervous system (Grabowski, 2008). Type I GD is classically defined as non-neuronopathic and is typically characterized by visceral and hematopoietic abnormalities. Types II and III are differentiated from type I by neurodegeneration with either rapid (type II) or chronic progression (type III). A common feature is accumulation of GlcCer in the affected tissues, but the reasons for variability of GD are not known (Bultron et al, 2010).

A subgroup of type I GD with parkinsonism suggest a possible link between the two disorders (Neudorfer et al., 1996). Neuropathology revealed α -synuclein-positive Lewy bodies (Wong et al., 2004), suggesting the involvement of α -synuclein aggregation. Several genetic studies in large patient cohorts demonstrated that patients with parkinsonism have an increased incidence of *GBA1* mutations (Sidransky et al., 2009), making *GBA1* the most common known genetic risk factor for PD to date.

GBA1 mutations have also been identified in patients with the diagnosis of DLB (Goker-Alpan et al., 2006). The complete mechanism of the linkage between GlcCer and α -synuclein has not been explained.

Metabolism of GlcCer and α -synuclein show a reverse control, with the elevation of α -synuclein contributing to a pathogenic cycle by inhibiting the maturation of normal GCase and the GlcCer accumulation providing α -synuclein oligomer formation (Mazzulli et al., 2011).

It's not completely clear why patients carrying one *GBA1* mutation have increased risk of PD, although many of these patients have well above 50% of control GCase activity.

Thus, it is quite likely that multiple factors, many of which are impacted by the aging process, play a role with deficient GCase to cause parkinsonism (Aflaki et al., 2017).

Additional Lysosomal Storage Disorders

Mucopolysaccharidoses

In the mucopolysaccharidoses (MPS), glycosaminoglycans (mucopolysaccharides) accumulate as a result of impaired function of any of 11 lysosomal enzymes that degrade the glycosaminoglycans (Vitner et al., 2010).

In human MPS IIIB brain tissue, neuronal loss was observed only in the SN, but no Lewy Body-like inclusions were detected. In both MPS IIIB and MPS II brains, α -synuclein was phosphorylated mainly in neurons containing high levels of storage materials (Hamano et al., 2008). In an MPS IIIA mouse model the block in autophagy can lead to accumulation of polyubiquitinated proteins and mutant α -synuclein (Settembre et al., 2008).

Tay-Sachs and Sandhoff Diseases

The GM2 gangliosidosis (including Tay–Sachs and Sandhoff diseases) are caused by defective b-hexosaminidase activity (Vitner et al., 2010) with numerous case reports linking with PD. Adults patients with GM2 gangliosidosis can present various parkinsonian symptoms including bradykinesia, rigidity, bilateral tremor and resting tremor (Argov and Navon, 1984).

Immunohistochemical analyses in neurons of a Sandhoff disease mouse model demonstrated accumulation of α -synuclein and b-synuclein in striatal terminals and in the SN. Neurons that accumulated α -synuclein were negative for ubiquitin, suggesting a different form from that in classical Lewy bodies' disease (Suzuki et al., 2003). α -Synuclein is also observed in the cerebrum, cerebellum, and brain stem of both Sandhoff and Tay–Sachs disease human patients (as young as 2–3 years old) (Suzuki et al., 2007).

A biochemical association between GM2 gangliosidosis and PD has been demonstrated through the reduced levels of UCH-L1 in cultured fibroblasts from brain extracts from a Sandhoff disease mouse model, as seen in PD (Leroy et al., 1998; Liu et al., 2002).

Niemann-Pick Type C

Niemann–Pick C (NPC) disease is caused by the defective activity of either NPC1 or NPC2, resulting in cholesterol and sphingolipid accumulation (Vitner et al., 2010).

 α -Synuclein was observed in the midbrain and amygdala, and classical Lewy bodies were found in the SN of NPC patients (Shachar et al., 2011).

Anecdotal reports linked NPC to sporadic parkinsonian syndrome in younger patients (Coleman et al., 1998).

Neurodegeneration With Brain Iron Accumulation

Neurodegeneration with brain iron accumulation (NBIA) is a group of ND characterized by iron accumulation in the basal ganglia, specifically in the globus pallidus (GP) and the substantia nigra (SN) and can be visualized with MRI. The cortex and the cerebellum can be affected, and cerebellar involvement correlates with the most severe NBIA subtypes (Wray et al, 2016).

 α -Synuclein pathology is not usual in this class. Authors presented the NBIA forms with α -synuclein presentation, described at Table 3.

PLA2G6-Associated Neurodegeneration

PLA2G6-associated neurodegeneration (PLAN) is the second core NBIA syndrome (NBIA type II, OMIM 256600 and 610217) and is associated to mutations in the PLA2G6 gene (Morgan et al., 2006). It can present as infantile neuroaxonal dystrophy (classic INAD) and as atypical neuroaxonal dystrophy (atypical INAD). INAD has an early onset presentation, usually with psychomotor regression and has a rapid progression with severe neurological deterioration.

The atypical INAD has a later onset and a less homogeneous presentation with gait problems, ataxia or speech difficulties being the most common symptoms (Gregory et al., 2008). The progression of the disease usually allows a longer life span than classic INAD.

Human Disorder	MRI	Symptomatology	Gene	α-Syn Pathology
Pantothenate kinase-associated neurodegeneration (PKAN) (NBIA1)	Hypointensity with central hyperintensity of the GP, referred to as 'eye of the tiger'	Dystonia, spasticity and parkinsonism cognition often spared	PANK2	No
COASY protein associated neurodegeneration (CoPAN)	Hypointensity with central hyperintensity of the GP	Spasticity, dystonia, dysarthria, parkinsonism and cognitive decline	COASY	Unknown
Mitochondrial membrane-associated neurodegeneration (MPAN) (NBIA4)	Hypointensity of the GP and SN plus hyperintensity in the GP	Spasticity, dystonia, dysarthria, parkinsonism and cognitive decline	C19orf12	Severe Lewy bodies and Lewy neuritis GP, SN, cortex, striatum
PLA2G6-associated neurodegeneration (PLAN) (NBIA2)	Hypointensity of the GP in a subset of patients Cerebellar atrophy. Some cortical atrophy	Hypotonia, spasticity, dystonia, parkinsonism and cerebellar ataxia Motor and mental retardation	PLA2G6	Severe, Lewy bodies and Lewy neurites Ubq, α-syn SNc, cortex
FA2H-associated neurodegeneration (FAHN)	Hypointensity of the GP Cerebellar and cortical atrophy	Spasticity, ataxia and dystonia	FA2H	Unknown
β-propeller-associated neurodegeneration (BPAN)	Hypointensity of the GP/SN with central hyperintense line Cerebral and cerebellar atrophy	Parkinsonism, dystonia and dementia, developmental delay, cognitive disturbances	WDR45	No
Kufor-Rakeb syndrome	General atrophy and hypointensity in the basal ganglia/caudo- putamen	Parkinsonism, dementia and some pyramidal signs	ATP13A2	Unknown
Woodhouse Sakati syndrome	Hypointensity of the GP and SN	Diabetes, alopecia, hypogonadism, deafness	DCAF17	Unknown
Aceruloplasminemia	Hypointense striatum, thalamus and dentate	Dystonia, dyskinesia and cerebellar ataxia, cognitive impairment	СР	Unknown
Neuroferritinopathy (NBIA3)	Hypointensity in basal ganglia, especially GP and SN. Also motor cortex	Dystonia, spasticity, rigidity and parkinsonism. Some cognitive impairment	FTL	Unknown

Table 3. Human forms of neurodegeneration with brain iron accumulation, genes and presence of α -synuclein pathology (Adapted from Wray et al., 2016).

The PLA2G6 gene encodes an 806 amino acid protein with predicted size of 88 kDa. (Mak et al., 2011). PLA2G6 has been shown to be expressed with protective function in the mitochondria (Saito et al, 2000).

Autosomal recessive mutations are found throughout PLA2G6 and lead to reduced enzyme activity, with the magnitude of enzymatic effect correlating with the disease severity (Morris et al., 1992).

PLA2G6 is classified as a PD gene (PARK14), and also, a reduction in PLA2G6 protein levels has been shown in the brains of Alzheimer's patients (Kupsch et al., 2003).

Pathology

The major pathological hallmark of PLAN is the presence of axonal swellings throughout the cortex, GP, striatum, cerebellum, brain stem and spinal cord. These are also present throughout the peripheral nervous system and can be diagnostic via peripheral nerve biopsies. There is evidence for PLA2G6 mutation-associated PLAN without spheroids (Morris et al., 1992). Peripheral spheroids are not observed in dystonia-parkinsonism subset of PLAN patients (Eidelberg et al., 1987).

 α -Synuclein positive Lewy bodies are found throughout the degenerating brain areas to a very high degree, and a proportion of the axonal swellings also stained positive for α -synuclein (Williams et al., 2012; Lee et al., 2006).

Atrophy is evident in the cortex, GP, white matter and cerebellar cells and was shown to be associated with gliosis in the cortex and the striatum (Lee and Andersen, 2006). Excess iron deposition was specifically observed in the GP and the SN pars reticulata and there is extensive cortical involvement in disease with Lewy bodies seen in all cases whereas tau deposition is rarer.

Mitochondrial Membrane-Associated Neurodegeneration

Mutations in the *C19orf12* gene are associated with mitochondrial membrane-associated neurodegeneration (MPAN), an autosomal recessive disorder that represents between 5 and 30% of NBIA cases (Hartig et al., 2011; Hogarth et al., 2013; Panteghini et al., 2012). These patients are characterized by pyramidal and extrapyramidal signs, cognitive decline, neuropsychiatric changes, optic atrophy, and upper and lower motor neuron signs (Hartig et al., 2011; Hogarth et al., 2013).

Great variability is present in disease onset (between 3 and 30 years) and progression. Gait instability or visual impairment are often the initial symptoms, then followed by muscular weakness and atrophy, dystonia and dysarthria. Almost constant is the cognitive decline leading to dementia as well as the appearance of neuropsychiatric symptoms (Arber et al, 2016).

Brain iron accumulation is the most significant sign at MRI. It involves both the GP and the substantia nigra; it can be accompanied by cortical and cerebellar atrophy. Neuropathologic assessment performed in two cases showed very similar results with iron accumulation, axonal spheroids, tau and Lewy pathology in basal ganglia, the archicortex, the neocortex, and the spinal cord. Iron was evident in the GP and the substantia nigra and associated with neuronal loss and gliosis. Eosinophilic axonal spheroids were evident and were strongly immunoreactive for ubiquitin, and less positive for tau or APP staining (Hogarth et al, 2013).

Lewy pathology was present in many brain regions. In one case the overall burden was greater than that observed in cases of sporadic Lewy body disease (Hogarth et al., 2013).

The *C19orf12* gene consists of three exons and codes for two alternative mRNA isoforms (NM_001031726.3 and NM_031448.4) and proteins that differ for the presence of a stretch of 11aminoacids at the N-terminus of the longer form (Arber et al, 2016). Different types of mutation have been identified and many of them lead to truncated, non-functional proteins. The missense mutation c.32C>T is the most common and affects tyrosine 11, exclusively present in the longer form. The protein has a hydrophobic domain that could represent a transmembrane region. Its expression appears to be involved with CoA and lipid metabolism and mitochondria. No other information is available to explain the connection between the protein function, the neurodegenerative process, and iron accumulation in the brain. Recently, a *Drosophila* model with impaired expression of the two orthologs of human *C19orf12*, was

obtained (Iuso et al., 2014); those flies showed reduced life span, signs of neurodegeneration (vacuoles) but not of iron accumulation (negative Prussian blue staining) (Arber et al, 2016).

THERAPEUTIC POSSIBILITIES

Reducing α-Synuclein Synthesis

Given that increased α -synuclein uclein expression causes familial PD, several studies have aimed to decrease its synthesis by using siRNA that targets α -synuclein mRNA. In mice, direct siRNA infusion decreases hippocampal and cortical α -synuclein levels for a week after infusion (Lewis et al, 2008), whereas in mice expressing a human form of α -synuclein, injecting siRNA-containing exosomes decreases protein aggregation in the SNc (Cooper et al, 2014).

Given that antisense oligonucleotide (ASO)-mediated therapies are being tested in clinical trials currently as potential therapeutics for other ND, ASO targeting of α -synuclein may also be effective.

Increasing α-Synuclein Degradation

Increasing the clearance could be achieved in many ways but basically consists of restoring the autophagy-lysosomal pathways that are impaired in PD. Overexpression of the lysosomal transcription factor EB (TFEB), which was reported to coordinate cellular clearance via macroautophagy and exocytosis, in rats expressing α -synuclein decreases α -synuclein oligomer levels and prevents lysosomal dysfunction decline and neurodegeneration. In addition, alteration in TFEB activity was reported in induced pluripotent stem cell-derived neurons from GD patients who present α -synuclein pathology. In parallel, academic and industrial laboratories are screening for clinically approved drugs that can enhance TFEB function (Dehay et al., 2016; Wong and Krainc, 2017).

In addition to attempting to boost lysosomal biogenesis, acidic NPs (aNPs) of poly DL-lactide*co*-glycolide (PLGA), which are FDA approved, have been reported to traffic to lysosomes and to act on lysosomal pH. However, whether PLGA-aNPs can acidify sick lysosomes and restore lysosomal/ autophagic function in pathological conditions remains to be determined. Reacidification of defective lysosomes following PLGA-aNP treatment restores lysosomal function in different pathological contexts (Dehay et al., 2016). An alternate protective strategy could be to prevent endosomal uptake of pathological α -synuclein into cells increased expression of other lysosomal proteins, including LIMP2 and ATP13A2, also promote α -synuclein degradation in various models, presenting additional targets for the acceleration of α -synuclein degradation (Wong and Krainc, 2017).

Reducing α-Synuclein Agregation

Given that α -synuclein oligomers and fibrils are implicated in α -synuclein toxicity, several strategies have been attempted to reduce their formation. The porphyrin phthalocyanine tetrasulfonate binds and stabilizes vesicle-associated α -synuclein, which thus delays its misfolding and aggregation (Fonseca-Ornelas et al., 2014).

Passive immunization using a protofibril-selective antibody decreases soluble and membrane-bound α -synuclein protofibrils in the spinal cord and decreases motor dysfunction in mice expressing PD-

associated mutant A30P α -synuclein (Lindstrom et al., 2014). In addition, if native α -synuclein exists predominantly as a tetramer, compounds that stabilize α -synuclein in this conformation—as has been applied for transthyretin, which misfolds in the multisystem disorder transthyretin-related amyloidosis—might be effective at combating α -synuclein toxicity (Dettmer et al., 2015).

Several clinical trials are currently using small molecules (Schneeberber et al., 2016) to inhibit either α -synuclein aggregation (glycerol phenylbutyrate (University of Colorado, Denver); nilotinib (George-town University) or α -synuclein oligomer formation (EGCG (University of Munich).

Blocking α-Synuclein Propagation

Because α -synuclein propagation may contribute to the spreading of α -synuclein toxicity, passive immunization studies to block spreading have also been performed. Antibodies against C-terminal truncated α -synuclein decrease its propagation in vitro and rescue motor and memory impairments in an α -synuclein mouse model (Games et al., 2014), and monoclonal α -synuclein antibodies prevent propagation and uptake of α -synuclein and rescue dopaminergic neuron loss and motor deficits in mice injected with α -synuclein pffs (Tran et al., 2014).

Importantly, understanding the role of propagation in disease will help to clarify whether these therapies will be successful for patients.

Passive and Active Immunization

The development of immunotherapeutic approaches targeting α -synuclein has received considerable attention in recent years. In this sense, both humoral (active and passive) and T-cell-based approaches have been explored. While active immunization stimulates the immune system to produce antibodies against target proteins (the response takes days/weeks to develop but may be long lasting—even lifelong), passive immunization consists in directly administering IgG antibodies that confer temporary protection against the disease (several weeks to few months) (Valera et al., 2016-b). Monoclonal antibodies against α -synuclein can regulate inflammation and facilitate the clearance of target protein via autophagy or microglia (Valera et al., 2016-b).

Immunotherapy has been shown to induce a physiological microglial response (M2 type) and reduce the production of proinflammatory cytokines, thus exerting an anti-inflammatory effect in neurodegenerative disorders (Valera et al., 2016-b). Among the cell types that can internalize α -synuclein aggregates, microglia exhibit the most rapid clearance of these proteins. Microglia became better scavengers for extracellular α -synuclein aggregates in the presence of specific antibodies against α -synuclein. Antibody- α -synuclein immune complexes entered microglia through the Fc γ receptors, which led to efficient delivery of these immune complexes to lysosomes, hence resulting in fast degradation. By clearing extracellular α -synuclein, antibody treatment significantly reduced the extent of cell-to-cell transmission of the protein in mouse models. Furthermore, clinical advantages might be expected if extracellular α -synuclein is selectively targeted by immunotherapy with the intraneuronal α -synuclein being left intact (Lee and Lee, 2016).

The first study vaccinated transgenic LBD-mice using full human α -synuclein, which cleared α -synuclein aggregates, potentially via lysosomal pathways (Maliash et al., 2005). These results suggested that α -synuclein vaccination is effective in reducing neuronal accumulation of α -synuclein aggregates and that further development of this approach might have a potential role in the treatment of synucle-

inopathies (Valera et al., 2016-b). Other active immunization approaches using a next-generation active vaccination technology with small peptides, or AFFITOPEs[®] (AFFiRiS AG) (Mandler et al., 2014). AFFITOPEs[®] that mimic the C-terminal region of α -synuclein are able to elicit an immune response specific to α -synuclein oligomers. Vaccination with one of these AFFITOPEs[®] (AFF 1) resulted in high antibody titers against α -synuclein aggregates, decreased accumulation of α -synuclein oligomers. When administered to a transgenic model of MSA, AFF 1 also induced a reduction in neurodegeneration and demyelination in neocortex, striatum, and corpus callosum. The clearance of α -synuclein induced by AFF 1 involved activation of microglia, increased anti-inflammatory cytokine production, and reduced spreading of α -synuclein to astroglial cells (Mandler et al., 2014).

Passive immunization approaches using antibodies against α -synuclein are also being actively pursued. Studies observed that antibodies that recognize an epitope in the C-terminal of α -synuclein are more effective at ameliorating the pathology in transgenic mouse models of PD, as they clear intracellular aggregates, inhibit α -synuclein propagation, and prevent C-terminal cleavage of the protein, which may lead to increased aggregation. The reports support the value of immunotherapy with monoclonal antibodies directed against α -synuclein for PD, and in this sense the C-terminal antibody PRX002 (Prothena, South San Francisco, CA, USA) and the antibody BIIB054 (Biogen, Cambridge, MA, USA) are currently being tested in phase I clinical trials (Valera et al., 2016; Schenk et al., 2017).

FUTURE RESEARCH DIRECTIONS

The main future expectations are related to the progress of treatments based on pathophysiology, mainly in the α -synuclein:

- Researchers and pharmaceutical companies are starting studies with immunotherapy for PD and MSA (Valera et al., 2016-b; Schenk et al., 2017). There is great expectation regarding immunological treatments for α-synucleinopathies.
- The knowledge of aggregation and clearance of α -synuclein will be the source of much research for many years towards better treatments for α -synucleinopathies (Schneeberber et al, 2016).
- The action of the gut and gut's flora on the pathophysiology of PD, as well as the involvement of the vagus nerve in the propagation of α -synuclein like the prions, will remain in the target of the scientists in the next years (Lee et al., 2011). It is hoped that new treatments may emerge from research in this area.

CONCLUSION

The studies to date have demonstrated the important participation of α -synuclein and Lewy bodies in the pathophysiology of several ND. However, a long road of discovery and understanding still awaits us. Research into the production of new drugs for the treatment of α -synucleinopathies has developed a lot, demonstrating new ways and unexpected and very interesting solutions. Regarding new treatments, the knowledge of recent years has brought many doors that need to be opened.

REFERENCES

Aarsland, D., Ballard, C. G., & Halliday, G. (2004). Are Parkinson's disease with dementia and dementia with Lewy bodies the same entity? *Journal of Geriatric Psychiatry and Neurology*, *17*(3), 137–145. doi:10.1177/0891988704267470 PMID:15312277

Aarsland, D., Brønnick, K., Ehrt, U., De Deyn, P. P., Tekin, S., Emre, M., & Cummings, J. L. (2007). Neuropsychiatric symptoms in patients with Parkinson's disease and dementia: Frequency, profile and associated care giver stress. *Journal of Neurology, Neurosurgery, and Psychiatry*, 78(1), 36–42. doi:10.1136/jnnp.2005.083113 PMID:16820421

Aarsland, D., Bronnick, K., Williams-Gray, C., Weintraub, D., Marder, K., Kulisevsky, J., ... Emre, M. (2010). Mild cognitive impairment in Parkinson disease: A multicenter pooled analysis. *Neurology*, 75(12), 1062–1069. doi:10.1212/WNL.0b013e3181f39d0e PMID:20855849

Aarsland, D., Marsh, L., & Schrag, A. (2009). Neuropsychiatric symptoms in Parkinson's disease. *Movement Disorders*, 24(15), 2175–2186. doi:10.1002/mds.22589 PMID:19768724

Aflaki, E., Westbroek, W., & Sidransky, E. (2017). The Complicated Relationship between Gaucher Disease and Parkinsonism: Insights from a Rare Disease. *Neuron*, *93*(4), 737–746. doi:10.1016/j.neuron.2017.01.018 PMID:28231462

Arber, C. E., Houlden, H., Li, A., & Wray, S. (2016). Review: Insights into molecular mechanism of disease in neurodegeneration with brain iron accumulation: unifying theories. *Neuropathology and Applied Neurobiology*, *42*(3), 200–241. doi:10.1111/nan.12242 PMID:25870938

Argov, Z., & Navon, R. (1984). Clinical and genetic variations in the syndrome of adult GM2 gangliosidosis resulting from hexosaminidase A deficiency. *Annals of Neurology*, *16*(1), 14–20. doi:10.1002/ ana.410160105 PMID:6235771

Beilina, A., Rudenko, I. N., Kaganovich, A., Civiero, L., Chau, H., Kalia, S. K., ... Cookson, M. R. (2014). Unbiased screen for interactors of leucine-rich repeat kinase 2 supports a common pathway for sporadic and familial Parkinson disease. *Proceedings of the National Academy of Sciences of the United States of America*, *111*(7), 2626–2631. doi:10.1073/pnas.1318306111 PMID:24510904

Beiske, A. G., Loge, J. H., Rønningen, A., & Svensson, E. (2009). Pain in Parkinson's disease: Prevalence and characteristics. Pain. *International Association for the Study of Pain*, *141*(1), 173–177. PMID:19100686

Bendor, J. T., Logan, T. P., & Edwards, R. H. (2013). The function of α-synuclein. *Neuron*, 79(6), 1044–1066. doi:10.1016/j.neuron.2013.09.004 PMID:24050397

Braak, H., Del Tredici, K., Rüb, U., Vos, R. A., Steur, E. N. J., & Braak, E. (2003). Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiology of Aging*, 24(2), 97–211. doi:10.1016/S0197-4580(02)00065-9 PMID:12498954

Brady, R. O., Kanfer, J., & Shapiro, D. (1965). The metabolism of glucocerebrosides.I. Purification and properties of a glucocerebroside-cleaving enzyme from spleen tissue. *The Journal of Biological Chemistry*, 240, 39–43. PMID:14253443

Brundin, P., Ma, J., & Kordower, J. H. (2016, August). How strong is the evidence that Parkinson's disease is a prion disorder? *Current Opinion in Neurology*, *29*(4), 459–466. doi:10.1097/WCO.000000000000349 PMID:27257944

Callesen, M. B., Hansen, K. V., Gjedde, A., Linnet, J., & Moller, A. (2013). Dopaminergic and clinical correlates of pathological gambling in Parkinson's disease: A case report. *Frontiers in Behavioral Neuroscience*, *7*, 1–8. doi:10.3389/fnbeh.2013.00095 PMID:23908610

Coleman, R. J., Robb, S. A., Lake, B. D., Brett, E. M., & Harding, A. E. (1998). The diverse neurological features of Niemann-Pick disease type C: A report of two cases. *Movement Disorders*, *3*(4), 295–299. doi:10.1002/mds.870030403 PMID:3145417

Cooper, J. M., Wiklander, P. B., Nordin, J. Z., Al-Shawi, R., Wood, M. J., Vithlani, M., ... Alvarez-Erviti, L. (2014). Systemic exosomal siRNA delivery reduced alpha-synuclein aggregates in brains of transgenic mice. *Movement Disorders*, *29*(12), 1476–1485. doi:10.1002/mds.25978 PMID:25112864

de la Fuente-Fernández, R. (2012). Frontostriatal cognitive staging in Parkinson's disease. *Parkinson's Disease*, 2012, 561046. doi:10.1155/2012/561046 PMID:22191070

de Lau, L. M., Schipper, C. M., Hofman, A., Koudstaal, P. J., & Breteler, M. M. (2005). Prognosis of Parkinson disease: risk of dementia and mortality: the Rotterdam Study. *Archives of Neurology*, 62(8), 1265–1269. doi:10.1001/archneur.62.8.1265 PMID:16087767

Defazio, G., Berardelli, A., Fabbrini, G., Martino, D., Fincati, E., Fiaschi, A., ... Tinazzi, M. (2008). Pain as a nonmotor symptom of Parkinson disease: Evidence from a case-control study. *Archives of Neurology*, *65*(9), 1191–1194. doi:10.1001/archneurol.2008.2 PMID:18779422

Dehay, B., Decressac, M., Bourdenx, M., Guadagnino, I., Fernagut, P.-O., Tamburrino, A., ... Bezard, E. (2016). Targeting α-synuclein: Therapeutic options. *Movement Disorders*, *31*(6), 882–888. doi:10.1002/mds.26568 PMID:26926119

Den Brok, M. G., van Dalen, J. W., van Gool, W. A., Moll van Charante, E. P., de Bie, R. M., & Richard, E. (2015). Apathy in Parkinson's disease: A systematic review and meta-analysis. *Movement Disorders*, *30*(6), 759–769. doi:10.1002/mds.26208 PMID:25787145

Dettmer, U., Newman, A. J., Soldner, F., Luth, E. S., Kim, N. C., von Saucken, V. E., ... Selkoe, D. (2015). Parkinson-causing α -synuclein missense mutations shift native tetramers to monomers as a mechanism for disease initiation. *Nature Communications*, *6*(1), 7314. doi:10.1038/ncomms8314 PMID:26076669

Dickson, D. W., Lin, W., Liu, W. K., & Yen, S. H. (1999). Multiple system atrophy: A sporadic synucleinopathy. *Brain Pathology (Zurich, Switzerland)*, 9(4), 721–732. doi:10.1111/j.1750-3639.1999.tb00553.x PMID:10517510

Dickson, D. W., Liu, W., Hardy, J., Farrer, M., Mehta, N., Uitti, R., ... Yen, S. H. (1999). Widespread alterations of alpha-synuclein in multiple system atrophy. *American Journal of Pathology*, *155*(4), 1241–1251. doi:10.1016/S0002-9440(10)65226-1 PMID:10514406

Alpha-Synucleinopathies

Donaghy, P. C., & McKeith, I. G. (2014). The clinical characteristics of dementia with Lewy bodies and a consideration of prodromal diagnosis. *Alzheimer's Research & Therapy*, *6*(4), 46. doi:10.1186/ alzrt274 PMID:25484925

Doty, R. L., Deems, D. A., & Stellar, S. (1988). Olfactory dysfunction in parkinsonism: A general deficit unrelated to neurologic signs, disease stage, or disease duration. *Neurology*, *38*(8), 1237–1244. doi:10.1212/WNL.38.8.1237 PMID:3399075

Dujardin, K., Defebvre, L., Krystkowiak, P., Degreef, J. F., & Destee, A. (2003). Executive function differences in multiple system atrophy and Parkinson's disease. *Parkinsonism & Related Disorders*, *9*(4), 205–211. doi:10.1016/S1353-8020(02)00050-0 PMID:12618055

Eidelberg, D., Sotrel, A., Joachim, C., Selkoe, D., Forman, A., Perl, D. P., & Pendlebury, W. W. (1987). Adult onset Hallervorden-Spatz disease with neurofibrillary pathology. A discrete clinico pathological entity. *Brain*, *110*(Pt 4), 993–1013. doi:10.1093/brain/110.4.993 PMID:2888513

Emre, M., Aarsland, D., Brown, R., Burn, D. J., Duyckaerts, C., Mizuno, Y., ... Dubois, B. (2007). Clinical diagnostic criteria for dementia associated with Parkinson's disease. *Movement Disorders*, 22(12), 1689–1707. doi:10.1002/mds.21507 PMID:17542011

Fonseca-Ornelas, L., Eisbach, S. E., Paulat, M., Giller, K., Fernández, C. O., Outeiro, T. F., ... Zweckstetter, M. (2014). Small molecule-mediated stabilization of vesicle-associated helical α-synuclein inhibits pathogenic misfolding and aggregation. *Nature Communications*, *5*, 5857. doi:10.1038/ncomms6857 PMID:25524885

Funayama, M., Ohe, K., Amo, T., Furuya, N., Yamaguchi, J., Saiki, S., ... Hattori, N. (2015). CHCHD2 mutations in autosomal dominant late-onset Parkinson's disease: A genome-wide linkage and sequencing study. *Lancet Neurology*, *14*(3), 274–282. doi:10.1016/S1474-4422(14)70266-2 PMID:25662902

Games, D., Valera, E., Spencer, B., Rockenstein, E., Mante, M., Adame, A., ... Masliah, E. (2014). Reducing C-terminal-truncated alpha-synuclein by immunotherapy attenuates neurodegeneration and propagation in Parkinson's disease-like models. *The Journal of Neuroscience*, *34*(28), 9441–9454. doi:10.1523/JNEUROSCI.5314-13.2014 PMID:25009275

Gilman, S., Wenning, G. K., Low, P. A., Brooks, D. J., Mathias, C. J., Trojanowski, J. Q., ... Vidailhet, M. (2008). Second consensus statement on the diagnosis of multiple system atrophy. *Neurology*, *71*(9), 670–676. doi:10.1212/01.wnl.0000324625.00404.15 PMID:18725592

Goetz, C. G., Emre, M., & Dubois, B. (2008). Parkinson's disease dementia: Definitions, guidelines, and research perspectives in diagnosis. *Annals of Neurology*, 64(S2), S81–S92. doi:10.1002/ana.21455 PMID:19127578

Goker-Alpan, O., Giasson, B. I., Eblan, M. J., Nguyen, J., Hurtig, H. I., Lee, V. M., ... Sidransky, E. (2006). Glucocerebrosidase mutations are an important risk factor for Lewy body disorders. *Neurology*, *67*(5), 908–910. doi:10.1212/01.wnl.0000230215.41296.18 PMID:16790605

Goldman, J. G., & Weintraub, D. (2015). Advances in the treatment of cognitive impairment in Parkinson's disease. *Movement Disorders*, *30*(11), 1471–1489. doi:10.1002/mds.26352 PMID:26297863 Goldman, J. G., Williams-Gray, C., Barker, R. A., Duda, J. E., & Galvin, J. E. (2014). The spectrum of cognitive impairment in Lewy body diseases. *Movement Disorders*, 29(5), 608–621. doi:10.1002/mds.25866 PMID:24757110

Gomperts, S. N. (2016). Lewy Body Dementias: Dementia With Lewy Bodies and Parkinson Disease Dementia. *Continuum (Minneapolis, Minn.)*, 22(2), 435–463. doi:10.1212/CON.00000000000000009 PMID:27042903

Grabowski, G. A. (2008). Phenotype, diagnosis, and treatment of Gaucher's disease. *Lancet*, 372(9645), 1263–1271. doi:10.1016/S0140-6736(08)61522-6 PMID:19094956

Graham, J. G., & Oppenheimer, D. R. (1969). Orthostatic hypotension and nicotine sensitivity in a case of multiple system atrophy. *Journal of Neurology, Neurosurgery, and Psychiatry*, *32*(1), 28–34. doi:10.1136/jnnp.32.1.28 PMID:5774131

Greggio, E., Taymans, J. M., Zhen, E. Y., Ryder, J., Vancraenenbroeck, R., Beilina, A., ... Cookson, M. R. (2009). The Parkinson's disease kinase LRRK2 autophosphorylates its GTPase domain at multiple sites. *Biochemical and Biophysical Research Communications*, *389*(3), 449–454. doi:10.1016/j. bbrc.2009.08.163 PMID:19733152

Hamano, K., Hayashi, M., Shioda, K., Fukatsu, R., & Mizutani, S. (2008). Mechanisms of neurodegeneration in mucopolysaccharidoses II and IIIB: Analysis of human brain tissue. *Acta Neuropathologica*, *115*(5), 547–559. doi:10.100700401-007-0325-3 PMID:18060551

Hernandez, D. G., Reed, X., & Singleton, A. B. (2016). Genetics in Parkinson disease: Mendelian vs. non-Mendelian inheritance. *Journal of Neurochemistry*, *139*(Suppl 1), 59–74. doi:10.1111/jnc.13593 PMID:27090875

Hogarth, P., Gregory, A., Kruer, M. C., Sanford, L., Wagoner, W., Natowicz, M. R., ... Hayflick, S. J. (2013). New NBIA subtype: Genetic, clinical, pathologic, and radiographic features of MPAN. *Neurology*, *80*(3), 268–275. doi:10.1212/WNL.0b013e31827e07be PMID:23269600

Iwai, A., Masliah, E., Yoshimoto, M., Ge, N., Flanagan, L., de Silva, H. A., ... Saitoh, T. (1995). The precursor protein of non-A beta component of Alzheimer's disease amyloid is a presynaptic protein of the central nervous system. *Neuron*, *14*(2), 467–475. doi:10.1016/0896-6273(95)90302-X PMID:7857654

Jellinger, K. A. (2009). A critical evaluation of current staging of alpha-synucleinpathology in Lewy body disorders. *Biochimica et Biophysica Acta*, *1792*(7), 730–734. doi:10.1016/j.bbadis.2008.07.006 PMID:18718530

Josephs, K. A., Matsumoto, J. Y., & Lindor, N. M. (2004). Heterozygous Niemann- Pick disease type C presenting with tremor. *Neurology*, *63*(11), 2189–2190. doi:10.1212/01.WNL.0000145710.25588.2F PMID:15596783

Kahle, P. J. (2008). alpha-Synucleinopathy models and human neuropathology: Similarities and differences. *Acta Neuropathologica*, *115*(1), 87–95. doi:10.100700401-007-0302-x PMID:17932682

Alpha-Synucleinopathies

Kalinderi, K., Bostantjopoulou, S., & Fidani, L. (2016). The genetic background of Parkinson's disease: Current progress and future prospects. *Acta Neurologica Scandinavica*, *134*(5), 314–326. doi:10.1111/ ane.12563 PMID:26869347

Kim, C., Ho, D. H., Suk, J. E., You, S., Michael, S., Kang, J., ... Lee, S. J. (2013). Neuron-released oligomeric α-synuclein is an endogenous agonist of TLR2 for paracrine activation of microglia. *Nature Communications*, *4*, 1562. doi:10.1038/ncomms2534 PMID:23463005

Kumar, K. R., Djarmati-Westenberger, A., & Grunewald, A. (2011). Genetics of Parkinson's disease. *Seminars in Neurology*, *31*(5), 433–440. doi:10.1055-0031-1299782 PMID:22266881

Kupsch, A., Kuehn, A., Klaffke, S., Meissner, W., Harnack, D., Winter, C., ... Trottenberg, T. (2003). Deep brain stimulation in dystonia. *Journal of Neurology*, *250*(S1Suppl 1), I47–I52. doi:10.100700415-003-1110-2 PMID:12761637

Lee, D. W., & Andersen, J. K. (2006). Role of HIF-1 in iron regulation: Potential therapeutic strategy for neurodegenerative disorders. *Current Molecular Medicine*, 6(8), 883–893. doi:10.2174/156652406779010849 PMID:17168739

Lee, H. J., Suk, J. E., Lee, K. W., Park, S. H., Blumbergs, P. C., Gai, W. P., & Lee, S. J. (2011). Transmission of synucleinopathies in the enteric nervous system of A53T alpha-synuclein transgenic mice. *Experimental Neurobiology*, 20(4), 181–188. doi:10.5607/en.2011.20.4.181 PMID:22355263

Lee, J. S., & Lee, S. J. (2016). Mechanism of Anti-α-Synuclein Immunotherapy. *Journal of Movement Disorders*, 9(1), 14–19. doi:10.14802/jmd.15059 PMID:26828212

Lees, A. J., Hardy, J., & Revesz, T. (2009). Parkinson's disease. *Lancet*, 373(9680), 2055–2066. doi:10.1016/S0140-6736(09)60492-X PMID:19524782

Leroy, E., Boyer, R., Auburger, G., Leube, B., Ulm, G., Mezey, E., ... Polymeropoulos, M. H. (1998). The ubiquitin pathway in Parkinson's disease. *Nature*, *395*(6701), 451–452. doi:10.1038/26652 PMID:9774100

Lesage, S., Drouet, V., Majounie, E., Deramecourt, V., Jacoupy, M., Nicolas, A., ... Brice, A. (2016). Loss of VPS13C function in autosomal-recessive parkinsonism causes mitochondrial dysfunction and increases PINK1/Parkin-dependent mitophagy. *American Journal of Human Genetics*, *98*(3), 500–513. doi:10.1016/j.ajhg.2016.01.014 PMID:26942284

Lewis, J., Melrose, H., Bumcrot, D., Hope, A., Zehr, C., Lincoln, S., ... Farrer, M. J. (2008). In vivo silencing of alpha-synuclein using naked siRNA. *Molecular Neurodegeneration*, *3*(1), 19. doi:10.1186/1750-1326-3-19 PMID:18976489

Li, Y., Dunn, L., Greggio, E., Krumm, B., Jackson, G. S., Cookson, M. R., ... Deng, J. (2009). The R1441C mutation alters the folding properties of the ROC domain of LRRK2. *Biochimica et Biophysica Acta*, *1792*(12), 1194–1197. doi:10.1016/j.bbadis.2009.09.010 PMID:19781641

Lierberman, A. (2006). Are dementia and depression in Parkinson's disease related? *Journal of the Neurological Sciences*, 248(4), 138–142. doi:10.1016/j.jns.2006.05.022 PMID:16814323

Lin, C. H., Wu, R. M., Chang, H. Y., Chiang, Y. T., & Lin, H. H. (2013). Preceding pain symptoms and Parkinson's disease: A nationwide population-based cohort study. *European Journal of Neurology*, 20(10), 1398–1404. doi:10.1111/ene.12197 PMID:23679105

Lindström, V., Fagerqvist, T., Nordström, E., Eriksson, F., Lord, A., Tucker, S., ... Ingelsson, M. (2014). Immunotherapy targeting α-synuclein protofibrils reduced pathology in (Thy-1)-h[A30P] α-synuclein mice. *Neurobiology of Disease*, 69, 134–143. doi:10.1016/j.nbd.2014.05.009 PMID:24851801

Liu, Y., Fallon, L., Lashuel, H. A., Liu, Z., & Lansbury, P. T. Jr. (2002). The UCHL1 gene encodes two opposing enzymatic activities that affect alpha-synuclein degradation and Parkinson's disease susceptibility. *Cell*, *111*(2), 209–218. doi:10.1016/S0092-8674(02)01012-7 PMID:12408865

Mak, C. M., Sheng, B., Lee, H. H., Lau, K. K., Chan, W. T., Lam, C. W., & Chan, Y. W. (2011). Youngonset parkinsonism in a Hong Kong Chinese man with adult-onset Hallervorden-Spatz syndrome. *The International Journal of Neuroscience*, *121*(4), 224–227. doi:10.3109/00207454.2010.542843 PMID:21198414

Mandler, M., Valera, E., Rockenstein, E., Weninger, H., Patrick, C., Adame, A., ... Masliah, E. (2014). Next-generation active immunization approach for synucleinopathies: Implications for Parkinson's disease clinical trials. *Acta Neuropathologica*, *127*(6), 861–879. doi:10.100700401-014-1256-4 PMID:24525765

Maroteaux, L., Campanelli, J. T., & Scheller, R. H. (1988). Synuclein: A neuron-specific protein localized to the nucleus and presynaptic nerve terminal. *The Journal of Neuroscience*, *8*, 2804–2815. PMID:3411354

Masliah, E., Rockenstein, E., Adame, A., Alford, M., Crews, L., Hashimoto, M., ... Schenk, D. (2005). Effects of alpha-synuclein immunization in a mouse model of Parkinson's disease. *Neuron*, *46*(6), 857–868. doi:10.1016/j.neuron.2005.05.010 PMID:15953415

Mazzulli, J. R., Xu, Y. H., Sun, Y., Knight, A. L., McLean, P. J., Caldwell, G. A., ... Krainc, D. (2011). Gaucher disease glucocerebrosidase and α -synuclein form a bidirectional pathogenic loop in synucleinopathies. *Cell*, *146*(1), 37–52. doi:10.1016/j.cell.2011.06.001 PMID:21700325

McKeith, I. G., Dickson, D. W., Lowe, J., Emre, M., O'Brien, J. T., Feldman, H., ... Yamada, M. (2005). Diagnosis and management of dementia with Lewy bodies: Third report of the DLB Consortium. *Neurology*, *65*(12), 1863–1872. doi:10.1212/01.wnl.0000187889.17253.b1 PMID:16237129

Monchi, O., Petrides, M., Mejia-Constain, B., & Strafella, A. P. (2007). Cortical activity in Parkinson's disease during executive processing depends on striatal involvement. *Brain*, *130*(1), 233–244. doi:10.1093/brain/awl326 PMID:17121746

Morris, C. M., Candy, J. M., Oakley, A. E., Bloxham, C. A., & Edwardson, J. A. (1992). Histochemical distribution of non-haem iron in the human brain. *Acta Anatomica*, 144(3), 235–257. doi:10.1159/000147312 PMID:1529678

Nègre-Pagès, L., Regragui, W., Bouhassira, D., Grandjean, H., & Rascol, O. (2008). Chronic pain in Parkinson's disease: The cross-sectional French DoPaMiP survey. *Movement Disorders*, 23(10), 1361–1369. doi:10.1002/mds.22142 PMID:18546344 Neudorfer, O., Giladi, N., Elstein, D., Abrahamov, A., Turezkite, T., Aghai, E., ... Zimran, A. (1996). Occurrence of Parkinson's syndrome in type I Gaucher disease. *QJM: an International Journal of Medicine*, 89(9), 691–694. doi:10.1093/qjmed/89.9.691 PMID:8917744

Okazaki, H., Lipkin, L. E., & Aronson, S. M. (1961). Diffuse intracytoplasmic ganglionic inclusions (Lewy type) associated with progressive dementia and quadriparesis in flexion. *Journal of Neuropathology and Experimental Neurology*, 20(2), 237–244. doi:10.1097/00005072-196104000-00007 PMID:13730588

Parkinson, J. (1817). An Essay on the Shaking Palsy. London: Sherwood, Neely, and Jones.

Pfeiffer, R. F. (2007). Multiple system atrophy. *Handbook of Clinical Neurology*, 84, 305–326. doi:10.1016/S0072-9752(07)84046-2 PMID:18808955

Pigott, K., Rick, J., Xie, S. X., Hurtig, H., Chen-Plotkin, A., Duda, J. E., ... Weintraub, D. (2015). Longitudinal study of normal cognition in Parkinson disease. *Neurology*, *85*(15), 1276–1282. doi:10.1212/ WNL.0000000000000002001 PMID:26362285

Poewe, W. (2008). Non-motor symptoms in Parkinson's disease. *European Journal of Neurology*, 15(s1), 14–20. doi:10.1111/j.1468-1331.2008.02056.x PMID:18353132

Poewe, W., & Mahlknecht, P. (2009). The clinical progression of Parkinson's disease. *Parkinsonism & Related Disorders*, *15*, S28–S32. doi:10.1016/S1353-8020(09)70831-4 PMID:20123553

Polymeropoulos, M. H., Lavedan, C., Leroy, E., Ide, S. E., Dehejia, A., Dutra, A., ... Nussbaum, R. L. (1997). Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. *Science*, 276(5321), 2045–2047. doi:10.1126cience.276.5321.2045 PMID:9197268

Pont-Sunyer, C., Hotter, A., Gaig, C., Seppi, K., Compta, Y., Katzenschlager, R., ... Tolosa, E. (2015). The Onset of Nonmotor Symptoms in Parkinson's disease (The ONSET PD Study). *Movement Disorders*, *30*(2), 229–237. doi:10.1002/mds.26077 PMID:25449044

Postuma, R. B., Berg, D., Stern, M., Poewe, W., Olanow, C. W., Oertel, W., ... Deuschl, G. (2015). MDS clinical diagnostic criteria for Parkinson's disease. *Movement Disorders*, *30*(12), 1591–1601. doi:10.1002/mds.26424 PMID:26474316

Rajput, A. H., Rozdilsky, B., & Rajput, A. (1991). Accuracy of clinical diagnosis in Parkinsonism – a prospective study. *The Canadian Journal of Neurological Sciences*, *18*(3), 275–278. doi:10.1017/S0317167100031814 PMID:1913360

Reijnders, J. S., Scholtissen, B., Weber, W. E., Aalten, P., Verhey, F. R., & Leentjens, A. F. (2010). Neuroanatomical correlates of apathy in Parkinson's disease: A magnetic resonance imaging study using voxel-based morphometry. *Movement Disorders*, 25(14), 2318–2325. doi:10.1002/mds.23268 PMID:20669264

Remy, P., Doder, M., Lees, A., Turjanski, N., & Brooks, D. (2005). Depression in Parkinson's disease: Loss of dopamine and noradrenaline innervation in the limbic system. *Brain*, *128*(6), 1314–1322. doi:10.1093/brain/awh445 PMID:15716302

Salat, D., Noyce, A. J., Schrag, A., & Tolosa, E. (2016). Challenges of modifying disease progression in prediagnostic Parkinson's disease. *Lancet Neurology*, 1–12. PMID:26993435

Schenk, D. B., Koller, M., Ness, D. K., Griffith, S. G., Grundman, M., Zago, W., ... Kinney, G. G. (2017). First-in-human assessment of PRX002, an anti– α -synuclein monoclonal antibody, in healthy volunteers. *Movement Disorders*, *32*(2), 211–218. doi:10.1002/mds.26878 PMID:27886407

Scheperjans, F., Aho, V., Pereira, P. A., Koskinen, K., Paulin, L., Pekkonen, E., ... Auvinen, P. (2015, March). Gut microbiota are related to Parkinson's disease and clinical phenotype. *Movement Disorders*, *30*(3), 350–358. doi:10.1002/mds.26069 PMID:25476529

Schneeberger, A., Tierney, L., & Mandler, M. (2016). Active immunization therapies for Parkinson's disease and multiple system atrophy. *Movement Disorders*, *31*(2), 214–224. doi:10.1002/mds.26377 PMID:26260853

Selkoe, D. (1990). Amyloid β -protein deposition as a seminal pathogenic event in AD: An hypothesis. *Neurobiology of Aging*, *11*(3), 299. doi:10.1016/0197-4580(90)90746-M

Seppi, K., Weintraub, D., Coelho, M., Perez-Lloret, S., Fox, S. H., Katzenxchlager, R., ... Sampaio, C. (2011). The Movement Disorder Society Evidence-Based Medicine Review Update: Treatments for the non-Motor symptoms of Parkinson's disease. *Movement Disorders*, *26*(3), 42–80. doi:10.1002/mds.23884 PMID:22021174

Settembre, C., Fraldi, A., Jahreiss, L., Spampanato, C., Venturi, C., Medina, D., ... Ballabio, A. (2008). A block of autophagy in lysosomal storage disorders. *Human Molecular Genetics*, *17*(1), 119–129. doi:10.1093/hmg/ddm289 PMID:17913701

Shachar, T., Lo-Bianco, C., Recchia, A., Wiessner, C., Raas-Rosthschild, A., & Futerman, A. H. (2011). Lysosomal Storage Disorders and Parkinson's Disease: Gaucher Disease and Beyond. *Movement Disorders*, 29(9), 1593–1604. doi:10.1002/mds.23774 PMID:21618611

Simuni, T., Luo, S. T., Chou, K. L., Fernandez, H., He, B., & Parashos, S. (2013). Rankin scale as a potencial measure of global disability in early Parkinson's disease. *Journal of Clinical Neuroscience*, 20(9), 1200–1203. doi:10.1016/j.jocn.2012.10.030 PMID:23810387

Spillantini, M. G., Schmidt, M. L., Lee, V. M., Trojanowski, J. Q., Jakes, R., & Goedert, M. (1997). Alpha-synuclein in Lewy bodies. *Nature*, *388*(6645), 839–840. doi:10.1038/42166 PMID:9278044

Suzuki, K., Iseki, E., Katsuse, O., Yamaguchi, A., Katsuyama, K., Aoki, I., ... Kosaka, K. (2003). Neuronal accumulation of alpha- and beta-synucleins in the brain of a GM2 gangliosidosis mouse model. *Neuroreport*, *14*(4), 551–554. doi:10.1097/00001756-200303240-00004 PMID:12657883

Suzuki, K., Iseki, E., Togo, T., Yamaguchi, A., Katsuse, O., Katsuyama, K., ... Hirayasy, Y. (2007). Neuronal and glial accumulation of alpha- and beta-synucleins in human lipidoses. *Acta Neuropathologica*, *114*(5), 481–489. doi:10.100700401-007-0264-z PMID:17653558

Tran, H. T., Chung, C. H., Iba, M., Zhang, B., Trojanowski, J. Q., Luk, K. C., & Lee, V. M. Y. (2014). A-synuclein immunotherapy blocks uptake and templated propagation of misfolded α-synuclein and neurodegeneration. *Cell Reports*, 7(6), 2054–2065. doi:10.1016/j.celrep.2014.05.033 PMID:24931606

Ubhi, K., Low, P., & Masliah, E. (2011). Multiple System Atrophy: A Clinical and Neuropathological Perspective. *Trends in Neurosciences*, *34*(11), 581–590. doi:10.1016/j.tins.2011.08.003 PMID:21962754

Alpha-Synucleinopathies

Valera, E., Compagnoni, G. M., & Masliah, E. (2016, February). (2016-a). Novel treatment strategies targeting alpha-synuclein in multiple system atrophy as a model of synucleinopathy. *Neuropathology and Applied Neurobiology*, *42*(1), 95–106. doi:10.1111/nan.12312 PMID:26924723

Valera, E., Spencer, B., & Masliah, E. (2016, January). (2016-b). Immunotherapeutic Approaches Targeting Amyloid- β , α -Synuclein, and Tau for the Treatment of Neurodegenerative Disorders. *Neurotherapeutics; the Journal of the American Society for Experimental NeuroTherapeutics*, *13*(1), 179–189. doi:10.100713311-015-0397-z PMID:26494242

Vitner, E. B., Platt, F. M., & Futerman, A. H. (2010). Common and uncommon pathogenic cascades in lysosomal storage diseases. *The Journal of Biological Chemistry*, 285(27), 20423–20427. doi:10.1074/jbc.R110.134452 PMID:20430897

Vonsattel, J. P., Keller, C., & Cortes Ramirez, E. P. (2011). Huntington's disease – neuropathology. *Handbook of Clinical Neurology*, *100*, 83–100. doi:10.1016/B978-0-444-52014-2.00004-5 PMID:21496571

Wenning, G. K., Ben Shlomo, Y., Magalhaes, M., Daniel, S. E., & Quinn, N. P. (1994). Clinical features and natural history of multiple system atrophy. An analysis of 100 cases. *Brain*, *117*(Pt4), 835–845. doi:10.1093/brain/117.4.835 PMID:7922469

Wenning, G. K., Seppi, K., Scherfler, C., Stefanova, N., & Puschban, Z. (2001). Multiple system atrophy. *Seminars in Neurology*, *21*(01), 33–40. doi:10.1055-2001-13117 PMID:11346023

Withers, G. S., George, J. M., Banker, G. A., & Clayton, D. F. (1997). Delayed localization of synelfin (synuclein, NACP) to presynaptic terminals in cultured rat hippocampal neurons. *Brain Research*. *Developmental Brain Research*, 99(1), 87–94. doi:10.1016/S0165-3806(96)00210-6 PMID:9088569

Wong, K., Sidransky, E., Verma, A., Mixon, T., Sandberg, G. D., Wakefield, L. K., ... Schiffmann, R. (2004). Neuropathology provides clues to the pathophysiology of Gaucher disease. *Molecular Genetics and Metabolism*, 82(3), 192–207. doi:10.1016/j.ymgme.2004.04.011 PMID:15234332

Wong, Y. C., & Krainc, D. (2017). α-synuclein toxicity in neurodegeneration: Mechanism and therapeutic strategies. *Nature Medicine*, 23(2), 1–13. doi:10.1038/nm.4269 PMID:28170377

Wray, S., Arber, C. E., Li, A., & Houlden, H. (2016). Insights into molecular mechanisms of disease in Neurodegeneration with Brain Iron Accumulation, unifying theories. *Neuropathology and Applied Neurobiology*, *42*(3), 220–241. doi:10.1111/nan.12242 PMID:25870938

Yabe, I., Soma, H., Takei, A., Fujiki, N., Yanagihara, H., & Sasaki, H. (2006). MSA-C is the predominant clinical phenotype of MSA in Japan, analysis of 142 patients with probable MSA. *Journal of the Neurological Sciences*, *249*(2), 115–121. doi:10.1016/j.jns.2006.05.064 PMID:16828805

Zaccai, J., McCracken, C., & Brayne, C. (2005). A systematic review of prevalence and incidence studies of dementia with Lewy bodies. *Age and Ageing*, *34*(6), 561–566. doi:10.1093/ageing/afi190 PMID:16267179

Chapter 14 Lewy Body Disease: Point Towards Progressive Dementia

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ABSTRACT

Fritz Heinrich Lewy described the intracytoplasmic inclusions found in the neurons for the very first time. In 1919 these inclusions were termed as "LBs" by Tretiakoff. LBs were found in the brain of the patients suffering from Lewy body disease (LBD). LBD is characterized by the presence of Parkinsonian symptoms in the earlier stages and dementia in the later stages of the disease. LBs were classified on the basis of the region of the brain in which they are distributed and so is the case of the LBD means the type of the LBD depends on the anatomical areas of the brain involved. LBD is not a single disorder. It is a spectrum of disorders. This chapter addresses the entire profile of LBs, types, composition, formation, and various LB pathologies as well as diagnostic criteria and pharmacotherapy.

INTRODUCTION

Lewy bodies (LBs) are the eosinophilic intracytoplasmic inclusions found in the brain of the patients of Parkinson's disease (PD) and were first described by Fritz Heinrich Lewy. These inclusions were later called "LBs" by Tretiakoff in 1919 (Wakabayashi et al, 2007). The presence of LBs in the brain of PD patients suggesting the possible involvement of LBs in the process of neurodegeneration, but the recent reports suggested that the LBs formation is a response evoked by the body to scavenge or degrade the neurotoxic substances that is responsible for neuronal degeneration (Sathiyamoorthy et al., 2014). LBs occur in context of various distinct pathologies known as Lewy body disease (LBD) characterized by the presence of the parkinsonian symptoms in earlier stages and dementia in the later stages of disease. Various symptoms of LBD include parkinsonism, hallucination, cognitive impairment, fluctuations in mental status, and dementia. Different types of LBD shares some common characteristics however the marked symptomatic difference found in these pathologies is mainly due to the difference in anatomi-

DOI: 10.4018/978-1-5225-5282-6.ch014

cal region of the brain involved (in which the LBs formation occurs) (Kosaka, 2014). Further each of these pathologies is characterized by the presence of LBs, therefore the symptoms appears very similar and sometimes overlaps making the exact pathology difficult to diagnose. Thus the exact pathology so involved sometimes remains undiagnosed and often misdiagnosed (Galvin & Balasubramaniam, 2013) which often becomes troublesome to the patients and caregivers (Galvin et al, 2010). Therefore very precise diagnostic criteria's is required to avoid the false diagnosis and the wrong prescription that may prove fatal later. The difference in diagnosis largely depends on the order of symptomatic presentation such that if only the movement deficits are present then the person is said to be suffering from PD, the development of dementia develops before the onset of the PD or within one year of the PD then it is referred as dementia with LBs (DLB) (Galvin & Balasubramaniam, 2013). In the present chapter authors demonstrate LBD with their similar and differentiating characteristics and provides several recommendations that will prove beneficial in the diagnosis and the pharmacotherapeutics of such pathologies.

BACKGROUND

LBs are the intracytoplasmic inclusions found in the neurons of the patients suffering from LBD, however the exact type of the LBD which a patients exactly suffers from depends upon the region of brain in which the LBs were found predominantly (Kalra et al, 1996). Brainstem nuclei involvement is almost universal in the LBD while the involvement of limbic region and neocortex suggested the progression of disease (Braak et al, 2003). Further it is suggested that the brainstem is involved in the early stages of LBs pathologies whereas the different region of brain also involved when the disease progresses or in the later stages of the disease (Adler et al, 2010). LBs pathologies or the LBD represent a family of the disease that involved three distinct pathologies including PD, PDD and DLB. Before going in to the depth involved in the pathogenesis of these pathologies it remains of interest how LBs occurs and what is the reason of LBs formation. Recent reports suggested that the dopaminergic neurons isolated from the healthy individuals when implanted in the brain of the PD patients form LBs (Kordower et al, 2008; Li et al, 2008) suggesting LBs formations occurs to counteract the pathological insult. Thus LBs formation represents an epiphenomenon, or the scars of neurodegeneration (Popescu et al, 2004). In the present work authors demonstrate the different types of LBs, their formation and composition and the occurrence of LBs in various LBD.

CLASSIFICATION OF LEWY BODIES

LBs can be classified on the basis of their distribution in the different regions of brain.

Type 1 Lewy Bodies

Type 1 LBs are also known as brainstem LBs because of the predominant involvement of the brainstem nuclei (Braak & Braak, 2000; Dickson et al, 1987; 1996; Kosaka et al, 1984). Type 1 LBs are spherical in shape containing eosinophilic cores, lamellar bands and pale halos (Campbell et al, 2001).

Type 2 Lewy Bodies

Type 2 LBs are also known as cortical LBs and characterized by the major involvement of limbic and temporal region and lesser involvement of frontal and posterior cortical region of brain (Braak & Braak, 2000; Braak et al, 2003; Kosaka, 1990; Kosaka et al, 1984). Type 2 LBs are less eosinophilic and less circumscribed (Dickson et al, 1987) and typically lack pale halos (Gomez-Tortosa et al, 2000).

COMPOSITION OF LEWY BODIES

Alpha-synuclein (α -syn) is the major component of LBs (Spillantini et al, 1997). α -syn is a small acidic protein comprises of three domains: N-terminal lipid-binding domain, responsible for the helical structure and the interaction of α -syn with the lipid membranes (Sode et al, 2007); Amyloid-binding central domain (NAC), responsible for the aggregation of α -syn (Rajagopalan & Andersen, 2001) and C-terminal acidic tail, this domain inhibit the aggregation of α -syn (Kim et al, 2004). It has been reported that the mutations in the SNCA gene increase the chances of α -syn aggregation (Minton, 2005) and the aggregation of α -syn is responsible α -synucleinopathies (Spillantini et al, 1997). Aggregation of α -syn monomers leads to the formation of oligomers (Walsh et al, 1997) responsible for neuroinflammation, and neurodegeneration (Wolozin & Behl, 2000) by disrupting proteostasis (Hinault et al, 2010); opening of "mitochondrial permeability transition pore" (mPTP) (Luth et al, 2014) and disrupting mitochondrial respiration (Nakamura, 2013). Oligomers also acts a seeds to promote seeding in which the soluble oligomers acts as the nucleus and form the large insoluble aggregates form known as amyloid fibrils (Eichner & Radford, 2011) that further promotes neuronal death (Jucker & Walker, 2011).

GENESIS OF LEWY BODIES

The precise sequence of events involved in the formation of LBs is completely not known. But it is reported that the process of LBs formation begin with the aggregation of α -syn (Campbell et al, 2001) followed by the incorporation of pale bodies (Gomez-Tortosa et al, 2000) further followed by the incorporation of ubiquitin (Engelender, 2008) and the accumulation of additional proteins (Kuusisto et al, 2003; Orosz et al, 2004). It is reported that LBs distortion often followed by the reduced immunoreactivity (Katsuse et al, 2003).

LEWY BODY DISEASE: PARKINSON'S DISEASE, PARKINSON'S DISEASE DEMENTIA AND DEMENTIA WITH LEWY BODIES

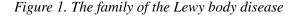
LBD often involves three distinct pathologies including PD, PDD and DLB suggesting LBD a generic term (Figure 1). LBD can be classified into three types (Type A, Type B, and Type C) according to the regional distributional of different LBs in the different regions of brain (Kosaka et al, 1984). Type A referred as brain stem type LBD; Type B referred as transitional type LBD and Type C referred as diffuse type LBD. In brainstem type, LBs occurs in the brainstem nuclei and diencephalon, this is often seen in the PD pathologies (Braak et al, 2003). In DLBD type, LBs are found in the cerebral cortex and

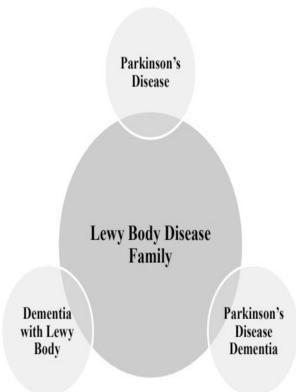
Lewy Body Disease

amygdale showing the rarely presence of LBs in brainstem and diencephalon nuclei thus the mesocortical dopaminergic system remains intact and dementia precedes parkinsonism as seen in DLBD (Kosaka et al, 1996). Further DLBD is a neurodegenerative disorder that occur either in common form (cognitive impairment followed by parkinsonism) or pure form (parkinsonism followed by dementia) (Kosaka, 2014); characterized by the symptoms including dementia, cognitive impairments, hallucinations, and parkinsonism (Zesiewicz et al, 2001). Transitional type is the transitional between brainstem type and diffuse LBD.

However besides these three forms of LBD there is another form of LBD which is known as incidental LBD (iLBD). LBs have been found in the patients suffering from the PD. However recent findings suggested LBs were found to be present in the brain of approx. 10% of the normal individuals above the age of 60 years, presenting a unique condition known as ILBD (Dickson et al, 2008). ILBD thus represents a condition either prior to or an early/intermediate stage leading to PD or DLB (Adler et al, 2010). Since the distribution of LBs in the brain in ILBD is similar to the PD, but the differentiating feature is that in case of ILBD there is no damage or loss of dopaminergic neurons. ILBD is thus considered as preclinical PD (Frigerio et al, 2011) but it remains to be clarified whether ILBD is pre-symptomatic PD or it is an age-related change which is unrelated to PD (Dickson et al, 2008).

LBD is not a distinct disorder but a continuum of disorders including disturbances of movement, cognition, behavior, sleep and autonomic function (Figure 2). PD, PDD and DLB, these pathologies are





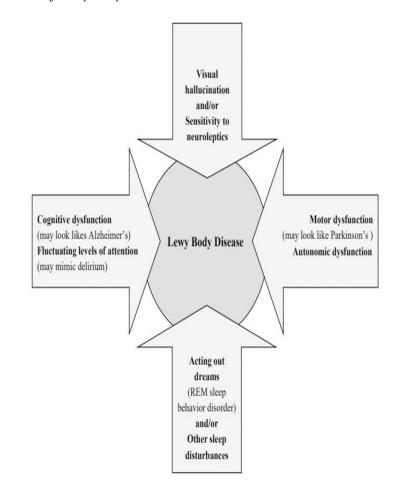


Figure 2. The spectrum of Lewy body disease

distinct actually but their symptoms overlap each other. In this section we describe each pathology and its characteristics in detail (Table 1).

Parkinson's Disease

The death of the dopaminergic neurons in the substantia nigra (SN) region of brain results in the pathology known as Parkinson's disease (PD). Since PD is characterized by the degeneration or the death or the damage of the dopaminergic neurons, however what is exact reason behind the damage of these neurons in the PD still remains unknown (Walia et al., 2014). The death of the dopaminergic neurons in the SN region give rise to the deficiency of the dopamine that further results in the motor symptoms of PD. Several genes have been targeted in context of the PD and these includes SNCA gene (Chartier-Harlin et al, 2004; Forno, 1996; Kruger et al, 1998; Nishioka et al, 2006; Polymeropoulos et al, 1997; Polymeropoulos et al, 1997; Singleton et al, 2003; Zarranz et al, 2004), ubiquitin C-terminal hydrolase L1 (UCH-L1), leucine-rich kinase 2 (LRRK 2), HtrA2/Omi DJ-1, PTEN-induced putative kinase 1

S. No.	Parkinson's Disease	Parkinson's Disease Dementia	Dementia with Lewy Bodies
1	PD results in the motor dysfunction.	PDD results in the motor and cognitive dysfunction.	DLB results in the motor and cognitive dysfunctions
2	Dementia may or may not be present in PD.	In PDD dementia develops more than one year after the diagnosis of PD.	In DLB dementia develops before or within one year of the PD diagnosis.
3	Executive difficulties in occurs in PD.	Visuospatial functioning and semantic memory deficits occurs in PDD.	Attentional deficits, executive and visuospatial deficits occur in DLB.
6	Resting tremor and rigidity are common.	Non-tremor dominant motor symptoms are present in PDD.	Resting tremor is uncommon, facial masking and axial rigidity occur in DLB.

Table 1. Differences amid Parkinson's disease, Parkinson's disease dementia and dementia with Lewy bodies

(PINK1) and parkin (PRKN) (Gosal et al, 2006; Hattori & Mizuno, 2004; Hedrich et al, 2004; Hernandez et al, 2005; Maraganore et al, 2004; Marx et al, 2003; Nishikawa et al, 2003; Strauss et al, 2005). It is suggested that the mutation in the SNCA gene results in the aggregation of the α -syn, which is the main component of the LBs. LBs formation has been reported in the brain of the PD patients. It is suggested that the LBs remain confined to the brainstem nuclei in the stage 1-2, progresses to the SN region in stage 3-4 and finally reaches to neocortex in the latter stages of PD or Stages 5-6 (for detail please refer Braak et al, 2002; 2003). It is suggested that the presence of LBs in the brain of PD patients represents the sign of neuronal degeneration but do not tell anything about the exact sequence of degenerative events, however the LBs formation nowadays is considered as the response of the body to counteracts the neuronal insults (Popescu et al, 2004).

Parkinson's Disease Dementia

The development of dementia more than one year after the PD onset is known as Parkinson's disease dementia (PDD) (McKeith et al, 2006) characterized by the presence of LBs in the cortical and limbic region of the brain (Irwin et al, 2012). Thus PDD presents the condition in which the motor symptom develops one year before the cognitive impairments (Kurtz & Kaufer, 2011). The prevalence rate for PDD in the population above 65 years is about 0.2%-0.5% (Aarsland et al, 2005a; 2007), suggesting aging accelerates the development of dementia, cognitive deficits and increase the severity of motor disturbances (Levy et al, 2002). The presence of PD further increased the risk of developing dementia (Aarsland et al, 2003; Emre et al, 2007; Levy et al, 2002; Williams-Gray et al, 2009) and about 80% of the patients suffering from PD pathology develops dementia (Aarsland et al, 2003; Halliday et al, 2008; Hely et al, 2008). The main difference between PD and PDD is that the PD is characterized by the resting tremor greatly, while PDD is non-tremor-dominant motor subtype (Aarsland et al, 2003; Jankovic et al, 1990; Jellinger et al, 2002; Selikhova et al, 2009). Presence of the cognitive impairment often worsens the motor dysfunction associated with PD (Braak et al, 2005) and increase the disability, morbidity and mortality in the PD patients (Lo et al, 2009; Rosenthal et al, 2010). Also the patients with PDD did not represents the intrinsic core language deficit (Emre et al, 2007) but the difficulties encounters by the patients might be due to the executive dysfunctioning (Grossman et al, 2012).

Dementia With Lewy Bodies

DLB is a neurodegenerative disorder arises due to cortical and subcortical impairments that results in the development of attentional, executive and visuospatial dysfunction (Manning, 2004). The core features include dementia, parkinsonism, fluctuating cognition and hallucinations. Fluctuating cognition includes excessive daytime drowsiness, marked confusion and disorganized speech. Parkinsonism is usually bilateral, resting tremor is uncommon, while axial rigidity and facial masking is often more prominent in DLB. Visual hallucinations in DLB are often powerful, strong, bright, clear and repetitive (McKeith et al, 2005). The recurrent hallucination in DLB might be due to the burden of LBs in the anterior and temporal region of the brain (Harding et al, 2002). Rapid eye movement (REM) disturbance, a suggestive clinical feature of DLB often precedes cognitive or motor symptoms. REM sleep behavioral disturbances are characterized by the intense dreams that might harm the patients since the dream carries the destructive elements, the patients often starts screaming or fighting while sleeping that might results in the injuries (Ferman et al, 2002). Additional supportive features include repeated falls, syncope, unconsciousness, hallucinations, delusions and autonomic dysfunction (McKeith et al, 2005). In DLB, LBs formation has been noticed in the different regions of the brain including brainstem, limbic, neocortex and brain stem (Jellinger, 2009) however why limbic areas are selectively vulnerable to LBs pathology still remains unclear (Rezaie et al, 1996).

PATHOGENESIS OF LEWY BODY DISEASE

PD is disease associated with the defects in the various regions of basal ganglia (BG) resulting in movement deficits (Rubin et al, 2012). Striatum is the receptive nucleus of BG that receives afferent projections from neocortex, thalamus and SNc (Rubin et al, 2012) and sends inhibitory projection to direct pathway and indirect pathway. In direct pathway striatum send projections to globus pallidus interna (GPi)/SN which in turn send projection to thalamus and in indirect pathway striatum send projections to GPe, which then send projection to STN, which further send projection to GPi/SNr. Direct pathway of BG initiate movement and the indirect pathway of BG suppress movements (Kreitzer & Malenka, 2008). The balance between direct and indirect pathway is essential for the normal movements and functioning (Sperlagh & Vizi, 2011). Direct pathway is modulated by D_1 receptors while indirect pathway is modulated by D_2 receptors (Fuentes et al, 2010). Since PD characterized by the death of the dopaminergic neurons often resulting in the dopaminergic deficits and the alteration in the functioning of direct and indirect pathway. It is reported that in PD patients the activity of direct pathway decreases while the activity of indirect pathway increases (Gerfen & Surmeier, 2011; Kreitzer & Malenka, 2008) resulting in the difficulty in initiating movement and excessive inhibition of movement in PD.

The occurrence of LBs in the brainstem nuclei is the characteristic feature of PD (Braak et al, 2003). In PD, LBs has also been found in the different cranial nerve nuclei including premotor oculomotor, precerebellar and vestibular brainstem nuclei (Seidel et al, 2015). Further the presence of LBs in the cortical region is the characteristics of dementia (Duda et al, 2002; Tsuboi et al, 2007). PPN is a brainstem locomotor center, maintain balance and therefore damage to it results in the impaired gait and posture control. In PD about 50% population of the cholinergic neurons of PPN degenerate (Gai et al, 1991; Hirsch et al, 1987; Jellinger, 1988; Karachi et al, 2010; Zweig et al, 1989) resulting in the impaired postural control and gait and DA-resistant akinesia in PD (Stein, 2009).

Lewy Body Disease

Since PD involves the degeneration of dopaminergic neurons in the brain but Frederic Lewy first identified LBs in the nbM (Holdorff, 2006). nbM is an open nucleus of basal forebrain characterized by extensive neuronal loss in PD pathology (Arendt et al, 1983; Candy et al, 1983; Chui et al, 1986; Perry et al, 1985) suggesting cholinergic deficits results in the cognitive impairment observed in the patients of PD (Kehagia et al, 2013). This cholinergic neuronal loss even become more prominent in the presence of dementia (Candy et al, 1983; Nakano and Hirano, 1984; Rogers et al, 1985; Tagliavini et al, 1984; Whitehouse et al, 1983) suggesting a greater impairment in the PDD patients. Further the dopaminergic neuronal destruction in the PD occurs concurrently with accumulation of α -syn in the cholinergic neurons of forebrain (BF) (Braak et al, 2003). It is reported that the denervation of cholinergic neurons of probabile marker of slowing of gait observed in the patients of PD (Bohnen et al, 2012) that might be due to the possible decrease in the cognitive processing abilities during ambulation (Woollacott & Shumway-Cook, 2002).

PDD refers to the development of dementia more than one year after the onset of PD pathology (Emre et al, 2007). The cognitive picture of PDD is commonly considered as a "subcortical" type and patients typically presents executive dysfunctions (Chui et al, 1986; Darvesh & Freedman, 1996). Both PDD and DLB involved the loss of cholinergic neurons (Klein et al, 2010; Shimada et al, 2009) resulting in the severe cortical acetylcholine deficits (Rolinski et al, 2012). LBs pathologies exist in brains of PDD and DLB (Jellinger & Attems, 2015). Approx. 78% patients suffering from PD develops dementia (Aarsland et al, 2003) which is similar to DLB (Aarsland et al, 2003; Ballard et al, 2002; Emre, 2003). Further pure DLB appear similar to PDD (Emre et al, 2007) and both of these pathologies involved narcolepsy and REM sleep behavior disorder (Kurtz and Kaufer, 2011).

DLB is the condition refers to the onset of the dementia before the onset of PD or with one year of PD onset (McKeith et al, 2006). The DLB and PDD distinguished on the basis of cognitive and motor symptoms (McKeith et al 1996). Cortical cholinergic loss is greater in DLB (Bohnen et al, 2012; Bohnen et al, 2009), while the dopaminergic neuronal loss is more severe in PDD than DLB (Tsuboi & Dickson, 2005). Thus nigrostriatal pathological changes occur in PD, subcortical pathological changes in PDD and the cortical changes occur in DLB (Petrova et al, 2015). Bilateral frontal atrophy occurs in PDD while parietal and occipital atrophy occurs in DLB (Borroni et al, 2015). Further PDD tends to affect younger patients than does DLB (Horimoto et al, 2003; Richard et al, 2002) and this might be due to its specific topography so involved (Revuelta & Lippa, 2009). These findings present the similarities and the dissimilarities in the PD, PDD and DLB.

DIAGNOSTIC CRITERIA FOR LEWY BODY DISEASE

Diagnosis of Parkinson's Disease

The degeneration of dopaminergic neurons in the SN and the presence of LBs in the brainstem nuclei of brain are the main characteristics of PD (Lees et al, 2009; Marsden, 1994). The careful assessment of patient history is very important and therefore thorough questioning should be performed and, inquiry should be made about the presence of premotor symptoms (Massano & Bhatia, 2012). The clinician should note resting tremor, rigidity, postural instability, and akinesia and further should perform CT scan and MRI scan (Massano et al, 2008; Sitburana & Ondo, 2009) the former is used to identify the calcium deposits while the later is used to confirm degeneration (Kagi et al, 2010).

Diagnosis of Parkinson's Disease Dementia

Dementia is the primary feature of the PDD patients (Dubois et al, 2007; Emre et al, 2007). For PDD diagnosis, two core features must be present: first is the presence of PD and second is, PD developed prior to the onset of dementia. Associated clinical features of PDD include deficits in the cognition and behavioral impairments (Dubois et al, 2007; Emre et al, 2007). A patient is said to be suffering from the probable PDD if the impairment is noted in atleast two of the four cognitive domains supported by the presence of atleast one behavioral symptom. A patient is said to be suffer from the possible PDD, if dementia and PD exist together, however the associated clinical features are not considered typical (Dubois et al, 2007; Emre et al, 2007).

Diagnosis of Dementia With Lewy Bodies

According to McKeith et al. DLB is characterized by the presence of central, core, suggestive, and supportive features (McKeith et al, 2005). Like the PDD, central feature of the DLB is dementia, however the early prominent symptoms that can be seen in the patients suffering from DLB includes attention deficits, visual and executive dysfunction. Fluctuating cognition, recurrent visual hallucinations and spontaneous Parkinsonism, represent the core features, REM sleep disorder, severe neuroleptic sensitivity and low dopamine transporter uptake in BG represents the suggestive features and repeated falls, syncope, unexplained loss of consciousness; severe autonomic dysfunction, and non-visual hallucinations constitute the supportive features of DLB. Presence of one core feature with the presence of a suggestive feature suggests the probable DLB while the absence of any core features with the presence of a suggestive feature suggests possible DLB (McKeith et al, 2005).

PHARMACOTHERAPEUTICS OF LEWY BODY DISEASE

Various pharmacotherapeutics agents have been used in the symptomatic treatment of the LBD. Following section defines the selection, benefits, and restrictions of the different pharmacotherapeutics agents for the symptomatic treatment of PD.

Treatment of Cognitive Symptoms

Cognitive symptoms of LBD should be treated with cholinesterase inhibitors (Aarsland et al, 2004; McKeith et al, 2000; Mori et al, 2012; Samuel et al, 2000; Wild et al, 2003). However cholinesterase inhibitors can exacerbate parkinsonism, and increase tremor in the prescribed patients (McKeith et al, 2000).

Treatment of Motor Symptoms

Levodopa/carbidopa might prove beneficial (Goldman et al, 2008). But the efficacy is lesser in DLB than in PD and the risk of development of psychosis increases (Lucetti et al, 2010; Molloy et al, 2005). Further the administration of oral dopamine agonists frequently increases the incidence of hallucinations, abnormal repetitive behaviors (Hassan et al, 2011) and compulsive behavior (Wesnes et al, 2005).

Treatment of Neuropsychiatric Symptoms

SSRIs and SNRIs might prove beneficial (McKeith et al, 2005). However SSRIs increased the risk of suicidality in the prescribed patients (Walia, 2017).

Treatment of Hallucinations and Delusions

Hallucinations and delusions should left untreated if possible (McKeith et al, 2000). DLB patients have increased risk of antipsychotics toxicity (Aarsland et al, 2005b) and a single dose of antipsychotics in these patients increased sedation, confusion, parkinsonism, rigidity, dysautonomia, and mortality (Aarsland et al, 2005b; Hassan et al, 2011; Rochon et al, 2005).

Treatment of Sleep Dysfunction

The presence of sleep dysfunction should be confirmed by polysomnography (Boeve et al, 2013). Treatment with caffeine is useful (Malhotra & Avidan, 2012) while the other therapeutic options may include methylphenidate, dextroamphetamine, and modafinil (Hogl et al, 2002; Seppi et al, 2011).

Treatment of Agitation and Behavioral Disturbance

Pain is often a trigger for agitation and therefore acetaminophen might prove beneficial (Husebo et al, 2011). The incidence of agitation and behavioral disturbances occurs generally in the later stages of disease (Husebo et al, 2011). Agitation and behavioral disturbance can often be treated by the simple measures that include and increased social interaction, training to the caregiver and removal of fear triggers (Rhodes-Kropf et al, 2011).

Treatment of Restless Leg Syndrome

Levodopa/carbidopa, clonazepam, and other GABA analogues might prove beneficial for the treatment of these symptoms (Hening et al, 2004).

Treatment of Autonomic Dysfunction

Autonomic dysfunction includes constipation or diarrhea. (Jost, 1997; Lawrence et al, 2013) as a consequence of DLB (Boot et al, 2013; Postuma et al, 2013; Savica et al, 2010) treatment should include high-fiber diet, exercise, stool softeners, psyllium (Ashraf et al, 1997) and polyethylene glycol (Zangaglia et al, 2007).

ISSUES AND PROBLEMS RELATED TO LEWY BODY DISEASE

LBD are characterized by the presence of LBs in various cortical and subcortical regions (Jellinger, 2009). Presence of LBs in the cortical and limbic regions of the brain correlates dementia in patients with PD or PDD (Irwin et al, 2012). Further the pathological assessment of DLB reveals the presence

of LBs primarily in limbic, and secondly in neocortical regions (Rezaie et al, 1996). The complexity of full spectrum of LBD symptoms (Galvin et al, 2008) and the similarities in clinical features, and neuropathology of LBD, make the diagnosis and management becomes so difficult (Gomperts, 2016) that sometimes clinicians made the wrong diagnosis (Zweig & Galvin, 2014). Also the clinicians view all dementias as the dementia develops due to AD pathology (Galvin et al, 2008). However there are some questions that remains still unresolved in context of LBD includes why the limbic region is more vulnerable to the LBs formation in DLB (Rezaie et al, 1996); why DLB and PDD have similar symptoms despite the fact they are distinct pathologies (Berg et al, 2014) and why the discrimination of DLB from other types of dementia is difficult (Sakamoto et al, 2014).

FUTURE RESEARCH DIRECTIONS

LBD imposed a great burden worldwide. The major limitation is that, till now the complete pathophysiology and the mechanism implicated in the pathogenesis of the LBD is not known completely. Further LBD is often misdiagnosed or underdiagnosed (Zweig & Galvin, 2014) resulting either in the delay in the pharmacotherapeutics of the disease or the wrong prescription that may prove fatal. Epidemiological studies should be done to provide the better understanding and prevalence of LBD (Galvin et al, 2008). Further the development of mouse models and an appropriate biomarker for LBD might prove beneficial in the area of the drug development for LBD (Galvin et al, 2008). Also newer diagnostic techniques and methods should be developed to avoid the incidence of misdiagnosis and under diagnosis that further worsens the conditions of patients (Zweig & Galvin, 2014).

CONCLUSION

It is concluded that the LBD put a great burden worldwide and is responsible for the disability. The greatest problem is that the symptoms are very similar and often led to wrong diagnosis. Therefore a great care should be taken while diagnosing and treating LBD because wrong treatment may further worsens the condition. Since the key step involved in the formation of LBs is the aggregation of α -syn. Therefore the interventional strategy might include the agents that interfere with α -syn aggregation. Further the interfering in either the process of α -syn secretion or neuronal uptake of α -syn might prove beneficial.

REFERENCES

Aarsland, D., Andersen, K., Larsen, J. P., Lolk, A., & Kragh-Sorensen, P. (2003). Prevalence and characteristics of dementia in Parkinson disease: An 8-year prospective study. *Archives of Neurology*, *60*(3), 387–392. doi:10.1001/archneur.60.3.387 PMID:12633150

Aarsland, D., Kvaløy, J. T., Andersen, K., Larsen, J. P., Tang, M. X., Lolk, A., ... Marder, K. (2007). The effect of age of onset of PD on risk of dementia. *Journal of Neurology*, 254(1), 38–45. doi:10.100700415-006-0234-8 PMID:17508138

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Aarsland, D., Mosimann, U. P., & McKeith, I. G. (2004). Role of cholinesterase inhibitors in Parkinson's disease and dementia with Lewy bodies. *Journal of Geriatric Psychiatry and Neurology*, *17*(3), 164–171. doi:10.1177/0891988704267463 PMID:15312280

Aarsland, D., Perry, R., Larsen, J. P., McKeith, I. G., O'Brien, J. T., Perry, E. K., ... Ballard, C. G. (2005b). Neuroleptic sensitivity in Parkinson's disease and parkinsonian dementias. *The Journal of Clinical Psychiatry*, *66*(05), 633–637. doi:10.4088/JCP.v66n0514 PMID:15889951

Aarsland, D., Zaccai, J., & Brayne, C. (2005a). A systematic review of prevalence studies of dementia in Parkinson's disease. *Movement Disorders*, 20(10), 1255–1263. doi:10.1002/mds.20527 PMID:16041803

Adler, C. H., Connor, D. J., Hentz, J. G., Sabbagh, M. N., Caviness, J. N., Shill, H. A., ... Beach, T. G. (2010). Incidental Lewy body disease: Clinical comparison to a control cohort. *Movement Disorders*, *25*(5), 642–646. doi:10.1002/mds.22971 PMID:20175211

Arendt, T., Bigl, V., Arendt, A., & Tennstedt, A. (1983). Loss of neurons in the nucleus basalis of Meynert in Alzheimer's disease, paralysis agitans and Korsakoff's disease. *Acta Neuropathologica*, *61*(2), 101–108. doi:10.1007/BF00697388 PMID:6637393

Ashraf, W., Pfeiffer, R. F., Park, F., Lof, J., & Quigley, E. M. (1997). Constipation in Parkinson's disease: Objective assessment and response to psyllium. *Movement Disorders*, *12*(6), 946–951. doi:10.1002/ mds.870120617 PMID:9399219

Ballard, C. G., Aarsland, D., McKeith, I., O'brien, J., Gray, A., Cormack, F., ... Brown, R. (2002). Fluctuations in attention PD dementia vs DLB with parkinsonism. *Neurology*, *59*(11), 1714–1720. doi:10.1212/01.WNL.0000036908.39696.FD PMID:12473758

Bartus, R. T., Dean, R., Beer, B., & Lippa, A. (1982). The cholinergic hypothesis of geriatric memory dysfunction. *Science*, *217*(4558), 408–414. doi:10.1126cience.7046051 PMID:7046051

Berg, D., Postuma, R. B., Bloem, B., Chan, P., Dubois, B., Gasser, T., ... Deuschl, G. (2014). Time to Redefine PD? Introductory Statement of the MDS Task Force on the Definition of Parkinson's Disease. *Movement Disorders*, 29(4), 454–462. doi:10.1002/mds.25844 PMID:24619848

Boeve, B. F., Silber, M. H., Ferman, T. J., Lin, S. C., Benarroch, E. E., Schmeichel, A. M., ... Dickson, D. W. (2013). Clinicopathologic correlations in 172 cases of rapid eye movement sleep behavior disorder with or without a coexisting neurologic disorder. *Sleep Medicine*, *14*(8), 754–762. doi:10.1016/j. sleep.2012.10.015 PMID:23474058

Bohnen, N. I., Müller, M. L. T. M., Koeppe, R. A., Studenski, S. A., Kilbourn, M. A., Frey, K. A., & Albin, R. L. (2009). History of falls in Parkinson disease is associated with reduced cholinergic activity. *Neurology*, *73*(20), 1670–1676. doi:10.1212/WNL.0b013e3181c1ded6 PMID:19917989

Bohnen, N. I., Müller, M. L. T. M., Kotagal, V., Koeppe, R. A., Kilbourn, M. R., Gilman, S., ... Frey, K. A. (2012). Heterogeneity of cholinergic denervation in Parkinson's disease without dementia. *Journal of Cerebral Blood Flow and Metabolism*, *32*(8), 1609–1617. doi:10.1038/jcbfm.2012.60 PMID:22569194

Boot, B. P., Orr, C. F., Ahlskog, J. E., Ferman, T. J., Roberts, R., Pankratz, V. S., ... Boeve, B. F. (2013). Risk factors for dementia with Lewy bodies: A case-control study. *Neurology*, *81*(9), 833–840. doi:10.1212/ WNL.0b013e3182a2cbd1 PMID:23892702

Borroni, B., Premi, E., Formenti, A., Turrone, R., Alberici, A., Cottini, E., ... Padovani, A. (2015). Structural and functional imaging study in dementia with Lewy bodies and Parkinson's disease dementia. *Parkinsonism & Related Disorders*, 21(9), 1049–1055. doi:10.1016/j.parkreldis.2015.06.013 PMID:26109553

Braak, H., & Braak, E. (2000). Pathoanatomy of Parkinson's disease. *Journal of Neurology*, 247(S2), II3–II10. doi:10.1007/PL00007758 PMID:10991663

Braak, H., Del Tredici, K., Bratzke, H., Hamm-Clement, J., Sandmann-Keil, D., & Rüb, U. (2002). Staging of the intracerebral inclusion body pathology associated with idiopathic Parkinson's disease (preclinical and clinical stages). *Journal of Neurology*, 249(0), III-1. doi:10.100700415-002-1301-4 PMID:12528692

Braak, H., Del Tredici, K., Rüb, U., de Vos, R. A., Steur, E. N. J., & Braak, E. (2003). Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiology of Aging*, 24(2), 197–211. doi:10.1016/S0197-4580(02)00065-9 PMID:12498954

Braak, H., Rüb, U., Gai, W. P., & Del Tredici, K. (2003). Idiopathic Parkinson's disease: Possible routes by which vulnerable neuronal types may be subject to neuroinvasion by an unknown pathogen. *Journal of Neural Transmission (Vienna, Austria)*, *110*(5), 517–536. doi:10.100700702-002-0808-2 PMID:12721813

Braak, H., Rüb, U., Steur, E. J., Del Tredici, K., & De Vos, R. A. I. (2005). Cognitive status correlates with neuropathologic stage in Parkinson disease. *Neurology*, *64*(8), 1404–1410. doi:10.1212/01. WNL.0000158422.41380.82 PMID:15851731

Campbell, B. C., McLean, C. A., Culvenor, J. G., Gai, W. P., Blumbergs, P. C., Jäkälä, P., ... Li, Q.-X. (2001). The solubility of α -synuclein in multiple system atrophy differs from that of dementia with Lewy bodies and Parkinson's disease. *Journal of Neurochemistry*, *76*(1), 87–96. doi:10.1046/j.1471-4159.2001.00021.x PMID:11145981

Candy, J., Perry, R. H., Perry, E. K., Irving, D., Blessed, G., Fairbairn, A. F., & Tomlinson, B. E. (1983). Pathological changes in the nucleus of Meynert in Alzheimer's and Parkinson's diseases. *Journal of the Neurological Sciences*, *59*(2), 277–289. doi:10.1016/0022-510X(83)90045-X PMID:6854353

Chartier-Harlin, M. C., Kachergus, J., Roumier, C., Mouroux, V., Douay, X., Lincoln, S., ... Waucquier, N. (2004). α-synuclein locus duplication as a cause of familial Parkinson's disease. *Lancet*, *364*(9440), 1167–1169. doi:10.1016/S0140-6736(04)17103-1 PMID:15451224

Chui, H. C., Mortimer, J. A., Slager, U., Zarow, C., Bondareff, W., & Webster, D. D. (1986). Pathologic correlates of dementia in Parkinson's disease. *Archives of Neurology*, *43*(10), 991–995. doi:10.1001/archneur.1986.00520100013007 PMID:3753274

Darvesh, S., & Freedman, M. (1996). Subcortical dementia: A neurobehavioral approach. *Brain and Cognition*, *31*(2), 230–249. doi:10.1006/brcg.1996.0043 PMID:8812000

Dickson, D. W., Davies, P., Mayeux, R., Crystal, H., Horoupian, D. S., Thompson, A., & Goldman, J. E. (1986). Diffuse Lewy body disease. Neuropathological and biochemical studies of six patients. *Acta Neuropathologica*, *75*(1), 8–15. doi:10.1007/BF00686786 PMID:3434218

Dickson, D. W., Feany, M. B., Yen, S. H., Mattiace, L. A., & Davies, P. (1996). Cytoskeletal pathology in non-Alzheimer degenerative dementia: new lesions in diffuse Lewy body disease, Pick's disease, and corticobasal degeneration. In *New Trends in the Diagnosis and Therapy of Non-Alzheimer's Dementia* (pp. 31–46). Springer Vienna. doi:10.1007/978-3-7091-6892-9_2

Dickson, D. W., Fujishiro, H., DelleDonne, A., Menke, J., Ahmed, Z., Klos, K. J., ... Ahlskog, J. E. (2008). Evidence that incidental Lewy body disease is pre-symptomatic Parkinson's disease. *Acta Neuropathologica*, *115*(4), 437–444. doi:10.100700401-008-0345-7 PMID:18264713

Dubois, B., Burn, D., Goetz, C., Aarsland, D., Brown, R. G., Broe, G. A., ... Korczyn, A. (2007). Diagnostic procedures for Parkinson's disease dementia: Recommendations from the movement disorder society task force. *Movement Disorders*, 22(16), 2314–2324. doi:10.1002/mds.21844 PMID:18098298

Duda, J. E., Giasson, B. I., Mabon, M. E., Lee, V. M. Y., & Trojanowski, J. Q. (2002). Novel antibodies to synuclein show abundant striatal pathology in Lewy body diseases. *Annals of Neurology*, 52(2), 205–210. doi:10.1002/ana.10279 PMID:12210791

Eichner, T., & Radford, S. E. (2011). A diversity of assembly mechanisms of a generic amyloid fold. *Molecular Cell*, 43(1), 8–18. doi:10.1016/j.molcel.2011.05.012 PMID:21726806

Emre, M. (2003). Dementia associated with Parkinson's disease. *Lancet Neurology*, 2(4), 229–237. doi:10.1016/S1474-4422(03)00351-X PMID:12849211

Emre, M., Aarsland, D., Brown, R., Burn, D. J., Duyckaerts, C., Mizuno, Y., ... Goldman, J. (2007). Clinical diagnostic criteria for dementia associated with Parkinson's disease. *Movement Disorders*, 22(12), 1689–1707. doi:10.1002/mds.21507 PMID:17542011

Engelender, S. (2008). Ubiquitination of α -synuclein and autophagy in Parkinson's disease. *Autophagy*, 4(3), 372–374. doi:10.4161/auto.5604 PMID:18216494

Ferman, T. J., Boeve, B. F., Smith, G. E., Silber, M. H., Lucas, J. A., Graff-Radford, N. R., ... Ivnik, R. J. (2002). Dementia with Lewy bodies may present as dementia and REM sleep behavior disorder without parkinsonism or hallucinations. *Journal of the International Neuropsychological Society*, 8(7), 907–914. doi:10.1017/S1355617702870047 PMID:12405541

Forno, L. S. (1996). Neuropathology of Parkinson's disease. *Journal of Neuropathology and Experimental Neurology*, 55(3), 259–272. doi:10.1097/00005072-199603000-00001 PMID:8786384

Frigerio, R., Fujishiro, H., Ahn, T.-B., Josephs, K. A., Maraganore, D. M., DelleDonne, A., ... Ahlskog, E. J. (2011). Incidental Lewy Body Disease: Do some cases represent a preclinical stage of Dementia with Lewy Bodies? *Neurobiology of Aging*, *32*(5), 857–863. doi:10.1016/j.neurobiolaging.2009.05.019 PMID:19560232

Fuentes, R., Petersson, P., & Nicolelis, M. A. (2010). Restoration of locomotive function in Parkinson's disease by spinal cord stimulation: Mechanistic approach. *The European Journal of Neuroscience*, *32*(7), 1100–1108. doi:10.1111/j.1460-9568.2010.07417.x PMID:21039949

Gai, W. P., Halliday, G. M., Blumbergs, P. C., Geffen, L. B., & Blessing, W. W. (1991). Substance P-containing neurons in the mesopontine tegmentum are severely affected in Parkinson's disease. *Brain*, *114*(5), 2253–2267. doi:10.1093/brain/114.5.2253 PMID:1718530

Galvin, J. E., & Balasubramaniam, M. (2013). Lewy Body Dementia: The Under-Recognized but Common FOE. *Cerebrum: The Dana Forum on Brain Science*, 2013, 13. PMID:24772233

Galvin, J. E., Duda, J. E., Kaufer, D. I., Lippa, C. F., Taylor, A., & Zarit, S. H. (2010). Lewy Body Dementia: Caregiver burden and unmet needs. *Alzheimer Disease and Associated Disorders*, 24(2), 177–181. doi:10.1097/WAD.0b013e3181c72b5d PMID:20505434

Galvin, J. E., Duda, J. E., Kaufer, D. I., Lippa, C. F., Taylor, A., & Zarit, S. H. (2010). Lewy Body Dementia: Caregiver Burden and Unmet Needs. *Alzheimer Disease and Associated Disorders*, 24(2), 177–181. doi:10.1097/WAD.0b013e3181c72b5d PMID:20505434

Galvin, J. E., Duda, J. E., Kaufer, D. I., Lippa, C. F., Taylor, A., & Zarit, S. H. (2010). Lewy body dementia: The caregiver experience of clinical care. *Parkinsonism & Related Disorders*, *16*(6), 388–392. doi:10.1016/j.parkreldis.2010.03.007 PMID:20434939

Gerfen, C. R., & Surmeier, D. J. (2011). Modulation of striatal projection systems by dopamine. *Annual Review of Neuroscience*, *34*(1), 441–466. doi:10.1146/annurev-neuro-061010-113641 PMID:21469956

Goldman, J. G., Goetz, C. G., Brandabur, M., Sanfilippo, M., & Stebbins, G. T. (2008). Effects of dopaminergic medications on psychosis and motor function in dementia with Lewy bodies. *Movement Disorders*, 23(15), 2248–2250. doi:10.1002/mds.22322 PMID:18823039

Gómez-Tortosa, E., Newell, K., Irizarry, M. C., Sanders, J. L., & Hyman, B. T. (2000). α-Synuclein immunoreactivity in dementia with Lewy bodies: Morphological staging and comparison with ubiquitin immunostaining. *Acta Neuropathologica*, 99(4), 352–357. doi:10.1007004010051135 PMID:10787032

Gomperts, S. N. (2016). Lewy Body Dementias: Dementia With Lewy Bodies and Parkinson Disease Dementia. *Continuum: Lifelong Learning in Neurology*, 22(2), 435-463.

Gosal, D., Ross, O. A., & Toft, M. (2006). Parkinson's disease: The genetics of a heterogeneous disorder. *European Journal of Neurology*, *13*(6), 616–627. doi:10.1111/j.1468-1331.2006.01336.x PMID:16796586

Grossman, M., Gross, R. G., Moore, P., Dreyfuss, M., McMillan, C. T., Cook, P. A., ... Siderowf, A. (2012). Difficulty processing temporary syntactic ambiguities in Lewy body spectrum disorder. *Brain and Language*, *120*(1), 52–60. doi:10.1016/j.bandl.2011.08.007 PMID:21962945

Halliday, G., Hely, M., Reid, W., & Morris, J. (2008). The progression of pathology in longitudinally followed patients with Parkinson's disease. *Acta Neuropathologica*, *115*(4), 409–415. doi:10.100700401-008-0344-8 PMID:18231798

Harding, A. J., Broe, G. A., & Halliday, G. M. (2002). Visual hallucinations in Lewy body disease relate to Lewy bodies in the temporal lobe. *Brain*, *125*(2), 391–403. doi:10.1093/brain/awf033 PMID:11844739

Hassan, I. (2011). Consultation-liaison psychiatry and prevention of severe neuroleptic sensitivity reactions in dementia with Lewy bodies. *Australasian Psychiatry*, *19*(6), 536–537. doi:10.3109/10398562. 2011.580750 PMID:22077306

Hattori, N., & Mizuno, Y. (2004). Pathogenetic mechanisms of parkin in Parkinson's disease. *Lancet*, *364*(9435), 722–724. doi:10.1016/S0140-6736(04)16901-8 PMID:15325839

Hedrich, K., Eskelson, C., Wilmot, B., Marder, K., Harris, J., Garrels, J., ... Lang, A. E. (2004). Distribution, type, and origin of Parkin mutations: Review and case studies. *Movement Disorders*, *19*(10), 1146–1157. doi:10.1002/mds.20234 PMID:15390068

Hely, M. A., Reid, W. G., Adena, M. A., Halliday, G. M., & Morris, J. G. (2008). The Sydney multicenter study of Parkinson's disease: The inevitability of dementia at 20 years. *Movement Disorders*, 23(6), 837–844. doi:10.1002/mds.21956 PMID:18307261

Hening, W. A., Allen, R. P., Earley, C. J., Picchietti, D. L., & Silber, M. H. (2004). An update on the dopaminergic treatment of restless legs syndrome and periodic limb movement disorder. *Sleep*, *27*(3), 560–583. doi:10.1093leep/27.3.560 PMID:15164915

Hernandez, D., Ruiz, C. P., Crawley, A., Malkani, R., Werner, J., Gwinn-Hardy, K., ... Singleton, A. (2005). The dardarin G2019S mutation is a common cause of Parkinson's disease but not other neurodegenerative diseases. *Neuroscience Letters*, *389*(3), 137–139. doi:10.1016/j.neulet.2005.07.044 PMID:16102903

Hinault, M. P., Cuendet, A. F. H., Mattoo, R. U., Mensi, M., Dietler, G., Lashuel, H. A., & Goloubinoff, P. (2010). Stable α -synuclein oligomers strongly inhibit chaperone activity of the Hsp70 system by weak interactions with J-domain co-chaperones. *The Journal of Biological Chemistry*, 285(49), 38173–38182. doi:10.1074/jbc.M110.127753 PMID:20847048

Hirsch, E. C., Graybiel, A. M., Duyckaerts, C., & Javoy-Agid, F. (1987). Neuronal loss in the pedunculopontine tegmental nucleus in Parkinson disease and in progressive supranuclear palsy. *Proceedings of the National Academy of Sciences of the United States of America*, 84(16), 5976–5980. doi:10.1073/ pnas.84.16.5976 PMID:3475716

Hogl, B., Saletu, M., Brandauer, E., Glatzl, S., Frauscher, B., Seppi, K., ... Poewe, W. (2002). Modafinil for the treatment of daytime sleepiness in Parkinson's disease: A double-blind, randomized, crossover, placebo-controlled polygraphic trial. *Sleep*, *25*(8), 905–909. doi:10.1093leep/25.8.62 PMID:12489899

Holdorff, B. (2006). Fritz heinrich lewy (1885-1950). *Journal of Neurology*, 253(5), 677–678. doi:10.100700415-006-0130-2 PMID:16767545

Horimoto, Y., Matsumoto, M., Nakazawa, H., Yuasa, H., Morishita, M., Akatsu, H., ... Kosaka, K. (2003). Cognitive conditions of pathologically confirmed dementia with Lewy bodies and Parkinson's disease with dementia. *Journal of the Neurological Sciences*, *216*(1), 105–108. doi:10.1016/S0022-510X(03)00220-X PMID:14607310

Husebo, B. S., Ballard, C., Sandvik, R., Nilsen, O. B., & Aarsland, D. (2011). Efficacy of treating pain to reduce behavioural disturbances in residents of nursing homes with dementia: Cluster randomised clinical trial. *BMJ (Clinical Research Ed.)*, *343*(jul151), d4065. doi:10.1136/bmj.d4065 PMID:21765198

Irwin, D. J., White, M. T., Toledo, J. B., Xie, S. X., Robinson, J. L., Van Deerlin, V., ... Hurtig, H. I. (2012). Neuropathologic substrates of Parkinson disease dementia. *Annals of Neurology*, 72(4), 587–598. doi:10.1002/ana.23659 PMID:23037886

Jankovic, J., McDermott, M., Carter, J., Gauthier, S., Goetz, C., Golbe, L., ... Stern, M. (1990). Variable expression of Parkinson's disease A base-line analysis of the DAT ATOP cohort. *Neurology*, 40(10), 1529–1529. doi:10.1212/WNL.40.10.1529 PMID:2215943

Jellinger, K. (1988). The pedunculopontine nucleus in Parkinson's disease, progressive supranuclear palsy and Alzheimer's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, *51*(4), 540–543. doi:10.1136/jnnp.51.4.540 PMID:3379428

Jellinger, K. A. (2009). Formation and development of Lewy pathology: A critical update. *Journal of Neurology*, 256(3), 270–279. doi:10.100700415-009-5243-y PMID:19711116

Jellinger, K. A., & Attems, J. (2015). Challenges of multimorbidity of the aging brain: A critical update. *Journal of Neural Transmission (Vienna, Austria)*, *122*(4), 505–521. doi:10.100700702-014-1288-x PMID:25091618

Jellinger, K. A., Seppi, K., Wenning, G. K., & Poewe, W. (2002). Impact of coexistent Alzheimer pathology on the natural history of Parkinson's disease. *Journal of Neural Transmission (Vienna, Austria)*, *109*(3), 329–339. doi:10.1007007020200027 PMID:11956955

Jost, W. H. (1997). Gastrointestinal motility problems in patients with Parkinson's disease. *Drugs & Aging*, *10*(4), 249–258. doi:10.2165/00002512-199710040-00002 PMID:9108986

Jucker, M., & Walker, L. C. (2011). Pathogenic protein seeding in Alzheimer disease and other neurodegenerative disorders. *Annals of Neurology*, 70(4), 532–540. doi:10.1002/ana.22615 PMID:22028219

Kalra, S., Bergeron, C., & Lang, A. E. (1996). Lewy body disease and dementia: A review. Archives of Internal Medicine, 156(5), 487–493. doi:10.1001/archinte.1996.00440050031004 PMID:8604954

Karachi, C., Grabli, D., Bernard, F. A., Tandé, D., Wattiez, N., Belaid, H., ... Hartmann, A. (2010). Cholinergic mesencephalic neurons are involved in gait and postural disorders in Parkinson disease. *The Journal of Clinical Investigation*, *120*(8), 2745–2754. doi:10.1172/JCI42642 PMID:20628197

Katsuse, O., Iseki, E., Marui, W., & Kosaka, K. (2003). Developmental stages of cortical Lewy bodies and their relation to axonal transport blockage in brains of patients with dementia with Lewy bodies. *Journal of the Neurological Sciences*, 211(1), 29–35. doi:10.1016/S0022-510X(03)00037-6 PMID:12767494

Kehagia, A. A., Barker, R. A., & Robbins, T. W. (2013). Cognitive impairment in Parkinson's disease: The dual syndrome hypothesis. *Neurodegenerative Diseases*, *11*(2), 79–92. doi:10.1159/000341998 PMID:23038420

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Kim, T. D., Choi, E., Rhim, H., Paik, S. R., & Yang, C. H. (2004). α-synuclein has structural and functional similarities to small heat shock proteins. *Biochemical and Biophysical Research Communications*, *324*(4), 1352–1359. doi:10.1016/j.bbrc.2004.09.208 PMID:15504363

Klein, J. C., Eggers, C., Kalbe, E., Weisenbach, S., Hohmann, C., Vollmar, S., ... Hilker, R. (2010). Neurotransmitter changes in dementia with Lewy bodies and Parkinson disease dementia in vivo. *Neurology*, 74(11), 885–892. doi:10.1212/WNL.0b013e3181d55f61 PMID:20181924

Kordower, J. H., Chu, Y., Hauser, R. A., Freeman, T. B., & Olanow, C. W. (2008). Lewy body-like pathology in long-term embryonic nigral transplants in Parkinson's disease. *Nature Medicine*, *14*(5), 504–506. doi:10.1038/nm1747 PMID:18391962

Kosaka, K. (1990). Diffuse Lewy body disease in Japan. *Journal of Neurology*, 237(3), 197–204. doi:10.1007/BF00314594 PMID:2196340

Kosaka, K. (2014). Lewy body disease and dementia with Lewy bodies. *Proceedings of the Japan Academy, Series B*, 90(8), 301–306. doi:10.2183/pjab.90.301 PMID:25311140

Kosaka, K., Iseki, E., Odawara, T., & Yamamoto, T. (1996). Cerebral type of Lewy body disease. *Neuropathology*, *16*(1), 32–35. doi:10.1111/j.1440-1789.1996.tb00152.x

Kosaka, K., Yoshimura, M., Ikeda, K., & Budka, H. (1983). Diffuse type of Lewy body disease: Progressive dementia with abundant cortical Lewy bodies and senile changes of varying degree--a new disease? *Clinical Neuropathology*, *3*(5), 185–192. PMID:6094067

Kreitzer, A. C., & Malenka, R. C. (2008). Striatal plasticity and basal ganglia circuit function. *Neuron*, 60(4), 543–554. doi:10.1016/j.neuron.2008.11.005 PMID:19038213

Krüger, R., Kuhn, W., Müller, T., Woitalla, D., Graeber, M., Kösel, S., ... Riess, O. (1998). AlaSOPro mutation in the gene encoding α -synuclein in Parkinson's disease. *Nature Genetics*, *18*(2), 106–108. doi:10.1038/ng0298-106 PMID:9462735

Kurtz, A. L., & Kaufer, D. I. (2011). Dementia in Parkinson's disease. *Current Treatment Options in Neurology*, *13*(3), 242–254. doi:10.100711940-011-0121-1 PMID:21461668

Kuusisto, E., Parkkinen, L., & Alafuzoff, I. (2003). Morphogenesis of Lewy bodies: Dissimilar incorporation of α -synuclein, ubiquitin, and p62. *Journal of Neuropathology and Experimental Neurology*, 62(12), 1241–1253. doi:10.1093/jnen/62.12.1241 PMID:14692700

Lawrence, J., Parmenter, T., & McDonald, T. (2013). Failure to manage constipation in Parkinson's disease. A review of medical services: A patients perspective. *Movement Disorders*, 28(1), 187.

Lees, A. J., Hardy, J., & Revesz, T. (2009). Parkinson's disease. *Lancet*, *373*(9680), 2055–2066. doi:10.1016/S0140-6736(09)60492-X PMID:19524782

Levy, G., Schupf, N., Tang, M. X., Cote, L. J., Louis, E. D., Mejia, H., ... Marder, K. (2002). Combined effect of age and severity on the risk of dementia in Parkinson's disease. *Annals of Neurology*, *51*(6), 722–729. doi:10.1002/ana.10219 PMID:12112078

Li, J. Y., Englund, E., Holton, J. L., Soulet, D., Hagell, P., Lees, A. J., ... Widner, H. (2008). Lewy bodies in grafted neurons in subjects with Parkinson's disease suggest host-to-graft disease propagation. *Nature Medicine*, *14*(5), 501–503. doi:10.1038/nm1746 PMID:18391963

Lo, R. Y., Tanner, C. M., Albers, K. B., Leimpeter, A. D., Fross, R. D., Bernstein, A. L., ... Van Den Eeden, S. K. (2009). Clinical features in early Parkinson disease and survival. *Archives of Neurology*, *66*(11), 1353–1358. doi:10.1001/archneurol.2009.221 PMID:19901166

Lucetti, C., Logi, C., Del Dotto, P., Berti, C., Ceravolo, R., Baldacci, F., ... Bonuccelli, U. (2010). Levodopa response in dementia with lewy bodies: A 1-year follow-up study. *Parkinsonism & Related Disorders*, *16*(8), 522–526. doi:10.1016/j.parkreldis.2010.06.004 PMID:20615745

Luth, E. S., Stavrovskaya, I. G., Bartels, T., Kristal, B. S., & Selkoe, D. J. (2014). Soluble, prefibrillar α-synuclein oligomers promote complex I-dependent, Ca2+-induced mitochondrial dysfunction. *The Journal of Biological Chemistry*, 289(31), 21490–21507. doi:10.1074/jbc.M113.545749 PMID:24942732

Malhotra, R., & Avidan, A. Y. (2012). Neurodegenerative disease and REM behavior disorder. *Current Treatment Options in Neurology*, *14*(5), 474–492. doi:10.100711940-012-0194-5 PMID:22879077

Manning, C. (2004). Beyond memory: Neuropsychologic features in differential diagnosis of dementia. *Clinics in Geriatric Medicine*, 20(1), 45–58. doi:10.1016/j.cger.2003.10.002 PMID:15062486

Maraganore, D. M., Wilkes, K., Lesnick, T. G., Strain, K. J., De Andrade, M., Rocca, W. A., ... Farrer, M. J. (2004). A limited role for DJ1 in Parkinson disease susceptibility. *Neurology*, *63*(3), 550–553. doi:10.1212/01.WNL.0000133402.78621.AD PMID:15304593

Marsden, C. D. (1994). Parkinson's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 57(6), 672–681. doi:10.1136/jnnp.57.6.672 PMID:7755681

Marx, F. P., Holzmann, C., Strauss, K. M., Li, L., Eberhardt, O., Gerhardt, E., ... Engelender, S. (2003). Identification and functional characterization of a novel R621C mutation in the synphilin-1 gene in Parkinson's disease. *Human Molecular Genetics*, *12*(11), 1223–1231. doi:10.1093/hmg/ddg134PMID:12761037

Massano, J., & Bhatia, K. P. (2012). Clinical Approach to Parkinson's Disease: Features, Diagnosis, and Principles of Management. *Cold Spring Harbor Perspectives in Medicine*, 2(6), a008870. doi:10.1101/ cshperspect.a008870 PMID:22675666

Massano, J., Costa, F., & Nadais, G. (2008). Teaching neuroImage: MRI in multiple system atrophy: "Hot cross bun" sign and hyperintense rim bordering the putamina. *Neurology*, *71*(15), e38. doi:10.1212/01. wnl.0000327520.99034.28 PMID:18838658

McKeith, I., Del Ser, T., Spano, P., Emre, M., Wesnes, K., Anand, R., ... Spiegel, R. (2000). Efficacy of rivastigmine in dementia with Lewy bodies: A randomised, double-blind, placebo-controlled international study. *Lancet*, *356*(9247), 2031–2036. doi:10.1016/S0140-6736(00)03399-7 PMID:11145488

McKeith, I. G., Ballard, C. G., Perry, R. H., Ince, P. G., O'brien, J. T., Neill, D., ... Swann, A. (2000). Prospective validation of consensus criteria for the diagnosis of dementia with Lewy bodies. *Neurology*, *54*(5), 1050–1058. doi:10.1212/WNL.54.5.1050 PMID:10720273

Lewy Body Disease

McKeith, I. G., Dickson, D. W., Lowe, J., Emre, M., O'brien, J. T., Feldman, H., ... Aarsland, D. (2005). Diagnosis and management of dementia with Lewy bodies third report of the DLB consortium. *Neurology*, *65*(12), 1863–1872. doi:10.1212/01.wnl.0000187889.17253.b1 PMID:16237129

McKeith, I. G., Dickson, D. W., Lowe, J., Emre, M., O'brien, J. T., Feldman, H., ... Aarsland, D. (2005). Diagnosis and management of dementia with Lewy bodies third report of the DLB consortium. *Neurology*, *65*(12), 1863–1872. doi:10.1212/01.wnl.0000187889.17253.b1 PMID:16237129

McKeith, I. G., Galasko, D., Kosaka, K., Perry, E. K., Dickson, D. W., Hansen, L. A., ... Lennox, G. (1996). Consensus guidelines for the clinical and pathologic diagnosis of dementia with Lewy bodies (DLB) Report of the consortium on DLB international workshop. *Neurology*, *47*(5), 1113–1124. doi:10.1212/WNL.47.5.1113 PMID:8909416

Minton, A. P. (2005). Influence of macromolecular crowding upon the stability and state of association of proteins: Predictions and observations. *Journal of Pharmaceutical Sciences*, *94*(8), 1668–1675. doi:10.1002/jps.20417 PMID:15986476

Molloy, S., McKeith, I. G., O'brien, J. T., & Burn, D. J. (2005). The role of levodopa in the management of dementia with Lewy bodies. *Journal of Neurology, Neurosurgery, and Psychiatry*, 76(9), 1200–1203. doi:10.1136/jnnp.2004.052332 PMID:16107351

Mori, E., Ikeda, M., & Kosaka, K. (2012). Donepezil for dementia with Lewy bodies: A randomized, placebo-controlled trial. *Annals of Neurology*, 72(1), 41–52. doi:10.1002/ana.23557 PMID:22829268

Nakamura, K. (2013). α-Synuclein and mitochondria: Partners in crime? *Neurotherapeutics; the Journal of the American Society for Experimental NeuroTherapeutics, 10*(3), 391–399. doi:10.100713311-013-0182-9 PMID:23512373

Nakano, I., & Hirano, A. (1984). Parkinson's disease: Neuron loss in the nucleus basalis without concomitant Alzheimer's disease. *Annals of Neurology*, *15*(5), 415–418. doi:10.1002/ana.410150503 PMID:6732189

Nishikawa, K., Li, H., Kawamura, R., Osaka, H., Wang, Y. L., Hara, Y., ... Aoki, S. (2003). Alterations of structure and hydrolase activity of parkinsonism-associated human ubiquitin carboxyl-terminal hydrolase L1 variants. *Biochemical and Biophysical Research Communications*, *304*(1), 176–183. doi:10.1016/S0006-291X(03)00555-2 PMID:12705903

Nishioka, K., Hayashi, S., Farrer, M. J., Singleton, A. B., Yoshino, H., Imai, H., ... Mizoguchi, K. (2006). Clinical heterogeneity of α -synuclein gene duplication in Parkinson's disease. *Annals of Neurology*, *59*(2), 298–309. doi:10.1002/ana.20753 PMID:16358335

Orosz, F., Kovács, G. G., Lehotzky, A., Oláh, J., Vincze, O., & Ovádi, J. (2004). TPPP/p25: from unfolded protein to misfolding disease: prediction and experiments. *Biology of the Cell*, *96*(9), 701–711. doi:10.1016/j.biolcel.2004.08.002 PMID:15567525

Perry, E. K., Curtis, M., Dick, D. J., Candy, J. M., Atack, J. R., Bloxham, C. A., ... Perry, R. H. (1985). Cholinergic correlates of cognitive impairment in Parkinson's disease: Comparisons with Alzheimer's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 48(5), 413–421. doi:10.1136/jnnp.48.5.413 PMID:3998751

Petrova, M., Mehrabian-Spasova, S., Aarsland, D., Raycheva, M., & Traykov, L. (2015). Clinical and Neuropsychological Differences between Mild Parkinson's Disease Dementia and Dementia with Lewy Bodies. *Dementia and Geriatric Cognitive Disorders. Extra*, 5(2), 212–220. doi:10.1159/000375363 PMID:26195977

Polymeropoulos, M. H., Lavedan, C., Leroy, E., Ide, S. E., Dehejia, A., Dutra, A., ... Stenroos, E. S. (1997). Mutation in the α -synuclein gene identified in families with Parkinson's disease. *Science*, 276(5321), 2045–2047. doi:10.1126cience.276.5321.2045 PMID:9197268

Popescu, A., Lippa, C. F., Lee, V. M. Y., & Trojanowski, J. Q. (2004). Lewy bodies in the amygdala: Increase of α -synuclein aggregates in neurodegenerative diseases with tau-based inclusions. *Archives* of Neurology, 61(12), 1915–1919. doi:10.1001/archneur.61.12.1915 PMID:15596612

Postuma, R. B., Gagnon, J. F., Pelletier, A., & Montplaisir, J. (2013). Prodromal autonomic symptoms and signs in Parkinson's disease and dementia with Lewy bodies. *Movement Disorders*, 28(5), 597–604. doi:10.1002/mds.25445 PMID:23554107

Rajagopalan, S., & Andersen, J. K. (2001). Alpha synuclein aggregation: Is it the toxic gain of function responsible for neurodegeneration in Parkinson's disease? *Mechanisms of Ageing and Development*, *122*(14), 1499–1510. doi:10.1016/S0047-6374(01)00283-4 PMID:11511392

Revuelta, G. J., & Lippa, C. F. (2009). Dementia with Lewy bodies and Parkinson's disease dementia may best be viewed as two distinct entities. *International Psychogeriatrics*, *21*(2), 213. doi:10.1017/S1041610208008600 PMID:19173761

Rezaie, P., Cairns, N. J., Chadwick, A., & Lantos, P. L. (1996). Lewy bodies are located preferentially in limbic areas in diffuse Lewy body disease. *Neuroscience Letters*, *212*(2), 111–114. doi:10.1016/0304-3940(96)12775-0 PMID:8832651

Rhodes-Kropf, J., Cheng, H., Castillo, E. H., & Fulton, A. T. (2011). Managing the patient with dementia in long-term care. *Clinics in Geriatric Medicine*, *27*(2), 135–152. doi:10.1016/j.cger.2011.01.001 PMID:21641502

Richard, I. H., Papka, M., Rubio, A., & Kurlan, R. (2002). Parkinson's disease and dementia with Lewy bodies: One disease or two? *Movement Disorders*, *17*(6), 1161–1165. doi:10.1002/mds.10274 PMID:12465052

Rochon, P. A., Stukel, T. A., Sykora, K., Gill, S., Garfinkel, S., Anderson, G. M., ... Bronskill, S. E. (2005). Atypical antipsychotics and parkinsonism. *Archives of Internal Medicine*, *165*(16), 1882–1888. doi:10.1001/archinte.165.16.1882 PMID:16157833

Rogers, J. D., Brogan, D., & Mirra, S. S. (1985). The nucleus basalis of Meynert in neurological disease: A quantitative morphological study. *Annals of Neurology*, *17*(2), 163–170. doi:10.1002/ana.410170210 PMID:3883886

Rolinski, M., Fox, C., Maidment, I., & McShane, R. (2012). Cholinesterase inhibitors for dementia with Lewy bodies, Parkinson's disease dementia and cognitive impairment in Parkinson's disease. *The Cochrane Library*. PMID:22419314

Rosenthal, E., Brennan, L., Xie, S., Hurtig, H., Milber, J., Weintraub, D., ... Siderowf, A. (2010). Association between cognition and function in patients with Parkinson disease with and without dementia. *Movement Disorders*, *25*(9), 1170–1176. doi:10.1002/mds.23073 PMID:20310053

Rubin, J. E., McIntyre, C. C., Turner, R. S., & Wichmann, T. (2012). Basal ganglia activity patterns in parkinsonism and computational modeling of their downstream effects. *The European Journal of Neuroscience*, *36*(2), 2213–2228. doi:10.1111/j.1460-9568.2012.08108.x PMID:22805066

Sakamoto, F., Shiraishi, S., Yoshida, M., Tomiguchi, S., Hirai, T., Namimoto, T., ... Yamashita, Y. (2014). Diagnosis of dementia with Lewy bodies: Diagnostic performance of combined ¹²³I-IMP brain perfusion SPECT and ¹²³I-MIBG myocardial scintigraphy. *Annals of Nuclear Medicine*, *28*(3), 203–211. doi:10.100712149-013-0796-3 PMID:24363079

Samuel, W., Caligiuri, M., Galasko, D., Lacro, J., Marini, M., McClure, F. S., ... Jeste, D. V. (2000). Better cognitive and psychopathologic response to donepezil in patients prospectively diagnosed as dementia with Lewy bodies: A preliminary study. *International Journal of Geriatric Psychiatry*, *15*(9), 794–802. doi: PMID:10984725

Sathiyamoorthy, S., Tan, X., & Tan, E. K. (2014). Lewy Body in Parkinson's Disease: Causes or Scars of Neurodegeneration? *Austin Journal of Clinical Neurology*, *1*(3), 1011.

Savica, R., Rocca, W. A., & Ahlskog, J. E. (2010). When does Parkinson disease start? Archives of Neurology, 67(7), 798–801. doi:10.1001/archneurol.2010.135 PMID:20625084

Seidel, K., Mahlke, J., Siswanto, S., Krüger, R., Heinsen, H., Auburger, G., ... den Dunnen, W. (2015). The brainstem pathologies of Parkinson's disease and dementia with Lewy bodies. *Brain Pathology* (*Zurich, Switzerland*), 25(2), 121–135. doi:10.1111/bpa.12168 PMID:24995389

Selikhova, M., Williams, D. R., Kempster, P. A., Holton, J. L., Revesz, T., & Lees, A. J. (2009). A clinico-pathological study of subtypes in Parkinson's disease. *Brain*, *132*(11), 2947–2957. doi:10.1093/ brain/awp234 PMID:19759203

Seppi, K., Weintraub, D., Coelho, M., Perez-Lloret, S., Fox, S. H., Katzenschlager, R., ... Sampaio, C. (2011). The Movement Disorder Society Evidence-Based Medicine Review Update: Treatments for the non-motor symptoms of Parkinson's disease. *Movement Disorders*, *26*(S3), S42–S80. doi:10.1002/mds.23884 PMID:22021174

Shimada, H., Hirano, S., Shinotoh, H., Aotsuka, A., Sato, K., Tanaka, N., ... Hattori, T. (2009). Mapping of brain acetylcholinesterase alterations in Lewy body disease by PET. *Neurology*, *73*(4), 273–278. doi:10.1212/WNL.0b013e3181ab2b58 PMID:19474411

Singleton, A. B., Farrer, M., Johnson, J., Singleton, A., Hague, S., Kachergus, J., ... Lincoln, S. (2003). α -Synuclein locus triplication causes Parkinson's disease. *Science*, *302*(5646), 841–841. doi:10.1126cience.1090278 PMID:14593171

Sitburana, O., & Ondo, W. G. (2009). Brain magnetic resonance imaging (MRI) in parkinsonian disorders. *Parkinsonism & Related Disorders*, *15*(3), 165–174. doi:10.1016/j.parkreldis.2008.04.033 PMID:19059803

Sode, K., Ochiai, S., Kobayashi, N., & Usuzaka, E. (2007). Effect of reparation of repeat sequences in the human alpha-synuclein on fibrillation ability. *International Journal of Biological Sciences*, *3*(1), 1–7. doi:10.7150/ijbs.3.1 PMID:17200685

Sperlágh, B., & Sylvester Vizi, E. (2011). The role of extracellular adenosine in chemical neurotransmission in the hippocampus and Basal Ganglia: Pharmacological and clinical aspects. *Current Topics in Medicinal Chemistry*, *11*(8), 1034–1046. doi:10.2174/156802611795347564 PMID:21401497

Spillantini, M. G., Schmidt, M. L., Lee, V. M. Y., Trojanowski, J. Q., Jakes, R., & Goedert, M. (1997). α-Synuclein in Lewy bodies. *Nature*, *388*(6645), 839–840. doi:10.1038/42166 PMID:9278044

Stein, J. F. (2009). Akinesia, motor oscillations and the pedunculopontine nucleus in rats and men. *Experimental Neurology*, 215(1), 1–4. doi:10.1016/j.expneurol.2008.09.022 PMID:18977223

Strauss, K. M., Martins, L. M., Plun-Favreau, H., Marx, F. P., Kautzmann, S., Berg, D., ... Wolburg, H. (2005). Loss of function mutations in the gene encoding Omi/HtrA2 in Parkinson's disease. *Human Molecular Genetics*, *14*(15), 2099–2111. doi:10.1093/hmg/ddi215 PMID:15961413

Tagliavini, F., Pilleri, G., Bouras, C., & Constantinidis, J. (1984). The basal nucleus of Meynert in idiopathic Parkinson's disease. *Acta Neurologica Scandinavica*, 70(1), 20–28. doi:10.1111/j.1600-0404.1984. tb00798.x PMID:6475484

Tsuboi, Y., & Dickson, D. W. (2005). Dementia with Lewy bodies and Parkinson's disease with dementia: Are they different? *Parkinsonism & Related Disorders*, *11*, S47–S51. doi:10.1016/j.parkreldis.2004.10.014 PMID:15885629

Tsuboi, Y., Uchikado, H., & Dickson, D. W. (2007). Neuropathology of Parkinson's disease dementia and dementia with Lewy bodies with reference to striatal pathology. *Parkinsonism & Related Disorders*, *13*, S221–S224. doi:10.1016/S1353-8020(08)70005-1 PMID:18267239

Wakabayashi, K., Tanji, K., Mori, F., & Takahashi, H. (2007). The Lewy body in Parkinson's disease: Molecules implicated in the formation and degradation of α -synuclein aggregates. *Neuropathology*, 27(5), 494–506. doi:10.1111/j.1440-1789.2007.00803.x PMID:18018486

Walia, V. (2017). Possible Role of Serotonin and Selective Serotonin Reuptake Inhibitors in Suicidal Ideations and Attempts. *Journal of Pharmaceutical Sciences and Pharmacology*, *3*(1), 54–70. doi:10.1166/ jpsp.2017.1076

Walia, V., Sharma, A., Gahlawat, M., & Dube, O. P. (2014). Dual Role of Nitric Oxide in the Pathogenesis of Parkinson's Disease. *Journal of Pharmaceutical Sciences and Pharmacology*, *1*(4), 243–253. doi:10.1166/jpsp.2014.1038

Walsh, D. M., Lomakin, A., Benedek, G. B., Condron, M. M., & Teplow, D. B. (1997). Amyloid β-protein fibrillogenesis detection of a protofibrillar intermediate. *The Journal of Biological Chemistry*, 272(35), 22364–22372. doi:10.1074/jbc.272.35.22364 PMID:9268388

Wesnes, K. A., McKeith, I., Edgar, C., Emre, M., & Lane, R. (2005). Benefits of rivastigmine on attention in dementia associated with Parkinson disease. *Neurology*, *65*(10), 1654–1656. doi:10.1212/01. wnl.0000184517.69816.e9 PMID:16301500

Lewy Body Disease

Whitehouse, P. J., Hedreen, J. C., White, C. L., & Price, D. L. (1983). Basal forebrain neurons in the dementia of Parkinson disease. *Annals of Neurology*, *13*(3), 243–248. doi:10.1002/ana.410130304 PMID:6847136

Wild, R., Pettit, T. A., & Burns, A. (2003). Cholinesterase inhibitors for dementia with Lewy bodies. *The Cochrane Library*. PMID:12917981

Williams-Gray, C. H., Evans, J. R., Goris, A., Foltynie, T., Ban, M., Robbins, T. W., ... Barker, R. A. (2009). The distinct cognitive syndromes of Parkinson's disease: 5 year follow-up of the CamPaIGN cohort. *Brain*, *132*(11), 2958–2969. doi:10.1093/brain/awp245 PMID:19812213

Winner, B., Jappelli, R., Maji, S. K., Desplats, P. A., Boyer, L., Aigner, S., ... Tzitzilonis, C. (2011). In vivo demonstration that α-synuclein oligomers are toxic. *Proceedings of the National Academy of Sciences of the United States of America*, *108*(10), 4194–4199. doi:10.1073/pnas.1100976108 PMID:21325059

Wolozin, B., & Behl, C. (2000). Mechanisms of neurodegenerative disorders: Part 1: protein aggregates. *Archives of Neurology*, *57*(6), 793–796. doi:10.1001/archneur.57.6.793 PMID:10867775

Woollacott, M., & Shumway-Cook, A. (2002). Attention and the control of posture and gait: A review of an emerging area of research. *Gait & Posture*, *16*(1), 1–14. doi:10.1016/S0966-6362(01)00156-4 PMID:12127181

Zangaglia, R., Martignoni, E., Glorioso, M., Ossola, M., Riboldazzi, G., Calandrella, D., ... Pacchetti, C. (2007). Macrogol for the treatment of constipation in Parkinson's disease. A randomized placebocontrolled study. *Movement Disorders*, 22(9), 1239–1244. doi:10.1002/mds.21243 PMID:17566120

Zarranz, J. J., Alegre, J., Gómez-Esteban, J. C., Lezcano, E., Ros, R., Ampuero, I., ... Llorens, V. (2004). The new mutation, E46K, of α-synuclein causes parkinson and Lewy body dementia. *Annals of Neurology*, *55*(2), 164–173. doi:10.1002/ana.10795 PMID:14755719

Zesiewicz, T. A., Baker, M. J., Dunne, P. B., & Hauser, R. A. (2001). Diffuse Lewy body disease. *Current Treatment Options in Neurology*, 3(6), 507–518. doi:10.100711940-001-0013-x PMID:11581527

Zweig, R. M., Jankel, W. R., Hedreen, J. C., Mayeux, R., & Price, D. L. (1989). The pedunculopontine nucleus in Parkinson's disease. *Annals of Neurology*, *26*(1), 41–46. doi:10.1002/ana.410260106 PMID:2549845

Zweig, Y. R., & Galvin, J. E. (2014). Lewy body dementia: The impact on patients and caregivers. *Alzheimer's Research & Therapy*, 6(2), 21. doi:10.1186/alzrt251 PMID:25031635

Chapter 15 Amyotrophic Lateral Sclerosis: A Predominant Form of Degenerative Disease of the Motor Neuron System

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ABSTRACT

Amyotrophic lateral sclerosis (ALS) is a progressive neurodegenerative disorder (ND) that primarily comprises the neurons responsible for controlling voluntary muscle movement. The unique neuropathologic findings include anterior horn cell degeneration producing muscle atrophy or amyotrophy, degeneration, and sclerosis of the corticospinal tracts. It is a common neuromuscular disease worldwide and has been identified in people of all races. There seems to be neither identified risk factors nor family history associated with most of the documented ALS cases. There exists no treatment for ALS that can prevent neither its progression nor reverse its development. However, there are treatments available that can help control symptoms, prevent unnecessary complications, and make living with the disease easier. This chapter extensively discusses this neurodegenerative disorder based on the currently available knowledge on the condition.

DOI: 10.4018/978-1-5225-5282-6.ch015

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INTRODUCTION

ALS, also known as Charcot's or Lou Gehrig's disease, is a progressive and deadly neurodegenerative disorder (ND) which greatly impacts on patient's quality of life and results in death 2-5 years after diagnosis (Pasinelli & Brown, 2006). From its underlying pathophysiology, the disease was named, that is "amyotrophic" referring to atrophy or death of muscle fibers and "lateral sclerosis" referring to stiffness or hardness of the lateral column of the spinal cord as fibrous astrocytes replace degenerated axons of the upper motor neuron (Armon & Lorenzo, 2017). The disease is associated with the loss of motor neurons that regulate voluntary muscle activities which include walking, breathing, chewing, talking etc. Most importantly, neurons of the spinal cord, brainstem and those present in the motor cortex (Stamenkovic et al, 2017). The effect on the motor neurons accounts for it being also called motor neuron disease (MND).

Clinically, the hallmark of ALS is the presence of upper motor neuron (UMN) and lower motor neuron (LMN) features involving the brainstem and multiple spinal cord regions of innervation. Patients with ALS can present either with bulbar-onset disease (about 25%), limb-onset disease (about 70%), or initial trunk or respiratory involvement (5%), which subsequently spreads to involve other regions (Vucic et al, 2007). Atypical modes of presentation may include weight loss, which is indicative of poor prognosis, cramps and fasciculations in the absence of muscle weakness, emotional lability, and frontal lobe-type cognitive dysfunction (Ferguson & Elman, 2007).

Currently, there is no defined diagnostic test or biomarker for ALS hence experts only rely on its clinical presentation to diagnose. Henceforth, research to establish a novel biomarker that accurately assesses the progression of the condition is of paramount interest in improving therapeutic trial design while also decreasing cost of clinical trials. Lately, it is being progressively identified that population registries are important additions to improved clinical assessment techniques. These collaborative activities will certainly lead to a better understanding of ALS and its often random development, and will lead to the establishment of guidelines for better care of patients (Kiernan et al, 2011).

It is believed that ALS may differ based on clinical phenotype with longer survival reported in ALS without cognitive impairment (Montuschi et al, 2015). The chapter therefore discusses ALS as informed by currently available knowledge on this ND.

BACKGROUND

ALS is a clinically and genetically heterogeneous, devastating, rapidly progressive neurodegenerative motor disorder with cognitive and behavioral impairments as core features (Schmidt et al, 2016). Along-side behavioral features, executive dysfunction is present in up to 50% of ALS patients (Beeldman et al, 2016). Anatomical connectivity studies have revealed clear white matter impairments, mostly affecting tracts directly linked to the motor cortex (Schmidt et al, 2014).

Improvements in our knowledge of the glutamate neurotransmitter system coupled with the detection of causal genes associated with the progression of familial ALS (fALS) have inspired research interest (Kiernan et al, 2011). Whereas these research findings allow for vital comprehensions regarding the ultimate consequences of ALS on the brain, the causal pathogenic mechanism of ALS remains largely unknown (Schmidt et al, 2016).

Cronin et al (2007) reported that there is a low incidence of ALS in people with mixed ancestral origin than in people of Spanish origin. About 5–10% of ALS is familial, with a Mendelian pattern of inheritance. The remaining 90% of the ALS population is identified as having sporadic disease for which results from family aggregation studies have realized a connection between ALS and common NDs, indicative of the presence of susceptibility genes that might elevate the overall risk of neurodegeneration among relatives (Fallis & Hardiman, 2009). As such, attempts to identify susceptibility genes have had little influence on the establishment of a complex genetic basis for sporadic ALS.

SIGNS AND SYMPTOMS OF AMYOTROPHIC LATERAL SCLEROSIS

Muscle weakness and stiffness are the symptoms seen in the early stages of the disease which gradually affect muscles under voluntary control as the disease progresses (Javier & Rojas-garc, 2016).

UMN and LMN signs are observable on neurological examination due to the concomitant association of these neurons in the disease progression. Babinski sign is identified with UMN, as well as, spasticity and hyperreflexia while signs of LMN include cramps, muscle atrophy and fasciculation (Javier & Rojas-garc, 2016). These signs are seen in more advanced stages of the disease.

Bulbar onset ALS usually commences with symptoms such as slurred speech, croakiness or reduced speech volume as well as choking or aspiration during meals which later progresses to the development of strained, strangled vocal quality and eventual speech loss as well as swallowing difficulties and drooling (Armon & Lorenzo, 2017).

Some ALS patients may also experience symptoms such as depression, impaired executive function, maladaptive social behavior and involuntary laughing or crying due to emotional and special cognitive complications (Murphy et al, 2007). Due to atrophy of respiratory muscles, patients with ALS also suffer difficulty in breathing and eventually inability to breath, thereby depending on a ventilator. There is also an accelerated risk of pneumonia as well as painful neuropathy in these patients as the disease progresses (Oliveira & Pereira, 2009).

PATHOPHYSIOLOGICAL MECHANISMS OF MOTOR NEURON DEGENERATION IN AMYOTROPHIC LATERAL SCLEROSIS

There is a multifactorial pathophysiological mechanism underlying the development of ALS-associated motor neuron degeneration despite its exact molecular pathway not being known (Cozzolino et al, 2008; Vucic & Kiernan, 2009; Pasinelli & Brown, 2006). A probable primary mechanism is due to mutation in super oxide dismutase 1 (SOD1), such as anomalous protein aggregation, disorganization of intermediate filaments and glutamate-mediated excitotoxicity, and other abnormalities of intracellular calcium regulation in a process that may involve mitochondrial abnormalities and apoptosis (Rowland & Shneider, 2001). In addition to these, dysfunction of the sodium/potassium ion pump, autophagy and disrupted axonal transport systems have all been associated with the pathogenesis of ALS (Lederer et al, 2007; Ellis et al, 2003; Sasaki & Iwata, 1996)

Genetics

The discoveries of gene mutations causing familial forms of ALS and frontotemporal lobar dementia (FTLD) (microtubule-associated protein tau [*MAPT*]) in combination with neuropathological findings have provided crucial directions in initial research. The discovery that the first genetic causes identified for ALS (mutations in *SOD1*) and FTLD (Rosen et al, 1993, Hutton et al, 1998), each causes exclusively either ALS or FTLD was partially responsible for the delayed recognition of a linkage between ALS and FTLD (Polymenidou & Cleveland, 2017). The identification of a growing list of genetic ALS and FTLD causes and risk factors has led to the realization that errors in RNA metabolism and protein degradation are pivotal in disease initiation. Genetics and pathology of ALS and FTLD subdivide disease into *SOD1*, *TDP-43*, *FUS*, and *TDP-43/DPR* proteinopathies (Polymenidou & Cleveland, 2017).

Superoxide Dismutase 1

Discovery, Prevalence and Clinical Presentation

Superoxide dismutase 1 (SOD1) is an enzyme that in humans is encoded by the *SOD1* gene, located on chromosome 21. Mutations in *SOD1* were the first genetic causes identified in ALS (Rosen et al, 1993), and account for almost 20% of familial (Ling et al, 2013) and rare sporadic (Chio et al, 2008) ALS cases. The clinical presentation of *SOD1* mutation carriers is that of classical ALS (Hardiman et al, 2011), and although rare cases of ALS with cognitive dysfunction have been reported (Stewart et al, 2006), *SOD1*-linked ALS is typically not associated with FTLD symptoms (Wicks et al, 2009).

Mutation Distribution and Effect

More than 170 ALS-linked missense mutations spanning the entirety of the SOD1 polypeptide have been reported (Ling et al, 2013), with each mutation disrupting the physiological conformation of at least a proportion of the mutant subunits, thereby leading to the misfolding and buildup of aberrant forms. Although the misfolded protein is ubiquitinated, and thus directed for degradation (Basso et al, 2006), most mutant SOD1 escape rapid proteolysis, resulting in the development of ubiquitin-positive cytoplasmic inclusions (Kerman et al, 2010), the pathological hallmark of ALS.

Pathogenic Mechanism

Although some ALS-linked mutations decrease the enzymatic activity of SOD1, others do not (Borchelt et al, 1994, 1995). In combination with the observation that loss of SOD1 in knockout mice does not lead to motor neuron degeneration (Reaume et al, 1996) and that disease from a transgene-encoded, dismutase-inactive mutation is unaffected by reduction or elimination of endogenous SOD1 (Bruijn et al, 1998), the evidence argues strongly against a loss-of-function mechanism in SOD1-linked ALS. In contrast, expression of mutant SOD1 in transgenic mice (Gurney et al, 1994; Bruijn et al, 1997) and rats (Howland et al, 2002), or naturally occurring in dogs with canine degenerative myopathy (Crisp et al, 2013), leads to late-onset, progressive paralysis with gliosis and ubiquitinated misfolded SOD1 buildup.

TAR-DNA Binding Protein of 43 kDa

Discovery, Prevalence and Clinical Presentation

In sporadic ALS, histology has established the involvement of phosphorylated 43 kDa TAR DNAbinding protein aggregates in the disease pathology (Brettschneider et al, 2012, 2014; Neumann et al, 2006). The discovery of TDP-43 as the principal composition of the ubiquitin-positive cytoplasmic inclusions, which is in high levels in patients with sporadic ALS (Neumann et al, 2006), was a significant breakthrough. Mutations in *TARDBP* gene (encoding the TAR-DNA binding protein TDP-43) located on chromosome 1p36.22 have been associated with familial and sporadic ALS with the mutation found to cause ALS in 5% of familial cases (Gitcho et al, 2008; Van Deerlin et al, 2008; Yokoseki et al, 2008; Sreedharan et al, 2008). In most patients with TDP-43 mutations, there is a classical ALS phenotype marked by the absence of cognitive deficit, with some unevenness within families on the site of onset and age at which condition starts to develop. Although early studies were unsuccessful in establishing the existence of mutations in TDP-43 in FTLD patients, rare TDP-43 mutations in patients displaying FTLD, with or without motor neuron disease, were reported in 2009 (Benajiba et al, 2009; Borroni et al, 2009; Kovacs et al, 2009).

Mutation Distribution and Effect

The bulk of ALS-associated missense mutations in TDP-43 are restricted to the C-terminal, a glycinerich domain (Lagier-Tourenne et al, 2010; Ling et al, 2013), which contains a glutamine/asparagine-rich low complexity or prion-like domain that shares similarities with yeast prions (Fuentealba et al, 2010). The latter are proteins showing systematic, self-perpetuating aggregation, which are transmissible from an affected cell to its progeny (Chien et al, 2004; Shorter & Lindquist, 2005). It has been identified that, ALS-associated point mutations expressively increase the aggregation tendency of the already aggregation-prone prion-like domain of TDP-43 (Johnson et al, 2009; Guo et al, 2011; Molliex et al, 2015), indicating a causative link between TDP-43 aggregation and the development of ALS. However, expression of ALS-linked TDP-43 mutants in transgenic mice (Wegorzewska et al, 2009; Zhou et al, 2010; Huang et al, 2012; Arnold et al, 2013) has largely failed to reproduce the characteristic TDP-43 pathology seen in ALS patients, and although these mice develop various aberrations with aspects of motor neuron disease, they do not fully replicate the spectrum of ALS symptoms. Notwithstanding prior reports that mutant TDP-43 displays regular activity in controlling splicing of a reporter gene (D'Ambrogio et al, 2009), genome- wide analysis of mice expressing mutant TDP-43 showed broad aberrant splicing events (Arnold et al, 2013). The TDP-43 mutants moreover show defective transport of their RNA cargo across neuronal axons (Alami et al, 2014).

Pathogenic Mechanism

Two mechanisms have been implicated in TDP-43 induced toxicity. First is the loss of TDP-43 normal function following the redistribution and aggregation of the protein in the cytoplasm (Neumann et al, 2006). Secondly, it is thought that the misfolded protein assemblies in the cytoplasm are toxic to the motor neuron (Tai & Schuman, 2008). However, the relative input of each of these mechanisms is still not fully implicit. However, these two mechanisms are not independent of each other and likely cause

Amyotrophic Lateral Sclerosis

motor neuron degeneration in amalgamation (Lagier-Tourenne et al, 2010; Polymenidou et al, 2012; Ling et al, 2013). After an initiation phase, the formation of TDP-43 pre-inclusions assumes a dual effect in cells (Polymenidou & Cleveland, 2017).

Fused in Sarcoma/Translocated in Liposarcoma

Discovery and Prevalence

FUS or translocated in liposarcoma is a nuclear DNA/RNA binding protein that controls various stages of gene expression, including transcription, splicing and messenger RNA transport (Dormann & Haass, 2013; Vance et al, 2009). *FUS* has been associated with the progression of neurodegeneration since missense and nonsense mutations in the *FUS* gene contribute to approximately 4% of fALS cases and rare sporadic cases (Vance et al, 2009). In the absence of mutations, pathologic accumulations of wild type *FUS* occur in 10% of FTLD cases (Neumann et al, 2009).

Mutation Distribution and Effect

Majority of the ALS-linked *FUS* mutations are localized in the nuclear localization signal sequence (Lagier-Tourenne et al, 2010; Ling et al, 2013) and do not influence aggregation per se (Dormann et al, 2010) but instead enhance redistribution of *FUS* to the cytoplasm (Dormann et al, 2010; Ito et al, 2011), which consequently may initiate its aggregation and confer toxicity. Unlike TDP-43, *FUS* aggregation and toxicity require not just a prion-like domain but also other determinants in the RGG domain (Acton, 2012).

Pathogenic Mechanism

Mutations in the C-terminal domain affect the cytoplasmic localization of *FUS* and disrupts RNA processing and transport. The binding of a single strand DNA to the C-terminus of *FUS* is important for direct binding of *FUS* to promoters of target genes (Bronisz et al, 2014; Tan et al, 2012). The functional and structural parallels of TDP-43 and *FUS* (Lagier-Tourenne et al, 2010) indicate that the two proteins trigger ALS by an independent instigating actions, but likely via common downstream pathways, one of which involves their common RNA targets. Certainly, while TDP-43 and *FUS* bind a distinct spectrum of RNAs, they have high specificity for mRNAs produced from genes with very long introns (Polymenidou et al, 2011; Lagier-Tourenne et al, 2012). The high brain levels of these long intronic genes, tightly bound to both TDP-43 and *FUS* (Ameur et al, 2011; Polymenidou et al, 2011), presents a likely description for why neurons are typically prone to ALS pathogenic processes (Polymenidou & Cleveland, 2017).

Intronic Hexanucleotide Repeats in C9orf72

Discovery and Prevalence

Mutations in the *C9orf72* gene are the commonly associated factor for the development of amyotrophic lateral sclerosis and frontotemporal dementia (Jovicic et al, 2015). The identification of intronic hexanucleotide repeat expansions in the *C9orf72* gene of ALS patients was a significant advance in the study of ALS. These dipeptide repeat proteins (DPRs) fashioned from unusual translation of *C9orf72* repeat expansions are now identified as the most common genetic cause of ALS and FTLD, accounting for 40% of fALS and 30% of familial FTLD (fFTLD) cases (Renton et al, 2011; Gijselinck et al, 2012; Majounie et al, 2012). It has been identified that up to 90% of families with concurrent ALS and FTLD have the hexanucleotide repeat expansions in their *C9orf72* genes (Majounie et al, 2012; Rademakers & van Blitterswijk, 2013; Renton et al, 2014). However, the *C9orf72* hexanucleotide repeat expansions have also been found in approximately 7% or 5% of apparent sporadic ALS or FTLD cases, respectively, suggesting that the fraction of ALS and FTLD cases with genetic origin may be bigger than predicted (Polymenidou & Cleveland, 2017).

Mutation Distribution and Effect

In normal healthy controls, the intronic GGGGCC repeat in the *C9orf72* gene is smaller than 25 units, whereas in ALS or FTLD patients, it can increase up to 800–4400 units (Gijselinck et al, 2012), with deleterious effects such as neurodegeneration (Majounie et al, 2012; Rademakers & van Blitterswijk, 2013; Renton et al, 2014). First these large intronic expansions interfere with transcription of *C9orf72* RNA, thereby decreasing its levels (Gijselinck et al, 2012). In addition, the repeat RNA accumulates in the nuclear foci (Lagier- Tourenne et al, 2013; Zu et al, 2013; Haeusler et al, 2014) where they can be translated via the repeat-associated non-ATG (RAN) translation (Ash et al, 2013; Zu et al, 2013), resulting in the synthesis and accumulation of anomalous dipeptide proteins.

Pathogenic Mechanism

Despite several descriptions of the cytotoxic mechanisms of *C9orf72* hexanucleotide repeat expansions, their relative influence to the pathogenesis of ALS is yet to be established (Polymenidou & Cleveland, 2017). Firstly, patients with *C9orf72* expansions show typical TDP-43 cytoplasmic additions with nuclear clearance (Gijselinck et al, 2012), which suggests that all of the mechanisms described above for TDP-43 are relevant for the pathogenesis of this specific ALS phenotype. As a result of inactivation of the repeat-containing allele via promoter hypermethylation (Xi et al, 2013), or transcriptional termination of repeat RNAs (Haeusler et al, 2014), patients carrying hexanucleotide repeat expansions produce decreased levels of *C9orf72* RNA (Gijselinck et al, 2012) and, by extension, protein, which may reduce the protein's normal role, a mechanism referred to as haploinsufficiency (Polymenidou & Cleveland, 2017). In contrast, the lack of an ALS/FTLD-like phenotype in mice with heterozygous (Panda et al, 2013; Suzuki et al, 2013) or homozygous (Koppers et al, 2015) disruption of the mouse *C9orf72* homolog strongly argues against the role of haploinsufficiency in *C9orf72*-ALS/FTLD pathogenesis.

However, conflicting reports have proposed that poly-GP, a DPR, is present in UMN and LMN (Zu et al, 2013), which was subsequently shown to be in the cerebrospinal fluid of *C9orf72* patients (Su et al, 2014). On another hand, following exogenous application of the relatively short synthetic sense poly-GR and antisense poly-PR DPRs to cultured human astrocytes, these DPRs enter the nucleus where they bind to the nucleoli and induces cell destruction (Kwon et al, 2014). Lastly, expression of "protein-only" poly-GR and poly-PR constructs in *Drosophila* has been linked with progressive neuronal death *in vivo*, possibly due to the basic nature or a shared structural motif of these DPRs (Mizielinska et al, 2014). In addition to already described mechanisms, the toxic effect of these aberrant proteins can also be mediated via the compromise of the nucleocytoplasmic transport probably by interacting with karyopherin proteins or directly with the nuclear pore (Freibaum et al, 2015; Jovicic et al, 2015).

Impaired Axonal Transport

In addition to glutamate-induced excitotoxicity, mitochondrial dysfunction and oxidative stress, disrupted axonal transport systems have also been implicated in the pathogenesis of ALS (Sasaki & Iwata, 1996). Motor neuron axons may reach up to one meter in length in humans, and rely on effective intracellular transport systems which consist of anterograde (slow and fast) and retrograde transport systems. These systems depend on molecular 'motors', the kinesin complex of proteins (for anterograde) and the dynein-dynactin complex (for retrograde) (Grierson & Miller, 2006). Studies in SOD1 transgenic mouse models of ALS have yielded proof of slowed anterograde transport and retrograde transport in motor neurons (Williamson & Cleveland, 1999). Though no such observations have been realized in humans with ALS, mutations in the kinesin genes are known to cause neurodegenerative motor nerve diseases in humans which include hereditary spastic paraplegia and Type 2A Charcot-Marie-Tooth disease (Reid et al, 2002). Olney et al (2003) has reported that mutations in the dynactin complex cause a LMN disorder with vocal cord paralysis in humans.

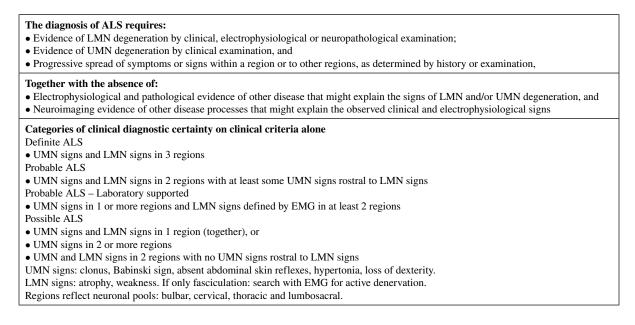
DIAGNOSIS OF AMYOTROPHIC LATERAL SCLEROSIS

The diagnosis of ALS depends on distinct clinical findings together with examinations to discount "ALSmimic" syndromes for example multifocal motor neuropathy, inclusion body myositis, conus lesions and lumbo-sacral radiculopathy (Davenport et al, 1996, Traynor et al, 2000). ALS is implicated when cerebrospinal fluid or serological studies or electrophysiological imaging reveals signs of concomitant UMN and LMN which cannot be linked with any other disease in addition to disease progression compatible with motor neuron degeneration. Isolated investigation result is therefore inadequate to arrive at a diagnosis and hence should be considered together with patient history and other clinical findings (Wijesekera & Leigh 2009).

In 1994, the World Federation of Neurology (WFN) Research Group on Motor Neuron Diseases developed the El Escorial diagnostic criteria (Maurer 2011), to help in diagnosis and classifying patients for research purposes and clinical trials (Wijesekera & Leigh 2009). However, this diagnostic criterion had several limitations which compromised its usefulness (Wilbourn 1998). The El Escorial standards can have poor sensitivity, especially in the early stages of ALS when patients are most likely to profit from therapeutic intervention (Traynor et al, 2000). Because of these drawbacks, the criteria have been revised (Table 1) to benefit early diagnosis (Ross et al, 1998) and in the clinical trial setting, improve levels of diagnostic confidence (Beghi et al, 2002).

MANAGEMENT OF AMYOTROPHIC LATERAL SCLEROSIS

The management of ALS has significantly transformed over the years, with current accent on coordinated multidisciplinary care between specialists, community based therapists and palliative care teams (Kiernan et al, 2011). This extent of survival amongst ALS patients has been found to be greater for management of patients in specialized multidisciplinary ALS clinics than in other settings (Traynor et al, 2003). In spite of the fact that ALS cannot be cured, its associated symptoms can be treated to help improve the quality of life of the patient. Advanced directives on end of life care, respiratory and nutriTable 1. A revised El Escorial research diagnostic criteria for ALS (Adapted from Brooks et al., 2000)



tional management during late stages of life are important issues, and should be discussed with patients and relatives at the earliest opportunity that they are willing to do this (Kiernan et al, 2011). Associated with ALS is psychological and emotional difficulties for the sufferer and their families (Averill et al, 2007; Wicks et al, 2007), which makes it important to offer psychological support and early palliative care to the patients and their families alike (Mitsumoto & Rabkin, 2007).

Pharmacotherapy

Riluzole

Was originally advanced as an antiepileptic drug, riluzole, currently is the only drug approved for use in slowing the progression of ALS (Bensimon et al, 1994; Kiernan et al, 2011). Although the exact mechanism of action of riluzole in modifying disease progression is not entirely elucidated, it is believed to have effect on N-methyl-D-aspartate (NMDA) receptor-mediated responses, stabilization of the inactivated state of voltage-dependent sodium channels, inhibition of glutamate release from pre-synaptic terminals, and elevating extracellular glutamate uptake (Distad et al, 2008). In two therapeutic trials, riluzole prolonged survival by three to six months (Bensimon et al, 1994; Lacomblez et al, 1996).

Lithium Carbonate

A trial of lithium carbonate in ALS compared 16 patients treated with riluzole and lithium carbonate with 28 patients treated with riluzole alone (Class III) (Fornai et al, 2008). Mortality was reduced and disease progression was curtailed in treated patients. Nonetheless, small sample size, lack of adequate blinding, and other design issues compromised the evidence provided by the trial.

Cytidine-5-Diphosphocholine

Cytidine-5-diphosphocholine (citicoline) is an endogenous nucleoside that has neuroprotective roles in certain central nervous system (CNS) injury models (Cakir et al, 2005; de Carvalho, 2008). The neuroprotective function of CDP-choline seems to be linked to its action on glutamate-mediated cell death. Citicoline might lower the extracellular level of glutamate by blockade of neuronal glutamate efflux and elevating astrocytic glutamate uptake. Matyja et al (2008) reports that the neuroprotective role of this compound is related to the blockade of the glutamate-induced apoptotic pathway of cell injury.

Erythropoietin

Erythropoietin (EPO) is believed to be one of the compounds that might play a potential neuroprotective role in ALS. There are many data supporting EPO's protective function of neurons exposed to damaging agents. As a neuroprotective agent, erythropoietin; antagonizes glutamate cytotoxic action, enhances antioxidant enzyme expression, lowers the free radical production rate and affects neurotransmitter release. EPO exerts a neuroprotective effect in the investigated model of chronic excitotoxicity mainly through inhibition of apoptotic neuronal alterations. Results from work by Nagańska et al (2010), Grasso et al (2007) and Liu et al (2008) show that EPO may be an important therapeutic drug in various neurological diseases, including ALS.

Nutrition

Results from nine studies indicate that enteral nutrition administered via PEG is probably effective in stabilizing body weight/body mass index (Desport et al, 2000; Desport et al, 2005). In patients with ALS with impaired oral food intake, enteral nutrition via PEG should be considered to stabilize body weight (Miller et al, 2009). There are no clear-cut indications for the timing of PEG insertion in ALS patients. However, Kasarskis and his colleagues demonstrated that, patients with dysphagia will possibly be exposed to reduced risk if PEG is placed when FVC is above 50% of predicted (Kasarskis et al, 1999). Risks of PEG placement include laryngeal spasm, localized infection, gastric hemorrhage, failure to place PEG due to technical difficulties, and death due to respiratory arrest (Mazzini et al, 1995).

Respiratory Support

The diagnosis and management of respiratory insufficiency is important because respiratory failure causes most deaths from ALS (Miller et al, 2009). The presenting symptoms of respiratory muscle weakness include dyspnea on exertion or talking, orthopneoa, disturbed sleep, excessive daytime somnolence, morning headaches, fatigue, anorexia, depression, poor concentration, vivid nightmares and nocturia (Leigh et al, 2003; Heffernan et al, 2006). Despite uncertainties in respiratory care of ALS patients, several controlled studies have provided evidence to guide its management (Figure 1).

Abbreviations: PFT: Pulmonary function tests; PCEF: Peak cough expiratory flow; NIV: Noninvasive ventilation; SNP: Sniff nasal pressure; MIP: Maximal inspiratory pressure; FVC: Forced vital capacity (supine or erect); Abnl.nocturnal oximetry: $pO_2 < 4\%$ from baseline.

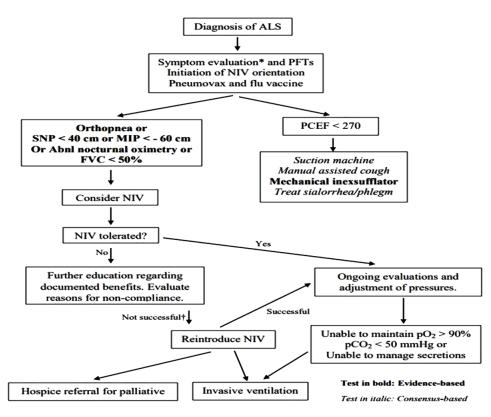


Figure 1. Algorithm for the airway management of ALS (Adapted from AAN Practice Parameter Update, 2009)

*Symptoms suggestive of nocturnal hypoventilation: frequent arousals, morning headaches, excessive daytime sleepiness, vivid dreams. †If NIV is not tolerated or accepted in the setting of advancing respiratory compromise, consider invasive ventilation or referral to hospice.

FUTURE RESEARCH DIRECTIONS

It is almost impossible to translate animal studies on ALS to humans though such studies have been ongoing for about three decades. This stems from the fact that ALS has not been identified in animals and currently studies only try to subject animals to artificial gene expressions which however differs from human ALS phenotypically (Clerc et al, 2016). Again, some drugs which had been studied to be effective in managing ALS in animals such as celecoxib (Drachman et al, 2002; Cudkpwicz et al, 2004), creatine (Klivenyi et al, 1999; Groeneveld et al, 2003), gabapentin (Gurney et al, 1996; Miller et al, 2001) and acetylcysteine (Kuther & Struppler, 1987; Andreassen et al, 2000) showed no therapeutic benefit in human ALS. This can be attributed to intraspecies differences that exist between humans and animals (Oliveira & Pereira, 2009). This makes these animal models not entirely valid.

However, with advances in molecular biology, human diseases biomonitoring and computer science, there is hope of improved research, potentially more relevant and more effective. These may include *in silico*, disease pathway approach and human-based studies. These offer ethically superior know how which are more succinct, robust, and more astute into human biology and disease hence speeding up transition from the bench to the bedside (Clerc et al, 2016).

A common mutation associated with ALS is an unstable repeated DNA sequence within the *C9orf72* gene that could reach into the thousands (Xi et al, 2015). Xi and his colleagues have also reported that less than 30 repeats of the *C9orf72* gene have been established to be common in most individuals. With further research, there could be utilization of this information to come up with genetic test(s) that can identify if parents are likely to pass a mutation that may cause ALS to their progeny. By increasing this research to include other multi-generational families, as well as study unaffected parents, there is a hope of identifying a range of *C9orf72* repeats that can be used to predict a genetic predisposition to ALS in children (Xi et al, 2015).

CONCLUSION

This chapter has constructively detailed ALS based on available sound literature on the condition. However, future advances will seek to better comprehend the symptomatic management of ALS and also provide an improved insight into the etiology of the condition. This will offer valuable piece in the research objective of finding effective therapy that does not just slow the progression but would also stop and/or reverse this deleterious neurodegenerative process.

REFERENCES

Abhinav, K., Stanton, B., Johnston, C., Hardstaff, J., Orrell, R. W., Howard, R., ... Shaw, C. E. (2007). Amyotrophic lateral sclerosis in South-East England: A population-based study. The South-East England register for amyotrophic lateral sclerosis (SEALS Registry). *Neuroepidemiology*, *29*(1-2), 44–48. doi:10.1159/000108917 PMID:17898523

Acton, Q. A. (2012). Amyotrophic Lateral Sclerosis: New insights for the healthcare professional. Scholarly Editions.

Alami, N. H., Smith, R. B., Carrasco, M. A., Williams, L. A., Winborn, C. S., Han, S. S., ... Taylor, J. P. (2014). Axonal transport of TDP-43 mRNA granules is impaired by ALS-causing mutations. *Neuron*, *81*(3), 536–543. doi:10.1016/j.neuron.2013.12.018 PMID:24507191

Alonso, A., Logroscino, G., Jick, S. S., & Hernan, M. A. (2010). Association of smoking with amyotrophic lateral sclerosis risk and survival in men and women: A prospective study. *BMC Neurology*, *10*(1), 6. doi:10.1186/1471-2377-10-6 PMID:20074360

Ameur, A., Zaghlool, A., Halvardson, J., Wetterbom, A., Gyllensten, U., Cavelier, L., & Feuk, L. (2011). Total RNA sequencing reveals nascent transcription and widespread co-transcriptional splicing in the human brain. *Nature Structural & Molecular Biology*, *18*(12), 1435–1440. doi:10.1038/nsmb.2143 PMID:22056773

Andersen, P. M., Borasio, G. D., Dengler, R., Hardiman, O., Kollewe, K., Leigh, P. N., ... Tomik, B. (2005). EFNS task force on management of amyotrophic lateral sclerosis: Guidelines for diagnosing and clinical care of patients and relatives. *European Journal of Neurology*, *12*(12), 921–938. doi:10.1111/j.1468-1331.2005.01351.x PMID:16324086

Andreassen, O. A., Dedeoglu, A., Klivenyi, P., Beal, M. F., & Bush, A. I. (2000). N-acetyl-L-cysteine improves survival and preserves motor performance in an animal model of familial amyotrophic lateral sclerosis. *Neuroreport*, *11*(11), 2491–2493. doi:10.1097/00001756-200008030-00029 PMID:10943709

Armon, C. (2003). An evidence-based medicine approach to the evaluation of the role of exogenous risk factors in sporadic amyotrophic lateral sclerosis. *Neuroepidemiology*, 22(4), 217–228. doi:10.1159/000070562 PMID:12792141

Armon, C. (2009). Smoking may be considered an established risk factor for sporadic ALS. *Neurology*, 73(20), 1693–1698. doi:10.1212/WNL.0b013e3181c1df48 PMID:19917993

Armon, C., & Lorenzo, N. (2017). *Amyotrophic lateral sclerosis*. *Practical essentials*. Available at http://emedicine.medscape.com/article/1170097-overview

Ash, P. E. A., Bieniek, K. F., Gendron, T. F., Caulfield, T., Lin, W. L., Dejesus-Hernandez, M., ... Joseph, W. (2013). Unconventional translation of C9ORF72 GGGGCC expansion generates insoluble polypeptides specific to c9FTD/ALS. *Neuron*, 77(4), 639–646. doi:10.1016/j.neuron.2013.02.004 PMID:23415312

Averill, A. J., Kasarskis, E. J., & Segerstrom, S. C. (2007). Psychological health in patients with amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases*, 8(4), 243–254. doi:10.1080/17482960701374643 PMID:17653923

Basso, M., Massignan, T., Samengo, G., Cheroni, C., De Biasi, S., Salmona, M., ... Bonetto, V. (2006). Insoluble mutant SOD1 is partly oligoubiquitinated in amyotrophic lateral sclerosis mice. *The Journal of Biological Chemistry*, *281*(44), 33325–33335. doi:10.1074/jbc.M603489200 PMID:16943203

Beeldman, E., Raaphorst, J., Twennaar, M. K., de Visser, M., Schmand, B. A., & de Haan, R. J. (2016). The cognitive profile of ALS: A systematic review and metaanalysis update. *Journal of Neurology, Neurosurgery, and Psychiatry*, 87(6), 611–619. doi:10.1136/jnnp-2015-310734 PMID:26283685

Beghi, E. (2013). Are professional soccer players at higher risk for ALS? *Amyotrophic Lateral Sclerosis & Frontotemporal Degeneration*, *14*(7-8), 501–506. doi:10.3109/21678421.2013.809764 PMID:23859483

Beghi, E., Balzarini, C., Bogliun, G., Logroscino, G., Manfredi, L., Mazzini, L., ... Vitelli, E. (2002). Reliability of the El Escorial diagnostic criteria for amyotrophic lateral sclerosis. *Neuroepidemiology*, *21*(6), 265–270. doi:10.1159/000065524 PMID:12411728

Beghi, E., Millul, A., Micheli, A., Vitelli, E., & Logroscino, G. (2007). Incidence of ALS in Lombardy, Italy. *Neurology*, *68*(2), 141–145. doi:10.1212/01.wnl.0000250339.14392.bb PMID:17210896

Benajiba, L., Le Ber, I., Camuzat, A., Lacoste, M., Thomas-Anterion, C., Couratier, P., ... Brice, A. (2009). TARDBP mutations in motoneuron disease with frontotemporal lobar degeneration. *Annals of Neurology*, 65(4), 470–473. doi:10.1002/ana.21612 PMID:19350673

Amyotrophic Lateral Sclerosis

Bensimon, G., Lacomblez, L., & Meininger, V. (1994). A controlled trial of riluzole in amyotrophic lateral sclerosis: ALS/Riluzole Study Group. *The New England Journal of Medicine*, *330*(9), 585–591. doi:10.1056/NEJM199403033300901 PMID:8302340

Bentmann, E., Neumann, M., Tahirovic, S., Rodde, R., Dormann, D., & Haass, C. (2012). Requirements for stress granule recruitment of fused in sarcoma (FUS) and TAR DNA-binding protein of 43 kDa (TDP-43). *The Journal of Biological Chemistry*, 287(27), 23079–23094. doi:10.1074/jbc.M111.328757 PMID:22563080

Borchelt, D. R., Guarnieri, M., Wong, P. C., Lee, M. K., Slunt, H. S., Xu, Z. S., ... Cleveland, D. W. (1995). Superoxide dismutase 1 subunits with mutations linked to familial amyotrophic lateral sclerosis do not affect wild-type subunit function. *The Journal of Biological Chemistry*, 270(7), 3234–3238. doi:10.1074/jbc.270.7.3234 PMID:7852409

Borchelt, D. R., Lee, M. K., Slunt, H. S., Guarnieri, M., Xu, Z. S., Wong, P. C., Brown, R. H., Jr., & Cleveland, D. W. (1994). Superoxide dismutase 1 with mutations linked to familial amyotrophic lateral sclerosis possesses significant activity. *Proceedings of the National Academy of Sciences*, *91*, 8292–8296.

Borroni, B., Bonvicini, C., Alberici, A., Buratti, E., Agosti, C., Archetti, S., ... Padovani, A. (2009). Mutation within TARDBP leads to frontotemporal dementia without motor neuron disease. *Human Mutation*, *30*(11), E974–E983. doi:10.1002/humu.21100 PMID:19655382

Brettschneider, J., Libon, D. J., Toledo, J. B., Xie, S. X., McCluskey, L., Elman, L., ... Trojanowski, J. Q. (2012). Microglial activation and TDP-43 pathology correlate with executive dysfunction in amyotrophic lateral sclerosis. *Acta Neuropathologica*, *123*(3), 395–407. doi:10.100700401-011-0932-x PMID:22210083

Bronisz, A., Carey, H. A., Godlewski, J., Sif, S., Ostrowski, M. C., & Sharma, S. M. (2014). The multifunctional protein Fused in Sarcoma (FUS) is a coactivator of Microphthalmia-associated Transcription Factor (MITF). *The Journal of Biological Chemistry*, *289*(1), 326–334. doi:10.1074/jbc.M113.493874 PMID:24257758

Brooks, B. R., Miller, R. G., Swash, M., & Munsat, T. L. (2000). El Escorial revisited: Revised criteria for the diagnosis of amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, *41*(5), 293–299. doi:10.1080/146608200300079536 PMID:11464847

Bruijn, L. I., Becher, M. W., Lee, M. K., Anderson, K. L., Jenkins, N. A., Copeland, N. G., ... Cleveland, D. W. (1997). ALS-linked SOD1 mutant G85R mediates damage to astrocytes and promotes rapidly progressive disease with SOD1-containing inclusions. *Neuron*, *18*(2), 327–338. doi:10.1016/S0896-6273(00)80272-X PMID:9052802

Bruijn, L. I., Houseweart, M. K., Kato, S., Anderson, K. L., Anderson, S. D., & Ohama, E. (1851–1854). ... Cleveland, D. W. (1998). Aggregation and motor neuron toxicity of an ALS-linked SOD1 mutant independent from wild-type SOD1. *Science*, 281.

Cakir, E., Usul, H., Peksoylu, B., Sayin, O. C., Alver, A., Topbas, M., ... Kuzeyli, K. (2005). Effects of citicoline on experimental spinal cord injury. *Journal of Clinical Neuroscience*, *12*(8), 923–926. doi:10.1016/j.jocn.2005.03.013 PMID:16257217

Chien, P., Weissman, J. S., & DePace, A. H. (2004). Emerging principles of conformation-based prion inheritance. *Annual Review of Biochemistry*, 73(1), 617–656. doi:10.1146/annurev.biochem.72.121801.161837 PMID:15189155

Chio, A., Traynor, B. J., Lombardo, F., Fimognari, M., Calvo, A., Ghiglione, P., ... Restagno, G. (2008). Prevalence of SOD1 mutations in the Italian ALS population. *Neurology*, *70*(7), 533–537. doi:10.1212/01. wnl.0000299187.90432.3f PMID:18268245

Clerc, P., Lipnick, S., & Willett, C. (2016). A look into the future of ALS research. *Drug Discovery Today*, 21(6), 939–949. doi:10.1016/j.drudis.2016.02.002 PMID:26861067

Cozzolino, M., Ferri, A., & Carri, M. T. (2008). Amyotrophic lateral sclerosis: From current developments in the laboratory to clinical implications. *Antioxidants & Redox Signalling*, *10*(3), 405–443. doi:10.1089/ars.2007.1760 PMID:18370853

Crisp, M. J., Beckett, J., Coates, J. R., & Miller, T. M. (2013). Canine degenerative myelopathy: Biochemical characterization of superoxide dismutase 1 in the first naturally occurring non-human amyotrophic lateral sclerosis model. *Experimental Neurology*, 248, 1–9. doi:10.1016/j.expneurol.2013.05.009 PMID:23707216

Cronin, S., Hardiman, O., & Traynor, B. J. (2007). Ethnic variation in the incidence of ALS: A systematic review. *Neurology*, 68(13), 1002–1007. doi:10.1212/01.wnl.0000258551.96893.6f PMID:17389304

Cudkowicz, M. E., Shefner, J. M., Schoenfeld, D. A., Zhang, H., Andreasson, K. I., Rothstein, J. D., & Drachman, D. B. (2004). Clinical trial of celecoxib in subjects with amyotrophic lateral sclerosis. *Annals of Neurology*, *5*, 25–26.

Davenport, R. J., Swingler, R. J., Chancellor, A. M., & Warlow, C. P. (1996). Avoiding false positive diagnoses of motor neuron disease: Lessons from the Scottish Motor Neuron Disease Register. *Journal of Neurology, Neurosurgery, and Psychiatry*, 60(2), 147–151. doi:10.1136/jnnp.60.2.147 PMID:8708642

de Carvalho, M., Dengler, R., Eisen, A., England, J. D., Kaji, R., Kimura, J., ... Swash, M. (2008). Electrodiagnostic criteria for diagnosis of ALS. *Clinical Neurophysiology*, *119*(3), 497–503. doi:10.1016/j. clinph.2007.09.143 PMID:18164242

de Jong, S. W., Huisman, M. H., Sutedja, N. A., van der Kooi, A. J., de Visser, M., Schelhaas, H. J., ... van den Berg, L. H. (2012). Smoking, alcohol consumption, and the risk of amyotrophic lateral sclerosis: A population-based study. *American Journal of Epidemiology*, *176*(3), 233–239. doi:10.1093/aje/kws015 PMID:22791740

Desport, J. C., Mabrouk, T., Bouillet, P., Perna, A., Preux, P. M., & Couratier, P. (2005). Complications and survival following radiologically and endoscopically-guided gastrostomy in patients with amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, 6(2), 88–93. doi:10.1080/14660820410021258a PMID:16036431

Desport, J. C., Preux, P. M., Truong, C. T., Courat, L., Vallat, J. M., & Couratier, P. (2000). Nutritional assessment and survival in ALS patients. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, *1*, 91–96. doi:10.1080/14660820050515386 PMID:11467055

Amyotrophic Lateral Sclerosis

Distad, B. J., Meekins, G. D., Liou, L. L., Weiss, M. D., Carter, G. T., & Miller, R. G. (2008). Drug Therapy in Amyotrophic Lateral Sclerosis. *Physical Medicine and Rehabilitation Clinics of North America*, *19*(3), 633–651. doi:10.1016/j.pmr.2008.04.005 PMID:18625421

Ditsworth, D., Maldonado, M., McAlonis-Downes, M., Sun, S., Seelman, A., Drenner, K., ... Da Cruz, S. (2017). Mutant TDP-43 within motor neurons drives disease onset but not progression in amyotrophic lateral sclerosis. *Acta Neuropathologica*, *133*(6), 907–922. doi:10.100700401-017-1698-6 PMID:28357566

Dormann, D., & Haass, C. (2013). Fused in sarcoma (FUS): An oncogene goes awry in neurodegeneration. *Molecular and Cellular Neurosciences*, 56, 475–286. doi:10.1016/j.mcn.2013.03.006 PMID:23557964

Dormann, D., Rodde, R., Edbauer, D., Bentmann, E., Fischer, I., Hruscha, A., ... Haass, C. (2010). ALSassociated fused in sarcoma (FUS) mutations disrupt Transportin-mediated nuclear import. *The EMBO Journal*, 29(16), 2841–2857. doi:10.1038/emboj.2010.143 PMID:20606625

Drachman, D. B., Frank, K., Dykes-Hoberg, M., Teismann, P., Almer, G., Przedborski, S., & Rothstein, J. D. (2002). Cyclooxygenase 2 inhibition protects motor neurons and prolongs survival in a transgenic mouse model of ALS. *Annals of Neurology*, *52*(6), 771–778. doi:10.1002/ana.10374 PMID:12447931

Ellis, D. Z., Rabe, J., & Sweadner, K. J. (2003). Global loss of Na,K-ATPase and its nitric oxide-mediated regulation in a transgenic mouse model of amyotrophic lateral sclerosis. *The Journal of Neuroscience*, 23, 43–51. PMID:12514200

Fallis, B. A., & Hardiman, O. (2009). Aggregation of neurodegenerative disease in ALS kindreds. *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases, 10*(2), 95–98. doi:10.1080/17482960802209664 PMID:18608094

Fang, F., Bellocco, R., Hernan, M. A., & Ye, W. (2006). Smoking, snuff dipping and the risk of amyotrophic lateral sclerosis—a prospective cohort study. *Neuroepidemiology*, 27(4), 217–221. doi:10.1159/000096956 PMID:17106211

Fang, F., Hallmarker, U., James, S., Ingre, C., Michaelsson, K., Ahlbom, A., & Feychting, M. (2016). Amyotrophic lateral sclerosis among cross-country skiers in Sweden. *European Journal of Epidemiology*, *31*(3), 247–253. doi:10.100710654-015-0077-7 PMID:26220522

Ferguson, T. A., & Elman, L. B. (2007). Clinical presentation and diagnosis of amyotrophic lateral sclerosis. *NeuroRehabilitation*, 22, 409–416. PMID:18198425

Fornai, F., Longone, P., Cafaro, L., Kastsiuchenka, O., Ferrucci, M., & Manca, M. L., ... Paparelli, A. (2008). Lithium delays progression of amyotrophic lateral sclerosis. *Proceedings of the National Academy of Sciences USA*, *105*, 2052–2057. 10.1073/pnas.0708022105

Freibaum, B. D., Lu, Y., Lopez-Gonzalez, R., Kim, N. C., Almeida, S., Lee, K. H., ... Wong, P. C. (2015). GGGGCC repeat expansion in C9orf72 compromises nucleocytoplasmic transport. *Nature*, *525*(7567), 129–133. doi:10.1038/nature14974 PMID:26308899

Gijselinck, I., Van Langenhove, T., van der Zee, J., Sleegers, K., Philtjens, S., Kleinberger, G., ... Van Broeckhoven, C. (2012). A C9orf72 promoter repeat expansion in a Flanders–Belgian cohort disorders of the frontotemporal lobar degeneration- amyotrophic lateral sclerosis spectrum: A gene identification study. *Lancet Neurology*, *11*(1), 54–65. doi:10.1016/S1474-4422(11)70261-7 PMID:22154785

Gitcho, M. A., Baloh, R. H., Chakraverty, S., Mayo, K., Norton, J. B., Levitch, D., ... Cairns, N. J. (2008). TDP-43 A315T mutation in familial motor neuron disease. *Annals of Neurology*, *63*(4), 535–538. doi:10.1002/ana.21344 PMID:18288693

Gouveia, L. O., & De Carvalho, M. (2007). Young-onset sporadic amyotrophic lateral sclerosis: A distinct nosological entity? *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation* of Neurology Research Group on Motor Neuron Diseases, 1–5. PMID:17852021

Grasso, G., Sfacteria, A., Meli, F., Passalacqua, M., Fodale, V., Buemi, M., ... Tomasello, F. (2007). The role of erythropoietin in neuroprotection: Therapeutic perspectives. *Drug News & Perspectives*, 20(5), 315–320. doi:10.1358/dnp.2007.20.5.1120219 PMID:17878959

Grierson, A. J., & Miller, C. (2006). Axonal transport and amyotrophic lateral sclerosis. In Amyotrophic Lateral Sclerosis (2nd ed.). Informa Healthcare Journal.

Groeneveld, G. J., Veldink, J. H., van der Tweel, I., Kalmijn, S., Beijer, C., de Visser, M., ... van den Berg, L. H. (2003). A randomized sequential trial of creatine in amyotrophic lateral sclerosis. *Annals of Neurology*, *53*(4), 437–445. doi:10.1002/ana.10554 PMID:12666111

Guo, W., Chen, Y., Zhou, X., Kar, A., Ray, P., Chen, X., ... Zhu, L. (2011). An ALS-associated mutation affecting TDP-43 enhances protein aggregation, fibril formation and neurotoxicity. *Nature Structural & Molecular Biology*, *18*(7), 822–830. doi:10.1038/nsmb.2053 PMID:21666678

Gurney, M. E., Cutting, F. B., Zhai, P., Doble, A., Taylor, C. P., Andrus, P. K., & Hall, E. D. (1996). Benefit of vitamin E, riluzole, and gabapentin in a transgenic model of familial amyotrophic lateral sclerosis. *Annals of Neurology*, *39*(2), 147–157. doi:10.1002/ana.410390203 PMID:8967745

Gurney, M. E., Pu, H., Chiu, A. Y., Dal Canto, M. C., Polchow, C. Y., & Alexander, D. D., ... Deng, H. X. (1994). Motor neuron degeneration in mice that express a human Cu,Zn superoxide dismutase mutation. *Science*, 264.

Haeusler, A. R., Donnelly, C. J., Periz, G., Simko, E. A., Shaw, P. G., Kim, M. S., ... Sattler, R. (2014). C9orf72 nucleotide repeat structures initiate molecular cascades of disease. *Nature*, *507*(7491), 195–200. doi:10.1038/nature13124 PMID:24598541

Hardiman, O., van den Berg, L. H., & Kiernan, M. C. (2011). Clinical diagnosis and management of amyotrophic lateral sclerosis. *Nature Reviews. Neurology*, 7(11), 639–649. doi:10.1038/nrneurol.2011.153 PMID:21989247

Harwood, C. A., McDermott, C. J., & Shaw, P. J. (2009). Physical activity as an exogenous risk factor in motor neuron disease (MND): A review of the evidence. *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases, 10*(4), 191–204. doi:10.1080/17482960802549739 PMID:19263258

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Amyotrophic Lateral Sclerosis

Heffernan, C., Jenkinson, C., Holmes, T., Macleod, H., Kinnear, W., Oliver, D., ... Ampong, M. A. (2006). Management of respiration in MND/ ALS patients: An evidence based review. *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases*, 7(1), 5–15. doi:10.1080/14660820510043235 PMID:16546753

Howland, D. S., Liu, J., She, Y., Goad, B., Maragakis, N. J., Kim, B., ... Rothstein, J. D. (2002). Focal loss of the glutamate transporter EAAT2 in a transgenic rat model of SOD1 mutant-mediated amyotrophic lateral sclerosis (ALS). *Proceedings of the National Academy of Sciences*, *99*, 1604–1609. 10.1073/ pnas.032539299

Huisman, M. H., Seelen, M., de Jong, S. W., Dorresteijn, K. R., van Doormaal, P. T., van der Kooi, A. J., ... Veldink, J. H. (2013). Lifetime physical activity and the risk of amyotrophic lateral sclerosis. *Journal of Neurology, Neurosurgery, and Psychiatry*, 84(9), 976–981. doi:10.1136/jnnp-2012-304724 PMID:23418211

Hutton, M., Lendon, C. L., Rizzu, P., Baker, M., Froelich, S., Houlden, H., ... Heutink, P. (1998). Association of missense and 5'-splice-site mutations in tau with the inherited dementia FTDP-17. *Nature*, *393*(6686), 702–705. doi:10.1038/31508 PMID:9641683

Ito, D., Seki, M., Tsunoda, Y., Uchiyama, H., & Suzuki, N. (2011). Nuclear transport impairment of amyotrophic lateral sclerosis-linked mutations in FUS/TLS. *Annals of Neurology*, 69(1), 152–162. doi:10.1002/ana.22246 PMID:21280085

Javier, F., & Rojas-garc, R. (2016). ALS: A bucket of genes, environment, metabolism and unknown ingredients. *Progress in Neurobiology*. doi:10.1016/j.pneurobio.2016.05.004 PMID:27236050

Jiang, J., Zhu, Q., Gendron, T. F., Saberi, S., McAlonis-Downes, M., Seelman, A., ... Schulte, D. (2016). Gain of toxicity from ALS/FTD-linked repeat expansions in C9ORF72 is alleviated by antisense oligonucleotides targeting GGGGCC-containing RNAs. *Neuron*, *90*(3), 535–550. doi:10.1016/j. neuron.2016.04.006 PMID:27112497

Johnson, B. S., Snead, D., Lee, J. J., McCaffery, J. M., Shorter, J., & Gitler, A. D. (2009). TDP-43 is intrinsically aggregation-prone, and amyotrophic lateral sclerosis-linked mutations accelerate aggregation and increase toxicity. *The Journal of Biological Chemistry*, 284(30), 20329–20339. doi:10.1074/jbc.M109.010264 PMID:19465477

Jovicic, A., Mertens, J., Boeynaems, S., Bogaert, E., Chai, N., Yamada, S. B., ... Gitler, A. D. (2015). Modifie.rs of C9orf72 dipeptide repeat toxicity connect nucleocytoplasmic transport defects to FTD/ ALS Nature. Neuroscience, 18, 1226–1229. PMID:26308983

Kasarskis, E. J., Scarlata, D., Hill, R., Fuller, C., Stambler, N., & Cedarbaum, J. M. (1999). A retrospective study of percutaneous endoscopic gastrostomy in ALS patients during the BDNF and CNTF trials. *Journal of the Neurological Sciences*, *169*(1-2), 118–125. doi:10.1016/S0022-510X(99)00230-0 PMID:10540019

Kerman, A., Liu, H. N., Croul, S., Bilbao, J., Rogaeva, E., Zinman, L., ... Chakrabartty, A. (2010). Amyotrophic lateral sclerosis is a non-amyloid disease in which extensive misfolding of SOD1 is unique to the familial form. *Acta Neuropathologica*, *119*(3), 335–344. doi:10.100700401-010-0646-5 PMID:20111867

Kiernan, M. C., Vucic, S., Cheah, B. C., Turner, M. R., Eisen, A., Hardiman, O., ... Zoing, M. C. (2011). Amyotrophic lateral sclerosis. *Lancet*, *377*(9769), 942–955. doi:10.1016/S0140-6736(10)61156-7 PMID:21296405

Klivenyi, P., Ferrante, R. J., Matthews, R. T., Bogdanov, M. B., Klein, A. M., Andreassen, O. A., ... Beal, M. F. (1999). Neuroprotective effects of creatine in a transgenic animal model of amyotrophic lateral sclerosis. *Nature Medicine*, *5*(3), 347–350. doi:10.1038/6568 PMID:10086395

Koppers, M., Blokhuis, A. M., Westeneng, H. J., Terpstra, M. L., Zundel, C. A., Vieira de Sa, R., ... Veldink, J. H. (2015). C9orf72 ablation in mice does not cause motor neuron degeneration or motor deficits. *Annals of Neurology*, 78(3), 426–438. doi:10.1002/ana.24453 PMID:26044557

Kovacs, G. G., Murrell, J. R., Horvath, S., Haraszti, L., Majtenyi, K., Molnar, M. J., ... Spina, S. (2009). TARDBP variation associated with frontotemporal dementia, supranuclear gaze palsy, and chorea. *Movement Disorders*, *24*(12), 1843–1847. doi:10.1002/mds.22697 PMID:19609911

Kuther, G., & Struppler, A. (1987). Therapeutic trial with N-acetylcysteine in amyotrophic lateral sclerosis. *Advances in Experimental Medicine and Biology*, 209, 281–284. PMID:3577918

Kwon, I., Xiang, S., Kato, M., Wu, L., Theodoropoulos, P., Wang, T., ... McKnight, S. L. (2014). Polydipeptides encoded by the C9orf72 repeats bind nucleoli, impede RNA biogenesis, and kill cells. *Science*, *345*(6201), 1139–1145. doi:10.1126cience.1254917 PMID:25081482

Lacomblez, L., Bensimon, G., Leigh, P. N., Guillet, P., Powe, L., Durrleman, S., ... Meininger, V. (1996). A confirmatory dose ranging study of riluzole in ALS. *Neurology*, *47*(4), S242–S250. doi:10.1212/WNL.47.6_Suppl_4.242S PMID:8959996

Lagier-Tourenne, C., Baughn, M., Rigo, F., Sun, S., Liu, P., Li, H. R., ... Katz, M. (2013). Targeted degradation of sense and antisense C9orf72 RNA foci as therapy for ALS and frontotemporal degeneration. *Proceedings of the National Academy of Sciences*, *110*, E4530–E4539.

Lagier-Tourenne, C., Polymenidou, M., & Cleveland, D. W. (2010). TDP-43 and FUS/TLS: Emerging roles in RNA processing and neurodegeneration. *Human Molecular Genetics*, *19*(R1), R46–R64. doi:10.1093/hmg/ddq137 PMID:20400460

Lagier-Tourenne, C., Polymenidou, M., Hutt, K. R., Vu, A. Q., Baughn, M., Huelga, S. C., ... Yeo, G. W. (2012). Divergent roles of ALS-linked proteins FUS/TLS and TDP-43 intersect in processing long pre-mRNAs. *Nature Neuroscience*, *15*(11), 1488–1497. doi:10.1038/nn.3230 PMID:23023293

Lederer, C. W., Torrisi, A., Pantelidou, M., Santama, N., & Cavallaro, S. (2007). Pathways and genes differentially expressed in the motor cortex of patients with sporadic amyotrophic lateral sclerosis. *BMC Genomics*, 8(1), 26. doi:10.1186/1471-2164-8-26 PMID:17244347

Leigh, P. N., Abrahams, S., Al-Chalabi, A., Ampong, M. A., Goldstein, L. H., Johnson, J., ... Rio, A. (2003). The management of motor neurone disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 74(4), 32–47. PMID:14645465

Amyotrophic Lateral Sclerosis

Ling, S. C., Polymenidou, M., & Cleveland, D. W. (2013). Converging mechanisms in ALS and FTD: Disrupted RNA and protein homeostasis. *Neuron*, *79*(3), 416–438. doi:10.1016/j.neuron.2013.07.033 PMID:23931993

Liu, X. B., Wang, J. A., Yu, S. P., Keogh, C. L. & Wei, L. (2008). Therapeutic strategy of erythropoietin in neurological disorders. *Central Nervous System and Neurological Disorders - Drug Targets*, 7, 227-234.

Luna, J., Logroscino, G., Couratier, P., & Marin, B. (2017). Current issues in ALS epidemiology: Variation of ALS occurrence between populations and physical activity as a risk factor. *Revue Neurologique*, *173*(5), 244–253. doi:10.1016/j.neurol.2017.03.035 PMID:28477849

Majounie, E., Renton, A. E., Mok, K., Dopper, E. G., Waite, A., Rollinson, S., ... Traynor, B. J. (2012). Frequency of the C9orf72 hexanucleotide repeat expansion in patients with amyotrophic lateral sclerosis and frontotemporal dementia: A crosssectional study. *Lancet Neurology*, *11*(4), 323–330. doi:10.1016/S1474-4422(12)70043-1 PMID:22406228

Matyja, E., Taraszewska, A., Nagańska, E., Grieb, P., & Rafałowska, J. (2008). CDP-choline protects motor neurons against apoptotic changes in a model of chronic glutamate excitotoxicity *in vitro*. *Folia Neuropathologica*, *46*, 139–148. PMID:18587708

Mazzini, L., Corra, T., Zaccala, M., Mora, G., Del Piano, M., & Galante, M. (1995). Percutaneous endoscopic gastrostomy and enteral nutrition in amyotrophic lateral sclerosis. *Journal of Neurology*, 242(10), 695–698. doi:10.1007/BF00866922 PMID:8568533

Miller, R. G., Jackson, C. E., Kasarskis, E. J., England, J. D., Forshew, D., Johnson, W., ... Wooley, S. C. (2009). Practice Parameter update: The care of the patient with amyotrophic lateral sclerosis: Drug, nutritional, and respiratory therapies (an evidence-based review): Report of the Quality Standards Subcommittee of the American Academy of Neurology. *Neurology*, *73*(15), 1218–1226. doi:10.1212/WNL.0b013e3181bc0141 PMID:19822872

Miller, R. G., Moore, D. H., Gelinas, D. F., Dronsky, V., Mendoza, M., Barohn, R. J., ... Olney, R. (2001). Phase III randomized trial of gabapentin in patients with amyotrophic lateral sclerosis. *Neurology*, *56*(7), 843–848. doi:10.1212/WNL.56.7.843 PMID:11294919

Mitsumoto, H., & Rabkin, J. G. (2007). Palliative care for patients with amyotrophic lateral sclerosis: Prepare for the worst and hope for the best. *Journal of the American Medical Association*, 298(2), 207–216. doi:10.1001/jama.298.2.207 PMID:17622602

Mizielinska, S., Gronke, S., Niccoli, T., Ridler, C. E., Clayton, E. L., Devoy, A., ... Pietrzyk, J. (2014). C9orf72 repeat expansions cause neurodegeneration in Drosophila through arginine-rich proteins. *Science*, *345*(6201), 1192–1194. doi:10.1126cience.1256800 PMID:25103406

Molliex, A., Temirov, J., Lee, J., Coughlin, M., Kanagaraj, A. P., Kim, H. J., ... Taylor, J. P. (2015). Phase separation by low complexity domains promotes stress granule assembly and drives pathological fibrillization. *Cell*, *163*(1), 123–133. doi:10.1016/j.cell.2015.09.015 PMID:26406374

Montuschi, A., Iazzolino, B., Calvo, A., Moglia, C., Lopiano, L., Restagno, G., ... Chiò, A. (2015). Cognitive correlates in amyotrophic lateral sclerosis: A population-based study in Italy. *Journal of Neurology, Neurosurgery, and Psychiatry*, *86*(2), 168–173. doi:10.1136/jnnp-2013-307223 PMID:24769471 Murphy, J., Henry, R., & Lomen-Hoerth, C. (2007). Establishing subtypes of the continuum of frontal lobe impairment in amyotrophic lateral sclerosis. *Archives of Neurology*, *64*(3), 330–334. doi:10.1001/ archneur.64.3.330 PMID:17353375

Nagańska, E., Taraszewska, A., Matyja, E., Grieb, P., & Rafałowska, J. (2010). Neuroprotective effect of erythropoietin in amyotrophic lateral sclerosis (ALS) model in vitro. Ultrastructural study. *Folia Neuropathologica*, *48*, 35–44. PMID:20383809

Nelson, L. M., McGuire, V., Longstreth, W. T., & Matkin, C. (2000). Population-based case-control study of amyotrophic lateral sclerosis in western Washington State. I. Cigarette smoking and alcohol consumption. *American Journal of Epidemiology*, *151*(2), 156–163. doi:10.1093/oxfordjournals.aje. a010183 PMID:10645818

Neumann, M., Rademakers, R., Roeber, S., Baker, M., Kretzschmar, H., & Mackenzie, I. R. (2009). A new subtype of frontotemporal lobar degeneration with FUS pathology. *Brain*, *132*(11), 2922–2931. doi:10.1093/brain/awp214 PMID:19674978

Neumann, M., Sampathu, D. M., Kwong, L. K., Truax, A. C., Micsenyi, M. C., Chou, T. T., ... Lee, V. M.-Y. (2006). Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science*, *314*(5796), 130–133. doi:10.1126cience.1134108 PMID:17023659

Oliveira, A. S. B., & Pereira, R. D. B. (2009). Amytrophic lateral Sclerosis (ALS): Three letters that change people's life. For ever. *Arquivos de Neuro-Psiquiatria*, 67(3a), 750–782. doi:10.1590/S0004-282X2009000400040 PMID:19722069

Panda, S. K., Wefers, B., Ortiz, O., Floss, T., Schmid, B., Haass, C., ... Kuhn, R. (2013). Highly efficient targeted mutagenesis in mice using TALENs. *Genetics*, *195*(3), 703–713. doi:10.1534/genet-ics.113.156570 PMID:23979585

Pasinelli, P., & Brown, R. H. (2006). Molecular biology of amyotrophic lateral sclerosis: Insights from genetics. *Nature Reviews. Neuroscience*, 7(9), 710–723. doi:10.1038/nrn1971 PMID:16924260

Piazza, O., Sirén, A. L., & Ehrenreich, H. (2004). Soccer, neurotrauma and amyotrophic lateral sclerosis: Is there a connection? *Current Medical Research and Opinion*, 20(4), 505–508. doi:10.1185/030079904125003296 PMID:15119987

Polymenidou, M., & Cleveland, D. W. (2017). Biological spectrum of Amyotrophic lateral sclerosis. *Cold Spring Harbor Perspectives in Medicine*, 7(11), a024133. doi:10.1101/cshperspect.a024133 PMID:28062558

Polymenidou, M., Lagier-Tourenne, C., Hutt, K. R., Bennett, C. F., Cleveland, D. W., & Yeo, G. W. (2012). Misregulated RNA processing in amyotrophic lateral sclerosis. *Brain Research*, *1462*, 3–15. doi:10.1016/j.brainres.2012.02.059 PMID:22444279

Polymenidou, M., Lagier-Tourenne, C., Hutt, K. R., Huelga, S. C., Moran, J., Liang, T. Y., ... Cleveland, D. W. (2011). Long pre-mRNA depletion and RNA missplicing contribute to neuronal vulnerability from loss of TDP-43. *Nature Neuroscience*, *14*(4), 459–468. doi:10.1038/nn.2779 PMID:21358643

Rabin, S. J., Kim, J. M., Baughn, M., Libby, R. T., Kim, Y. J., Fan, Y., ... Ravits, J. (2010). Sporadic ALS has compartment-specific aberrant exon splicing and altered cell-matrix adhesion biology. *Human Molecular Genetics*, *19*(2), 313–328. doi:10.1093/hmg/ddp498 PMID:19864493

Rademakers, R., & van Blitterswijk, M. (2013). Motor neuron disease in 2012: Novel causal genes and disease modifiers. *Nature Reviews. Neurology*, 9(2), 63–64. doi:10.1038/nrneurol.2012.276 PMID:23318296

Reaume, A. G., Elliott, J. L., Hoffman, E. K., Kowall, N. W., Ferrante, R. J., Siwek, D. F., ... Snider, W. D. (1996). Motor neurons in Cu/Zn superoxide dismutase-deficient mice develop normally but exhibit enhanced cell death after axonal injury. *Nature Genetics*, *13*(1), 43–47. doi:10.1038/ng0596-43 PMID:8673102

Reid, E., Kloos, M., Ashley-Koch, A., Hughes, L., Bevan, S., Svenson, I. K., ... Pericak-Vance, M. A. (2002). A kinesin heavy chain (KIF5A) mutation in hereditary spastic paraplegia (SPG10). *American Journal of Human Genetics*, *71*(5), 1189–1194. doi:10.1086/344210 PMID:12355402

Renton, A. E., Chio, A., & Traynor, B. J. (2014). State of play in amyotrophic lateral sclerosis genetics. *Nature Neuroscience*, *17*(1), 17–23. doi:10.1038/nn.3584 PMID:24369373

Renton, A. E., Majounie, E., Waite, A., Simón-Sánchez, J., Rollinson, S., Gibbs, J. R., ... Traynor, B. J. (2011). A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron*, *72*(2), 257–268. doi:10.1016/j.neuron.2011.09.010 PMID:21944779

Riggs, J. E. (1996). Amyotrophic lateral sclerosis, heterogeneous susceptibility, trauma, and epidemiology. *Archives of Neurology*, *53*(3), 225–227. doi:10.1001/archneur.1996.00550030031019 PMID:8651874

Rosen, D. R., Siddique, T., Patterson, D., Figlewicz, D. A., Sapp, P., Hentati, A., ... Deng, H. X. (1993). Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature*, *362*(6415), 59–62. doi:10.1038/362059a0 PMID:8446170

Ross, M. A., Miller, R. G., Berchert, L., Parry, G., Barohn, R. J., Armon, C., ... McGuire, D. (1998). Toward earlier diagnosis of amyotrophic lateral sclerosis: Revised criteria. rhCNTF ALS Study Group. *Neurology*, *50*(3), 768–772. doi:10.1212/WNL.50.3.768 PMID:9521272

Rowland, L. P., & Shneider, N. A. (2001). Amyotrophic Lateral Sclerosis. *The New England Journal of Medicine*, 344(22), 1688–1700. doi:10.1056/NEJM200105313442207 PMID:11386269

Sasaki, S., & Iwata, M. (1996). Impairment of fast axonal transport in the proximal axons of anterior horn neurons in amyotrophic lateral sclerosis. *Neurology*, *47*(2), 535–540. doi:10.1212/WNL.47.2.535 PMID:8757033

Schmidt, R., de Reus, M. A., Scholtens, S. H., van den Berg, L. H., & van den Heuvel, M. P. (2016). Simulating disease propagation across white matter connectome reveals anatomical substrate for neuropathology staging in amyotrophic lateral sclerosis. *NeuroImage*, *124*, 762–769. doi:10.1016/j.neuroimage.2015.04.005 PMID:25869856

Schmidt, R., Verstraete, E., de Reus, M. A., Veldink, J. H., van den Berg, L. H., & van den Heuvel, M. P. (2014). Correlation between structural and functional connectivity impairment in amyotrophic lateral sclerosis. *Human Brain Mapping*, *35*(9), 4386–4395. doi:10.1002/hbm.22481 PMID:24604691

Sejvar, J. J., Holman, R. C., Bresee, J. S., Kochanek, K. D., & Schonberger, L. B. (2005). Amyotrophic lateral sclerosis mortality in the United States, 1979–2001. *Neuroepidemiology*, 25(3), 144–152. doi:10.1159/000086679 PMID:15990445

Shaw, A. S., Ampong, M. A., Rio, A., McClure, J., Leigh, P. N., & Sidhu, P. S. (2004). Entristar skinlevel gastrostomy tube: Primary placement with radiologic guidance in patients with amyotrophic lateral sclerosis. *Radiology*, 233(2), 392–399. doi:10.1148/radiol.2332031487 PMID:15459322

Shorter, J., & Lindquist, S. (2005). Prions as adaptive conduits of memory and inheritance. *Nature Reviews. Genetics*, *6*(6), 435–450. doi:10.1038/nrg1616 PMID:15931169

Sreedharan, J., Blair, I. P., Tripathi, V. B., Hu, X., Vance, C., Rogelj, B., ... Shaw, C. E. (2008). TDP-43 mutations in familial and sporadic amyotrophic lateral sclerosis. *Science*, *319*(5870), 1668–1672. doi:10.1126cience.1154584 PMID:18309045

Stamenkovic, S., Ducic, T., Stamenkovic, V., Kranz, A., & Andjus, P. R. (2017). Imaging of glial cell morphology, SOD1 distribution and elemental composition in the brainstem and hippocampus of the ALS hSOD1 G93A rat. *Neuroscience*, *357*, 37–55. doi:10.1016/j.neuroscience.2017.05.041 PMID:28576725

Stewart, H. G., Andersen, P. M., Eisen, A., & Weber, M. (2006). Corticomotoneuronal dysfunction in ALS patients with different SOD1 mutations. *Clinical Neurophysiology*, *117*(8), 1850–1861. doi:10.1016/j. clinph.2006.04.004 PMID:16793335

Su, Z., Zhang, Y., Gendron, T. F., Bauer, P. O., Chew, J., Yang, W. Y., ... Desaro, P. (2014). Discovery of a biomarker and lead small molecules to target r(GGGGCC)-associated defects in c9FTD/ALS. *Neuron*, *83*(5), 1043–1050. doi:10.1016/j.neuron.2014.07.041 PMID:25132468

Suzuki, N., Maroof, A. M., Merkle, F. T., Koszka, K., Intoh, A., & Armstrong, I., ... Eggan, K. (2013). The mouse C9ORF72 ortholog is enriched in neurons known to degenerate in ALS and FTD. *Nature Neuroscience*, 16.

Tai, H. C., & Schuman, E. M. (2008). Ubiquitin, the proteasome and protein degradation in neuronal function and dysfunction. *Nature Reviews. Neuroscience*, 9(11), 826–838. doi:10.1038/nrn2499 PMID:18931696

Traynor, B. J., Alexander, M., Corr, B., Frost, E., & Hardiman, O. (2003). An outcome study of riluzole in amyotrophic lateral sclerosis-a population-based study in Ireland, 1996–2000. *Journal of Neurology*, *250*(4), 473–479. doi:10.100700415-003-1026-z PMID:12700914

Traynor, B. J., Codd, M. B., Corr, B., Forde, C., Frost, E., & Hardiman, O. (1999). Incidence and prevalence of ALS in Ireland, 1995-1997: A population-based study. *Neurology*, *52*(3), 504–509. doi:10.1212/ WNL.52.3.504 PMID:10025778

Traynor, B. J., Codd, M. B., Corr, B., Forde, C., Frost, E., & Hardiman, O. (2000). Amyotrophic lateral sclerosis mimic syndromes: A population-based study. *Archives of Neurology*, *57*(1), 109–113. doi:10.1001/archneur.57.1.109 PMID:10634456 Van Deerlin, V. M., Leverenz, J. B., Bekris, L. M., Bird, T. D., Yuan, W., Elman, L. B., ... Yu, C. E. (2008). TARDBP mutations in amyotrophic lateral sclerosis with TDP-43 neuropathology: A genetic and histopathological analysis. *Lancet Neurology*, *7*(5), 409–416. doi:10.1016/S1474-4422(08)70071-1 PMID:18396105

Vance, C., Rogelj, B., Hortoba'gyi, T., De Vos, K. J., Nishimura, A. L., Sreedharan, J., ... Shaw, C. E. (2009). Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science*, *323*(5918), 1208–1211. doi:10.1126cience.1165942 PMID:19251628

Vucic, S., Burke, D., & Kiernan, M. C. (2007). Diagnosis of motor neuron disease. In M. C. Kiernan (Ed.), *The Motor Neuron Disease Handbook* (pp. 89–115). Sydney: Australasian Medical Publishing Company Limited.

Vucic, S., & Kiernan, M. C. (2009). Pathophysiology of degeneration in familial amyotrophic lateral sclerosis. *Current Molecular Medicine*, *9*(3), 255–272. doi:10.2174/156652409787847173 PMID:19355908

Wang, H., O'Reilly, E. J., Weisskopf, M. G., Logroscino, G., McCullough, M. L., Thun, M. J., ... Ascherio, A. (2011). Smoking and risk of amyotrophic lateral sclerosis: A pooled analysis of 5 prospective cohorts. *Archives of Neurology*, *68*(2), 207–213. doi:10.1001/archneurol.2010.367 PMID:21320987

Wicks, P., Abrahams, S., Masi, D., Hejda-Forde, S., Leigh, P. N., & Goldstein, L. H. (2007). Prevalence of depression in a 12-month consecutive sample of patients with ALS. *European Journal of Neurology*, *14*(9), 993–1001. doi:10.1111/j.1468-1331.2007.01843.x PMID:17718691

Wicks, P., Abrahams, S., Papps, B., Al-Chalabi, A., Shaw, C. E., Leigh, P. N., & Goldstein, L. H. (2009). SOD1 and cognitive dysfunction in familial amyotrophic lateral sclerosis. *Journal of Neurology*, 256(2), 234–241. doi:10.100700415-009-0078-0 PMID:19252762

Wiedemann, F. R., Manfredi, G., Mawrin, C., Beal, M. E., & Schon, E. A. (2002). Mitochondrial DNA and respiratory chain function in spinal cords of ALS patients. *Journal of Neurochemistry*, 80(4), 616–625. doi:10.1046/j.0022-3042.2001.00731.x PMID:11841569

Wijesekera, L. C., & Leigh, P. N. (2009). Amyotrophic lateral sclerosis. Orphanet Journal of Rare Diseases, 4(1), 3. doi:10.1186/1750-1172-4-3 PMID:19192301

Wilbourn, A. J. (1998). Clinical neurophysiology in the diagnosis of amyotrophic lateral sclerosis : The Lambert and the El Escorial criteria. *Journal of the Neurological Sciences*, *160*, 25–29. doi:10.1016/S0022-510X(98)00194-4 PMID:9851644

Williamson, T. L., & Cleveland, D. W. (1999). Slowing of axonal transport is a very early event in the toxicity of ALS-linked SOD1 mutants to motor neurons. *Nature Neuroscience*, 2(1), 50–56. doi:10.1038/4553 PMID:10195180

Wingo, T. S., Cutler, D. J., Yarab, N., Kelly, C. M., & Glass, J. D. (2011). The Heritability of Amyotrophic Lateral Sclerosis in a Clinically Ascertained United States Research Registry. *Public Library of Science One*, 6(11), e27985. PMID:22132186

Worms, P. M. (2001). The epidemiology of motor neuron diseases: A review of recent studies. *Journal of the Neurological Sciences*, *191*(1-2), 3–9. doi:10.1016/S0022-510X(01)00630-X PMID:11676986

Xi, Z., van Blitterswijk, M., Zhang, M., McGoldrick, P., McLean, J. R., Yunusova, Y., ... Rogaeva, E. (2015). Jump from Pre-mutation to Pathologic Expansion in C9orf72. *American Journal of Human Genetics*, *96*(6), 962–970. doi:10.1016/j.ajhg.2015.04.016 PMID:26004200

Xi, Z., Zinman, L., Moreno, D., Schymick, J., Liang, Y., Sato, C., ... Rogaeva, E. (2013). Hypermethylation of the CpG island near the G4C2 repeat in ALS with a C9orf72 expansion. *American Journal of Human Genetics*, *92*(6), 981–989. doi:10.1016/j.ajhg.2013.04.017 PMID:23731538

Yokoseki, A., Shiga, A., Tan, C. F., Tagawa, A., Kaneko, H., Koyama, A., ... Onodera, O. (2008). TDP-43 mutation in familial amyotrophic lateral sclerosis. *Annals of Neurology*, *63*(4), 538–542. doi:10.1002/ana.21392 PMID:18438952

Zoccolella, S., Beghi, E., Palagano, G., Fraddosio, A., Guerra, V., Samarelli, V., ... Logroscino, G. (2008). Analysis of survival and prognostic factors in amyotrophic lateral sclerosis: A population based study. *Journal of Neurology, Neurosurgery, and Psychiatry*, 79(1), 33–37. doi:10.1136/jnnp.2007.118018 PMID:17550991

Zu, T., Liu, Y., Banez-Coronel, M., Reid, T., Pletnikova, O., Lewis, J., ... Subramony, S. H. (2013). RAN proteins and RNA foci from antisense transcripts in C9ORF72 ALS and frontotemporal dementia. *Proceedings of the National Academy of Sciences*, *110*, E4968–E4977.

Chapter 16 Spectrum of Neurodegeneration in Autism Spectrum Disorder

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ABSTRACT

Autism spectrum disorder (ASD) is neurodevelopmental disorder which is characterized by lack of social behaviors and impaired non-verbal interactions that start early in childhood. It can also lead to progressive neurodegeneration like schizophrenia disorder, Alzheimer's disease, Parkinson's disease, and dementia. Genetic studies of ASD have confirmed the mutations that interfere with neurodevelopment in mother's womb through childhood and these mutations are further involved in synaptogenesis and axon motility. Crucial role of amygdala is found to be deficit in ASD individuals whose association cognition with nucleus accumbens lead to impaired social behaviors and cognitive stimulus. Educational and behavioral treatments are considered the key steps used for its management along with pharmacological and interventional therapies. In this chapter, the author presents the etiology of ASD, proof of neurodegeneration in ASD, as well as the clinical feature and the management of ASD.

INTRODUCTION

ASD is a set of neurodevelopmental disorders characterized by a lack of social interaction, verbal and nonverbal communication which is noticed in the first 3 years of childhood. In some of cases, mental and health conditions are progressively deteriorates with the time if not diagnosed and treated at time. The distinctive social behaviors include an avoidance of eye contact, fluctuate emotional control and difficulties in understanding the emotions of other people (Mattila *et al.*, 2011). Environmental factors are also likely to interact with the genetic profile and cause aberrant changes in brain growth, neuronal development, and functional connectivity. Increase in prevalence of ASD has been found and higher rate is reported in males than females (Kim *et al.*, 2011; Elsabagh *et al.*, 2012; Fombone *et al.*, 2011). Various post-mortem evaluations of ASD individuals have been postulated that these individuals experienced a loss of neuron cells and pyramidal cells in their amygdala than control samples. These observations have supported that these microglia can be responsible for the dissolution of neurons that can also induce the

DOI: 10.4018/978-1-5225-5282-6.ch016

production of toxic cytokines that can damage neurons and lead to neurodegeneration in ASD individuals (Kern et al., 2013). In last decades, despite growing evidence for the involvement of endogenous biomarkers in the pathophysiology of ASD is reported and early detection of this disorder remains a big challenge that describes the main behavioral and cognitive features of ASD, as well as the symptoms that differentiate autism from other developmental disorders such as reduced brain connectivity, mirror neurons deficits, and inhibition-excitation imbalance in individuals with ASD. In ASD patients, the frontal and temporal lobes are the markedly affected brain areas. And the role of amygdala in cognition and ASD has been proved in numerous neuropathological and neuroimaging studies (London & Etzel, 2001; Kern & Jones, 2006). The amygdala is a major component of the limbic system and affective loop of the cortico-striato-thalamo-cortical circuit. The amygdala located the medial temporal lobe anterior to the hippocampal formation has been thought to have a strong association with social and aggressive behaviors ASD patients (London & Etzel, 2001; Kern & Jones, 2006). ASD is clinically diagnosed on the basis of the presence of its associated non-specific manifestation like individual abilities in intelligence and verbal domains. The onsets of nonspecific manifestation are noticed in infants or toddlers such as irritability, passivity, and difficulties with sleeping and eating, followed by delays in language and social involvement (Kolevzon et al., 2007). Previously, some of reported data of few ASD patients have been studied for experiencing the improvements after deep brain stimulation as one of the interventional treatments. The key bone of neurobiology of ASD development is still a target for laying out its treatment and clinical management that required broadening its clinical horizons to understand ASD (Kolevzon et al., 2007). Hence, ASD is noted as multifactorial disorder and still unpredictable for investigators for drawing its fate to postulate its correlation with neural loss and progressive neurodegeneration in ASD individuals. So, the information provided in this chapter can be helpful for the researchers to postulate the effective therapies and treatments for ASD individuals. As well as, to provide points know more about the role of various genetic, environmental and epigenetic factors that have deleterious effects in ASD individuals such as cognitive impairment and immune homeostasis.

BACKGROUND

ASD affect the brain with the time that leads to neurodegeneration in ASD individual if left untreated or neglected so, many clinicians and researchers are involved to understand the ASD for long decades. However, pinpointing ASD's root cause may be aided by postulated all previous findings and clinical studies into a neurodegeneration hypothesis. This hypothesis can suggests that ASD can be regressive if ASD children are diagnosed and treated at time, so that they can acquire certain skill abilities before any serious neurological impairments happens to them. This ASD regression is said to helpful in 15% to 65% of ASD cases and hence, whether ASD can be considered a neurodegenerative disorder has remained under clinical critics and debate (Kern *et al.*, 2013). Previously, the development of the brain in individuals with ASD is discussed to be a complex neurodevelopmental disorders which is followed by interaction of various genetic and environmental factors and their interactions. Genetic studies of ASD have identified mutations in genes that interfere with typical neurodevelopment in childhood that involved in synaptogenesis and axon motility. An altered ratio of short to long diameter axons and disorganization of cortical layers are also observed along with MRI studies assessing brain volume in ASD individuals. Hence, as a result of the observed altered pattern of axons and MRI studies are found to responsible for altering the socioemotional networks (Mattila *et al.*, 2011). Characteristics of normal

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children in the sample which used for criteria of autism based on their caregivers reports were similar to those reported in the general population of individuals with ASDs. Even when taking into account the disproportionate percentage of males in the sample, the male-female ratio in children which used for autism was 3–4: 1 (Burd, Severud, Kerbeshian, & Klug, 1999; Fombonne, 1999, 2003; Steffenburg & Gillberg, 1986). In addition, although, developmentally delayed children who met criteria for ASD is tended to have more severe cognitive delays is considered key line path in past research findings that has suggested that ASD is often co-morbid with severe to profound cognitive impairments (Fombonne, 1999, 2003; Gillberg *et al.*, 1990; Lord & Volkar, 2002; Sigman & Capps, 1997).

ETIOLOGY OF AUTISM SPECTRUM DISORDER

ASD is broadly considered to be a multi-factorial disorder resulting from genetic and non-genetic risk factors and its respective interaction along with various diverse environmental causative elements including pre-natal, peri-natal, and post-natal factors that contribute ASD. This diagnostic bias towards males might result from under-recognition of females with ASD (Kern & Jones, 2001; Kolevzon et al., 2007; Baron et al., 2011). Also, some clinicians have suggested the possibility that the female-specific protective effects against ASD might exist (Robinson et al., 2013). Previously, various prenatal factors are found to related with ASD such as exposure to teratogens, thalidomide, certain viral infections (congenital rubella syndrome), maternal anticonvulsants (valproic acid) as well as low birth weight, abnormally short gestation length and birth asphyxia are considered the peri-natal factors (Davidson et al., 2004; Gorlin et al., 2016). The lesion of the amygdala results in fear-processing, modulation of memory with emotional content, and eye gaze when looking at human face which found to reported in ASD patients (Adolphs et al., 2005; Spezio et al., 2007). The amygdala receives highly processed somatosensory, visual, auditory, and all types of visceral inputs that sends efferent through two major pathways, the stria terminalis and the ventral amygdalofugal pathway. Several studies proposed the use of an animal model to confirm the evidence for the association between amygdala and ASD (Emery & Clayton, 2001; Sweeten et al., 2002). Previously, monkeys with the Kluver-Bucy syndrome have been reported ASD like disorder that show absence of social chattering, lack of facial expression, absence of emotional reactions, repetitive abnormal movement patterns, and increased aggression (Soydyk & Shekhar, 2000). Recently, the development of functional neuroimaging also provided some evidence for the correlation between amygdala deficit and ASD. A study using Technetium-99m (Tc-99m) singlephoton emission computed tomography (SPECT) is found that regional cerebral blood flow (rCBF) was decreased in the bilateral insula, superior temporal gyri, and left prefrontal cortices in individuals with ASD compared to age- and gender-matched controls with mental retardation (Quirk & Mueller, 2008; Stores-Bayon & Quirk, 2010; Ohnishi et al., 2000). Medial prefrontal cortex (mpFC) involves in fear learning and extinction by reciprocal synaptic connections with the basolateral amygdala (Bishop et al., 2007; Fuchs et al., 2007). Disturbed communication within amygdala-mPFC circuitry caused deficits in memory processing this information provides support for a role of the mpFC in the development of ASD. Besides amygdala, nucleus accumbens (NAc) is also considered as the key structure which is related with the social reward response in ASD and NAc borders ventrally on the anterior limb of the internal capsule, and the lateral subventricular fundus of the NAc is permeated in rostral sections by internal capsule fiber bundles (Mashhoom et al., 2010).

NEURODEGENERATION AND AUTISM SPECTRUM DISORDER

Impact of Oxidative Stress in Neurodegeneration

Over production of reactive oxygen species (ROS) due to oxidative stress is a central feature of neurodegenerative disorders which is confirmed with postmortem study done in brain tissues from ASD individuals diagnosed (Anderson, 2004). Free radicals and other ROS are harmful because the unpaired electron oxidatively reacts with other ions and molecules lead to progressive neural damage (Gutman, 2002). The oxidative degradation of the lipid membrane called lipid peroxidation that results in loss of membrane integrity and fluidity, which ultimately leads to neural cell death. ROS also react with proteins and nucleic acids which can lead to neural cell death via apoptosis or necrosis which further lead to neurodegenerative disorders (Kannan & Jain, 2000). The brain is highly vulnerable to oxidative stress when antioxidant capacity is halted due to any brain inflammatory diseases like Alzheimer's diseases, Parkinson's disease and dementia (Granot & Kohen, 2004). Several post-mortem studies have been performed in ASD individuals who revealed that affected areas of the brain associated with accelerated cell death under conditions of oxidative stress. It is also observed with this study that the density of lipofuscin, a matrix of oxidized lipid and cross-linked protein is increased in cortical brain areas that usually formed as a result of oxidative injury in the neural tissues (Evans et al., 2008). From earlier findings, it is postulated that brain regions are observed with the highest levels of the oxidative stress marker, 3-nitrotyrosine (3-NT) in the orbitofrontal cortex, Wernicke's area, cerebellar vermis, cerebellar hemisphere, and pons in ASD individuals. This elevated oxidative stress markers in the brains of ASD individuals is found to associate with evaluating the level of oxidative stress metabolites of carboxyethyl pyrrole (CEP) and iso[4]levuglandin (iso[4]LG)E2-protein adducts in cortical brain tissues when compared to neurotypical samples (Sajdel-Sukouska, 2008). DNA oxidation and glutathione redox status have been also studied in postmortem brain samples from the cerebellum and frontal, temporal, parietal and occipital cortex from ASD individuals when compared with age-matched neurotypical controls. These observations were confirmed that DNA oxidation was significantly increased by two-fold in the frontal cortex, temporal cortex, and cerebellum in ASD individuals compared to healthy controls (Chauhan et al., 2011). The levels of reduced glutathione were significantly reduced and inversely proportional to increased levels of oxidized glutathione in samples of the cerebellum and temporal cortex from ASD individuals as compared to the corresponding levels in the control brain samples. It was also confirmed that significant increase in the levels of lipid hydroperoxides (oxidative stress marker) in the cerebellum and temporal cortex in ASD individuals as compared to controls (Wyss-Coray & Muke, 2002).

Impact of Neuronal Loss in Neurodegeneration

Earlier clinical studies of ASD is characterized by neuronal cell loss which occurred due to significant reductions of neural-cerebellar Purkinje cells count (PCs) in ASD individuals when compared to neurotypical controls according to a postmortem study (Whitney *et al.*, 2008). Other ASD studies also supported the neuronal pathology found in the brain of ASD individuals that is suggestive tool of ASD associated neurodegenerative process when compares to neurotypical subjects. It has been also reported that degeneration in PCs and also microglial activation associated with gliosis. These finding are indicated that these changes are acquired rather than neurodevelopmental because gliosis is proliferation of neuroglial tissue that leads to progressive neural damage (Casanova, 2007 & Vajda, 2002). From earlier reports, it has been postulated that neuronal numbers in fusiform gyrus were found to be significantly reduced in the amygdala of individuals diagnosed with ASD individuals than controls, especially, in neuron densities of neural layer III (Schumanon & Amaral, 2006).

Impact of Microglia Activation in Neurodegeneration

Microglial reactivity is found to reported in neurodegenerative diseases that induces inflammation in infected or damaged brain tissues which also important for maintaining homeostasis in non-infected regions (Vargas et al., 2005). So, microglial cells act as key cellular mediators of the neuroinflammatory processes and are associated with the pathogenesis of many neurodegenerative disorders such as Alzheimer's diseases, Parkinson's diseases, stroke, spinal cord injury, encephalitis, and multiple sclerosis (Carson, 2007). Although the role of microglia in neurodegeneration is clearly understood yet but, many evidence indicates that microglia can become transiently activated to an amoeboid phenotype responsible for the phagocytosis of living neurons (Huuskonen *et al.*, 2005). It is also reported that during long-term or sustained neuroimmune activation, microglia produce cytokines that are toxic to neurons and this neurotoxicity plays a potential role in collateral neurodegeneration processed in many brain inflammatory diseases or disorders (Rock et al., 2004). SH neurons (i.e., SH-SY5Y) are also found to have their over expressive pattern followed by uncoupling of protein-2 that exhibited an increase in neuron-microglia interactions which represented an early step in microglial phagocytosis of neurons. Although, neurodegeneration also found to associated with degeneration of presynaptic neurons. Hence, some clinical debate is still with this type of involvement of microglia in synaptic stripping and synapse degeneration (Perry & O'Connor, 2010). Some of documented evidences from post-mortem brain tissue have been postulated for activated microglia and astrocytes responses in ASD individuals (Vargas et al., 2005; Enstrom et al., 2005). Previous findings are consistently supported with an active and ongoing postnatal process of neurodegeneration and neuroinflammation that leads to elevated level of proinflammatory chemokine, monocyte chemotactic protein-1 (MCP-1) in neural regions in individuals diagnosed with an ASD (Margan et al., 2010). As well as immunocytochemically identified microglia and stereologically quantification has been proposed for studying the microglial densities in the fronto-insular and visual cortex from autopsies of the brain in ASD individuals in comparison to controls by using positron emission tomography (PET) and a radiotracer. These observations are determined that increased microglial activation in the cerebellum, midbrain, pons, fusiform gyri, and the anterior cingulate and orbitofrontal cortices in ASD individuals as compared to controls (Suzuki et al., 2013).

Impact of Proinflammatory Cytokines in Neurodegeneration

Activated microglia are reported to induce the release of many potentially neurotoxic substances, such as reactive oxygen species, nitric oxide, and various proinflammatory cytokines that evidenced for implication of neuroinflammation and overproduction of proinflammatory cytokines which are potent contributor to pathophysiology of chronic neurodegenerative disorders (Vagas *et al.*, 2005; Chez *et al.*, 2007). It is also proved from previous clinical observation that proinflammatory cytokines (tumor

necrosis factor (TNF)- α , interleukin (IL)-6 and granulocyte-macrophage colonystimulating factor (GM-CSF), Th1 cytokine (interferon (IFN)- γ) and chemokine (IL-8) were significantly increased in the brains tissues of ASD compared with controls by using a protein array technique, it is also confirmed that that MCP-1, IL-6, IL-8 and IFN- γ were significantly increased in the cerebrospinal fluid (CSF) in individuals diagnosed with an ASD individuals when compared to healthy controls (Li X, 2009).

CLINICAL FEATURES OF AUTISM SPECTRUM DISORDER

ASD in children is also manifested with non-specific symptoms such as unusual sensory perception skills, motor clumsiness, disturbed emotional control and insomnia along with associated phenomena like mental retardation, emotional indifference, hyperactivity, aggression, self-injury and repetitive/ stereotyped behaviors such as body rocking or hand flapping. It is characterized by complex behavioral phenotype with eficits in both social and cognitive functions. Although the main findings of ASD emphasize the role of genetic and environmental factors in the development of autistic behavior. As well as, environmental factors are also likely to interact with the genetic profile and cause aberrant changes in brain growth, neuronal development, and functional connectivity. Despite growing evidence for the involvement of endogenous biomarkers in the pathophysiology of ASD, early detection of this disorder remains a big challenge. An attempt will be made to integrate all the available evidence which made the reduced brain connectivity, mirror neurons deficits, and inhibition-excitation imbalance in individuals with ASD (Fakhoury et al., 2015). Increased recognition and awareness of autism in the last few decades have been driven by the significant growth in research evidence. Well controlled cohort studies following-up pregnant mothers are likely to clarify the effects of some pre- and perinatal risk factors implicated in autism. Significant strides have also contributed towards developing and validating screening and diagnostic instruments to reduce heterogeneity in clinical characterization in research studies (Elsabbagh et al., 2012). However, developed countries have more improved and effective provisions for screening, diagnosis, and intervention are highly variable and many cases absent in community settings. Repetitive or stereotyped behaviors are often accompanied by cognitive impairment, seizures or epilepsy, gastrointestinal complaints, lack of sleep and other problems whose differential diagnosis reported for various associated disorders such as childhood schizophrenia, learning disability and deafness. Previously, longitudinal study of high-risk ASD infants have been initiated with 150 infant siblings, including 65 followed to age 24 months (Zwaigenbaum et al., 2005). Preliminary results of the proposed study are indicated that by 12 months of age, siblings who are later diagnosed with autism may be distinguished from other siblings which are low-risk controls on the basis of:

- Including atypicalities in eye contact, visual tracking, disengagement of visual attention, orienting to name, imitation, social smiling, reactivity, social interest and affect, and sensory-oriented behaviors (Zwaigenbaum *et al.*, 2005);
- Prolonged latency to disengage visual attention (Zwaigenbaum *et al.*, 2005);
- Delayed expressive (Zwaigenbaum *et al.*, 2005);
- Receptive language (Zwaigenbaum *et al.*, 2005);
- Poor visual orienting (Zwaigenbaum *et al.*, 2005).

DIAGNOSIS OF AUTISM SPECTRUM DISORDER

ASD is diagnosed on the basis of the presence of its associated core symptoms especially non-specific manifestation of individual abilities in intelligence and verbal domains (Elsabbagh et al., 2012). The onsets of nonspecific symptoms are recognized in infants or toddlers include irritability, passivity, and difficulties with sleeping and eating, followed by delays in language and social engagement. However, many researchers have reported that about 50% of infants showed behavioral abnormalities or atypical behaviors including extremes of temperament, impaired visual contact, fluctuated emotional control, lack of response to parental voices or interaction, imitation, social responses and motor control (Zwaigenbaum et al., 2005; Sab et al., 2003). The frontal and temporal lobes are the markedly affected brain areas in ASD patients especially the deficit of amygdala in cognition and ASD that have been observed under numerous neuropathological and neuroimaging clinical studies. These reports are confirmed for abrupt social and aggressive behavior (Sab et al., 2003; Alexander et al., 1986). Prefrontal lobe has been considered as playing an important role in higher-level control and a key structure lined with autism and its deficit demonstrate higher-order cognitive, language, social, and emotion dysfunction, which is deficient in ASD patients (Stuss & Knight, 2013; Mashhoon et al., 2010). One of the more ubiquitous behavioral findings in ASD population has been found prevalence of social skills difficulties (Niklasson, Rasmussen, Oskarsdottir, & Gillberg, 2001, 2002; Swillen et al., 1999; Woodin et al., 2001; Shprintzen, 2000). Several earlier studies and report evidence of social skills deficits including withdrawn and shy behaviors, difficulties initiating interactions, and anecdotal evidence of a limited range of facial expressions (Gerdes et al., 1999; Niklasson et al., 2002; Swillen et al., 1999). Despite the evidence that a large percentage of individuals with the 22q11.2 deletion experience psychiatric, social and communication difficulties which used for diagnosis of an autism spectrum disorder (ASD) with greater frequency than the general population.

ASD are atypical developmental delays characterized by impairments in communication, social skills, and restricted or stereotyped patterns of behaviors and interests with diagnosis of Asperger's syndrome in ASD individuals. This diagnosis is reserved for those with impaired social behavior, including inability to read nonverbal cues, and stereotyped and restricted patterns of interest, but without concomitant delayed language and cognitive ability (Yergin-Allsop *et al.*, 2003).

Children diagnosed with an ASD experience marked social impairment characterized by a lack of relatedness and lack of emotional connections with other surrounding people. For example, they exhibit inappropriate or constricted ranges of facial affect when compared to normal children without ASDs (Yirmiya *et al.*, 1992). Similar behaviors have been reported in some studies of individuals with the 22q11.2 deletion (e.g., Roubertie *et al.*, 2001), but other findings are mixed in terms of the prevalence of ASD symptoms and disorders in this population. Some researchers have reported that the occurrence of ASDs in children with 22q11.2 deletion is relatively uncommon (e.g., Kozma, 1998). Using the Autism Diagnostic Interview to confirm a diagnosis of autism based on strict criteria, one research group reported that none of the 103 autistic children in their sample had a chromosome 22 deletion (Ogilvie, Moore, Daker, Palferman, & Docherty, 2000). Although the Ogilvie *et al.* (2000) was proposed a clinical observations where samples was drawn entirely from multiplex families, which limits the generalizability of their findings, they concluded that behavioral and psychiatric symptoms observed in the 22q11.2 deletion population were likely inconsistent with autism when strict criteria for diagnosis is used for ASD genetic studies (Ogilvie *et al.* (2000).

Besides amygdala and nucleus accumbens (NAc) is also considered as the key point structure which is found to deficit in ASD patients to have impaired social reward and pleasure response in ASD (Knuston *et al.*, 2001; Stingler *et al.*, 2011).

Limitation of this diagnostic tool is the use of the ADI-R to detect ASD within this sample of children with an established genetic diagnosis. The ADI-R has been revised to discriminate ASD from other genetic disorders (e.g., Fragile X). However, that the estimated prevalence of ASD in this sample of children with 22q11.2 deletion may have been somewhat inflated due to a measurement artifact. ADI-R may not be able to reliably discriminate behaviors that characterize children with ASD from behaviors of children with 22q11.2 deletion. This potential weakness in the ADI-R which mostly applies to other measures such as the Vineland "Adaptive Behavior Scales", highlights the importance of using multiple methods to carefully and thoroughly assess ASD, particularly in children who have genetic diagnoses. Certain characteristics of the sample may also limit the plot of these findings. There is a possibility of ascertainment bias, in that all children described in the present study had experienced manifestations of the 22q11.2 deletion that were apparent enough to challenge the genetic testing. This characteristic suggests that they may have been more seriously affected than other children who have this kind of deletion but have not experienced the structural anomalies such as congenital heart defects or the other health problems such as hypocalcemia associated with the deletion. Further research focusing on linkages between specific genes and ASDs is greatly needed, as phenotype research on family characteristics and personality types remains imprecise and relies upon clinical interviews that require further testing and standardization in ASD individuals (e.g., M-PAS, FHI; Bolton et al., 1994; Folstein et al., 1999; Piven, Palmer et al., 1997; Szatmari et la., 1995; Szatmari et al., 2000).

It is also possible that parents of children who were more seriously affected by the 22q11.2 deletion were more motivated to respond to the invitation to participate than parents of children with fewer difficulties (Folstein *et al.*, 1999). As a result, the findings may represent an overestimate of the prevalence of ASD in the population of children with 22q11.2 deletion. Moreover, participating families had the time and resources to allow their children to travel to The Children's Hospital of Philadelphia, in some cases from considerable distances, to participate in the larger study and they were further able to take the time to participate in the current study over the telephone which may represent a slightly more economically privileged group. Given these sample characteristics, findings should be interpreted with caution, as this sample may not be fully representative of all individuals with the 22q11.2 chromosomal deletion (Bolton *et al.*, 1994; Szatmari *et al.*, 2000).

TREATMENT OF AUTISM SPECTRUM DISORDER

In order to assess behaviors and symptoms that are characteristic of ASDs, caregivers completed a measure that screens for these behaviors. Caregivers of children ages four years and older (n = 78) completed the Social Communication Questionnaire, Lifetime Version (SCQ, previously known as the Autism Screening Questionnaire. So, correct line of treatment can be given to ASD individuals (Berument *et al.*, 1999).

Many treatments (Txs) have been proposed for ASD with the most effective being combined Tx involving specialized and supportive educational programming, communication training (e.g., speech/language therapy), social skills support, and behavioral intervention (Lord & McGee, 2001: Mayer *et al.*, 2007). Occupational and physical therapy also may promote progress by addressing comorbid difficulties of motor coordination and sensory deficits (Levy & Hyman, 2008). Behavior modification

(e.g., applied behavior analysis [ABA]) has the most empirical support for a single Tx, with documented improvements in language, social, play, and academic skills, and reduction in severe behavioral problems (Schreibman et al., 2000). However, behavioral Txs are time and staff intensive, requiring up to 30-40 hours of Tx per week for several years by trained staff working directly with the child and typically focusing on one or a few behaviors at a time. Risperidone and aripiprazole are the only FDA-approved medications for ASD and they are approved only for the Tx of irritability in 5–16 year olds with ASD. Still no fixed medications are currently established to treat ASD core symptoms. "Off-label" medications are often prescribed for various cognitive behaviors such as inattention, impulsivity/hyperactivity, sleep problems, repetitive/perseverative behaviors, anxiety, mood, agitation, aggression, and disruptive and self-injurious behaviors but may have significant side effects (Finding et al., 2005). Previous survey on ASD individuals has estimated the utilization of psychotropic medication for youth with ASD as high as 47% (Witwer & Lecavalier, 2005), but there is ongoing debate about the role of such agents (Bryson et al., 2003). Response rates to medication for comorbid diagnoses in ASD children may be lower than for children without ASD e.g., the response rate of methylphenidate for typically developing children with attention deficit hyperactivity disorder (ADHD) is 70% (Spancer et al., 1996). Complementary and alternative medicines are also commonly reported, but their effectiveness remains unproven (Newschaffer et al., 2007). Therefore, given the limitations of available Txs for ASD and the severe and chronic nature of ASD, there is a large public health need for additional interventions.

Because these interventions can include both ingestible via oral administration and noningestible when externally administered Txs is referred to them collectively as complementary and alternative Txs (CATs). They are complementary when these practices are used together with conventional medicine and alternative when used in place of conventional medicine. However, newly designated complimentary Tx, incremental effects when added to conventional Tx should be empirically demonstrated (Hanson *et al.*, 2007; Harrigton *et al.*, 2006).

ASD medical management criterion includes typical antipsychotics, atypical antipsychotics, antidepressants, selective serotonin reuptake inhibitors, α^2 -adrenergic agonists, β -adrenergic antagonist, mood stabilizers and anticonvulsants (Farmer et al., 2013). Antidepressants are most commonly prescribed agents followed by other stimulants and antipsychotics for ASD patients. The high prevalence of comorbidities is found to reflect in the rates of psychotropic medication used in some of ASD patients. So, antipsychotics are found more effective in treating the repetitive behaviors in children with ASD (Kerbeshain et al., 2001). Music therapy is also reported for ASD children involving playing and/or listening to music that has been used to Tx ASD because of its potential for assisting communication, joint attention, expression, engagement, and relationships with the environment (Wigram & Gold, 2006). Most research on music therapy is still in progress for ASD case studies with only two randomized single-blind, repeated measures, within-subject comparison designs (Buday, 1995; Kim et al., 2008). These studies had a total of 20, 3-to-9 year-olds with ASD, with varied Tx presentations, given 1–20X/week for 1–12 weeks for 30 minutes. Significant results and potential clinical outcomes include improvement in imitating signs and words, longer and more eye contact and turn-taking, joint attention, nonverbal communication, longer and more joy, emotional synchronicity, initiating engagement and compliant behavior. Research on music therapy for ASD lacks evidence-based assessment of ASD, large samples, randomized controlled trials (RCTs), standardized protocols, double-blind, sham, use of standard Tx outcome measures, follow-up, monitoring of adverse-effects, or concomitant Txs. However, it appears safe, seems sensible, easy and cheap and is therefore acceptable. There is limited scientific evidence of efficacy for some CATs, but research on CATs for ASD is imperative because key safety and efficacy questions remain for the majority (Kerbeshain et al., 2001). Berman and Straus (McPheeters et al., 2011) observed that many CAT studies assume that Txs are well defined, including optimal dose/duration/intensity, that the sample has been correctly diagnosed and selected, and that the Tx is consistent from one practitioner to another. They note that CATs should meet the same fundamental requirements as for conventional Txs, using the same tools and techniques as those for conventional research to isolate the specific effects from the nonspecific effects of Tx as much as possible. However, to be a truly designated complimentary Tx, incremental effects when added to conventional Tx should be empirically demonstrated. Likewise, to be truly designated an alternative Tx, similar effects when compared to conventional Tx should be demonstrated. Few CATs for any psychiatric condition (and none for ASD) fulfill these requirements, so the majorities are not valid CATs. Such controls include rigorous protocols, RCTs with, where possible, placebo/sham control conditions with double-blind designs, and careful diagnosis. Such kind of controlled and well programmed research activities are vital because, even though people often assume CATs, particularly natural ones are safe as well as their use without supportive evidence is risky because they may have dangerous, sometimes life-threatening and irreversible side-effects which fail to reduce symptoms or improve functioning in ASD patients with severely impairing disorders accompanied with delay use of other more established Txs, waste of time, waste of energy and waste of money. These days, there are also alternative medication which are chosen by some of physicians includes opiate antagonist, immunotherapy, hormonal agents, megavitamins and other dietary supplements to treat ASD patients (McPheeters et al., 2011).

FUTURE RESEARCH DIRECTIONS

Findings from the present study may directly impact current practices and medical care for ASD children with a 22q11.2 deletion. In this study, hospital stays and previous diagnoses might have delayed the detection of ASDs (Folstein *et al.*, 1999; Piven, Palmer *et al.*, 1997; Szatmari *et al.*, 1995). Parents may not be mentally prepared to recognize behaviors that did not confirm the primary diagnosis or may have ignored these behaviors as the result of the trauma of their child's hospitalization. These factors suggest that certain situations, such as prematurity or regular hospital stays, require careful attention and consideration of missed diagnoses. Future studies should investigate methods to prevent biased interpretations of symptoms in the context of the 22q11.2 deletion. In addition, it might be fruitful to provide all children with a 22q11.2 deletion with more stringent developmental assessments that specifically rule out or confirm ASDs.

Given the host of difficulties faced by many children with a 22q11.2 deletion, such as chronic medical conditions, learning disabilities, and other psychiatric issues, some may question the incremental value to families of formally diagnosing an ASD. However, the existence of empirically supported treatments and interventions for individuals with autism means that children who are diagnosed can have access to early intervention and ongoing special services that can improve social, behavioral, and language functioning (Goldstein, 2002; Horner *et al.*, 2002; McConnell, 2002). Complementary and alternative medical (CAM) therapies are commonly recommended by physician for ASD children who have ADHD. The use of these therapies is well documented but, evidence of its safety and efficacy of these treatments in children is limited. This article describes the current evidence-based CAM therapies for ADHD and autism that focusing on nutritional interventions; natural health products, including essential fatty acids, vitamins, minerals along with other health supplements (Geraghty *et al.*, 2011; Huffman,

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2011; Rossignol *et al.*, 2009; Schechtman, 2007; Weber & Newmark, 2007). Diagnosis that combat with understanding to parents and informs their attributions about their children's behaviors. In addition, this type of ASD diagnostic measure prepares parents for areas of difficulty that develop in the later years of autism, including depression among higher functioning individuals (Ghaziuddin *et al.*, 2002; Volkmar *et al.*, 1999) and also have the need of residential living and preparation for sheltered occupations (Frith, 1991). An accurate diagnosis of ASD may expedite acquisition of services and be particularly empowering for families of children with a 22q11.2 deletion which alarmed the medical and educational professionals about this little known fact of ASD but relatively common genetic disorder, about which we still have so much to learn.

CONCLUSION

So, this chapter is helpful to sort the outline of neurobiology of ASD and its associated disorders and non-specific manifestations which is considered a neurodevelopment disorder. The causes of ASD are not still known and various conducted researches suggest that both genes and environment play important roles to develop this neurodevelopment disorder. Although many children with ASDs are currently treated with medical interventions, strikingly little evidence exists to support benefit for most treatments. Risperidone and aripiprazole have shown benefit for challenging and repetitive behaviors, but associated adverse effects limit their use to patients with severe impairment or risk of injury. Its respective early diagnosis and medical management is important to improve the mental and physical health of ASD affected individuals. Otherwise, with the time, neurodegeneration happens followed by dementia and schizophrenia in some of ASD patients with the increased age. There is currently no one standard medical treatment used for ASD in affected population. Although, proposed treatments are including behavior and communication therapies, skills training, and use of opiate antagonist, immunotherapy, hormonal agents, megavitamins, other dietary supplements or medicines that proposed to increase affected patient's ability to grow and learn new skills to achieve better results to control its associated symptoms.

REFERENCES

Adolphs, R., Gosselin, F., Buchanan, T. W., Tranel, D., Schyns, P., & Damasio, A. R. (2005). A mechanism for impaired fear recognition after amygdala damage. *Nature*, *433*(7021), 68–72. doi:10.1038/ nature03086 PMID:15635411

Anderson, J. K. (2004). Oxidative stress in neurodegeneration: Cause or consequence? *Nat Neurosci Rev*, 5(7), S18–S25. doi:10.1038/nrn1434 PMID:15298006

Arnold, P. D., Siegel-Bartelt, J., Cytrynbaum, C., Teshima, I., & Schachar, R. (2001). Velo-cardio-facial syndrome: Implications of microdeletion 22q11 for schizophrenia and mood disorders. *American Journal of Medical Genetics*, *105*(4), 354–362. doi:10.1002/ajmg.1359 PMID:11378850

Baron-Cohen, S., Lombardo, M. V., Auyeung, B., Ashwin, E., Chakrabarti, B., & Knickmeyer, R. (2011). Why are autism spectrum conditions more prevalent in males? *PLoS Biology*, *9*(6), 1001081. doi:10.1371/journal.pbio.1001081 PMID:21695109

Bassett, A. S., & Chow, E. W. (1999). 22q11 deletion syndrome: A genetic subtype of schizophrenia. *Biological Psychiatry*, *46*(7), 882–891. doi:10.1016/S0006-3223(99)00114-6 PMID:10509171

Berument, S. K., Rutter, M., Lord, C., Pickles, A., & Bailey, A. (1999). Autism screening questionnaire: Diagnostic validity. *The British Journal of Psychiatry*, *175*(05), 444–451. doi:10.1192/bjp.175.5.444 PMID:10789276

Bishop, S. J. (2007). Neurocognitive mechanisms of anxiety: An integrative account. *Trends in Cognitive Sciences*, *11*(7), 307–316. doi:10.1016/j.tics.2007.05.008 PMID:17553730

Bolton, P., Macdonald, H., Pickles, A., Rios, P., Goode, S., Crowson, M. A., ... Rutter, M. (1994). Case-control family history study of autism. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, *35*(5), 877–900. doi:10.1111/j.1469-7610.1994.tb02300.x PMID:7962246

Bryson, S. E., Rogers, S. J., & Fombonne, E. (2003). Autism spectrum disorders: Early detection, intervention, education, and psychopharmacological management. *Canadian Journal of Psychiatry*, 48(8), 506–516. doi:10.1177/070674370304800802 PMID:14574826

Buday, E. (1995). The effects of signed and spoken words taught with music on sign and speech imitation by children with autism. *Journal of Music Therapy*, *32*(3), 189–202. doi:10.1093/jmt/32.3.189

Burd, L., Severud, R., Kerbeshian, J., & Klug, M. (1999). Prenatal and perinatal risk factors for autism. *Journal of Perinatal Medicine*, *27*, 441–450. PMID:10732302

Carson, M. J., Bilousova, T. V., Puntambekar, S. S., Melchior, B., Doose, J. M., & Ethell, I. M. (2007). A rose by any other name: the potential consequences of microglial heterogeneity during CNS health and disease. *Neurotherapeutics; the Journal of the American Society for Experimental NeuroTherapeutics, 4*(4), 571–579. doi:10.1016/j.nurt.2007.07.002 PMID:17920538

Casanova, M. F. (2007). The neuropathology of autism. *Brain Pathology (Zurich, Switzerland)*, 417(4), 422–433. doi:10.1111/j.1750-3639.2007.00100.x PMID:17919128

Chauhan, A., Gu, F., Essa, M. M., Wegiel, J., Kaur, K., Brown, W. T., & Chauhan, V. (2011). Brain region-specific deficit in mitochondrial electron transport chain complexes in children with autism. *Journal of Neurochemistry*, *117*(2), 209–220. doi:10.1111/j.1471-4159.2011.07189.x PMID:21250997

Chez, M. G., Dowling, T., Patel, P. B., Khanna, P., & Kominsky, M. (2007). Elevation of tumor necrosis factor-alpha in cerebrospinal fluid of autistic children. *Pediatric Neurology*, *36*(6), 361–365. doi:10.1016/j. pediatrneurol.2007.01.012 PMID:17560496

Davidson, P. W., Myers, G. J., & Weiss, B. (2004). Mercury exposure and child development outcomes. *Pediatrics*, *113*, 1023–1029. PMID:15060195

Ehlers, S., & Gillberg, C. (1993). The epidemiology of Asperger syndrome: A total population study. *Journal of Child Psychology and Psychiatry, and Allied Disciplines, 34*(8), 1327–1350. doi:10.1111/j.1469-7610.1993.tb02094.x PMID:8294522

Elsabbagh, M., Divan, G., Koh, Y. J., Kim, Y. S., Kauchali, S., Marcín, C., ... Fombonne, E. (2012). Global prevalence of autism and other pervasive developmental disorders. *Autism Research*, *5*(3), 160–179. doi:10.1002/aur.239 PMID:22495912

Elsabbagh, M., Divan, G., Koh, Y. J., Kim, Y. S., Kauchali, S., Marcín, C., ... Fombonne, E. (2012). Global prevalence of autism and other pervasive developmental disorders. *Autism Research*, *5*(3), 160–179. doi:10.1002/aur.239 PMID:22495912

Emery, N. J., & Clayton, N. S. (2001). Effects of experience and social context on prospective caching strategies by scrub jays. *Nature*, *414*(6862), 443–446. doi:10.1038/35106560 PMID:11719804

Enstrom, A. M., Lit, L., Onore, C. E., Gregg, J. P., Hansen, R. L., Pessah, I. N., ... Zimmerman, A. W. (2005). Immunity, neuroglia and neuroinflammation in autism. *International Review of Psychiatry* (*Abingdon, England*), *17*(6), 485–495. doi:10.1080/02646830500381930 PMID:16401547

Evans, T. A., Siedlak, S. L., Lu, L., Fu, X., Wang, Z., McGinnis, W. R., ... Zhu, X. (2008). The autistic phenotype exhibits a remarkably localized modification of brain protein by products of free radicalinduced lipid oxidation. *American Journal of Biochemistry and Biotechnology*, *4*(2), 61–72. doi:10.3844/ ajbbsp.2008.61.72

Fakhoury, M. (2015). Autistic spectrum disorders: A review of clinical features, theories and diagnosis. *International Journal of Developmental Neuroscience*, *43*, 70–77. doi:10.1016/j.ijdevneu.2015.04.003 PMID:25862937

Farmer, C., Thurm, A., & Grant, P. (2013). Pharmacotherapy for the core symptoms in autistic disorder: Current status of the research. *Drugs*, 73(4), 303–314. doi:10.100740265-013-0021-7 PMID:23504356

Findling, R. L. (2005). Pharmacologic treatment of behavioral symptoms in autism and pervasive developmental disorders. *The Journal of Clinical Psychiatry*, *66*(supplement 10), S26–S31. PMID:16401147

Folstein, S. E., Santangelo, S. L., Gilman, S. E., Piven, J., Landa, R. R., Lainhart, J., ... Wzorek, M. (1999). Predictors of cognitive test patterns in autism families. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, 40(7), 1117–1128. doi:10.1111/1469-7610.00528 PMID:10576540

Fombonne, E. (1999). The epidemiology of autism: A review. *Psychological Medicine*, 29(4), 769–786. doi:10.1017/S0033291799008508 PMID:10473304

Fombonne, E. (2003). Epidemiological surveys of autism and other pervasive developmental disorders: An update. *Journal of Autism and Developmental Disorders*, *33*(4), 365–382. doi:10.1023/A:1025054610557 PMID:12959416

Fombonne, E. (2011). Incidence and prevalence of pervasive developmental disorders. In E. Hollander, A. Kolevzon, & J. T. Coyle (Eds.), *Textbook of autism spectrum disorders* (pp. 117–136). Washington, DC: American Psychiatric Publishing, Inc. doi:10.1093/med/9780195371826.003.0007

Fuchs, R. A., Eaddy, J. L., Su, Z. I., & Bell, G. H. (2007). Interactions of the basolateral amygdala with the dorsal hippocampus and dorsomedial prefrontal cortex regulate drug context-induced reinstatement of cocaine-seeking in rats. *The European Journal of Neuroscience*, *26*(2), 487–498. doi:10.1111/j.1460-9568.2007.05674.x PMID:17650119

Geraghty, M. E., Bates-Wall, J., Ratliff-Schaub, K., & Lane, A. E. (2010). Nutritional interventions and therapies in autism: a spectrum of what we know: part 2. *Infant, Child & Adolescent Nutrition*, 2(2), 120–133. doi:10.1177/1941406410366848

Gerdes, M., Solot, C., Wand, P. P., Moss, E., LaRossa, D., & Randall, P. (1999). Cognitive and behavior profile of pre-school children with chromosome 22q11.2 deletion. *American Journal of Medical Genetics*, 85(2), 127–133. doi: PMID:10406665

Ghaziuddin, M., Ghaziuddin, N., & Greden, J. (2002). Depression in persons with autism: Implications for research and clinical care. *Journal of Autism and Developmental Disorders*, *32*(4), 299–306. doi:10.1023/A:1016330802348 PMID:12199134

Gillberg, C., Ehlers, S., Schaumann, H., Jakobsson, G., Dahlgren, S. O., Lindblom, R., ... Blidner, E. (1990). Autism under age 4 years: A clinical study of 28 cases referred for autistic symptoms in infancy. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, *31*(6), 921–934. doi:10.1111/j.1469-7610.1990. tb00834.x PMID:2246342

Gillham, J. E., Carter, A. S., Volkmar, F. R., & Sparrow, S. S. (2000). Toward a developmental operational definition of autism. *Journal of Autism and Developmental Disorders*, *30*(4), 269–278. doi:10.1023/A:1005571115268 PMID:11039854

Goldstein, H. (2002). Communication intervention for children with autism: A review of treatment efficacy. *Journal of Autism and Developmental Disorders*, *32*(5), 373–396. doi:10.1023/A:1020589821992 PMID:12463516

Gorlin, B. J., McAlpine, C.P, Garwick, A, Wieling, E. (2016). Severe Childhood Autism: The Family Lived Experience *J Pediatr Nurs.*, *31*(6), 580-597. doi: . 2016.09.00210.1016/j.pedn

Granot, E., & Kohen, R. (2004). Oxidative stress in childhood – in health and disease states. *Clinical Nutrition (Edinburgh, Lothian)*, 23(1), 3–11. doi:10.1016/S0261-5614(03)00097-9 PMID:14757387

Gutman, J. (2002). *Glutathione- Your Bodies Most Powerful Protector* (3rd ed.). Montreal: Communications Kudoca Inc.

Hanson, E., Kalish, L. A., Bunce, E., Curtis, C., McDaniel, S., Ware, J., & Petry, J. (2007). Use of complementary and alternative medicine among children diagnosed with autism spectrum disorder. *Journal of Autism and Developmental Disorders*, *37*(4), 628–636. doi:10.100710803-006-0192-0 PMID:16977497

Harrington, J. W., Rosen, L., Garnecho, A., & Patrick, P. A. (2006). Parental perceptions and use of complementary and alternative medicine practices for children with autistic spectrum disorders in private practice. *Journal of Developmental and Behavioral Pediatrics*, 27(2Supplement 2), S156–S161. doi:10.1097/00004703-200604002-00014 PMID:16685182

Horner, R., Carr, E., Strain, P., Todd, A., & Reed, H. (2002). Problem behavior interventions for young children with autism: A research synthesis. *Journal of Autism and Developmental Disorders*, *32*(5), 423–446. doi:10.1023/A:1020593922901 PMID:12463518

Huffman, L. C., Sutcliffe, T. L., Tanner, I. S. D., & Feldman, H. M. (2011). Management of symptoms in children with autism spectrum disorders: A comprehensive review of pharmacologic and complementaryalternative medicine treatments. *Journal of Developmental and Behavioral Pediatrics*, *32*(1), 56–68. doi:10.1097/DBP.0b013e3182040acf PMID:21160435

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Huuskonen, J., Suuronen, T., Miettinen, R., van Groen, T., & Salminen, A. (2005). A refined in vitro model to study inflammatory responses in organotypic membrane culture of postnatal rat hippocampal slices. *Journal of Neuroinflammation*, 2(1), 25. doi:10.1186/1742-2094-2-25 PMID:16285888

Kannan, K., & Jain, S. K. (2000). Oxidative stress and apoptosis. *Pathophysiology*, 7(3), 153–163. doi:10.1016/S0928-4680(00)00053-5 PMID:10996508

Kerbeshian, J., Burd, L., & Avery, K. (2011). Pharmacotherapy of autism: A review and clinical approach. *Journal of Developmental and Physical Disabilities*, *13*(3), 199–228. doi:10.1023/A:1016686802786

Kern, J. K., Geier, D. A., Sykes, C. K., & Geier, M. R. (2013). Evidences of neurodegeneration in autisum spectrum disorder. *Translational Neurodegeneration*, 2(1), 17. doi:10.1186/2047-9158-2-17 PMID:23925007

Kern, J. K., & Jones, A. M. (2006). Evidence of toxicity, oxidative stress, and neuronal insult in autism. *Journal of Toxicology and Environmental Health. Part B, Critical Reviews*, 9(6), 485–499. doi:10.1080/10937400600882079 PMID:17090484

Kim, J., Wigram, T. C., & Gold, C. (2008). The effects of improvisational music therapy on joint attention behaviors in autistic children: A randomized controlled study. *Journal of Autism and Developmental Disorders*, *38*(9), 1758–1766. doi:10.100710803-008-0566-6 PMID:18592368

Kim, Y. S., Leventhal, B. L., Koh, Y. J., Fombonne, E., Laska, E., Lim, E. C., ... Grinker, R. R. (2011). Prevalence of autism spectrum disorders in a total population sample. *The American Journal of Psychiatry*, *168*(9), 904–912. doi:10.1176/appi.ajp.2011.10101532 PMID:21558103

Knutson, B., Adams, C. M., Fong, G. W., & Hommer, D. (2001). Anticipation of increasing monetary reward selectively recruits nucleus accumbens. *The Journal of Neuroscience*, 21, RC159. PMID:11459880

Kolevzon, A., Gross, R., & Reichenberg, A. (2007). Prenatal and perinatal risk factors for autism: A review and integration of findings. *Archives of Pediatrics & Adolescent Medicine*, *161*(4), 326–333. doi:10.1001/archpedi.161.4.326 PMID:17404128

Kozma, C. (1998). On cognitive variability in velocardiofacial syndrome: Profound mental retardation and autism. *American Journal of Medical Genetics*, *81*(3), 269–270. doi: PMID:9603617

Levy, S. E., & Hyman, S. L. (2003). Use of complementary and alternative treatments: For children with autistic spectrum disorders is increasing. *Pediatric Annals*, *32*(10), 685–691. doi:10.3928/0090-4481-20031001-10 PMID:14606219

Levy, S. E., & Hyman, S. L. (2008). Levy., Hyman SL. Complementary and alternative medicine treatments for children with autism spectrum disorders. *Child and Adolescent Psychiatric Clinics of North America*, *17*(4), 803–820. doi:10.1016/j.chc.2008.06.004 PMID:18775371

Li, X., Chauhan, A., Sheikh, A. M., Patil, S., Chauhan, V., Li, X. M., ... Malik, M. (2009). Elevated immune response in the brain of autistic patients. *Journal of Neuroimmunology*, 207(1-2), 111–116. doi:10.1016/j.jneuroim.2008.12.002 PMID:19157572

London, E. A., & Etzel, R. A. (2000). The environment as an etiologic factor in autism: A new direction for research. *Environmental Health Perspectives*, *108*(s3Suppl 3), 401–404. doi:10.1289/ehp.00108s3401 PMID:10852835

Lord, C. (1995). Follow up of two year olds referred for possible autism. *Journal of Child Psychology* and Psychiatry, and Allied Disciplines, 36(8), 1365–1382. doi:10.1111/j.1469-7610.1995.tb01669.x PMID:8988272

Lord, C., & McGee, J. P. (2001). *Educating Children With Autism*. Washington, DC, USA: National Academy Press.

Lord, C., Risi, S., Lambrecht, L. E. H., Leventhal, B. L., & DiLavore, P. C. (2000). The Autism Diagnostic Observation Schedule-Generic: A standard measure of social and communication deficits associated with the spectrum of autism. *Journal of Autism and Developmental Disorders*, *30*(3), 205–223. doi:10.1023/A:1005592401947 PMID:11055457

Lord, C., Rutter, M., & Le Couteur, A. (1994). Autism Diagnostic Interview-Revised. A revised version of a diagnostic interview for caregivers of individuals with possible pervasive developmental disorders. *Journal of Autism and Developmental Disorders*, 24(5), 659–685. doi:10.1007/BF02172145 PMID:7814313

Lord, C., Volkmar, F., & Lombroso, P. J. (2002). Genetics of childhood disorders: XLII. Autism, Part 1: Diagnosis and assessment in autistic spectrum disorders (Development and Neurobiology). *Journal of the American Academy of Child and Adolescent Psychiatry*, *41*(9), 1134–1136. doi:10.1097/00004583-200209000-00015 PMID:12218436

Mashhoon, Y., Wells, A. M., & Kantak, K. M. (2010). Interaction of the rostral basolateral amygdala and prelimbic prefrontal cortex in regulating reinstatement of cocaine-seeking behavior. *Pharmacology, Biochemistry, and Behavior, 96*(3), 347–353. doi:10.1016/j.pbb.2010.06.005 PMID:20600250

Mattila, M. L., Kielinen, M., Linna, S. L., Jussila, K., Ebeling, H., Bloigu, R., ... Moilanen, I. (2011). Autism spectrum disorders according to DSM-IV-TR and comparison with DSM-5 draft criteria: An epidemiological study. *Journal of the American Academy of Child and Adolescent Psychiatry*, *50*(6), 583–592. doi:10.1016/j.jaac.2011.04.001 PMID:21621142

McConnell, S. (2002). Interventions to facilitate social interaction for young children with autism: Review of available research and recommendations for educational intervention and future research. *Journal of Autism and Developmental Disorders*, *32*(5), 351–372. doi:10.1023/A:1020537805154 PMID:12463515

McDonald-McGinn, D. M., Kirschner, R., Goldmuntz, E., Sullivan, K., Eicher, P., & Gerdes, M. (1999). The Philadelphia story: The 22q11.2 deletion: Report on 250 patients. *Genetic Counseling (Geneva, Switzerland)*, *10*, 11–24. PMID:10191425

McDonald-McGinn, D. M., Tonnesen, M. K., Laufer-Cahana, A., Finucane, B., Driscoll, D. A., Emanuel, B. S., & Zackai, E. H. (2001). Phenotype of the 22q11.2 deletion in individuals identified through an affected relative: Cast a wide FISHing net! *Genetics in Medicine*, *3*(1), 23–29. doi:10.1097/00125817-200101000-00006 PMID:11339373

Spectrum of Neurodegeneration in Autism Spectrum Disorder

McPheeters, M. L., Warren, Z., Sathe, N., Bruzek, J. L., Krishnaswami, S., Jerome, R. N., & Veenstra-Vanderweele, J. (2011). A systematic review of medical treatments for children with autism spectrum disorders. *Pediatrics*, *127*(5), e1312–e1321. doi:10.1542/peds.2011-0427 PMID:21464191

Morgan, J. T., Chana, G., Pardo, C. A., Achim, C., Semendeferi, K., Buckwalter, J., ... Everall, I. P. (2010). Microglial activation and increased microglial density observed in the dorsolateral prefrontal cortex in autism. *Biological Psychiatry*, *68*(4), 368–376. doi:10.1016/j.biopsych.2010.05.024 PMID:20674603

Myers, S. M., Johnson, C. P., & Lipkin, P. H. (2007). Management of children with autism spectrum disorders. *Pediatrics*, *120*(5), 1162–1182. doi:10.1542/peds.2007-2362 PMID:17967921

Newschaffer, C. J., Croen, L. A., Daniels, J., Giarelli, E., Grether, J. K., Levy, S. E., ... Windham, G. C. (2007). The epidemiology of autism spectrum disorders. *Annual Review of Public Health*, 28(1), 235–258. doi:10.1146/annurev.publhealth.28.021406.144007 PMID:17367287

Niklasso, L., Rasmussen, P., Oskarsdottir, S., & Gillberg, C. (2001). Neuropsychiatric disorders in the 22q11 deletion syndrome. *Genetics in Medicine*, *3*(1), 79–84. doi:10.1097/00125817-200101000-00017 PMID:11339385

Niklasson, L., Rasmussen, P., Oskarsdottir, S., & Gillberg, C. (2002). Chromosome 22q11 deletion syndrome (CATCH 22): Neuropsychiatric and neuropsychological aspects. *Developmental Medicine and Child Neurology*, 44(01), 44–50. doi:10.1017/S0012162201001645 PMID:11811651

Ogilvie, C. M., Moore, J., Daker, M., Palferman, S., & Docherty, Z. (2009). Chromosome 22q11 deletions are not found in autistic patients identified using strict diagnostic criteria. International Molecular Genetics Study of Autism Consortium. *American Journal of Medical Genetics*, *96*(1), 15–17. doi: PMID:10686546

Ohnishi, T., Matsuda, H., Hashimoto, T., Kunihiro, T., Nishikawa, M., Uema, T., & Sasaki, M. (2000). Abnormal regional cerebral blood flow in childhood autism. *Brain*, *123*(9), 1838–1844. doi:10.1093/brain/123.9.1838 PMID:10960047

Perry, V. H., & O'Connor, V. (2010). The role of microglia in synaptic stripping and synaptic degeneration: A revised perspective. *ASN Neuro*, *2*(5), e00047. doi:10.1042/AN20100024 PMID:20967131

Piven, J., Palmer, P., Jacobi, D., Childress, D., & Arndt, S. (1997). Broader autism phenotype: Evidence from a family history study of multiple-incidence autism families. *The American Journal of Psychiatry*, *154*(2), 185–190. doi:10.1176/ajp.154.2.185 PMID:9016266

Quirk, G. J., & Bee, J. S. (2006). Prefrontal involvement in the regulation of emotion: Convergence of rat and human studies. *Current Opinion in Neurobiology*, *16*(6), 723–727. doi:10.1016/j.conb.2006.07.004 PMID:17084617

Quirk, G. J., & Mueller, D. (2008). Neural mechanisms of extinction learning and retrieval. *Neuropsychopharmacology*, *33*(1), 56–72. doi:10.1038j.npp.1301555 PMID:17882236

Rineer, S., Finucane, B., & Simon, E. W. (1998). Autistic symptoms among children and young adults with isodicentric chromosome 15. *American Journal of Medical Genetics*, 74, 121–128. PMID:9754629

Robins, D. L., Fein, D., Barton, M. L., & Green, J. A. (2001). The Modified Checklist for Autism in Toddlers: An initial study investigating the early detection of autism and pervasive developmental disorders. *Journal of Autism and Developmental Disorders*, *31*(2), 131–144. doi:10.1023/A:1010738829569 PMID:11450812

Robinson, E. B., Lichtenstein, P., Anckarsäter, H., Happé, F., & Ronald, A. (2013). Examining and interpreting the female protective effect against autistic behavior. *Proceedings of the National Academy of Sciences of the United States of America*, *110*(13), 5258–5262. doi:10.1073/pnas.1211070110 PMID:23431162

Rock, R. B., Gekker, G., Hu, S., Sheng, W. S., Cheeran, M., Lokensgard, J. R., & Peterson, P. K. (2004). Role of microglia in central nervous system infections. *Clinical Microbiology Reviews*, *17*(4), 942–964. doi:10.1128/CMR.17.4.942-964.2004 PMID:15489356

Rossignol, D. A. (2009). Novel and emerging treatments for autism spectrum disorders: A systematic review. *Annals of Clinical Psychiatry*, 21(4), 213–236. PMID:19917212

Roubertie, A., Semprino, M., Chaze, A. M., Rivier, F., Humbertclaude, V., Cheminal, R., ... Echenne, B. (2001). Neurological presentation of three patients with 22q11 deletion (CATCH 22q11.2 deletion syndrome). *Brain & Development*, 23(8), 810–814. doi:10.1016/S0387-7604(01)00258-3 PMID:11720799

Rutter, M., Le Couteur, A., & Lord, C. (2003). *Autism Diagnostic Interview-Revised*. Los Angeles, CA: Western Psychological Services.

Sæmundsen, E., Magnusson, P., Smari, J., & Sigurdardottir, S. (2003). Autism Diagnostic Interview-Revised and the Childhood Autism Rating Scale: Convergence and discrepancy in diagnosing autism. *Journal of Autism and Developmental Disorders*, *33*(3), 319–327. doi:10.1023/A:1024410702242 PMID:12908834

Sah, P., Faber, E. S., Lopez De Armentia, M., & Power, J. (2003). The amygdaloid complex: Anatomy and physiology. *Physiological Reviews*, 83(3), 803–834. doi:10.1152/physrev.00002.2003 PMID:12843409

Sajdel-Sulkowska, E. M., Xu, M., & Koibuchi, N. (2009). Increase in cerebellar neurotrophin-3 and oxidative stress markers in autism. *Cerebellum (London, England)*, 8(3), 366–372. doi:10.100712311-009-0105-9 PMID:19357934

Sajdyk, T. J., & Shekhar, A. (2000). Sodium lactate elicits anxiety in rats after repeated GABA receptor blockade in the basolateral amygdala. *European Journal of Pharmacology*, *394*(2-3), 265–273. doi:10.1016/S0014-2999(00)00128-X PMID:10771292

Schechtman, M. A. (2007). Scientifically unsupported therapies in the treatment of young children with autism spectrum disorders. *Pediatric Annals*, *36*(8), 497–505. doi:10.3928/0090-4481-20070801-12 PMID:17849608

Schreibman, L. (2000). Intensive behavioral/psychoeducational treatments for autism: Research needs and future directions. *Journal of Autism and Developmental Disorders*, *30*(5), 373–378. doi:10.1023/A:1005535120023 PMID:11098871

Spectrum of Neurodegeneration in Autism Spectrum Disorder

Schumanon, C.M., & Amaral, D.G. (2006). Stereological analysis of amygdala neuron number in autism. *J Neurosci*, *26*, 7674–7679.

Shprintzen, R. J. (2006). Velo-cardio-facial syndrome: A distinctive behavioral phenotype. *Mental Retardation and Developmental Disabilities Research Reviews*, 6(2), 142–147. doi: PMID:10899808

Sigman, M., & Capps, L. (1997). *Children with autism: A developmental perspective*. Boston: Harvard University Press.

Sotres-Bayon, F., & Quirk, G. J. (2010). Prefrontal control of fear: More than just extinction. *Current Opinion in Neurobiology*, 20(2), 231–235. doi:10.1016/j.conb.2010.02.005 PMID:20303254

Spencer, T., Biederman, J., Wilens, T., Harding, M., O'Donneel, D., & Griffin, S. (1996). Pharmacotherapy of attention-deficit hyperactivity disorder across the life cycle. *Journal of the American Academy of Child and Adolescent Psychiatry*, 35(4), 409–432. doi:10.1097/00004583-199604000-00008 PMID:8919704

Spezio, M. L., Huang, P. Y., Castelli, F., & Adolphs, R. (2007). Amygdala damage impairs eye contact during conversations with real people. *The Journal of Neuroscience*, *27*(15), 3994–3997. doi:10.1523/JNEUROSCI.3789-06.2007 PMID:17428974

Steffenburg, S., & Gillberg, C. (1986). Autism and autistic like condition in Swedish rural and urban areas: A population study. *The British Journal of Psychiatry*, *149*(1), 81–87. doi:10.1192/bjp.149.1.81 PMID:3779317

Stigler, K. A., McDonald, B. C., Anand, A., Saykin, A. J., & McDougle, C. J. (2011). Structural and functional magnetic resonance imaging of autism spectrum disorders. *Brain Research*, *1380*, 146–161. doi:10.1016/j.brainres.2010.11.076 PMID:21130750

Stuss, D. T., & Knight, R. T. (2013). *Principles of frontal lobe function*. Oxford, UK: Oxford University Press. doi:10.1093/med/9780199837755.001.0001

Suzuki, K., Sugihara, G., Ouchi, Y., Nakamura, K., Futatsubashi, M., Takebayashi, K., ... Mori, N. (2013). Microglial activation in young adults with autism spectrum disorder. *JAMA Psychiatry*, *70*(1), 49–58. doi:10.1001/jamapsychiatry.2013.272 PMID:23404112

Sweeten, T. L., Posey, D. J., Shekhar, A., & McDougle, C. J. (2002). The amygdala and related structures in the pathophysiology of autism. *Pharmacology, Biochemistry, and Behavior*, 71(3), 449–455. doi:10.1016/S0091-3057(01)00697-9 PMID:11830179

Swillen, A., Devriendt, K., Legius, E., Prinzie, P., Vogels, A., & Ghesquiere, P. (1999). The behavioral phenotype in velo-cardio-facial syndrome (VCFS): From infancy to adolescence. *Genetic Counseling* (*Geneva, Switzerland*), *10*, 79–88. PMID:10191433

Szatmari, P., Jones, M. B., Fisman, S. F., Tuff, L., Bartolucci, G., Mahoney, W. J., & Bryson, S. E. (1995). Parents and collateral relatives of children with pervasive developmental disorders: A family history study. *American Journal of Medical Genetics*, *60*(4), 282–289. doi:10.1002/ajmg.1320600405 PMID:7485262

Szatmari, P., MacLean, J. E., Jones, M. B., Bryson, S. E., Zwaigenbaum, L., Bartolucci, G., ... Tuff, L. (2000). The familial aggretion of the lesser varient in biological and non-biological relatives of PDD probands: A family history study. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, *41*(5), 579–586. doi:10.1111/1469-7610.00644 PMID:10946750

Tezenas, D., Montcel, S., Mendizibal, H., Ayme, S., Levy, A., & Philip, N. (1996). Prevalence of 22q11 microdeletion. *Journal of Medical Genetics*, *33*, 719.

Vajda, F. J. (2002). Neuroprotection and neurodegenerative disease. *Journal of Clinical Neuroscience*, *9*(1), 4–8. doi:10.1054/jocn.2001.1027 PMID:11749009

Van-Kooten, I. A., Palmen, S. J., von Cappeln, P., Steinbusch, H. W., Korr, H., Heinsen, H., ... Schmitz, C. (2008). Neurons in the fusiform gyrus are fewer and smaller in autism. *Brain*, *131*(Pt 4), 987–999. doi:10.1093/brain/awn033 PMID:18332073

Vargas, D. L., Nascimbene, C., Krishnan, C., Zimmerman, A. W., & Pardo, C. A. (2005). Neuroglial activation and neuroinflammation in the brain of patients with autism. *Annals of Neurology*, *57*(1), 304. doi:10.1002/ana.20315 PMID:15546155

Volkmar, F., Cook, E., Pomeroy, J., Realmuto, G., & Tanguay, P. (1999). the Work Group on Quality Issues, AACAP Practice parameters for the assessment and treatment of children, adolescents, and adults with autism and other pervasive developmental disorders. *Journal of the American Academy of Child and Adolescent Psychiatry*, *38*(Suppl 12), 32S–54S. doi:10.1016/S0890-8567(99)80003-3 PMID:10624084

Vrancic, D., Nanclares, V., Soares, D., Kulesz, A., Mordzinski, C., Plebst, C., & Starkstein, S. (2002). Sensitivity and specificity of the Autism Diagnostic Inventory-telephone screening in Spanish. *Journal of Autism and Developmental Disorders*, *32*(4), 313–320. doi:10.1023/A:1016335003256 PMID:12199136

Weber, W., & Newmark, S. (2007). Complementary and alternative medical therapies for attention-deficit/ hyperactivity disorder and autism. *Pediatric Clinics of North America*, 54(6), 983–1006. doi:10.1016/j. pcl.2007.09.006 PMID:18061787

Whitney, E. R., Kemper, T. L., Bauman, M. L., Rosene, D. L., & Blatt, G. J. (2008). Cerebellar Purkinje cells are reduced in a subpopulation of autistic brains: A stereological experiment using calbindin-D28k. *Cerebellum (London, England)*, 7(3), 406–416. doi:10.100712311-008-0043-y PMID:18587625

Wigram, T., & Gold, C. (2006). Music therapy in the assessment and treatment of autistic spectrum disorder: Clinical application and research evidence. *Child: Care, Health and Development, 32*(5), 535–542. doi:10.1111/j.1365-2214.2006.00615.x PMID:16919132

Witwer, A., & Lecavalier, L. (2005). Treatment incidence and patterns in children and adolescents with autism spectrum disorders. *Journal of Child and Adolescent Psychopharmacology*, *15*(4), 671–681. doi:10.1089/cap.2005.15.671 PMID:16190798

Woodin, M., Wang, P. P., Aleman, D., McDonald-McGinn, D., Zackai, E., & Moss, E. (2001). Neuropsychological profile of children and adolescents with the 22q11.2 microdeletion. *Genetics in Medicine*, *3*(1), 34–39. doi:10.1097/00125817-200101000-00008 PMID:11339375

Spectrum of Neurodegeneration in Autism Spectrum Disorder

Wyss-Coray, T., & Mucke, L. (2002). Inflammation in neurodegenerative disease–a double-edged sword. *Neuron*, *35*(3), 419–432. doi:10.1016/S0896-6273(02)00794-8 PMID:12165466

Yergin-Allsop, M., Rice, C., Karapurkar, T., Doemberg, N., Boyle, C., & Murphy, C. (2003). Prevalence of autism in a US metropolitan community. *Journal of the American Medical Association*, 289, 49–55. PMID:12503976

Yirmiya, N., Sigman, M., Kasari, C., & Mundy, P. (1992). Empathy and cognition in high-functioning children with autism. *Child Development*, *63*(1), 150–160. doi:10.2307/1130909 PMID:1551323

Zwaigenbaum, L., Bryson, S., Rogers, T., Roberts, W., Brian, J., & Szatmari, P. (2005). Behavioral manifestations of autism in the first year of life. *International Journal of Developmental Neuroscience*, 23(2-3), 143–152. doi:10.1016/j.ijdevneu.2004.05.001 PMID:15749241

Chapter 17 Creutzfeldt-Jakob Disease: A Prion-Related Neurodegenerative Disorder

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ABSTRACT

Creutzfeldt-Jakob disease (CJD) is a rare disease associated with neurodegeneration mostly characterized by damage to the neurons. CJD is caused by aggregation of misfolded proteins known as prions; thus, CJD is said to be a prion-related illness. CJD and other prion-related illnesses such as Kuru and Gerstmann-Sträussler-Scheinker disease (GSS) have been reported to have complex mechanisms due to their association with the brain and the nervous system in general. A lot of questions have been raised about the mechanism, diagnosis, and pathogenesis of this disease. The complexity of prion proteins themselves have contributed to more questions about the complications of CJD, whether misfolding of the prions are responsible for neurodegeneration or the misfolding are mere symptoms of the disease. This chapter attempts to explore some details about CJD and answers most related questions about the disease's mechanism. The author finally attempts to explore recent development in pathogenesis, diagnosis, and treatment of CJD.

INTRODUCTION

Human NDs have been one of the most deadly disorders with complex mechanisms and complicated treatment processes (Montie & Durcan, 2013).Particularly, thehumanprion diseases are one of the most rare human neurodegenerative diseases widely studied. These are associated with misfolding of proteins which accumulated and cause destruction of cells of the system (Friedman-Levi *et al.*, 2011). These proteins otherwise called prions, are misfolded leading to their aggregation into opaque amyloid structures. The pathogenesis of these diseases are mostly related to accumulation of the amyloid aggregates which in most cases, is associated with neural damage (Holman *et al.*, 2010). Prion diseases are therefore a group of progressive but fatal neurodegenerative disorders affecting both humans and animals, they are otherwise referred to as transmissible spongiform encephalopathies (TSEs) (Jackson &

DOI: 10.4018/978-1-5225-5282-6.ch017

Creutzfeldt-Jakob Disease

Krost, 2014). Some of these prion diseases including CJD, Alzheimer's disease (AD), and Huntington's disease (HD) are having rare occurrence with complicated pathogenesis (Kovacs & Budka, 2008). CJD is a prion related illness characterized by aggregation of misfolded proteins or prions. It belongs to a family of human diseases associated to and or characterized by neurodegenerative conditions and they include; Gerstmann-Sträussler-Scheinker syndrome (GSS), fatal familial insomnia etc. These diseases are thought to be having similar characteristics with animals spongiform encephalopathies such as ''cow madness'' also known as bovine spongiform encephalopathies (BSEs) (Lang, Heckmann, & Neundörfer, 1998). These could have both sporadic or spontaneous occurrence or could be genetical depending on occurring cases. As such as in BSEs, prions could also be transmitted resulting in the spreading of the disease (States *et al.*, 2005). Most researches in this area are focusing towards establishing a link between variant phenotypes of CJD and with the molecular features studied in wide ranges of CJD conditions.

This chapter attempts to explore details on CJD including the basic description of disease, its pathogenesis, classification, diagnosis, and epidemiology. The author objectively attempts to explain different perspectives of the current researches on Creutzfeldt-Jakob and a future sight as well recommendation on the treatment of the disease.

BACKGROUND

Neurons being the major cells of the nervous system could be damaged progressively by one cause or another leading to neurodegeneration and consequently diseases, such diseases are known as NDs (Aguzzi & Zhu, 2012). NDs are diseases associated with damage in structure and functions of the neurons, one of the important cells of the nervous system (Aguzzi & Zhu, 2012). In neurobiology, neurons form the complex network of communication around the central nervous system. Damage to neurons due to a number of factors has been reported to be lethal and associated with a number of diseases and disorders (Murray & Davis, 2003).

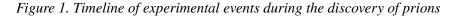
CJD was discovered by the German neurologist, Hans Gerhard Creutzfeldt and Alfons Maria Jakob in 1921 (Cordery *et al.*, 2003). Creutzfeldt was the first to describe the disease characterized with neurodegeneration and Jakob proposed the disease to be associated with prions (Holman *et al.*, 2010). However some of descriptions of the disease made by Jakob and Creutzfeldt did not match the current description of the disease (Creutzfeldt, 2002). This including being transmissible as well as currently being observed to be part of a class of both human and animal diseases called TSEs.

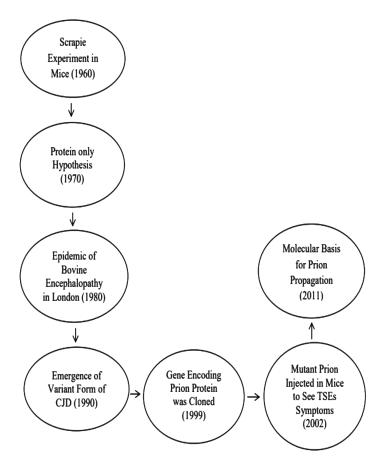
CJD as a TSE is associated with and mostly caused by diseased or misfolded prions (Aguzzi & Zhu, 2012). Prions are proteins that often misfold into amyloid structures and can possibly infect other proteins resulting into spreading or transmission (Jackson & Krost, 2014). In other words, prion proteins affect other neighboring proteins converting them into misfolded prion proteins and initiate the pathogenesis of TSEs including CJD (Lang, Heckmann, & Neundörfer). Spongiform refers to the characteristic appearance of infected brains, which become filled with holes until they resemble sponges under a microscope (Aguzzi & Zhu, 2012) . Transmission of the prion proteins in CJD was not initially established (Belay, 2004). The idea that prion proteins can be transmitted in CJD was established in 1986 after the London epidermic (Garske & Ghani, 2010). The incidence of the epidermic has stimulated interest in study of CJD with major aspects on the molecular mechanisms of its pathogenesis (Wilesmith *et al.*, 1988). Studies on prion proteins in fungi has already established the transmissible property of normal prions in the cell (Anderson *et al.*, 1996). Similar prion studies on vCJD was carried on human patients sample which

confirmed the spreading ability of misfolded prions in CJD (Griffith, 1967). Protein only hypothesis was established and served as a model for supporting transmissibility of prion proteins (Soto, 2011). Consequently, studies were on to provide experimental basis for the hypothesis. Particularly, transgenic animals expressing mutant prion protein that develop clinical and pathological signs of TSE, were produced (Soto, 2004). Figure 1 shows a brief timeline of the major occurrences and findings related to TSEs.

MISFOLDED PROTEINS' AGGREGATION AND THEIR ROLES IN PROTEINOPATHIES

Misfolded proteins existing together with unfolded proteins in the cell are re-folded by molecular chaperones (Kim, *et al.*, 2013). Proteins that refused to fold properly are immediately degraded however in pathologic cases, this protein resist degradation and accumulate (Halasi *et al.*, 2017). The ability to resist degradation has been attributed to folding into specific abnormal forms which are the infectious prion forms (Currais *et al.*, 2017). During biosynthesis cells continuously form stream of misfolded proteins and are refolded, degraded, or sequestered into specified cellular compartments such as aggresomes (Hartl, Bracher, & Hayer-Hartl, 2011). Many of such proteins have been seen to be misfolded and consequently





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aggregates to form insoluble structures in the cell (Stefani, 2004). These faulty or abnormally aggregates of proteins have been seen in many neurodegenerative diseases. Table 1 shows a brief summary of the misfolded proteins in some NDs. Molecular chaperones are proteins that bind assist in protein folding and remaining infolded form. They bind nascent polypeptides as they emerge from ribosomes and participate in every step in the handling of misfolded proteins. These proteins play crucial roles by monitoring the quality of the folded chains and like highlighted earliar, refold misfolded proteins (Kim *et al.*, 2013). Chaperones also target the misfolded proteins that couldn't be refolded/repaird for degradation.

PRION MISFOLDING AND CREUTZFELDT-JAKOB DISEASE

Prions are thought to be found all over the body with yet to be known function (Friedman-Levi *et al.*, 2011). In TSEs, when a prion enters a healthy organism, it induces an healthy protein to misfold into anonymous abnormal form through a chain of unknown processes (Aguzzi & Zhu, 2012). Certain proteins in cells of both lower and higher organisms such as bacteria and humans, promote or assist folding of proteins into various forms and conformations, these are known as chaperons. Thus as a prion promote misfolding of a normal protein into its own prion form, it results into spread of the prion protein which consequently form amyloid aggregate forms or plagues (Fania *et al.*, 2017). The amyloid form refers to an aggregate of misfolded or prion proteins with characteristic opaque sheets (Jackson & Krost, 2014).

Prion aggregates or amyloids are characteristically stable and show resistant to denaturation processes caused by temperature and chemical changes and as such stay for prolonged periods in the infected cell consequently resulting into its death (Kovacs & Budka, 2008). One question that is yet to be answered is, "misfolding of the prions causes the damage to the cell or other external damages to the cell causes the initiation of the misfolding resulting into the prion aggregation" (Kovacs & Budka, 2008). However some scientists believed mutations in some genes initiated prion development in HD, a prion related disease (Mercado *et al.*, 2015). However in most TSE, infection of a prion resulted into transmission and spreading of neighboring prions resulting into their misfolding and aggregation into amyloid prion forms (Jackson & Krost, 2014).

Proteinopathy	Aggregating Protein(s)
Spongiform Encephalopathies	Prion protein
Parkinson's Disease	α-Synuclein
Cerebral β-amyloid angiopathy	Amyloid β peptide
Frontotemporal lobar degeneration	TAR DNA-binding protein 43 (TDP-43)
Alzheimer's disease	Tau protein
Huntington's disease	Huntingtin protein
Amyotrophic lateral sclerosis	Superoxide dismutase 1
Multiple tauopathies	Tau protein (microtubule associated)
Alexander's disease	Glial fibrillary acidic protein
Enfuvirtide amyloidosis	Enfuvirtide protein

Table 1. Some proteins Misfolded and the consequencing proteinopathies

The ability of misfolded prions (mostly from bovine encephalopathies) to infect neighboring normal prions was initially a not accepted (Murray & Davis, 2003). With the ability to infect prion to cause disease upon incubation, it resembles the commonly accepted processes of pathogenesis in microbes including bacteria, viruses and fungi (Nuvolone et al., 2017). Critics of infectious prion pathogenesis argued that bacteria and viruses, major causative agents of diseases are mostly living organisms with delicate and distinct structures (Whalley, 2017). These organisms either secret toxins that temper normal cellular processes and or insert their genomes into the host cell thereby destroying the hosting cells and continue to spread and transmit to other cells by similar mechanism (Jackson & Krost, 2014). According to these critics, a protein would not be able to be transmitted in a similar fashion. However researches in various prion-related illnesses including CJD described the mechanism of prion transmission especially in TSEs (States et al., 2005). Normal prion proteins being found all over the cell can be infected by an infectious prion mostly coming from contaminated food (e.g meat from cattle suffering from BSEs, also known as cow madness). These infectious agents infect the normal prion by facilitating the folding its into a similar abnormal form hence becoming a potential infectious prion. The infected prion infects another and the process continues all over the neighboring prions consequently aggregating into amyloid forms (Nuvolone et al., 2017). However it is not exactly known how it facilitates the misfolding process, some hypothesized that by acting in a similar way as molecular chaperons (Whalley, 2017). In Figure 2, the underlying mechanism of prion misfolding is mentioned.

In CJD, aggregation of misfolded prions has been thought as the cause of neurodegeneration due to increased destruction of neuronal cells (Lang et al., 1998). However the actual mechanism by which these aggregates cause cell damage has not been clearly established and appears to be variable within each of the disease (Montie & Durcan, 2013). Some scholars hypothesized that misfolded disease proteins appear to act primarily by toxic gain-of-function and/or dominant-negative effects, although loss-of-function effects have also been observed (Whalley, 2017).

Although most misfolding events might be associated with toxicity leading to disease but not all events are responsible for toxicity (Valastyan, 2014). In the fungal organism, *Saccharomyces cerevisiae*, the presence and availability of prions are regulated and most of these prions utilized for various adaptive purposes (Chernova, Wilkinson, & Chernoff, 2014). Prion proteins in CJD are infectious and are associated with progressive neural damage hence dementia and other symptoms and conditions of neurodegeneration (Kovacs & Budka, 2008).

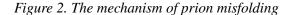
CLASSES OF CREUTZFELDT-JAKOB DISEASE

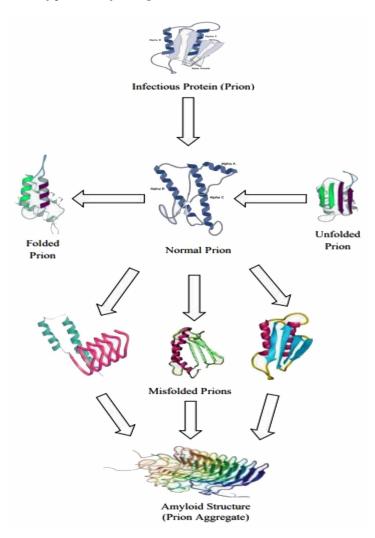
Different studies have suggested variety of classification of CJD. Some of the classifications were made based on the neuropathology of the disease (Hill *et al.*, 2003). However, based on the occurrence and cause of CJD, the disease can be classified into two main groups.

Sporadic Creutzfeldt-Jakob Disease

Sporadic CJD (sCJD) refers to the rarest yet the most common form of the disease. Its major cause has been established to be genetically associated (Brandt, 2003). sCJD is reported to possess variability in its clinical and neuropathological phenotype (Mead *et al.*, 2013). However, It remains to be seen whether the variability might be related to variations in the causative TSE agent strains, or to the influence of

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the methionine/valine polymorphism at codon 129 of the *PRNP* gene. The polymorphism on the PRNP gene encoding either methionine or valine remain the major susceptibility factor of sCJD with mutant methionine prototype dominating in most cases than valine (Rubenstein, Petersen, & Wisniewski, 2016). The germline mutation in the *PRNP* gene can be inherited in an autosomal dominant fashion leading to harboring a homozygous allele (Mead *et al.*, 2001). Key phenomenon in the molecular mechanism of sCJD is the conversion of PrP^C (normal prion) to the pathological PrP^{Sc} (diseased Prion) which is the pathological form associated with the disease. However the conversion is caused initially by mutation as earlier described (Bilheude *et al.*, 2008).

sCJD constitute about 85% of all recognized prion diseases in humans (Safar *et al.*, 2005). It has been reported that 1 person in every million of people suffers from sCJD every year (Chen & Dong, 2016). It usally affects people between the ages of 40-60 and mostly takes upto 6-8 weeks while taking upto

a year in some minority (Mastrianni, 2010). Similarly Brown et al. (1984) reported that about 10% of cases of sCJD have prolonged clinical course with the duration of the disease taking upto 2 years. Major symptoms associated but peculiar to sCJD are less pronounced and are mostly seen in other forms of the disease, however, a significant percentage of sCJD has been associated with cerebellar ataxia (Cooper *et al.*, 2006). Other symptoms including fatigue, weight loss, immediate loss of coordination, and Halucination. Later stage symptoms include depression and sensory disturbances are common in sCJD that consequently progress to profound dementia (Geschwind *et al.*, 2013).

Variant Creutzfeldt-Jakob Disease

Variant CJD is the most widely known form of CJD as it can be transmitted between individuals via blood transfusion and can be acquired through ingestion of food contaminated with the agent of BSE (Maheshwari *et al.*, 2015). BSE is a TSEs in cattle and bovine family (Creutzfeldt, 2002). Consumption of a meat of cattle infected with BSE results into infection and transmission of infectious into the body of the healthy consuming individual resulting into infection of normal prions in the individual (Friedman-Levi *et al.*, 2011) The mechanism of conversion or transmission of a normal prion to an infectious one has been argued (Jackson & Collinge, 2001). However certain researchers have described key mechanisms involved. A Normal prion (PrP^{C}) is converted to an infected or diseased prion (PrP^{Sc}) during a post-translation modification process involving structural modifications of the protein and resulting in a higher β -sheet content (Riesner, 2003). PrP^{C} is completely degraded after controlled digestion with proteinase K (PK) in the presence of detergents. PrP^{Sc} is N-terminally truncated under such conditions, resulting in a PK resistant core, termed PrP^{RES} (States *et al.*, 2005). Most symptoms associated with the vCJD are similar to vCJD. However, in vCJD cognitive disorders appeared at much earliar stages than sCJD with severe behavioural disturbances (Mastrianni, 2010).

EPIDEMIOLOGY OF CREUTZFELDT-JAKOB DISEASE

Before the identification of vCJD in 1996, only three forms of CJD were thought to be existing and represent the most common prion related illness in humans. The three forms include:

- Familial CJD, comprising of about 5-15% of CJD cases. It has been found to be with one or more mutations on the *PRNP* gene (Babi *et al.*, 2016, WHO, 2010).
- Iatrogenic CJD, representing about 5% of CJD cases and mostly occurs as result of contamination from used surgical equipment (WHO, 2010).
- Sporadic CJD, most common form of CJD consisting of about 85% of CJD cases of CJD. It is also associated with mutation of the *PRNP* gene and was reported to occur throughout the world at the rate of about one per million people (Bonda *et al.*, 2016, WHO, 2010).

CJD occurs world and among every population with an average of 1.5-2.5 cases in every million per years (Brandel *et al.*, 2015). According to the centers for disease control and prevention (2017), the average case occurrence of CJD is 285.19 from 1979-2015 with the cases increasing with time progress.

PATHOGENESIS OF CREUTZFELDT-JAKOB DISEASE

Destruction of neurons due to an accumulation of these proteins consequently resulted in neurodegeneration (Whalley, 2017). A major characteristic of CJD and by extension most TSEs is formation of sponge-like appearance in the brain (Acevedo-Morantes & Wille, 2014). The conversion of the normal prion protein PrP^{c} to $PrP^{s_{c}}$ initiates the pathogenesis of the disease (Riesner, 2003). Normal PrP^{c} is a membraned surfaced protein of whose clear function has been unknown. Structurally, it has a predominantly α -helical structure that is highly conserved between species (Trevitt & Singh, 2003). The pathogenic prion form, $PrP^{s_{c}}$ is formed by posttranslational modification of the normal PrP^{c} form without the alteration of the primary structure of the normal protein. $PrP^{s_{c}}$ is characterized by highly β -plated structures and being protease resistant (Kovacs & Budka, 2008). In vCJD, prion infection is caused eating meat or any contaminated food from cattle with ''cow madness'' also known as bovine spongiform ESEs (Billette de Villemeur *et al.*, 1996). The infected prion facilitates the misfolding of the normal prion PrP^{c} into the pathogenic prion $PrP^{s_{c}}$ (Figure 3). Most biochemical evidences have shown that both BSE and vCJD were caused by invariably similar prion strain (Riesner, 2003). Distinct prion related diseases or illnesses can be differentiated by their biological properties including incubation periods and patterns of neuropathology (Trevitt & Singh, 2003).

Major symptoms of associated generally with CJD include loss of memory, coordination, sight and dementia. At early stage symptoms include memory failure and behavioral changes. As the disease progress, individual suffer from involuntary movements, loss of sight and even coma (World Health Organization [WHO], 2010).

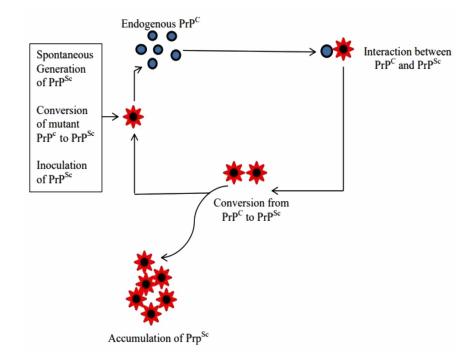


Figure 3. The pathogenesis of Creutzfeldt-Jakob disease

DIAGNOSIS OF CREUTZFELDT-JAKOB DISEASE

Clinical symptoms associated with CJD are more are less similar in most neurological impairment (Van Everbroeck *et al.*, 2004). Neurological signs characterizing CJD include;lack of coordination, difficulty in walking and involuntary movements. These signs progress with each developmental stage of the illness as it progresses and, by the time of death, patients become completely immobile and mute (Poser *et al.*, 2000). In diagnosis of CJD, disorders with similarl features with CJD can be ruled out using several tests, one of which is biopsy test. Brain biopsy is the most accurate test in CJD diagnosis. It involves a procedure where a small piece of brain tissue is removed for study (Heinemann *et al.*, 2008). However, any brain surgery carries risk of damage to the brain and a tissue might be removed from a non-infected area thus will likely show an inaccurate representation of the disease (Creutzfeldt, 2002). In such case, the biopsy cannot be a reliable test to confirm or rule out the diagnosis. Similarly, autopsies on the whole brain can be used in given cases of CJD diagnosis. Autopsy is the study carried out on dead bodies to analyse different systems of the body (Heinemann *et al.*, 2008). It also is useful for research. Other tests commonly used for CJD diagnosis include:

- A brain magnetic resonance imaging (MRI) which uses magnetic and radio waves to capture a detailed image of the whole or portion of the brain in other to have a detailed picture and observe defects in the brain peculiar to CJD (Morgan *et al.*, 2009) present
- A brain computed tomography (CT) scan, which provides much more details on the structural components of the brain tissues and can be used to observe patient's symptoms related to stroke or brain tumor (Rubenstein *et al.*, 2016).
- A blood and fluid test, where different techniques can be used on blood of patients' sample to rule out other forms of dementia that may be treated (Zerr *et al.*, 1998). Cerebrospinal fluid tests such as 14-3-3 protein test, are particularly carried out to see if proteins such as the 14-3-3 protein that is associated with dementia or CJD are present (Zanusso *et al.*, 2016).

TREATMENT OF CREUTZFELDT-JAKOB DISEASE

In general, there is no treatment available to cure CJD. Treatment of prion diseases remains supportive; no specific therapy has been shown to stop the progression of these diseases (Belcastro *et al.*, 2010). Treatments are mostly done to alleviate the symptoms and to make the patient as comfortable as possible (Pocchiari & Ladogana, 2015). However for patients with CJD at initial stages, therapeutic measures can be take via use of many some drugs such as amantadine and acyclovir that target misfolding of a normal prion (Appleby & Lyketsos, 2011). Similarly, some pentosan polysulfate were used to target protease resistance of the diseased or misfolded prions and thus increasing their susceptibility to protease attack (Zuber, Ludewigs, & Weiss, 2007). Another target initial accumulation of the misfolded prions into an amyloid structure and promotes its clearance (Gauczynski *et al.*, 2006). Although some of the drugs have been successfully used for those purposes, their use has not been consistent in subsequent CJD patients (Vetrugno *et al.*, 2015). Researchers are now targeting protein regions on the misfolded proteins and designing specific antibodies to target them for destruction (Panegyres & Armari, 2013).

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Due to the circumstances associated with treatment of CJD, the World Health Organization and other supporting agencies set up measures to alert the public about CJD (WHO, 2008). Some measures are taken to protect public health include:

- As researches established links between vCJD and BSE, governments through Health departments enlist CJD as a major health concern for the public. Example government of the United Kingdom made CJD a notifiable disease in 1988 (Ponte, 2006).
- The UK government similarly placed a ban on individuals that have lived in countries with high risk of BSE as a means to prevent a possible spread of infections to patients (Food and Drug Administration, 2002).

World Health Organization's Recommendations

The World Health Organization in its effort to provide measures that could decrease spread of BSEs and TSEs including CJD, made the following recommendations:

- WHO recommended that countries should ban use of ruminant tissues in food supplement preparation of domestic animals particularly members of the bovine family. All tissues potentially containing BSE agent, nor part or product of any animal which has shown signs of a TSE should be prevented from entering the (human or animal) food chain. (Centers for Disease Control and Prevention, 1996).
- WHO also recommended ban on pharmaceutical industries on the use of materials and product from the bovine family in which TSEs naturally occur. Id such materials are to be necessary be used, they should be important from countries with 0% reported cases of BSEs (WHO, 2003).
- The guidelines on tissue infectivity distribution in TSEs in 2006 provide information and assist national regulatory authorities in conducting risk assessments of vCJD. The tables on tissue infected were later updated in 2010 with the aim of providing current knowledge of tissue infectivity providing information on the possible transmissions in TSEs (WHO, 2010).

FUTURE RESEARCH DIRECTIONS

Previously researchers have been towards understanding mainly the pathogenesis and spread of the disease as being transmissible (Kovacs & Budka, 2008). Current and future researches focus on elucidation of the process of prion proteins associated with CJD (Aguzzi & Zhu, 2012). Targeting abnormal prions is crucial in treatment of CJD (Pocchiari & Ladogana, 2015). Recent development of techniques in immunotherapy and proteomics are used to understand structural organization of prion proteins and how they aggregate to amyloid plaques (Biasini *et al.*, 2012). Other researches focus on stimulating physiological processes to eliminate prion aggregates (Karapetyan *et al.*, 2013). For example astemizole is an antihistamine drug which has been found to stimulate autophagy through its action of being an antagonist of H1 receptor, leading to generation of reactive oxygen species and activation of autophagy (Jakhar *et al.*, 2017). Prion-based screening assays have been used in model organisms such as yeast to

identify infectious prions and allowed identification of drugs active against mammalian prions (Voisset *et al.*, 2009). Similarly researches are also focusing on ways to develop agents that will block transmission of infectious prions as ways to provide cure to CJD. Future researches should stress the following:

- With the progress made in immunotherapy, directing immune cells towards recognizing specifically abnormal prions and destroying them should be a focal research area to be exploited.
- Similarly designing specific agents that target molecular chaperones that aid abnormal folding of prions can similarly be exploited.
- Different types of chaperones are associated with re-folding of abnormal prions back into normal prions. Researches should focus on these chaperones as a potential target in therapy.
- Although biopsy has been used for period in treatment of CJD, advanced technique in radiotherapy can be used to target and eliminate aggregates of misfolded proteins. Researches can be focus in neuroimaging to study the structures of these aggregates. This will ensure success of the radiotherapy.

CONCLUSION

CJD is a major concern as most researchers believed neurodegeneration progressively becomes irreversible and the disease could not be cured. The chapter described the pathogenesis of different types of CJD and their distribution. It also revealed how a number of drugs have been unsuccessfully used in treatment of CJD. As the use of most therapeutic agents has proven to be consistently unsuccessful, there is a need to change focus towards the studies to find a possible cure to the disease. Studies are now focused more towards the molecular mechanisms underlying the development and spread of the different types of the disease. With advanced techniques currently in used, it is believed that a final cure for any type of CJD will be developed.

REFERENCES

Acevedo-Morantes, C. Y., & Wille, H. (2014). The Structure of Human Prions: From Biology to Structural Models—Considerations and Pitfalls. *Viruses*, 6(10), 3875–3892. doi:10.3390/v6103875 PMID:25333467

Aguzzi, A., & Zhu, C. (2012). Five Questions on Prion Diseases. *PLoS Pathogens*, 8(5). doi:10.1371/journal.ppat.1002651 PMID:22570608

Anderson, R. A., Donnelly, C. A., Ferguson, N. M., Woolhouse, M. E. J., Watt, C. J., Udy, H. J., ... Wells, G. A. H. (1996). Transmission dynamics and epidemiology of BSE in British cattle. *Nature*, *382*(6594), 779–788. doi:10.1038/382779a0 PMID:8752271

Babi, M. A., Kraft, B. D., Sengupta, S., Peterson, H., Orgel, R., Wegermann, Z., & Luedke, M. W. (2016). Related or not? Development of spontaneous Creutzfeldt–Jakob disease in a patient with chronic, well-controlled HIV: A case report and review of the literature. *SAGE Open Medical Case Reports, 4*.

Creutzfeldt-Jakob Disease

Belay, E. D. (2004). Prions and Prion Diseases: Current Perspectives. *Emerging Infectious Diseases*, 10(12), 2265–2266. doi:10.3201/eid1012.3040847

Biasini, E., Turnbaugh, J. A., Unterberger, U., & Harris, D. A. (2012). Prion protein at the crossroads of physiology and disease. *Trends in Neurosciences*, *35*(2), 92–103. doi:10.1016/j.tins.2011.10.002 PMID:22137337

Bilheude, J., Uro-coste, E., Basset-leobon, C., Perret-liaudet, A., Ironside, J. W., Peoch, K., & Andre, O. (2008). Beyond PrP res Type 1 / Type 2 Dichotomy in Creutzfeldt- Jakob Disease. *PLoS Pathogens*, *4*(3), 1–9. PMID:18383623

Billette de Villemeur, T., Deslys, J. P., Pradel, A., Soubrié, C., Alpérovitch, A., Tardieu, M., & Agid, Y. (1996). Creutzfeldt-Jakob disease from contaminated growth hormone extracts in France. *Neurology*, *47*(3), 690–695. doi:10.1212/WNL.47.3.690 PMID:8797466

Bonda, D. J., Manjila, S., Mehndiratta, P., Khan, F., Miller, B. R., Onwuzulike, K., & Cali, I. (2016). Human prion diseases: Surgical lessons learned from iatrogenic prion transmission. *Neurosurgical Focus*, *41*(1), E10. doi:10.3171/2016.5.FOCUS15126 PMID:27364252

Brandel, J. P., Salomon, D., Capek, I., Vaillant, V., & Alpérovitch, A. (2009). Epidemiological surveillance of Creutzfeldt-Jakob in France. *Revue Neurologique*, *165*(8-9), 684–693. doi:10.1016/j.neurol.2009.04.006 PMID:19467685

Brandt, T. (Ed.). (2003). Neurological disorders: course and treatment. Gulf Professional Publishing.

Brown, P., Rodgers-Johnson, P., Cathala, F., Gibbs, C. J., & Gajdusek, D. C. (1984). Creutzfeldt-Jakob disease of long duration: Clinicopathological characteristics, transmissibility, and differential diagnosis. *Annals of Neurology*, *16*(3), 295–304. doi:10.1002/ana.410160305 PMID:6385823

Centers for Disease Control and Prevention. (1996). World Health Organization consultation on public health issues related to bovine spongiform encephalopathy and the emergence of a new variant of Creutzfeldt-Jakob disease. *MMWR*. *Morbidity and Mortality Weekly Report*, 45(14), 295. PMID:8598828

Centers for Disease Control and Prevention (CDC). (2015). Creutzfeldt-Jakob Disease Surveillance and Diagnosis. *MMWR. Morbidity and Mortality Weekly Report*, 45(31), 665.

Chen, C., & Dong, X. P. (2016). Epidemiological characteristics of human prion diseases. *Infectious Diseases of Poverty*, 5(1), 47. doi:10.118640249-016-0143-8 PMID:27251305

Chernova, T. A., Wilkinson, K. D., & Chernoff, Y. O. (2014). Physiological and environmental control of yeast prions. *FEMS Microbiology Reviews*, *38*(2), 326–344. doi:10.1111/1574-6976.12053 PMID:24236638

Concha-Marambio, Pritzkow, Moda, Tagliavini, & Ironside. (2016). New detection method could lead to noninvasive diagnosis of Creutzfeldt-Jakob disease. *Science Translational Medicine*, 8(370), 1–2.

Cooper, S. A., Murray, K. L., Heath, C. A., Will, R. G., & Knight, R. S. (2006). Sporadic Creutzfeldt–Jakob disease with cerebellar ataxia at onset in the UK. *Journal of Neurology, Neurosurgery, and Psychiatry*, 77(11), 1273–1275. doi:10.1136/jnnp.2006.088930 PMID:16835290

Cordery, R. J., Hall, M., Cipolotti, L., Davidson, L., Adlard, P., & Rossor, M. N. (2003). Early cognitive decline in Creutzfeldt-Jakob disease associated with human growth hormone treatment. *JOURNAL OF Neurology. Neurosurgery and Psychiatry*, *10*(74), 1412–1416. doi:10.1136/jnnp.74.10.1412

Creutzfeldt, H. G. (2002). Clinical diagnosis and differential diagnosis of CJD and vCJD With special emphasis on laboratory tests. *Journal of Neurology, Neurosurgery and Psychiatry*, 1(3), 88–98.

Currais, A., Fischer, W., Maher, P., & Schubert, D. (2017). Intraneuronal protein aggregation as a trigger for inflammation and neurodegeneration in the aging brain. *The FASEB Journal*, *31*(1), 5–10. doi:10.1096/fj.201601184 PMID:28049155

Fania, C., Arosio, B., Capitanio, D., Torretta, E., Gussago, C., Ferri, E., & Gelfi, C. (2017). Protein signature in cerebrospinal fluid and serum of Alzheimer's disease patients : The case of apolipoprotein A-1 proteoforms. *PLoS One*, *12*(6), 1–19. doi:10.1371/journal.pone.0179280 PMID:28628634

Food and Drug Administration. (2002). Revised preventive measures to reduce the possible risk of transmission of Creutzfeldt-Jakob (CJD) disease and variant Creutzfeldt-Jakob disease (vCJD) by blood and blood products. FDA.

Friedman-Levi, Y., Meiner, Z., Canello, T., Frid, K., Kovacs, G. G., Budka, H., & Gabizon, R. (2011). Fatal Prion Disease in a Mouse Model of Genetic E200K Creutzfeldt-Jakob Disease. *PLoS Pathogens*, 7(11), e1002350. doi:10.1371/journal.ppat.1002350 PMID:22072968

Garske, T., & Ghani, A. C. (2010). Uncertainty in the tail of the variant Creutzfeldt-Jakob disease epidemic in the UK. *PLoS One*, *5*(12), e15626. doi:10.1371/journal.pone.0015626 PMID:21203419

Gauczynski, S., Nikles, D., El-Gogo, S., Papy-Garcia, D., Rey, C., Alban, S., & Weiss, S. (2006). The 37-kDa/67-kDa laminin receptor acts as a receptor for infectious prions and is inhibited by polysulfated glycanes. *The Journal of Infectious Diseases*, *194*(5), 702–709. doi:10.1086/505914 PMID:16897671

Geschwind, M. D., Kuo, A. L., Wong, K. S., Haman, A., Devereux, G., Raudabaugh, B. J., & Thai, J. N. (2013). Quinacrine treatment trial for sporadic Creutzfeldt-Jakob disease. *Neurology*, *81*(23), 2015–2023. doi:10.1212/WNL.0b013e3182a9f3b4 PMID:24122181

Gomori, A. J., Partnow, M. J., Horoupian, D. S., & Hirano, A. (1973). The ataxic form of Creutzfeldt-Jakob disease. *Archives of Neurology*, *29*(5), 318–323. doi:10.1001/archneur.1973.00490290058006 PMID:4582819

Griffith, J. S. (1967). Self-replication and scrapie. *Nature*, 215(5105), 1043–1044. doi:10.1038/2151043a0 PMID:4964084

Halasi, M., Váraljai, R., Benevolenskaya, E., & Gartel, A. L. (2016). A Novel Function of Molecular Chaperone HSP70 suppression of oncogenic foxm1 after proteotoxic stress. *The Journal of Biological Chemistry*, 291(1), 142–148. doi:10.1074/jbc.M115.678227 PMID:26559972

Harris, D. (2017). Study unravels mystery of how nerve cells are damaged in neurodegenerative diseases. *Medical and Life Sciences*, 1–2.

Creutzfeldt-Jakob Disease

Hartl, F. U., Bracher, A., & Hayer-Hartl, M. (2011). Molecular chaperones in protein folding and proteostasis. *Nature*, 475(7356), 324–332. doi:10.1038/nature10317 PMID:21776078

Heinemann, U., Krasnianski, A., Meissner, B., Kallenberg, K., Kretzschmar, H. A., Schulz-Schaeffer, W., & Zerr, I. (2008). Brain biopsy in patients with suspected Creutzfeldt-Jakob disease. *Journal of Neurosurgery*, *109*(4), 735–741. doi:10.3171/JNS/2008/109/10/0735 PMID:18826363

Hill, A. F., Joiner, S., Wadsworth, J. D., Sidle, K. C., Bell, J. E., Budka, H., & Collinge, J. (2003). Molecular classification of sporadic Creutzfeldt–Jakob disease. *Brain*, *126*(6), 1333–1346. doi:10.1093/ brain/awg125 PMID:12764055

Holman, R. C., Belay, E. D., Christensen, K. Y., Maddox, R. A., Minino, A. M., Folkema, A. M., & Schonberger, L. B. (2010). Human Prion Diseases in the United States. *PLoS One*, *5*(1), 1–8. doi:10.1371/journal.pone.0008521 PMID:20049325

Jackson, G. S., & Collinge, J. (2001). The molecular pathology of CJD: Old and new variants. *Molecular Pathology*, *54*(6), 393. PMID:11724914

Jackson, W. S., & Krost, C. (2014). Peculiarities of Prion Diseases. *PLoS Pathogens*, *10*(11), e1004451. doi:10.1371/journal.ppat.1004451 PMID:25411777

Jakhar, R., Paul, S., Bhardwaj, M., & Kang, S. C. (2017). Astemizole–Histamine induces Beclin-1independent autophagy by targeting p53-dependent crosstalk between autophagy and apoptosis. *Cancer Letters*, *372*(1), 89–100. doi:10.1016/j.canlet.2015.12.024 PMID:26739061

Karapetyan, Y. E., Sferrazza, G. F., Zhou, M., Ottenberg, G., Spicer, T., Chase, P., & Lasmezas, C. I. (2013). Unique drug screening approach for prion diseases identifies tacrolimus and astemizole as antiprion agents. *Proceedings of the National Academy of Sciences of the United States of America*, *110*(17), 7044–7049. doi:10.1073/pnas.1303510110 PMID:23576755

Kim, Y. E., Hipp, M. S., Bracher, A., Hayer-Hartl, M., & Ulrich Hartl, F. (2013). Molecular chaperone functions in protein folding and proteostasis. *Annual Review of Biochemistry*, 82(1), 323–355. doi:10.1146/ annurev-biochem-060208-092442 PMID:23746257

Kovacs, G. G., & Budka, H. (2008). Prion diseases: From protein to cell pathology. *American Journal of Pathology*, *172*(3), 555–565. doi:10.2353/ajpath.2008.070442 PMID:18245809

Lang, C. J., Heckmann, J. G., & Neundörfer, B. (1998). Creutzfeldt-Jakob disease via dural and corneal transplants. *Journal of the Neurological Sciences*, *160*(2), 128–139. doi:10.1016/S0022-510X(98)00226-3 PMID:9849795

Lolekha, P., Rasheed, A., & Yotsarawat, C. (2015). *Creutzfeldt-Jakob Disease in a Tertiary Care Hospital in Thailand: A Case Series and Review of the Literature*. Academic Press.

Maheshwari, A., Fischer, M., Gambetti, P., Parker, A., Ram, A., Soto, C., & Mead, S. (2015). Recent US case of variant Creutzfeldt-Jakob disease—global implications. *Emerging Infectious Diseases*, 21(5), 750–759. doi:10.3201/eid2105.142017 PMID:25897712

Mastrianni, J. A. (2010). The genetics of prion diseases. *Genetics in Medicine*, 12(4), 187–195. doi:10.1097/GIM.0b013e3181cd7374 PMID:20216075

Mead, S., Mahal, S. P., Beck, J., Campbell, T., Farrall, M., Fisher, E., & Collinge, J. (2001). Sporadic but not variant—Creutzfeldt-Jakob disease is associated with polymorphisms upstream of PRNP exon 1. *American Journal of Human Genetics*, *69*(6), 1225–1235. doi:10.1086/324710 PMID:11704923

Mead, S., Mahal, S. P., Beck, J., Campbell, T., Farrall, M., Fisher, E., & Collinge, J. (2001). Sporadic but not variant—Creutzfeldt-Jakob disease is associated with polymorphisms upstream of PRNP exon 1. *American Journal of Human Genetics*, *69*(6), 1225–1235. doi:10.1086/324710 PMID:11704923

Mead, S., Poulter, M., Uphill, J., Beck, J., Webb, T., Campbell, T., & Collinge, J. (2013). Identifies Genetic Risk Factors For Plaques In Variant Creutzfeldt-Jakob Plaques In Sporadic Creutzfeldt-Jakob. *Neurology*, *2*(4), 127–1135.

Mercado, G., Castillo, V., Vidal, R., & Hetz, C. (2015). ER proteostasis disturbances in Parkinson's disease : Novel insights. *Frontiers in Aging Neuroscience*, 7(March), 1–5. PMID:25870559

Montie, H. L., & Durcan, T. M. (2013). The cell and molecular biology of neurodegenerative diseases : An overview. *Frontiers in Neurology*, 4(November), 1–2. PMID:24348458

Morgan, C., Gupta, M., El-Feky, W., Shamim, S., & Opatowsky, M. (2009). Creutzfeldt-Jakob disease: Case discussion and imaging review. *Proceedings - Baylor University. Medical Center*, 22(1), 69–71. doi:10.1080/08998280.2009.11928476 PMID:19169404

Murray, R. K., & Davis, J. C. (2003). Harper's Illustrated Biochemistry. New York: McGraw-Hill Medical.

Nuvolone, M., Schmid, N., Miele, G., Sorce, S., Moos, R., Schori, C., & Aguzzi, A. (2017). Cystatin F is a biomarker of prion pathogenesis in mice. *PLoS One*, *1*(10), 1–25. PMID:28178353

Panegyres, P. K., & Armari, E. (2013). Therapies for human prion diseases. *American Journal of Neurodegenerative Disease*, 2(3), 176. PMID:24093082

Pocchiari, M., & Ladogana, A. (2015). Rethinking of doxycycline therapy in Creutzfeldt-Jakob disease. *JOURNAL OF Neurology. Neurosurgery and Psychiatry*, 0(0), 2014–2015.

Ponte, M. L. (2006). Insights into the Management of Emerging Infections : Regulating Variant Creutzfeldt-Jakob Disease Transfusion Risk in the UK and the US. Academic Press.

Poser, S., Zerr, I., Schroeter, A., Otto, M., Giese, A., Steinhoff, B. J., & Kretzschmar, H. A. (2000). Clinical and differential diagnosis of Creutzfeldt-Jakob disease. *Archives of Virology. Supplementum*, 2(16), 153–159. PMID:11214918

Riesner, D. (2003). Biochemistry and structure of PrPC and PrPSc. *British Medical Bulletin*, 66(1), 21–33. doi:10.1093/bmb/66.1.21 PMID:14522846

Rosenbloom, M. H., Tartaglia, M. C., Forner, S. A., Wong, K. K., Kuo, A., Johnson, D. Y., & Geschwind, M. D. (2015). Metabolic disorders with clinical and radiologic features of sporadic Creutzfeldt-Jakob disease. *Neurology. Clinical Practice*, *5*(2), 108–115. doi:10.1212/CPJ.000000000000114 PMID:26137419

Creutzfeldt-Jakob Disease

Rubenstein, R., Petersen, R. B., & Wisniewski, T. (2016). Diagnosis of Prion Diseases. In *Manual of Molecular and Clinical Laboratory Immunology* (8th ed.; pp. 682–702). American Society of Microbiology.

Safar, J. G., Geschwind, M. D., Deering, C., Didorenko, S., Sattavat, M., Sanchez, H., & Miller, B. L. (2005). Diagnosis of human prion disease. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(9), 3501–3506. doi:10.1073/pnas.0409651102 PMID:15741275

Soto, C. (2011). Prion hypothesis: The end of the controversy? *Trends in Biochemical Sciences*, *36*(3), 151–158. doi:10.1016/j.tibs.2010.11.001 PMID:21130657

Soto, C., & Castilla, J. (2004). The controversial protein-only hypothesis of prion propagation. *Nature Medicine*, *10*(July), 63–67. doi:10.1038/nm1069 PMID:15272271

States, U., Gambetti, P., Hunter, S., Maddox, R. A., Crockett, L., Zaki, S. R., & Schonberger, L. B. (2005). Variant Creutzfeldt-Jakob Disease. *Centres for Disease Control and Prevention*, *11*(9), 9–12.

Stefani, M. (2004). Protein misfolding and aggregation: New examples in medicine and biology of the dark side of the protein world. *Biochimica et Biophysica Acta (BBA)- Molecular Basis of Disease*, *1739*(1), 5–25. doi:10.1016/j.bbadis.2004.08.004 PMID:15607113

Trevitt, C. R., & Singh, P. N. (2003). Variant Creutzfeldt-Jakob disease : Pathology epidemiology, and public health implications 1 – 4. *The American Journal of Clinical Nutrition*, *1986*(3), 651–656. doi:10.1093/ajcn/78.3.651S PMID:12600856

Valastyan, J. S., & Lindquist, S. (2014). Mechanisms of protein-folding diseases at a glance. *Disease Models & Mechanisms*, 7(1), 9–14. doi:10.1242/dmm.013474 PMID:24396149

Van Everbroeck, B., Dobbeleir, I., De Waele, M., De Deyn, P., Martin, J.-J., & Cras, P. (2004). Differential diagnosis of 201 possible Creutzfeldt-Jakob disease patients. *Journal of Neurology*, 251(3), 298–304. doi:10.100700415-004-0311-9 PMID:15015009

Vetrugno, V., Puopolo, M., Cardone, F., Capozzoli, F., Ladogana, A., & Pocchiari, M. (2015). The future for treating Creutzfeldt–Jakob disease. *Expert Opinion on Orphan Drugs*, *3*(1), 57–74. doi:10.1517/21 678707.2015.994605

Voisset, C., Saupe, S. J., Galons, H., & Blondel, M. (2009). Procedure for identification and characterization of drugs efficient against mammalian prion: From a yeast-based antiprion drug screening assay to in vivo mouse models. *Infectious Disorders Drug Targets*, 9(1), 31–39. doi:10.2174/1871526510909010031 PMID:19200013

Whalley, K. (2017). Neurodegenerative disease: Probing prions. *Nature Reviews. Neuroscience*, *18*(1), 5–5. PMID:27974838

Wilesmith, J. W., Wells, G. A., Cranwell, M. P., & Ryan, J. B. (1988). Bovine spongiform encephalopathy: Epidemiological studies. *The Veterinary Record*, *123*(25), 638–644. PMID:3218047

World Health Organization. (2003). WHO manual for surveillance of human transmissible spongiform encephalopathies, including variant Creutzfeldt-Jakob disease. WHO.

World Health Organization. (2008). International Health Regulations (2005). World Health Organization.

World Health Organization. (2010). WHO Tables on tissue infectivity distribution in transmissible spongiform encephalopathies. Updated 2010. World Health Organization.

Zanusso, G., Monaco, S., Pocchiari, M., & Caughey, B. (2016). Advanced tests for early and accurate diagnosis of Creutzfeldt-Jakob disease. *Nature Reviews. Neurology*, *12*(6), 325–333. doi:10.1038/nr-neurol.2016.65 PMID:27174240

Zerr, I., Bodemer, M., Gefeller, O., Otto, M., Poser, S., Wiltfang, J., & Weber, T. (1998). Detection of 14-3-3 protein in the cerebrospinal fluid supports the diagnosis of Creutzfeldt-Jakob disease. *Annals of Neurology*, *43*(1), 32–40. doi:10.1002/ana.410430109 PMID:9450766

Zuber, C., Ludewigs, H., & Weiss, S. (2007). Therapeutic approaches targeting the prion receptor LRP/ LR. *Veterinary Microbiology*, *123*(4), 387–393. doi:10.1016/j.vetmic.2007.04.005 PMID:17498894

Chapter 18 Childhood Neurodegenerative Disorders

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ABSTRACT

Neurodegenerative disorders (NDs) are a group of disorders with the deterioration of attained skills often with no solutions, usually ending with death or crippling disabilities. This chapter contains a classification for childhood NDs as well as an algorithmic approach for easy management of these disorders. Genetic defects and pathophysiology of disorders like Canavan, Krabbe, subacute sclerosing panencephalitis (SSPE), etc. are written in detail. Suggestions regarding management of some of these conditions are described as a lifecycle approach, from birth to death, to enable those who are taking care of such kids. Allogeneic hematopoietic stem cell transplantation (HSCT), gene therapy, combination therapy, and other experimental therapies have enlarged the scope of diagnosis and treatment options for these disorders. The author aims to brush up the existing and latest possibilities in NDs, including those at the experimental stage, for an easy understanding and for further research, especially as treatment options.

INTRODUCTION

Childhood NDs are neglected group of central nervous system (CNS) disorders by clinicians, as much of these disorders have no answers and hence any investigations to find out cause of a developmental problem due to NDs are mainly of academic interest. Fresh interest among clinicians in this is kindled by recent advances in molecular diagnosis and neurosciences, in addition to newer modalities on treatment. Childhood NDs are mainly genetic mutations resulting in either enzyme deficiencies or formation of abnormal products in nervous system (Marder, K et al,1998). Most of the disorders, where definitive mutations and abnormal products have been identified in CNS, have significant variations in their clinical presentations. The knowledge about the cause of this individual variations is however limited. Individuals who carry the same mutation in the same disease-causing gene may display a range of different clinical symptoms (Kwon, 2016). This chapter gives a bird's eye view of the common NDs in children and tries

DOI: 10.4018/978-1-5225-5282-6.ch018

to bridge the gap between research and possible application of the knowledge in real life. It shall be the authors endeavor to put in approaches where ever such new knowledge can be inserted to find new ways of analyzing these disorders clinically. In this process author tries to bring in an algorithmic model utilizing the existing and new knowledge to find easy clinical approaches that also serves as a clinical classification. Where ever possible the author puts in life cycle approach, i.e., birth to death, to solve the difficulties these children undergo. Recent advances and possible areas of new researches are also added in this chapter, along with each disorder. The main objective of this chapter is to consolidate the existing knowledge from diverse sources to help in easy understanding to facilitate clinical management and bring out to fine tune the ideas and possibilities from the new options emerging out of researches.

BACKGROUND

NDs are a group of disorders often associated with constellations of findings suggesting loss of acquired skills, loss of memory; impairment of intellect; personality and behavioral changes, in addition to definite neurological signs like vision and hearing loss, changes in tone, seizures etc. However quite many NDs are due to identifiable causes like genetic, biochemical defects, chronic viral infections. With the advent of better imaging techniques and biochemical and molecular markers, a specific diagnosis is possible in many case; but thorough history and clinical examination still give chances to suggest a possible diagnosis (Kwon, 2016).

Most of the NDs of childhood are neurometabolic disorders. Often multisystem involvement with CNS signs and regression of attained milestones points to a neurometabolic disorder affecting the CNS. The age of onset, progression of the disease process and primary finding helps one to determine whether we are dealing with white matter or grey matter disorders (GM) (Kwon, 2016). Increasing spasticity and Upper Motor Neuron signs is a pointer towards white matter (WM) disease while seizures, visual impairment, loss of memory and executive functions are pointers to grey matter injury. This over simplification cannot explain the reason behind behavioral changes occurring in WM lesions. In fact, WM lesion often results in neurobehavioral disconnection syndromes due to disruption in nerve bundle connecting 2 cortical areas (Schwachman et al, 2008).

With the differentiation between presentation of GM and WM disorders decreasing and newer magnetic resonance imaging (MRI) evidences blurring these distinctions, a newer method of classification based on abnormal molecules or mutations is warranted. Even when options are limited, work up for a diagnosis is still needed for the promotion of preventive care in subsequent pregnancies. Advances in enzyme therapies can be of use only if diagnosed at an early age. This scenario necessitates the consolidation of emerging knowledge at different domains, including animal research, and linking it with existing ones.

CLASSIFICATION OF CHILDHOOD NEURODEGENERATIVE DISORDERS

The classification offered in some textbooks based on site involved is too simplistic and often confusing as mixed presentations are seen (Shu et al, 2017).

Clinical Classification

Another way of classification suggested is the clinically useful algorithmic way based on age of presentation and clinical features:

NDs With Regression Below 2 Years

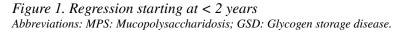
A simple approach is to look out for hepatomegaly and presence of extra neural manifestations (Kwon, 2016) (Figure 1 and 2).

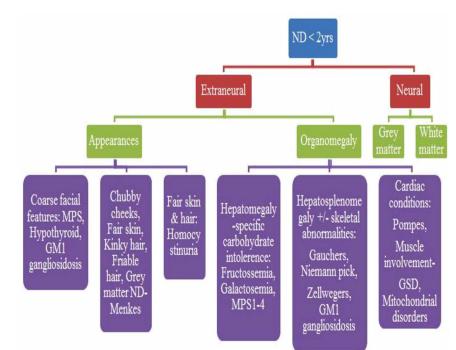
Regression in Children Between 2-5 Years

Progressive neurological deterioration between 2 and 5 years may be classified as follows given in Figure 3. These may be addressed by having a clear look at history and clinical examination and grouping accordingly.

Regression in Children Between 5-18 Years

History and clinical examination gives clear clues to classify the disorders in this age group. They may be subdivided into predominantly white matter, grey matter, extrapyramidal or ataxic group of disorders. Disorders like SSPE usually make their appearance in this age group (Figure 4).





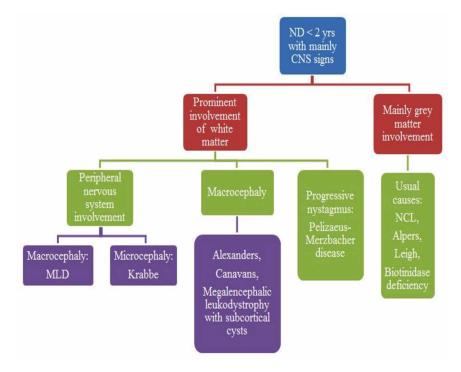
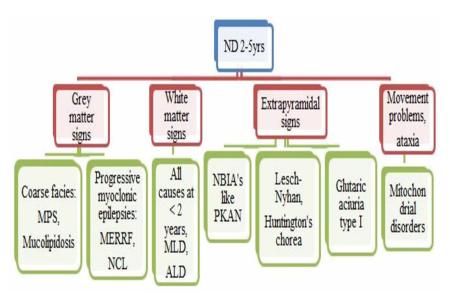


Figure 2. Regression starting at < 2 years with mainly neuronal involvement Abbreviations: NCL: Neuronal ceroid lipofuscinosis; MLD: Metachromatic leukodystrophy.

Figure 3. Regression starting between 2-5 years old

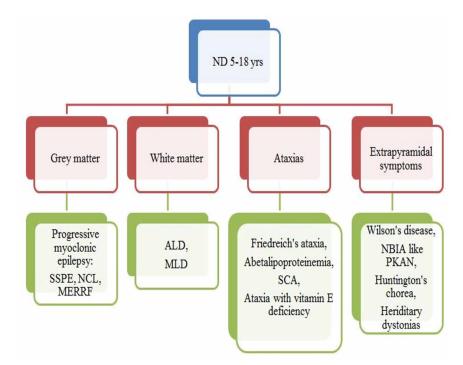
Abbreviations: MPS: Mucopolysaccharidosis; PKAN: Pantothenate kinase associated neurodegeneration; NBIA: Neurodegeneration with brain iron accumulation; ALD: Adrenoleukodystrophy; MLD: Metachromatic leukodystrophy; NCL: Neuronal ceroid lipofuscinosis; MERRF: Mitochondrial ragged red fibers.



Childhood Neurodegenerative Disorders

Figure 4. Neuroregression starting between 5-18 years

Abbreviations: PKAN: Pantothenate kinase associated neurodegeneration; NBIA: Neurodegeneration with brain iron accumulation; SCA: Spinocerebellar ataxia; ALD: Adrenoleukodystrophy; MLD: Metachromatic leukodystrophy; SSPE: Subacute sclerosing pan encephalitis; NCL: Neuronal ceroid lipofuscinosis; MERRF: Mitochondrial ragged red fibers.



NEURODEGENERATIVE DISORDERS OF CHILDHOOD

Canavan's Disease

This autosomal recessive (AR) disorder was first described by Myrtelle Canavan (1879-1953), an American neuropathologist, in her 1931 seminal paper (Canavan, 1931). Though Canavan's disease (CD) or cerebral spongy degeneration is seen pan ethnically (Zeng et al, 2002), it is most often noted in Ashkenazi Jews. The incidence of the disease is 1: 6,400 if both parents are carriers and 1: 13,500 if only one parent is carrier with no sexual predilection. Non-Jew's are mostly Europeans, Turks, African-Americans, Middle East people and Japanese. In these non-Jew population, the incidence is 1:1,00,000 (Makalon et al, 2015).

High quantities of N-acetyl-L-aspartic acid (NAA) produced and stored by the neurons are pushed into the extracellular fluid on its depolarization. NAA is further degraded into acetate and aspartate by asparto-acylase (ASPA) present in oligodendrocytes. Mutations in the *ASPA* gene in17p13.2 reduce the function of ASPA (Baslow et al, 2015). Since *ASPA* gene expresses in oligodendrocytes in the brain, this mutation results in elevated levels of NAA in the brain and cerebrospinal fluid (CSF). Over 40 mutations have been identified, main ones are listed in Table 1.

Spongiform changes occur in cerebrum, cerebellum and brain stem with loss of demarcation between cortical white and grey matter and multiple vacuoles throughout the white matter (WM). There is ac-

Mutation	Population	Percentage	Reference
E285A; Y231X	Jews	98	Makalon et al, 2015
P.Ala.305Glu	Non-Jews	57	Feigenbaum et al,2004; Marlon,2015
A305E missense mutation	Western European	48	
32deltaT	African American		
876delta AGAA	England		
D114E, R168 C	Turkey		
116T	Netherlands		Kaul et al, 1996
G27	Germany		
C152Y	Ireland]	
G123E	Canada		

Table 1. Mutations in Canavan's disease

cumulation of extracellular fluid between myelin lamellae of oligodendrocytes and astrocytes (Gambetti et al,1969). There is subcortical spongy WM degeneration with swelling and abnormal astrocyte mitochondria (Matalon et al,1988). NAA produced by neurons in high concentration increases near axons and may cause myelin injury by osmotic (Leone et al, 2012) and metabolic effects. This results in intact cell bodies of nerve cells but degeneration of myelin sheath.

Clinically, they are normal till 3-6 months when symptoms like floppiness (hypotonia), developmental delay, macrocephaly (91%) develop with mental changes like apathy, irritability or lethargy. Some, especially congenital form, have dysphagia, with delay in GM mile stones and progressive psychomotor regression. In addition, they may have seizures, sleep disorders, feeding difficulties, nasal regurgitation and reflux with vomiting and optic atrophy causing reduced visual responsiveness (Rodriguez, 2013). Eventually spasticity and dystonia develop and end up in decerebrate rigidity with admixture of different seizure types.

Alexander's Disease

The first recognized patient with Alexander's disease (AD) was a 16-month-old child with megalencephaly, hydrocephalus and developmental regression by Alexander (Alexander,1949). In a survey conducted in Japan, an incidence of 1:2.7 million was noted (Yoshida et al, 2011). Conventionally AD was classified based on age of onset into Infantile (0-2years), juvenile (3-11years) and adult (>12 years). Currently it is classified as 2 forms-those which begin before 4 years- (type1) and those that can present at any age but mainly after 4 years (type 2) (Prust et al, 2011). 42% of AD is infantile form with Glial fibrillary acidic protein (GFAP) mutation, 22% are juvenile forms and the adult form accounts for 33% (Srivastava & Naidu, 2002).

AD is an autosomal dominant leukodystrophy resulting from a mutation in the gene for glial fibrillary acidic protein (GFAP) in chromosome17 q 21.31. 95% of the patients have mutations for *GFAP* gene (Brenner et al, 2009). Mutations are mostly de-novo without any gender bias and occur in type I; but in late onset AD, often autosomal dominant inheritance is seen with penetrance of nearly100%. All the mutations are located in the rod domain of *GFAP* and the clinical severity correlated well with the affected amino acids (Rodriguez et al, 2001). GFAP gene expresses mainly on astrocytes and hence AD is an astrocytopathy. Pathologically, abundant astrocytic accumulations of eosinophilic cytoplasmic inclusions, recognized by neuropathologists as Rosenthal fibers (after the 19th century German pathologist who first described them in the context of an old astrocyte scar; Rosenthal, 1898) accumulate in cerebral cortex, in the white WM of the brain, brainstem and the spinal cord (Wippold et al, 2006). Abnormal GFAP appear mainly under the meninges (pia mater), sub-ependymal in ventricles, and in the perivascular regions. It is the overexpression of GFAP mutant gene that causes formation of these inclusion bodies in astrocytes (Li et al, 2002). A higher level of mutant proteins requires higher expression of GFAP; hence Rosenthal fibers are predominantly seen in sub-pial and white matter CNS regions, which are areas of high GFAP content, when compared with gray matter areas. AD astrocytes have excessive amounts of the small heat shock protein, α B-crystallin indicating role of stress (Tang et al, 2010). Accumulation of abnormal GFAP oligomers appears to cause of dysfunctional proteasomes resulting in an inability to break up substrates facilitating Rosenthal body formation. Another feature noticed in AD astrocyte is the progressively decreasing levels of GLT-1, the major glutamate transporter in astrocytes, exposing the cells to damage due to excitotoxic cell death (Tian et al, 2010). This explains the seizures seen in type I AD, where the major inhibitor viz, Glutamate action is less making grey matter neurons more excitable. However, this role is not seen in white matter where glutaminergic transmission has lesser role. Here the over expression of GFPA in axons and myelin and astrocytes of white matter causes clinical signs due to white matter diseases in late onset AD (Tian et al, 2010).

Clinically, infantile form (type 1) is typically spastic with developmental delay, seizures, megalencephaly and intellectual disability. Common problems in juvenile and adult forms (type 2) of AD include seizures, bulbar/pseudobulbar signs (speech abnormalities, swallowing difficulties -50%), ataxia (75%) and about 33% spasticity, delay or regression of development, macrocephaly, autonomic dysfunction, sleep apnea and psychomotor decline (Springer et al, 2000). Rare forms may present in the neonatal period with developmental delay, hydrocephalus, and seizures (Springer et al, 2000). Sometimes significant accumulation of altered GFAP sub-ependymally results in obstructive hydrocephalus (Prust et al, 2011).

Krabbe Disease

Krabbe disease (KD) or globoid cell leukodystrophy, named after Knud Haraldsen Krabbe, a Danish neurologist (1885-1961), is a lysosomal storage disorder. This affects both the CNS and peripheral nervous system. The approximate incidence of AD is 1 in 1,00,000–2,50,000 individuals worldwide (Haneef & Doss, 2016). KD is an AR disorder due to mutations in chromosome 14q24.3-q32.1 resulting in deficiency of β -galactocerebrosidase (GALC) leading to accumulation of galactocerebroside (galactosylcreamide) and psychosine (galactosyl sphingosine) in oligodendroglial cells. Though elevation of galactosyl ceramide is not detrimental in KD; the byproduct of galactosyl-ceramide metabolism, psychosine, gets accumulated resulting in extensive damage to oligodendroglia leading to demyelination (Kohlschütter, 2013). The architecture of regions in cell membrane called lipid rafts, which contain high concentration of sphingolipids and cholesterol, gets disrupted by psychosine resulting in damage to oligodendrocytes and schwann cells (White, 2009). More than 145 GALC mutations have been known to cause KD; but it is difficult to predict the age of onset based on mutation analysis alone (Wenger et al, 2011). These mutations result in abnormal enzyme folding in endoplasmic epithelium resulting in loss of functions of enzyme in lysosomes (Samantha et al, 2016).

Depending on age of onset of clinical manifestation, they may be classified as 4 subtypes- type-1(infantile onset), type 2 (late infantile onset), type 3 (juvenile onset), type 4 (adult onset) (Suzuki, 2009). Born normal they develop symptoms as early as 6 months like unexplained cry, irritability, vomiting, feeding difficulty and hyperpyrexia prompting many to change the feeds or treat as colic. Early seizures and hypersensitivity to stimuli including sound develops (mimicking Taysach's disease). Soon rigidity, opisthotonus spasms and ataxia sets in. Optic atrophy leading to blindness, sensorineural deafness, absent Deep Tendon Reflexes along with decerebrate rigidity appears later. Late onset (type 2to 4) presents as optic atrophy and cortical blindness with both spasticity and ataxia resulting in slowly progressive gait disturbances; hence confused with Adrenoleuko**d**ystrophy. They may present as spastic paraparesis or with impaired control of voluntary movements, vision loss and poly neuropathy. Childhood onset and adolescent onset KD is more likely to have optic atrophy (54, 33% respectively) and cerebellar ataxia (57 and 50%) (Rabab et al, 2013)

Metachromatic Leukodystrophy

Metachromatic leukodystrophy (MLD) is due to arylsulphatase A deficiency (ARSA) in the lysosomes, which is inherited as an AR trait. Incidence is 1 in 40,000-160,000 (Wang et al, 2011). *ARSA* gene is in chromosome 22q13-13qtr. Cerebroside sulfate accumulates in the neurons- both CNS and peripheral nervous system (PNS), due to partial or complete deficiency of ARSA enzyme leading to demyelination. The name MLD is given because of the peculiar appearance of the cells with an accumulation of galactosyl-3-sulfate ceramide, when stained with crystal violet produces granules known as metachromatic staining, since they stain differently from their surrounding cellular material as golden brown (Gomez-Ospina, N.,2017).

Based on when it presents, MLD is classified as late infantile, juvenile and adult. 50 to 60% of all individuals with MLD are infantile form presenting at 2nd year of life, usually beginning as gait disturbances between 1-2 years of age with hypotonia, weak deep tendon reflexes, clumsy gait and frequent falls (Fluharty, 2014). Intellectual deterioration, dysarthria, diminished visual fixations and nystagmus with optic atrophy, initial hypotonia with rigidity and finally decorticates posturing sets in. Feeding difficulties due to pseudobulbar palsy, peripheral neuropathy and death happens by 6 years due to aspiration pneumonia or bronchopneumonia. Both the lower and upper motor neuron involvement are seen in patients with MLD; hence absence or exaggeration of DTR should not deter one from making the diagnosis.20-30% are juvenile forms of MLD presenting between 4-10 years as behavioral issues or deterioration in scholastic performance, gait incoordination, urinary incontinence, hypertonia, ataxia, dysarthria and dystonia/tremors with difficult to control generalized tonic-clonic seizures (GTCS).

Adrenoleukodystrophy

This rapidly progressive X-linked ND affecting the white matter of children, also known as Schilder's disease, was first recognized in 1923. This disease affecting CNS, adrenals and testicles affects almost all races with an incidence of 1:20000 (Kwon, 2011). Being XLR disorder it affects boys before 10 years of age and girls are carriers. Mean age of appearance is around 7 years at which a previously normally developing male child develops signs of white matter involvement.

X-linked adrenoleukodystrophy (ALD) results in impaired peroxisomal β -oxidation of very longchain fatty acids (VLCFA), which is reduced to about 30% of control levels due to a mutation of ABCD1 gene located in X chromosome-Xq28. Mutation causes absence or deranged function of ALD protein (a peroxisomal transmembrane protein transporting VLC acyl-co-esters from cytosol to peroxisomes. These VLCF acyl-CoA esters are incorporated into different lipid fractions and act as substrate for further elongation to even longer fatty acids (Ofman et al, 2010). Very long-chain fatty acids (VLCFA) inside cells results in oxidative stress and oxidative damage to proteins, microglial activation and apoptosis (Fourcade et l, 2008). This may lead to destabilization of the myelin sheath and impairment of function of astrocytes and microglia which helps in myelin integrity. Not all males with X-ALD develop cerebral ALD, pointing to the fact that other factors like genetic, epigenetic and/or environmental may modify this process. ABCD1 mutation, resulting in less/no production of ALDP1 protein. Though plasma levels of VLCFA are elevated, no correlation between neurological phenotypes, blood levels of VLCFA or biochemical defects and type of mutation exists. The severity of disease correlates with the level of perivascular infiltration and inflammation. In about 80% of children with ALD, there will be demyelination of splenium of corpus callosum. Lesions may also be seen involving pyramidal tracts including inside pons and internal capsule (van der Knaap et al, 2005).

The presentation may vary significantly even with in the family with one male having the childhood form and his brother manifesting as adult form. The 7 phenotypes identified are the childhood cerebral form, adrenomyeloneuropathy (AMN), adult cerebral, adolescent, adrenal insufficiency without neurologic disease, asymptomatic, and heterozygotes. The childhood form mainly manifests as behavioral issues like inattention, hyperactivity, emotional labiality and school difficulties in a child who was normal till at least 3-4 years. Gum and mucosal hyperpigmentation often precedes hyperpigmentation of skin. Its progression into seizures, visual symptoms, auditory processing difficulties (auditory discrimination) and motor in-coordination proceeds rapidly and in 2 years' time they become vegetative. The difficulty with auditory discrimination results in difficulty in using telephones and low scores in verbal part of intelligence quotient tests. Adolescent form of ALD manifests around 10 years with slow disease progression and may have status epilepticus.

Lafora Disease

This fatal progressive myoclonic epileptic disorder was first described by Lafora and Glueck (1911). Prevalence of Lafora disease (LD) is 1-9/10,00,000 with AR inheritance manifesting towards adolescence. *EPM2A* mutations in 6q24.2, which codes for Laforin- a dual-specificity phosphatase or *NHLRC1* gene in 6p22.3 coding for an E3 ubiquitin ligase called Malin, together results in production of glycogen (Spuch et al, 2012). This glycogen accumulates inside the cytoplasm producing "lafora bodies" which is damaging to the neurons. For the functional activation of Malin in endoplasmic reticulum of neurons, Laforin is required. Gentry found that Malin binds and interacts with the laforin protein in HEK293T cells in vivo (Gentry et al, 2005). This results in Laforin getting polyubiquitinated in a Malin-dependent manner leading to Laforin degradation. Laforin polyubiquitination and degradation gets abolished due to mutations in the *NHLRC1* gene. This led them to conclude that malin regulates laforin protein concentrations and that mutation in the *NHLRC1* gene results in reduced E3 ligase activity of malin leading to Lafora disease (Gentry et al, 2005). Disease progression is slower in *NHLRC1* mutations and hence

they live longer. Lafora bodies (LBs) develop in the cytoplasm of cells from brain, kidney, skin, liver, and cardiac and skeletal muscle. LBs are composed of α 1,4-glycosidic linkages between glucose residues with α 1,6 branches, but with lesser number of branches (Worby et al, 2008). Polyglucosans, due to unknown reasons, accumulate in dendrites, initiating severe progressive myoclonus epilepsy after a threshold amount of accumulation is reached (Turnbull et al, 2011).

They are normal during their early childhood with few having minor learning problems. The earliest manifestation in children are- a decrease in scholastic performance with headaches and seizures (Minassian et al, 2001). Seizures in LD may be symmetric or generalized myoclonus or GTCS, occipital seizures resulting in visual hallucinations; progressive neurodegeneration results in deteriorated cognition and/ or behavior, dysarthria and ataxia (Monaghan & Delanty, 2010). Over time they become intractable and recurrent may present as Status epilepticus. Though dysarthria and ataxia occur early, spasticity appears late. Initially children may have visual hallucinations, depressed mood and cognitive deficits. Later on, increasing hallucination, dementia and agitation along with frontal executive dysfunction and cognitive impairment will interfere with performance ability.

Mucolipidosis

A child looking similar to mucopolysaccharidosis (MPS), but with developmental delay and dysostosis multiplex is usually a mucolipidosis (ML). The name ML was given after their phenotypical resemblance to both mucopolysaccharidosis and sphingolipidosis. Though originally classified as type1 to 4, currently type 1(sialidosis) is a glycoproteinosis and type 4 is under gangliosidosis.

ML III is a rare AR disorder, estimated to occur in about 1 in 100,000 to 400,000 individuals worldwide. Type 2 or I cell disease and type 3 or pseudo hurler polydystrophy are due to the deficiency of N-acetyl glucosamine -1- phosphotransferase in varying grades resulting in appearance similar to Hurlers syndrome. Type 2 is due to a defective phosphotransferase, whereas type 3 is due to partial deficiency (Kudo et al, 2006). Hence clinical features are not recognizable in type III till about 3-5 years, whereas type II may be appreciated early.

Phosphotransferases present in Golgi apparatus transfer phosphates to mannose on specific proteins so that these proteins are marked to be transported to lysosomes. If this doesn't happen then those proteins will be transported out of the cell, similar to the pathway taken by all proteins as they move through Golgi apparatus. Inside lysosomes these proteins actually function as enzymes for breaking down oligosaccharides, lipids, and glycosaminoglycans, failing which the substrates accumulate resulting in inclusion bodies inside lysosomes– hence called inclusion cell diseases (I cell disease)/ MPS-II. Mutations in *GNPTAB* gene (12q23.3) coding for α and β subunits of the N-acetylglucosamine phosphotransferase complex leads to a decreased synthesis of mannose-6-phosphate, which targets the enzymes to the lysosomes of connective tissue cells. This results in decreased functioning of lysosomes rather than the true storage of undigested products. This is the true pathogenesis of ML II (Cathey et al, 2010).

ML II usually presents at birth with hypotonia, intra uterine growth retardation and gradually increasing skeletal deformities like ulnar deviation of hands, broadened and campomelic fingers, kyphosis, dislocation of hip, and deformation of long bones. Range of movement of shoulder joints is limited from birth itself. Some severe children may have neonatal transient hyperparathyroidism. In the neonatal period they have flat facial profile with shallow orbits resulting in proptosis, depressed nasal bridge, prominent mouth and gingival hypertrophy. There is progressive coarsening of facial features with thickening of skin, noticeably around the earlobes. They have delay in early motor as well as in cognitive domains.

Postnatal growth usually ceases at 2nd year of life and contractures in all joints prevent many from walking (Leroy et al, 2012). Most have mitral and aortic valve thickening and regurgitation. Progressive narrowing of airways results in hoarse voice and noisy breathing. Mucosal thickening and stiffening of connective tissues together with cardiac involvement, results in cardiorespiratory failure.

Neuronal Ceroid Lipofuscinosis

Neuronal ceroid lipofuscinosis (NCLs) or Batten disease are a genetically and phenotypically heterogenous group of lysosomal storage disorders associated with progressive visual loss, motor deterioration, dementia, seizures and early mortality. NCLs are the most common hereditary progressive neurodegenerative disease with a prevalence of approximately 1.5 to 9/ million population. The incidence varies from1.3 to 7/100,000 live births in different countries (Mole et al, 2011). Based on age of onset, they are subdivided into- congenital, infantile (age of onset-6-24 month), late infantile (onset-2-8 years), juvenile (4-10 years). There are at least 9 genetically different types. Accumulation of autofluorescent storage material-ceroid and lipofuscin happen in NCL and hence the name. Most are AR except adult onset-caused by mutations in CLN4/DNAJC5 which is AD. The NCL genes codes for proteins in the secretary and or endosomal or lysosomal pathways. The main lysosomal proteins are encoded by CLN1, CLN2, CLN5, CLN10, CLN13 and transmembrane proteins are encoded by CLN3, CLN6, CLN7, MFSD8/, CLN12/ ATP13A2. The transmembrane proteins CLN6 and CLN8 directed to the endoplasmic reticulum (ER); and progranulin, the product of CLN11/GRN, are localized in compartments in the secretory pathway (Ryan et al, 2009).

CLN1 disease presents between 6-24 months with delayed development, myoclonic jerks, and/or seizures, speech problems, loose interest in playing and in toys, but remain interested in their surroundings. Some may have stereotypic hand movements of autistics with moderate motor dysfunction. By 4 years many have retinal blindness. CLN2 disease appears between 2-4 years usually starting with epilepsy. GTCS, clonic, partial and myoclonic seizures are seen. After seizure onset there is regression of cognitive and motor milestones and blindness ensues by 6 years. CLN2 mutations may also produce atypical Juvenile type (Wisniewski, 2017). CLN-3 (Juvenile NCL) has an onset at 5 years as rapidly progressing vision loss as the first clinical sign; it may be the only sign for two to five years. Soon they go into permanent vision loss within 2-5 years on its onset with initial macular changes soon progressing to pan-retinal degeneration with pigmentary changes in the retinal periphery, vascular attenuation and optic nerve pallor. Behavioral problems, extrapyramidal signs, and sleep disturbance occur in second decade. Speech disturbances and slow cognitive decline occur around the time of onset of seizures (Backman, 2005).

Subacute Sclerosing Pan encephalitis

SSPE is a chronic, demyelinating disease affecting brain of children and young adults, caused by a chronic infection with defective measles virus. It is also known as Dawson's encephalitis after Dawson who first described it and suggested a viral etiology (Dawson, 1934). Green field coined the word SSPE. Boutillee et al. (1965) demonstrated in 1965 the presence of measles virus in the brain tissue using electron microscope.

After the introduction of measles vaccination, though the incidence of SSPE in developed countries is negligible, in countries like India despite higher coverage of measles vaccination, it is still the most common cause of ND (Abajirao et al, 2013). Following an infection with Measles, after a latent period

of 6-8 years, progressive neurological deterioration develops. In immunocompetent children it causes SSPE; where as in immunocompromised it causes measles inclusion body encephalitis. Unlike in adult forms, the presentation in childhood is different with personality change and ophthalmic manifestations as common presenting features. Occurrence of SSPE was noticed among vaccinated children. This may be due to previous subclinical infection prior to measles vaccination or poor maintenance of cold chain. Since, in none of the cases vaccine strain has been isolated, SSPE in vaccinated may not be due to vaccine associated SSPE. Wide spread immunization had reduced the incidence of SSPE in developed countries. A recent study in Germany found that the risk of developing SSPE for children infected with measles below 5 years of age as 1:1700 to 1:3300. This risk is in the same order of magnitude as the risk of a fatal acute measles infection (Schönberger et al, 2013). The risk following measles infection under 1 year of age was 18/100 000 compared with 1.1/100 000 after 5 years of age in the UK (Campbell et al, 2007).

Measles demyelinates oligodendrocytes and at later stages of disease develop atrophy of grey and white matter due to infiltration of brain parenchyma with B cells and perivascular cuffing by CD4+ T cells. 2 types of inclusion bodies are seen in nucleus and cytoplasm- Cowdry-A saw in neurons and oligodendroglia and indicate fatal disease. Cowdry-B is small inclusions seen in brain stem. These inclusion bodies contain viral antigens (Gadoth, 2011; Scully, 1969).

Clinical Stages of SSPE

The progression of SSPE and stages was first proposed by Dr. J.T. Jabbour, a neurologist and hence known as Jabbour classification given in Table 2 (Jabbour, 1969).

DIAGNOSIS OF CHILDHOOD NEURODEGENERATIVE DISORDERS

Canavan's Disease

Gas chromatography-mass spectrometry, detects elevated levels of NAA in the urine which is diagnostic (Shoji Tsuji, 2007). CSF shows elevated levels of NAA (Granata, 2012). APSA deficiency can be identi-

Table 2. Jabbour	classification	of subacute	sclerosing pan	encephalitis ((<i>Jabbour</i> , 1969)
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Stages	Jabbour Classification	
Stage1	IA. Behavioral, mental, and personality changes IB. Myoclonic spasms; focal and non-periodic	
Stage 2	IIA. Additional mental deterioration; Myoclonus becomes periodic and generalized, Myoclonus may cause "drop" (falling) attacks when attempting to walk IIB. Apraxia, agnosia, language difficulties, motor dysfunction; ataxia, spasticity, inability to walk unaided, EEG; periodic synchronous discharges coincident with myoclonic spasms	
Stage 3	IIIA. Patient speaks less, vision deterioration, seizures may occur, myoclonic spasms every 3-5s IIIB. Patient is bedridden and has swallowing difficulties, choreiform movements, ballismus (both are involuntary movements), EEG background delta	
Stage 4	Myoclonus stops; akinetic mutism (coma/persistent vegetative state) EEG low voltage without periodic slow wave complexes	

Ophthalmology findings- most have optic atrophy, chorioretinitis and macular pigmentation; in 20% early in the course, papilledema is seen.

Childhood Neurodegenerative Disorders

fied from white blood cells and cultured skin fibroblasts. Preventive measures can be taken by prenatal diagnosis at 16 to18 weeks by measuring amniotic fluid NAA levels using amniocentesis (Matalon et al, 2011).

MRI confirms the megalencephalic appearance and shows white matter affected as diffuse, bilateral, and involving the sub cortical U-fibers" (Kantor, 2014). In T1weighted image- low signal in white matter; T2 weighted image- high signal in white matter mainly in cerebrum, Globus pallidus and thalamus; no enhancement of affected regions is seen on either computerized tomography (CT) or MRI (Singer, 2010). MR spectroscopy shows markedly elevated NAA and NAA: creatine ratio (Karim Zadeh et al, 2014).

Visual evoked potentials may be low showing cortical blindness; impairment in BAER may be due to problems in cochlear development (Kantor et al, 2014).

Alexander's Disease

MRI shows frontal leukodystrophy more common in the younger patients and occipital lesions, with occasional atrophy of the medulla oblongata and cervical spinal cord, in type-II patients. A multicentric study identified that four of the following five MR imaging criteria is required for diagnosis: extensive cerebral white matter changes with frontal predominance, a periventricular rim with high signal on T1-weighted images and low signal on T2-weighted images, abnormalities of basal ganglia and thalami(elevated signal intensity and swelling, atrophy, elevated or decreased signal intensity on T2-weighted images), brain stem abnormalities particularly involving the medulla and midbrain (T2 hyper intensities in caudate nucleus > globus pallidus > thalamus > brain stem), and contrast enhancement (gadolinium) of particular gray and white matter structures of 1/>(ventricular lining, periventricular rim, frontal white matter, optic chiasm, fornix, basal ganglia, thalamus, dentate nucleus, brain stem) (vanderKnapp et al, 2001). In type 2 cases there may not be any MRI abnormalities. Sometimes atypical MRI abnormalities may be seen like predominant or isolated involvement of posterior fossa structures, multifocal tumor-like brain stem lesions and brain stem atrophy, slight and garland-like feature along the ventricular wall, more likely in juvenile /adult onset AD (Knaap et al, 2006).

Sequence analysis shows 98% of proband with GFAP mutation in 17q 21, failing which exome sequencing studies have to be performed.

Krabbe Disease

In KD, the abnormal Visual Evoked Potential (VEP) response (decrease gradually with no response in later stages of disease- absent P100 wave) but normal ERG is characteristic. (Al-Essa et al, 2000). BAER shows only waves1 & 2. Marked delay in nerve conduction velocity (NCV) due to segmental demyelination obtained by nerve conduction studies (NCS) appears to correlate with clinical severity. NCS shows prolongation of distal latencies, low amplitude and absent evoked responses of prolonged F wave latencies. These findings may be the first signs in KD even before any symptoms develop even in newborns (Siddiqi, 2006). NCS along with brain MRI may be the most sensitive tool in the evaluating the severity and classification of globoid cell leukodystrophy.

CT shows hyper dense areas symmetrically involving the thalami, cerebellum, caudate nuclei, posterior limbs of the internal capsule, and brainstem (Loes et al, 1999). Changes may also extend into the centrum semiovale/corona radiata. There may be marked reduction in white matter in cerebellum and centrum semiovale. MRI on T2 imaging shows hyper intensity involving periventricular white matter centrum

semiovale and deep gray matter; resulting in marked reduction of white-matter with relative sparing of subcortical U-fiber until late in the course of the disease. T1C with Gadolinium enhancement shows no contrast enhancements. MR spectroscopy shows abnormal choline elevation in centrum semiovale.

In infantile form, after testing the enzyme levels of the proband, a targeted analysis for a 30kb deletion is tested. If 2 of the pathogenic variants are not there, then sequence analysis followed by deletion/ duplication analysis should be performed. For late onset forms, targeted analysis for c.857G>A -2variant- should be first performed; if this is negative then search for 2 pathogenic variants for 30kb deletions should be followed. If this is also negative then sequence analysis followed by deletion/duplication analysis should be performed (Wenger et al, 2000).

Metachromatic Leukodystrophy

An ARSA-A enzyme level blood test with a confirming urinary sulfatide test using toluidine blue is the best biochemical test for MLD. The urine sediments have metachromatic galactosphingosulfatides which accumulates in the renal epithelial cells and finally gets excreted through urine. A 24-hour urine samples collection gives ten- to 100-fold higher values than controls and appears as red after staining using thin layer chromatography/mass spectroscopy. The conforming urinary sulfatide is important to distinguish between MLD and pseudo-MLD blood results. In Baun type assay where aryl sulfatase is incompletely blocked, the ARSA enzyme levels should be < 10% of normal controls. When ARSA enzyme activity in WBC is 5-20% of normal controls, are seen in otherwise healthy individuals, then the term "pseudo-deficiency is used. CSF proteins are also elevated (Fluharty, 2014).

NCS shows demyelination; all evoked potentials (VEP, BAER, SSEP) are abnormal.

Electromyographgraphy (EMG) is abnormal with demyelinating sensorimotor neuropathy pattern. CT/MRI shows diffuse atrophy of WM of cerebrum, cerebellum with arcuate fibers sparing and "subcortical U" rim in WM and periventricular involvement. Axial T2WIMR shows the typical butterfly-shaped pattern of white matter involvement in MLD (Groeschel et al, 2011).

Targeted analysis for pathogenic variants - p.Arg84Gln, p.Ser96Phe, c.459+1G>A, p.Ile179Ser, p.Ala212Val, c.1204+1G>A, p.Pro426Leu, and c.1401_1411del- identifies up to 50% in late infantile and juvenile MLD and 73-90% of adult MLD. (Lugowska et al, 2005).

Prenatal diagnosis is possible by doing ARS-A assay levels in cells of either chorionic villi cells or amniotic fluid.

Adrenoleukodystrophy

Impaired cortisol challenge test by injecting ACTH250mcgI/V (low rise of cortisol) is seen, especially Addison only form. Elevated ACTH is seen in 85% of children with childhood form (Kown, 2016). In males with neurological symptoms raised VLCFA levels gives the diagnosis. In females who are carriers, mutation analysis needs to be done. MRI shows symmetric periventricular white matter involvement in parietal and occipital lobes in most cases. The perivascular lymphocytic infiltration, on T2-weighed and FLAIR sequences in the parieto-occipital region and the splenium, results in increased signal intensity in 50% of ALD resulting in a garland of contrast material adjacent to the posterior hypodense lesion (vander Knap & Valk, 2005). Newborn screening done with dried blood spots obtained in special paper is assessed for C26:0 lysophosphatidylcholine (26:0-lyso-PC).

Lafora Disease

Though initial EEG's may be normal, at later stages, focal abnormalities in occipital area or generalized irregular spike-wave discharges with occipital predominance against a slow background are seen. The most characteristic seizures include myoclonic and occipital lobe seizures with visual hallucinations, scotomata, and photo convulsions.

Visual Evoked potential (VEP) - initial stage- high voltage potentials. Prolonged P110- absent in some and in some it may be normal. BAER, SSEP- shows high voltage somatosensory evoked potentials (14-170 μ V) in initial stages. Patients with LD have varied cortical relay times. In response to afferent stimuli, a delayed and prolonged facilitation, evidenced as sustained hyperexcitability of the sensorimotor cortex, happens. The findings are suggestive of an impaired inhibitory mechanism in LD. Later in disease, these giant waves become normal with some prolongation of central and brain stem latencies. NCS and MRI are normal throughout (Jansen, A.C,2015)

Diagnosis is made by axillary skin biopsy in which pathognomic finding of Lafora bodies (polyglucosans) are seen in myoepithelial cells surrounding axillary apocrine (odoriferous) glands. In other areas outside the axilla, Lafora bodies (PAS positive) are found in the cells composing the ducts of the eccrine (perspiration) glands (Jansen, 2010).

Mucolipidosis

X-rays- shows progressive osteopenia and dysostosis multiplex (diaphyseal widening and shortening of tubular bones; long pubic and ischial bones and anterior-inferior hook configuration of 1^{st} and/or 2^{nd} lumbar vertebra). Plasma enzyme assays-A 5-20 times high levels of lysosomal hydrolase and hydrolases in other body fluids are seen (normal 0-200 pg/mL) (Leroy et al, 2012). Confirmation may be made by molecular mutation screening of the *GNPTAB* gene by single-gene testing, use of a multi-gene panel, or more comprehensive genomic testing.

MRI brain- shows typically, corpus callosal abnormalities like-corpus callosal hypoplasia, absent rostrum and a dysplastic or absent splenium; white matter on T1 weighted images shows signal abnormalities and thalamus and basal ganglia shows increased ferritin deposition. Cerebellar atrophy is seen in older. EEG- shows epileptiform discharges.

Neuronal Ceroid Lipofuscinosis

The ERG is not recordable by four years in CLN-1. In CLN-2, ERG is usually abnormal at presentation and becomes undetectable soon thereafter. Visual evoked potentials (VEPs) are enhanced for a long period and diminish in the final stage of the disease. In CLN-3, ERG shows loss of photoreceptor function early on (Weleber, 1998).

From 18 months onwards, EEG changes may be seen in CLN-1. In CLN-2 it shows spikes in the occipital region on photic stimulation at 1-2 Hz. In CLN-3, EEG shows nonspecific disorganization and spike-and-slow-wave complexes.

MRI findings in CLN1 are variable cerebral atrophy; signal change in the thalami and basal ganglia; and thin, hyperintense, periventricular high-signal rims of white matter (Riikonen et al, 2000). MRI in CLN-2 shows progressive cerebellar and cerebral atrophy with normal basal ganglia and thalami. CT and MRI in CLN-3 reveal cerebral, and to a lesser degree, cerebellar atrophy after 15 years of age.

Since the finding of 13 different mutations in genes, NCL is classified based on genes as seen in Table 3 (Williams & Mole, 2012).

Subacute Sclerosing Panencephalitis

The diagnosis of SSPE is made by Dyken's criteria. The diagnosis of SSPE can be established, if the patient fulfills any three of the following five criteria (Dyken, 1985):

- 1. Typical clinical presentation with progressive intellectual deterioration with signs of myoclonus;
- 2. Characteristic electroencephalographic (EEG) pattern;
- 3. Elevated cerebrospinal fluid (CSF) globulin levels, presence of oligoclonal IgG bands;
- 4. Elevated CSF measles antibody titers; and
- 5. Brain biopsy suggestive of measles-perivascular inflammatory cuffing, cortical and subcortical white matter astro-microgliosis, neurophagia and Cowdry type A, eosinophilic intaranuclear inclusion bodies.

Most characteristic EEG abnormality is bilaterally symmetrical, periodic (4-10seconds), stereotyped, high voltage delta wave discharges which appear along with the myoclonic jerks-type 1 waves. In early stages, bursts of high voltage delta waves are interspersed with a normal background at constant interval. The periodic pattern emerges with disease progression first during sleep, much before myoclonic jerk come. Later the characteristic periodic discharge pattern can be seen also during wakefulness, happening along with myoclonus (Garg, 2002).

CT/MRI shows cerebral edema and atrophy in all stages of the disease and the signal intensity changes in stages II-III. In stage II-Parieto-occipital white matter was affected, while diffuse frontoparietal changes were more common in stage II-III. There is no correlation between the imaging features and focal EEG changes or the abnormal neurological findings.

Gene Involved	Wide Spread Common Mutation	Phenotype - Clinical Features
CLN10/CTSD	Not known	Congenital - severe seizure, rigidity, blindness,
PPT1/CLN1	p.Arg122Trp; p.Arg151X- over 67 mutations reported	Infantile - early seizures, psycho- motor regression, vision loss. PPT1 has juvenile like presentation
CLN2/TPP1 CLN-5-8	c.509-1G>C; p.Arg208X; 116 mutations are identified- CLN-2 resulting in abnormalities in tripeptidyl peptidase, ; others result in abnormalities in membrane proteins	Late infantile - presentation as myoclonic seizures in 2-4 years; microcephaly, dementia, ataxia, loss of visual acuity. Black bony spicule like in Retinitis pigmentosa, optic atrophy, brown pigment in macula.
CLN3	1 kb intragenic deletion	Juvenile- most common form; first 5 years normal, then visual loss. Retinal pigmentary changes-RP; the rapid decline in psychomotor, motor incoordination, seizures. Later parkinsonism develops; die in twenties.
CLN12/ATP13A2		Juvenile
CLN14/KCTD7		Infantile

Table 3. Genetic mutations and presentation of neuronal ceroid lipofuscinosis

TREATMENT OF CHILDHOOD NEURODEGENERATIVE DISORDERS

Canavan's Disease

Management is limited currently to supportive measures and management of complications. Physical therapy aiming at maintaining postures and early intervention for communication skills are advised. Insertion of feeding tube for swallowing difficulties and G-tube if at risk of aspiration improves nutrition and hydration. Often multiple anticonvulsants for seizure control and oral supplementation with acetate is advised (Leone et al,1999). Survival beyond adolescence depends on medical and supportive care given; those with congenital forms will die in first weeks of life. Juvenile forms have often very protracted course (Kantor, 2014).

Alexander's Disease

Neurosurgical intervention should be considered on an individual basis, if need arises. For type-2, management of sleep apnea and autonomic symptoms alleviates encephalopathy and increase the quality of life. Life span of AD children is based on the type of disorder; the median age of survival of type -I is 14 years and of type -II is 24 years (Prust et al, 2011).

Krabbe Disease

HSCT is currently the standard of care for KD patients; however, it serves only to delay disease progression and is not an effective cure (Sakai, 2009). It is effective only when performed early during the disease course. If performed early in pre-symptomatic child, umbilical cord blood (UCB) transplantation can preserve the cognition and prolong life; however most experience a decline in language, motor functions and adaptive behavior. There are evidences that combination therapies that target multiple pathogenic mechanisms/pathways (CNS-directed gene therapy, substrate reduction therapy, and BMT to target GALC deficiency) have been more synergic. Hence, they were more effective at reducing histological signs of disease, delaying onset, prolongs life span, and improve the behavioral/cognitive functions in rodent models of KD. Supportive care to control irritability and spasticity may be given for KD in later stages of disease (Table 4).

Metachromatic Leukodystrophy

Enzyme -replacement therapy, bone marrow/UCB transplant, *ex vivo* transplantation of genetically modified hemopoietic stem cells, all have been tried with minimal benefits during clinical trials. Gene therapy with adeno associated viruses AAVrh10 serotype for clinical development as a treatment for MLD has been proposed recently as a possible ideal treatment (Rosenberg, 2016). The 5-year survival for metachromatic leukodystrophy is reported to be-late infantile: 52%, juvenile: 100% and adult: 95%. The late infantile subtype was the worst survivor than any other subtypes (Asif Mahmood, et al, 2010).

Complications	Management	
Vomiting/ reflux	Upright position during and after feeding; > 1 year-use Lanzoprazole; Nissen's fundoplication for reflux	
Constipation	Sufficient fluids; PEG, lactulose	
Dysphagia	Modify thickness of food. If not feeding orally, swallowing ability can be improved by providing juices many times in a day. The gastrostomy tube for terminally ill to be inserted for feeding while the child is still able to swallow rather than waiting until the child is undernourished and dehydrated. Caloric intake provided-to be based on the child's length than age or weight (7-9 calories/cm/day, adjusted to the child's activity level).	
Spasticity; contractures	Wedges, cushions and pillows for seating; gentle stretching, use of orthotics, splints and adaptive devices for standing etc. Botox injections	
Neuropathic pain	Gabapentin	
Seizures	Diazepam for breakthrough seizures. Monotherapy for seizures	
Urogenic bladder in later stages	Crede manavoure for complete emptying.	
Respiratory infection	Chest physiotherapy, postural drainage, frequent suctioning; annual influenza vaccine	

Table 4. Management of Krabbe disease complications

Adrenoleukodystrophy

Early start of intervention gives a better prognosis. Measure the adrenal insufficiency, document adrenocortical hypofunction and measure plasma VLCFA levels in all children and also in all male patients with Addison's disease as in > 25% of boys with Addison's disease, ALD causes adrenal crisis. Corticosteroid replacement is started for adrenal insufficiency as it is lifesaving, though it does not alter the course of disease. Coordination between the treating physician and school authorities helps in recognizing appearance of subtle behavioral issue and attentional issues early in course of disease to give proper counseling. Melatonin may be used for sleep cycle problems.

An initial and 6 monthly MRI should be done, especially in neurologically asymptomatic male children and adolescents. Usually MRI abnormalities precede the neuropsychological manifestation. Positive MRI should be followed by repeating every 3 months to see the progression of disease. Performance (nonverbal) IQ scores are shown to correlate well with MRI findings and hence can be used as a monitoring tool. Both MRI and performance IQ can be used as a predictive tool for assessing the need for early bone marrow transplantation in those who have not developed neuro/psychological manifestations (Suzuki, et. al, 2000).

Lorenzo oil therapy (4:1 mixture of glyceryl trioleate and glyceryl tri crucate) with dietary therapy has been used in asymptomatic boys with normal MRI and is < 8-year-old. Upon deterioration, they are considered for hemopoietic stem cell therapy. It has been found to reduce the childhood form of ALD by a factor 2 or more. However, in those who are already having neurological manifestation, Lorenzo's oil therapy is of no use. Bone marrow transplantation is indicated in boys with rapidly progressive neurological inflammatory demyelination seen by MRI. BMT cells are found to produce ALDP, the protein which is deficient in ALD oligodendroglial cells. However, BMT is not effective in those who have established brain damage and in fact may increase the progression in such cases. Hence careful selection of case is important as BMT itself has 10-20% mortality. Also 50% of those ALD who are not treated will not develop inflammatory demyelination and hence transplantation is not needed (Orchard, J., P,2010).

Gene modification of stem cells by inserting normal copy of ALD gene via a virus, after ablating the residual bone marrow of the child was found to be favorable. Infusion of their own genetically corrected stem cells with normal gene showed 15% of their cells produced ALDP with a 38% reduction of VLCFA in plasma with arrest of demyelination as measured by MRI and clinical findings (Cartier et al, 2009).

Another investigational study (Star beam study) to determine the safety and tolerability of Lenti-D (autologous hematopoietic cells that were genetically corrected with a lentiviral vector) while determining if the one-time treatment can stop the progression of cerebral ALD is under way. Interim results of the study are encouraging.

Lafora Disease

Valproate, levetiracetam, topiramate, and benzodiazepines are used with modest efficacy on seizure frequency. Phenytoin, carbamazepine, oxcarbazepine, vigabatrin, tiagabine, gabapentin and lamotrigine may be avoided as they aggravate the myoclonus and even precipitate GTCS. There are recent reports suggesting that a new drug, Perampanel, given as monotherapy produces dramatic reduction in seizure frequency with a striking improvement in cognition, behavior, and cerebellar function (Dirani et al, 2014).

Perampanel's anticonvulsant effect is because of its selective noncompetitive antagonistic action on AMPA type glutamate receptors. A recent study suggested that loss of GABAergic cortical neurons and the resulting imbalance in GABAergic: Glutamatergic neurons might be pathophysiologically related to Lafora's disease (Ortolano et al, 2014). This probably explains the mechanism by which Perampanel, by blocking the AMPA receptors, partially normalizes the imbalances in the inhibitory: excitatory neurotransmitters in cortex and cerebellum and thereby reducing seizures and restoring the manifestations to near normalcy. Vagus nerve stimulation may also help as an adjuvant in seizure control.

Mucolipidosis

Treatment options are limited, minimal exercises to avoid joint and tendon strain, like aqua therapy, cognitive stimulation through interactive programs; gingivectomy - for oral health; myringotomy tube placement-recurrent ear infections.

Neuronal Creroid Lipofuscinosis

Symptomatic treatment for the management of seizure, gastroesophageal reflux, sleep issues, drooling, behavioral issues has to be given. Lamotrigene has been found to be useful in this case as monotherapy with 100% control vs. valproic acid (60%) control (Aberg et al, 2000). For feeding issues when it becomes riskier, G-tube may be inserted. Trihexphenidyl may be used for drooling and dystonia.

Subacute Sclerosing Panencephalitis

Mostly they die in 2-3 years from presentation. A relative better result was achieved with either daily oral Isoprinosine or in combination with weekly intrathecal INF- α . (Garg, 2002). Isoprinosine (Inosiplex), improves immunity by its action on natural killer cell function; by increasing the number of CD4+ cells and by increasing the production of interleukin-1 and interleukin-2. Currently, the most effective treatment is the combined treatment of oral inosiplex for life and intraventricular interferon- α . Spontaneous

remission may occur during any stage of the disease only to relapse later. Age of onset < 12 years, disappearance of periodic complexes, normalization of the background of follow up EEGs, and a progressive increase in measles antibody titers in cerebrospinal fluid are associated with favorable outcome in SSPE.

FUTURE RESEARCH DIRECTIONS

In the management of NDs like CD, new treatment options are emerging like the introduction of a vector expressing *ASPA* gene by injection into 6 brain parenchymal sites resulting in reduction in seizures, slowed the progression with reduction in brain NAA, no severe long-term adverse effects, with slowing of atrophy of brain, significant improvement in motor functions and reduction in seizure frequency (Leone et al, 2012). Further studies are required to confirm these beneficial effects. Further studies using drugs which reduce the NAA levels in brain or increases the acetate levels in humans, after adequate trials, may give an idea about managing CD better. However, in a recent study *ASPA* gene replacement therapy targeted against oligodendrocytes resulted in reversing of preexisting neural pathology and lasting neurological benefits in rats (von Jonquieres et al, 2017). Similarly, Mohri et al. (2006) using twitcher mouse model, similar to KD, found that hematopoietic prostaglandin D synthase inhibitors may have a role as anti-neuroinflammatory therapy. This is based on the finding that activated microglia and astrocytes expresses H-PGDS and DP1 receptors respectively, in the brain of the mice and PGD2 is a key neuroinflammatory molecule involved in demyelination in twitcher mice.

Pharmacological chaperone therapy (PCT) where chaperons bind directly to a partially folded biosynthetic intermediate, stabilize the protein and allow completion of the folding process to yield a functional protein, is emerging as useful idea in many NDs. This may be used for selectively correcting defective protein folding and trafficking and for enhancing enzyme activity by small molecules in Krabbes disease. Potential therapeutic benefits of chaperons in Krabbes disease noted in preclinical studies is worth taking to next level of research (Graziano et al, 2016). Mutations leading to misfolding, giving rise to lack of secretion, processing and ER co-localization, may be good targets for pharmacological chaperone therapies. Those that retain significant capacity for correct trafficking and processing will require future enzyme replacement approaches. But the extend of misfolding caused by specific mutations will alter the effectiveness of pharmacological chaperones. Specifically, the misfolding caused by some mutations may be too severe to respond to these approaches and hence will require enzyme replacement strategies (Spratley et al, 2016).

In many diseases like ALD, the effect of several factors like epigenetics and environmental modifiers are still not known fully. There is a need to identify such factors /modifiers of human gene expression so that they can be used constructively and may also be used to predict the clinical outcome.

CONCLUSION

The genetic basis of NDs, deranged gene products and its effects on children, gives an option to classification of these disorders in the future. The newer advances at cellular level/molecular levels especially as treatment options are at experimental level in many disorders. Soon those options may become operationalized like, interventions aimed at substrate replacement/substitute (enzyme replacement therapy) or may be an alternate one in which abnormal molecule is prevented from forming like in chaperon therapy. In many cases, prompt recognition and early treatment favor a better therapeutic response. Next generation sequencing technologies have helped to fill gaps in diagnosis. Finding the biomarkers of many of these disorders has made it possible to think in terms of finding solutions for treatment for many of these disorders. Future researches focusing on the biomarkers may be the right direction for finding solutions for treatment.

REFERENCES

Abajirao, S. A., Nair, M., & Kambale, H. (2013). Subacute sclerosing panencephalitis: A clinical appraisal. *Annals of Indian Academy of Neurology*, *16*(4), 631. doi:10.4103/0972-2327.120497 PMID:24339595

Aberg, L. E., Backman, M., Kirveskari, E., & Santavuori, P. (2000). Epilepsy and Antiepileptic Drug Therapy in Juvenile Neuronal Ceroid Lipofuscinosis. *Epilepsia*, *41*(10), 1296–1302. doi:10.1111/j.1528-1157.2000. tb04608.x PMID:11051125

Al-Essa, M. A., Bakheet, S. M., Patay, Z. J., Powe, J. E., & Ozand, P. T. (2000). Clinical and cerebral fdg pet scan in a patient with krabbe's disease. *Pediatric Neurology*, 22(1), 44–47. doi:10.1016/S0887-8994(99)00107-1 PMID:10669205

Alexander, W. S. (1949). Progressive Fibrinoid Degeneration of Fibrillary Astrocytes Associated with Mental Retardation in a Hydrocephalic Infant. *Brain*, 72(3), 373–381. doi:10.1093/brain/72.3.373 PMID:15409268

Backman, M. L., Santavuori, P. R., Aberg, L. E., & Aronen, E. T. (2005). Psychiatric symptoms of children and adolescents with juvenile neuronal ceroid lipofuscinosis. *Journal of Intellectual Disability Research*, *49*(1), 25–32. doi:10.1111/j.1365-2788.2005.00659.x PMID:15634309

Baslow, M. H., & Guilfoyle, D. N. (2013). Canavan disease, a rare early-onset human spongiform leukodystrophy: Insights into its genesis and possible clinical interventions. *Biochimie*, *95*(4), 946–956. doi:10.1016/j.biochi.2012.10.023 PMID:23151389

Bouteille, M., Fontaine, C., & Vedrenne, C. L. (1965). Sur uncas d'encephalite subaiguea inclusions. Etude anatomoclinique et ultra structurale. *Revue Neurologique*, *113*, 454–458.

Brenner, M., Goldman, J. E., Quinlan, R. A., & Messing, A. (2009). Alexander Disease: A Genetic Disorder of Astrocytes. In P. Haydon & V. Parpura (Eds.), *Astrocytes in (Patho)Physiology of the Nervous System* (pp. 591–648). Boston, MA: Springer. doi:10.1007/978-0-387-79492-1_24

Campbell, H., Andrews, N., Brown, K. E., & Miller, E. (2007, December). Review of the effect of measles vaccination on the epidemiology of SSPE. *International Journal of Epidemiology*, *36*(6), 1334–1348. doi:10.1093/ije/dym207 PMID:18037676

Canavan, M. M. (1931). Schilder's encephalitis periaxialis diffusa - Report or a case in a child aged sixteen and one-half months. *Archives of Neurology and Psychiatry*, 25(2), 299–308. doi:10.1001/archneurpsyc.1931.02230020085005

Cartier, N., Hacein-Bey-Abina, S., Bartholomae, C. C., Veres, G., Schmidt, M., Kutschera, I., ... Aubourg, P. (2009). Hematopoietic Stem Cell Gene Therapy with a Lentiviral Vector in X-Linked Adrenoleukodystrophy. *Science*, *326*(5954), 818–823. doi:10.1126cience.1171242 PMID:19892975

Cathey, S. S., Leroy, J. G., Wood, T., Eaves, K., Simensen, R. J., Kudo, M., ... Friez, M. J. (2010). Phenotype and genotype in mucolipidoses II and III alpha/beta: A study of 61 probands. *Journal of Medical Genetics*, 47(1), 38–48. doi:10.1136/jmg.2009.067736 PMID:19617216

Dawson, J. R. (1933). Cellular Inclusions in Cerebral Lesions of Lethargic Encephalitis. *The American Journal of Pathology*, 9(1), 7–16.3.

Dirani, M., Nasreddine, W., Abdulla, F., & Beydoun, A. (2014). Seizure control and improvement of neurological dysfunction in Lafora disease with perampanel. *Epilepsy & Behavior Case Reports*, 2, 164–166. doi:10.1016/j.ebcr.2014.09.003 PMID:25667898

Dyken, P., & Krawiecki, N. (1983). Neurodegenerative diseases of infancy and childhood. *Annals of Neurology*, *13*(4), 351–364. doi:10.1002/ana.410130402 PMID:6301358

Dyken, P. R. (1985). Subacute sclerosing panencephalitis. Current status. *Neurologic Clinics*, 3(1), 179–196. PMID:2581121

Fourcade, S., López-Erauskin, J., Galino, J., Duval, C., Naudi, A., Jove, M., ... Pujol, A. (2008). Early oxidative damage underlying neurodegeneration in X-adrenoleukodystrophy. *Human Molecular Genetics*, *17*(12), 1762–1773. doi:10.1093/hmg/ddn085 PMID:18344354

Gadoth, N. (2011). Subacute Sclerosing Pan-Encephalitis (SSPE) – Past and Present. *Pathogenesis of Encephalitis*, 135-149.

Gambetti, P., Mellman, W. J., & Gonatas, N. K. (1969). Familial spongy degeneration of the central nervous system(vanBogaert-Bertrant disease). *Acta Neuropathologica*, *12*(2), 103–115. doi:10.1007/BF00692500 PMID:5789730

Garg, R. (2002). Subacute sclerosing panencephalitis. *Postgraduate Medical Journal*, 78(916), 63–70. doi:10.1136/pmj.78.916.63 PMID:11807185

Gentry, M. S., Worby, C. A., & Dixon, J. E. (2005). Insights into Lafora disease: Malin is an E3 ubiquitin ligase that ubiquitinates and promotes the degradation of laforin. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(24), 8501–8506. doi:10.1073/pnas.0503285102 PMID:15930137

Gomez-Ospina, N. (2006 May 30). Arylsulfatase A Deficiency. In M. P. Adam, H. H. Ardinger, & R. A. Pagon (Eds.), *GeneReviews*® (pp. 1993–2017). Seattle, WA: University of Washington, Seattle. Available from https://www.ncbi.nlm.nih.gov/books/NBK1130/

Granata, T. (2012). Metabolic and degenerative disorders. *Handbook of Clinical Neurology*, *108*, 485–511. doi:10.1016/B978-0-444-52899-5.00045-9 PMID:22939050

Graziano, A. C. E., Pannuzzo, G., Avola, R., & Cardile, V. (2016, November 1). Chaperones as potential therapeutics for Krabbe disease. *Journal of Neuroscience Research*, 94(11), 1220–1230. doi:10.1002/jnr.23755 PMID:27638605

Groeschel, S., Kehrer, C., Engel, C., & Dali, Í. (2011). Metachromatic leukodystrophy: Natural course of cerebral MRI changes in relation to clinical course. *Journal of Inherited Metabolic Disease*, *34*(5), 1095–1102. doi:10.100710545-011-9361-1 PMID:21698385

Haneef, S. S., & Doss, C. G. (2016). Personalized Pharmacoperones for Lysosomal Storage Disorder. *Advances in Protein Chemistry and Structural Biology Personalized Medicine*, *102*, 225–265. doi:10.1016/ bs.apcsb.2015.10.001 PMID:26827607

Jabbour, J. T., Garcia, J. H., Lemmi, H., Ragland, J., Duenas, D. A., & Sever, J. L. (1969). Subacute sclerosing pan encephalitis. A multidisciplinary study of eight cases. *Journal of the American Medical Association*, 207(12), 2248–2254. doi:10.1001/jama.1969.03150250078007 PMID:5818397

Jansen, A. C. (2010). Lafora disease. In K. Kompoliti & V. L. Metman (Eds.), *Encyclopedia of move*ment disorders (pp. 113–116). Oxford, UK: Academic Press. doi:10.1016/B978-0-12-374105-9.00338-5

Jansen, A. C., & Andermann, E. (2007, Dec. 28). Progressive Myoclonus Epilepsy, Lafora Type. In *Gene Reviews* (pp. 1993-2017). Seattle, WA: University of Washington. Available from: https://www.ncbi.nlm.nih.gov/books/NBK1389/

Kantor, B., McCown, T., Leone, P., & Gray, S. J. (2014). Clinical Applications Involving CNS Gene Transfer. *Advances in Genetics*, 87, 71–124. doi:10.1016/B978-0-12-800149-3.00002-0 PMID:25311921

Karimzadeh, P., & Jafari, N., Nejad biglari, H., Rahimian, E., Ahmadabadi, F., Nemati, H., ... Mollamohammadi, M. (2014). The Clinical Features and Diagnosis of Canavan's Disease: A Case Series of Iranian Patients. *Iranian Journal of Child Neurology*, 8(4), 66–71. PMID:25657773

Kaul, R., Gao, G. P., Matalon, R., Aloya, M., Su, Q., Jin, M., ... Clarke, J. T. (1996). Identification and expression of eight novel mutations among non-Jewish patients with Canavan disease. *American Journal of Human Genetics*, *59*(1), 95–102. PMID:8659549

Knaap, M. S., Ramesh, V., Schiffmann, R., Blaser, S., Kyllerman, M., Gholkar, A., ... Salomons, G. S. (2006). Alexander disease: Ventricular garlands and abnormalities of the medulla and spinal cord. *Neurology*, *66*(4), 494–498. doi:10.1212/01.wnl.0000198770.80743.37 PMID:16505300

Kohlschütter, A. (2013). Lysosomal leukodystrophies. Handbook of Clinical Neurology Pediatric Neurology Part III, 1611-1618.

Kudo, M., Brem, M. S., & Canfield, W. M. (2006). Mucolipidosis II (I-Cell Disease) and Mucolipidosis IIIA (Classical Pseudo-Hurler Polydystrophy) Are Caused by Mutations in the GlcNAc-Phosphotransferase α/β –Subunits Precursor Gene. *American Journal of Human Genetics*, 78(3), 451–463. doi:10.1086/500849 PMID:16465621

Kwon, M. J. (2016). Neurodegenerative disorders of childhood. In R. Kliegman, B. Stanton, J. W. St. Geme, N. F. Schor, & R. E. Behrman (Eds.), *Nelson text book of Pediatrics* (20th ed.; pp. 2910–2918). Philadelphia, PA: Elsevier.

Leone, P., Janson, C. G., McPhee, S. J., & During, M. J. (1999). Global CNS gene transfer for a childhood neurogenetic enzyme deficiency: Canavan disease. *Current Opinion in Molecular Therapeutics*, *1*(4), 487–492. PMID:11713764

Leone, P., Shera, D., McPhee, S. W. J., Francis, J. S., Kolodny, E. H., Bilaniuk, L. T., ... Janson, C. G. (2012). Long-Term Follow-Up After Gene Therapy for Canavan Disease. *Science Translational Medicine*, *4*(165).

Leroy, J. G., Cathey, S., & Friez, M. J. (2008 Aug 26). Mucolipidosis II. In M. P. Adam, H. H. Ardinger, & R. A. Pagon (Eds.), GeneReviews (pp. 1993–2017). Academic Press.

Li, R., Messing, A., Goldman, J. E., & Brenner, M. (2002). GFAP mutations in Alexander disease. *International Journal of Developmental Neuroscience*, 20(3–5), 259–268. doi:10.1016/S0736-5748(02)00019-9 PMID:12175861

Loes, D. J., Peters, C., & Krivit, W. (1999, February). Globoid Cell Leukodystrophy: Distinguishing Early- Onset from Late-Onset Disease Using a Brain MR Imaging Scoring Method. *AJNR. American Journal of Neuroradiology*, 316–323. PMID:10094363

Lugowska, A., Berger, J., Tylki-Szymańska, A., Löschl, B., Molzer, B., Zobel, M., & Czartoryska, B. (2005). Molecular and phenotypic characteristics of metachromatic leukodystrophy patients from Poland. *Clinical Genetics*, 68(1), 48–54. doi:10.1111/j.1399-0004.2005.00451.x PMID:15952986

Mahmood, A., Berry, J., Wenger, D. A., Escolar, M., Sobeih, M., Raymond, G., & Eichler, F. S. (2009). Metachromatic Leukodystrophy: A Case of Triplets with the Late Infantile Variant and a Systematic Review of the Literature. *Journal of Child Neurology*, 25(5), 572–580. doi:10.1177/0883073809341669 PMID:20038527

Marder, K., Logroscino, G., Alfaro, B., Mejia, H., Halim, A., Louis, E., ... Mayeux, R. (1998). Environmental risk factors for Parkinson's disease in an urban multiethnic community. *Neurology*, *50*(1), 279–281. doi:10.1212/WNL.50.1.279 PMID:9443493

Matalon, R., & Matalon, K. M. (2015). Canavan Disease. In Rosenberg's Molecular and Genetic Basis of Neurological and Psychiatric Disease (5th ed.; pp. 695-701). Boston: Academic.

Matalon, R., & Michals-Matalon, K. (1999 Sep 16). Canavan Disease. In M. P. Adam, H. H. Ardinger, R. A. Pagon, & ... (Eds.), *GeneReviews* (pp. 1993–2017). Seattle, WA: University of Washington. Available from https://www.ncbi.nlm.nih.gov/books/NBK1234/

Messing, A., Brenner, M., Feany, M. B., Nedergaard, M., & Goldman, J. E. (2012). Alexander Disease. *The Journal of Neuroscience*, *32*(15), 5017–5023. doi:10.1523/JNEUROSCI.5384-11.2012 PMID:22496548

Minassian, B. A. (2001). Lafora's disease: Towards a clinical, pathologic, and molecular synthesis. *Pediatric Neurology*, 25(1), 21–29. doi:10.1016/S0887-8994(00)00276-9 PMID:11483392

Childhood Neurodegenerative Disorders

Mohri, I., Taniike, M., Taniguchi, H., Kanekiyo, T., Aritake, K., Inui, T., ... Urade, Y. (2006). *Prosta*glandin D 2 -Mediated Microglia / Astrocyte Interaction Enhances Astrogliosis and Demyelination in twitcher. Academic Press.

Monaghan, T. S., & Delanty, N. (2010). Lafora disease: Epidemiology, pathophysiology and management. *CNS Drugs*, 24(7), 549–561. doi:10.2165/11319250-000000000-00000 PMID:20527995

Ofman, R., Dijkstra, I. M. E., Van Roermund, C. W. T., Burger, N., Turkenburg, M., Van Cruchten, A., ... Kemp, S. (2010). The role of ELOVL1 in very long-chain fatty acid homeostasis and X-linked adrenoleu-kodystrophy. *EMBO Molecular Medicine*, *2*(3), 90–97. doi:10.1002/emmm.201000061 PMID:20166112

Orchard, J. P., & Tolar, J. (2010, January). Transplant Outcomes in Leukodystrophies. *Seminars in Hematology*, 47(1), 70–78. doi:10.1053/j.seminhematol.2009.10.006 PMID:20109614

Ortolano, S., Vieitez, I., Agis-Balboa, R. C., & Spuch, C. (2014). Loss of GABAergic cortical neurons underlies the neuropathology of Lafora disease. *Molecular Brain*, 7(1), 7. doi:10.1186/1756-6606-7-7 PMID:24472629

Prust, M., Wang, J., Morizono, H., Messing, A., Brenner, M., Gordon, E., ... Vanderver, A. (2011). *GFAP* mutations, age at onset, and clinical subtypes in Alexander disease. *Neurology*, 77(13), 1287–1294. doi:10.1212/WNL.0b013e3182309f72 PMID:21917775

Riikonen, R. (2017). Insulin-Like Growth Factors in the Pathogenesis of Neurological Diseases in Children. *International Journal of Molecular Sciences*, *18*(10), 2056. doi:10.3390/ijms18102056 PMID:28954393

Rodriguez, D. (2013). Leukodystrophies with astrocytic dysfunction. Handbook of Clinical Neurology Pediatric Neurology Part III, 1619-1628.

Rodriguez, D., Gauthier, F., Bertini, E., Bugiani, M., Brenner, M., N'guyen, S., ... Boespflug-Tanguy, O. (2001). Infantile Alexander Disease: Spectrum of GFAP Mutations and Genotype-Phenotype Correlation. *American Journal of Human Genetics*, 69(5), 1134–1140. doi:10.1086/323799 PMID:11567214

Rosenberg, J. B., Kaminsky, S. M., Aubourg, P., Crystal, R. G., & Sondhi, D. (2016, November 1). Gene therapy for metachromatic leukodystrophy. *Journal of Neuroscience Research*, *94*(11), 1169–1179. doi:10.1002/jnr.23792 PMID:27638601

Ryan, C. L., Baranowski, D. C., Chitramuthu, B. P., Malik, S., Li, Z., Cao, M., ... Bateman, A. (2009). Progranulin is expressed within motor neurons and promotes neuronal cell survival. *BMC Neuroscience*, *10*(1), 130. doi:10.1186/1471-2202-10-130 PMID:19860916

Sakai, N. (2009, August). Pathogenesis of leukodystrophy for Krabbe disease: Molecular mechanism and clinical treatment. *Brain & Development*, *31*(7), 485–487. doi:10.1016/j.braindev.2009.03.001 PMID:19332366

Schmahmann, J. D., Smith, E. E., Eichler, F. S., & Filley, C. M. (2008, October). Cerebral white matter: Neuroanatomy, clinical neurology, and neurobehavioral correlates. *Annals of the New York Academy of Sciences*, *1142*(1), 266–309. doi:10.1196/annals.1444.017 PMID:18990132

Schönberger, K., Ludwig, M. S., Wildner, M., & Weissbrich, B. (2013). Epidemiology of Subacute Sclerosing Panencephalitis (SSPE) in Germany from 2003 to 2009: A Risk Estimation. *PLoS One*, 8(7), e68909. doi:10.1371/journal.pone.0068909 PMID:23874807

Shu, S. K., Michelson, D. J., & Ashwal, S. (2017). Cognitive and Motor Regression. In Swaiman's Pediatric Neurology: Principles &Practice (6th ed.; pp. 424-430). Elsevier Inc. doi:10.1016/B978-0-323-37101-8.00052-7

Siddiqi, Z. A., Sanders, D. B., & Massey, J. M. (2006). Peripheral neuropathy in Krabbe disease: Electrodiagnostic findings. *Neurology*, 67(2), 263–267. doi:10.1212/01.wnl.0000230153.34613.84 PMID:16864819

Singer, H. S., Mink, J. W., Gilbert, D. L., & Jankovic, J. (2010). Inherited Metabolic Disorders Associated with Extrapyramidal Symptoms. *Movement Disorders in Childhood*, 164-204.

Spratley, S. J., & Deane, J. E. (2016). New therapeutic approaches for Krabbe disease: The potential of pharmacological chaperones. *Journal of Neuroscience Research*, *94*(11), 1203–1219. doi:10.1002/jnr.23762 PMID:27638604

Spratley, S. J., Hill, C. H., Viuff, A. H., Edgar, J. R., Skjødt, K., & Deane, J. E. (2016). Molecular Mechanisms of Disease Pathogenesis Differ in Krabbe Disease Variants. *Traffic (Copenhagen, Denmark)*, *17*(8), 908–922. doi:10.1111/tra.12404 PMID:27126738

Springer, S., Erlewin, R., Nagaele, T., Becker, I., Auer, D., Grodd, W., & Krägeloh-Mann, I. (2000). Alexander Disease - Classification revisited and isolation of a neonatal form. *Neuropediatrics*, *31*(2), 86–92. doi:10.1055-2000-7479 PMID:10832583

Spuch, C., Ortolano, S., & Navarro, C. (2012, May). Lafora progressive myoclonus epilepsy: Recent insights into cell degeneration. *Recent Patents on Endocrine, Metabolic & Immune Drug Discovery*, *6*(2), 99–107. doi:10.2174/187221412800604617 PMID:22369717

Srivasthava, N. S. (2002, Nov. 15). Alexander disease. In Gene Reviews. Seattle, WA: University of Washington.

Suzuki, K. (2003). Globoid Cell Leukodystrophy (Krabbes Disease): Update. *Journal of Child Neurology*, *18*(9), 595–603. doi:10.1177/08830738030180090201 PMID:14572137

Suzuki, Y., Isogai, K., Teramoto, T., Tashita, H., Shimozawa, N., Nishimura, M., & Kondo, N. (2000). JULY). Bone marrow transplantation for the treatment of X-linked adrenoleukodystrophy. *Journal of Inherited Metabolic Disease*, *23*(5), 453–458. doi:10.1023/A:1005656029200 PMID:10947199

Tang, G., Perng, M. D., Wilk, S., Quinlan, R., & Goldman, J. E. (2010). Oligomers of Mutant Glial Fibrillary Acidic Protein (GFAP) Inhibit the Proteasome System in Alexander Disease Astrocytes, and the Small Heat Shock Protein αB-Crystallin Reverses the Inhibition. *The Journal of Biological Chemistry*, 285(14), 10527–10537. doi:10.1074/jbc.M109.067975 PMID:20110364

Tian, R., Wu, X., Hagemann, T. L., Sosunov, A. A., Messing, A., McKhann, G. M., & Goldman, J. E. (2010). Alexander disease mutant GFAP compromises glutamate transport in astrocytes. *Journal of Neuropathology and Experimental Neurology*, *69*(4), 335–345. doi:10.1097/NEN.0b013e3181d3cb52 PMID:20448479

Tsuji, S. (2007). 4 - Leukodystrophies. In S. Gilman (Ed.), *Neurobiology of Disease* (pp. 43–49). Burlington: Academic Press. doi:10.1016/B978-012088592-3/50006-2

Turnbull, J., DePaoli-Roach, A. A., Zhao, X., Cortez, M. A., Pencea, N., Tiberia, E., ... Minassian, B. A. (2011). PTG depletion removes lafora bodies and rescues the fatal epilepsy of lafora disease. *PLOS Genetics*, *7*(4), e1002037. doi:10.1371/journal.pgen.1002037 PMID:21552327

van der Knaap, M.S., Naidu, S., & Breiter, S. N., Blaser, S., Stroink, H., Springer, S., ... Powers, J.M. (2001). Alexander disease: Diagnosis with MR imaging. *Ameican Journal of Neuroradiology.*, 22, 541–552. PMID:11237983

van der Knaap, M. S., & Valk, J. (2005). X-linked adrenoleukodystrophy. In U. Heilmann (Ed.), *Magnetic Resonance of Myelination and Myelin Disorders* (3rd ed.; pp. 176–190). Berlin: Springer. doi:10.1007/3-540-27660-2_21

von Jonquieres, G., Spencer, Z. H. T., & Rowlands, B. D. (2017). Uncoupling *N*-acetylaspartate from brain pathology: Implications for Canavan disease gene therapy. *Acta Neuropathologica*, *134*(627), 1–19. doi:10.100700401-017-1784-9 PMID:29116375

Wang, R. Y., Bodamer, O. A., Watson, M. S., & Wilcox, W. R. (2011). Lysosomal storage diseases: Diagnostic confirmation and management of presymptomatic individuals. *Genetics in Medicine*, *13*(5), 457–484. doi:10.1097/GIM.0b013e318211a7e1 PMID:21502868

Weleber, R. G. (1998). The dystrophic retina in multisystem disorders: The electroretinogram in neuronal ceroid lipofuscinoses. *Eye (London, England)*, *12*(3), 580–590. doi:10.1038/eye.1998.148 PMID:9775220

Wenger, D. A. (2000 Jun 19). Krabbe Dsease. In M. P. Adam, H. H. Ardinger, & R. A. Pagon (Eds.), *GeneReviews* (pp. 1993–2017). Seattle, WA: University of Washington.

White, A. B., Givogri, M. I., Lopez-Rosas, A., Cao, H., Breemen, R. V., Thinakaran, G., & Bongarzone, E. R. (2009). Psychosine Accumulates in Membrane Microdomains in the Brain of Krabbe Patients, Disrupting the Raft Architecture. *The Journal of Neuroscience*, *29*(19), 6068–6077. doi:10.1523/JNEU-ROSCI.5597-08.2009 PMID:19439584

Williams, R. E., & Mole, S. E. (2012). New nomenclature and classification scheme for the neuronal ceroid lipofuscinoses. *Neurology*, *79*(2), 183–191. doi:10.1212/WNL.0b013e31825f0547 PMID:22778232

Wippold, F. J., Perry, A., & Lennerz, J. (2006, May). Neuropathology for the Neuroradiologist: Rosenthal Fibers. *AJNR. American Journal of Neuroradiology*, 27(5), 958–961. PMID:16687524

Wisniewski, K. E., Kaczmarski, A., Kida, E., Connell, F., Kaczmarski, W., Michalewski, M., ... Zhong, N. (2017). Reevaluation of Neuronal Ceroid Lipofuscinoses: Atypical Juvenile Onset May Be the Result of CLN2 Mutations. *Molecular Genetics and Metabolism*, 66(4), 248–252. doi:10.1006/mgme.1999.2814 PMID:10191110

Worby, C. A., Gentry, M. S., & Dixon, J. E. (2008). Malin decreases glycogen accumulation by promoting the degradation of protein targeting to glycogen (PTG). *The Journal of Biological Chemistry*, 283(7), 4069–4076. doi:10.1074/jbc.M708712200 PMID:18070875

Yoshida, T., Sasaki, M., Yoshida, M., Namekawa, M., Okamoto, Y., Tsujino, S., ... Nakagawa, M. (2011). Nationwide survey of Alexander disease in Japan and proposed new guidelines for diagnosis. *Journal of Neurology*, 258(11), 1998–2008. doi:10.100700415-011-6056-3 PMID:21533827

Zeng, B. J., Pastores, M. G., Leone, P., Raghavan, S., Wang, H. Z., Ribeiro, A. L., Torres, P., ... Kolodny, H. E. (2006). Mutation Analysis of the Aspartoacylase Gene in Non-Jewish Patients with Canavan Disease. Academic Press.

Section 3

Therapeutic Interventions for Neurodegenerative Disorders

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ABSTRACT

Functional neurosurgery consists of procedures that either promotes judicious destruction or chronic stimulation of the nervous system in order to treat disordered behavior or aberrant function, as it is expected in neurodegenerative disorders ([NDs], e.g., movement disorders [Parkinson's disease, Tourette's syndrome, essential tremor, ballism, and dystonia]). Over the past 20 years, approximately 100,000 deep brain stimulation implant procedures have been performed worldwide. Neurosurgery is also a well-established therapeutic option for people with epilepsy whose seizures are not controlled by anti-epilepsy drugs. The most common pathological finding in patients with drug-resistant mesial temporal lobe epilepsy is hippocampal sclerosis. The aim of this chapter is to present the main NDs that can be treated through surgical procedures, and to describe the surgeries with a focus on the pathophysiology of diseases.

INTRODUCTION

Researchers have directed their efforts towards finding effective treatments for neurodegenerative diseases. The concept that neurological diseases are not treatable has been dissipated with the advances of the last decades. There is a great success with clinical therapeutics, genetic findings and immunology.

DOI: 10.4018/978-1-5225-5282-6.ch019

This chapter will show movement disorders and epilepsies that had their limiters and disabling results on patients altered by surgical procedures. Examples of diseases that can be treated by functional neurosurgery comprise movement disorders (Parkinson's disease [PD], Tourette's syndrome [TS], essential tremor [ET] and dystonia), spasticity, chronic pain, epilepsy and psychiatric disturbances (obsessivecompulsive disorders [OCD] and depression).

Over the past 20 years, approximately 100,000 deep brain stimulation implant procedures have been performed worldwide (Strauss et al., 2014). The efficacy of these functional procedures in the treatment of Parkinson's disease was verified up to 10 years after surgery (Moro et al., 2010-b). For patients who are compromised by such seizures, referral to an epilepsy surgery center should be strongly considered (Engel et al., 2003). Surgery is a well-established therapeutic option for people with epilepsy whose seizures are not controlled by anti-epilepsy drugs.

The aims of this chapter are to present the main NDs that can be treated through surgical procedures and to describe the surgeries with a focus on the pathophysiology of diseases.

BACKGROUND

The first image about neurosurgery is of a specialty that utilizes surgical techniques aimed at correcting structural / anatomical problems of the nervous system such as aneurysms, brain and spine tumors, fractures and herniated disks. For several years, this type of surgery was the only treatment option available for PD and movement disorders. These procedures were ablative in nature and consisted of surgeries performed on both the peripheral nervous system, including rhizotomies and sympathectomies (Teixeira and Fonoff, 2004), and in the brain, such as corticectomies, trans-ventricular accesses to the basal ganglia nuclei and even section of the cerebral peduncle, procedures that were performed under direct vision and not by a stereotactic approach (Speelman and Bosch, 1998), as the first stereotactic procedures for the treatment of movement disorders would only be performed by Spiegel and Wycis in 1947 (Speelman and Bosch, 1998).

These new perspectives, the "Functional Neurosurgery", are completely different. Functional neurosurgery consist of procedures that either promote judicious, localized destruction of a target area within the central nervous system, or chronic stimulation of specific structures in order to treat disordered behavior or aberrant function of the nervous system, as it is expected in NDs (Teixeira and Fonoff, 2004).

NEURODEGENERATIVE DISEASES WITH POSSIBLE SURGICAL TREATMENT

Movement Disorders

Parkinson's Disease

PD is a ND, characterized by movement disorders and non-motors symptoms (Salat et al., 2016), associated mainly with the death of dopaminergic neurons located in the substantia nigra, leading to progressive depletion of dopaminergic nigrostriatal and mesocorticolimbic neurons (Callesen et al., 2013), but also there is abnormal deposition of α -synuclein in the remaining cells and gliosis in specific areas of the nervous system (Lees et al., 2009; Salat et al., 2016). However, such changes are not restricted to these brain regions and can be found in other nuclei of the brainstem, in the cerebral cortex and even in peripheral neurons, such as those in the myenteric plexus. Non-dopaminergic pathways are also involved, including the serotoninergic and cholinergic neurons located in the spinal cord and peripheral nervous system, correlating mainly with the non-motor symptoms of the disease (Poewe, 2008).

Since its original description, the clinical diagnosis of PD has centered on a defined motor syndrome. The prerequisite for applying the MDS-PD criteria is the diagnosis of parkinsonism, which is based on three cardinal motor manifestations. Parkinsonism itself is defined as bradykinesia, in combination with either rest tremor, rigidity, or both (Postuma et al., 2015). However, over the years the non-motor manifestations became better characterized and can now be identified in most, if not all, patients (Seppi et al., 2011). The non-motor manifestations of PD comprise autonomic, neuropsychiatric dysfunctions, changes in the sleep-wakefulness cycle and in pain perception (Poewe, 2008; Goldman et al., 2015).

Dopamine replacement therapy, currently the most efficacious treatment for PD, so much so that it's considered the gold standard, mainly comprises the use of the dopamine precursor levodopa, either isolated or in combination with dopamine agonists, monoamine oxidase B inhibitors, and catechol-O-methyltransferase inhibitors. Long-term use of levodopa may result in several complications, including motor fluctuations and levodopa-induced dyskinesia, which cause serious distress to patients. Moreover, symptoms that appear at the later stages of PD are often not responsive to dopaminergic treatments. (Du and Chen, 2017).

Dystonia

In 1984 an *ad hoc* committee of Scientific Advisory Board of the Dystonia Medical Research Foundation developed the modern concept of dystonia (Teive et al., 2001-b; Albanese et al., 2013). Although the 1984 definition was seminal, over the following decades several shortcomings have been recognized. Therefore, in view of these limitations of the 1984 definition, an international Consensus Committee proposed in 2013 the following revised definition: Dystonia is a movement disorder characterized by sustained or intermittent muscle contractions causing abnormal, often repetitive, movements, postures, or both. Dystonic movements are typically patterned, twisting, and may be tremulous. Dystonia is often initiated or worsened by voluntary action and associated with overflow muscle activation (Albanese et al., 2013).

If defining dystonia is difficult and controversial, classifying the various forms of dystonias is a much more complex task, primarily because the term dystonia can mean not only a disease, but also a symptom that can be part of many disorders with a wide range of causes. In an attempt to clarify the term dystonia, three "surnames" for dystonia were proposed: "symptom", "movement" and "disorder". A patient may complain of dystonia if, for example, he has a twisted neck. Therefore, the patient has a dystonia symptom (dystonia^{Sx}) and on examination the signs of dystonia may be confirmed. This patient then has a dystonia movement (dystonia^{Mov}). Finally, dystonia as a disorder (dystonia^{Dx}) requires a clinicopathologic understanding of the underlying etiology of the disease: genetic, late-onset, post-traumatic, etc (Frucht, 2013; Camargo et al., 2015). These new concepts led to the replacement of the 1998 dystonia classification (Albanese et al., 2013) with the one proposed in 2013. In the 2013 classification, dystonias are subdivided according to clinical features and etiology.

Dystonia remains a challenging field in both diagnostic and therapeutic aspects. Further understanding of its pathophysiology may shed light on more specific therapies. Three main approaches are employed in the treatment of dystonia: pharmacological therapies, botulinum toxin injection (BoNT) and surgical

interventions (Termsarasab et al., 2016). Treatment of dystonic symptoms with botulinum toxins is now a major area of medical practice. Botulinum toxins are established as the first line of treatment for cervical dystonia, blepharospasm, and laryngeal dystonia and are considered a major treatment modality for limb dystonias, including task-specific dystonias (Jabbari, 2016). Four major categories of medications are most commonly used: anticholinergics (particularly trihexyphenidyl), baclofen, benzodiazepines (particularly clonazepam), and dopamine-related medications. Symptomatic medical therapy can improve quality of life and should not be overlooked (Termsarasab et al., 2016).

Tourette's Syndrome and Tics

There is no specific diagnostic test for TS and diagnosis is made solely on clinical grounds. According to the Diagnostic and Statistical Manual of Mental Disorders (DSM-V) it is defined by:

- 1. Multiple motor tics and one or more vocal tics.
- 2. Tics occur several times a day, almost every day or intermittently for one year or more.
- 3. Onset of tics must occur before the patient is 18 years-old.
- 4. The disorder is not due to the use of drugs nor to another disease (APA, 2013).

It is a quite complex neurobehavioral disorder in which patients may present with coexistent obsessive-compulsive disorder (OCD), attention deficit hyperactivity disorder (ADHD), and other behavioral comorbidities (Teive et al., 2001-a). It has been much maligned amid laypeople as one of its most striking features, the occurrence of coprophenomena (coprolalia and/or copropraxia), has been described even in the first historical accounts, regardless of its low prevalence (around 14 - 19%), but carries great social impact. (Germiniani et al. 2012).

A meta-analysis of antipsychotics for the treatment of TS found no difference in efficacy among risperidone, haloperidol, pimozide, and ziprasidone (Weisman et al., 2013). Side effects such as sedation, weight gain, and hyperprolactinemia have limited their tolerability (Thenganatt and Jankovic, 2016). A systematic review of aripiprazole for TS found a similar efficacy of aripiprazole and haloperidol (Yoo et al., 2013). Tetrabenazine has been shown to be effective in open-label trials for the treatment of TS. Alpha agonists such as clonidine have been found to be useful in the treatment of mild tics and may have a particular benefit in patients with co-existing ADHD and impulse control disorder. Botulinum toxin may be helpful in the treatment of focal motor tics (Thenganatt and Jankovic, 2016).

Ballism

Jakob (1923) was the first to demonstrate the relationship between hemiballism and injury to the contralateral subthalamic nucleus, coining the term Luy's body syndrome (Shannon, 1998). Ballism is a rare hyperkinetic movement disorder characterized by an abnormal, choreic, large-amplitude, violent, pitching involuntary movement using the proximal appendicular and associated axial musculature (Vidakovid et al., 1994; Shannon, 1998). A distinction must be made between ballism and other hyperkinetic movement disorders, mainly chorea (Coral et al., 2000).

Treatment is based on neuroleptics, mainly haloperidol, and in selected patients chlorpromazine and pimozide. Other drugs such as tetrabenazine, reserpine and clonazepam can be used. Clinical treatment

is supported by few open-label studies with small samples (Dewey and Jankovic, 1989; Vidakovid et al., 1994).

Essential Tremor

Essential tremor (ET) is among the most common movement disorders, and the most prevalent tremor disorder (Louis, 2014). Tremor itself is not a featureless, nondescript action tremor; rather, it is characterized by a specific pattern of features:

- 1. Kinetic tremor is usually greater in amplitude than postural tremor.
- 2. Tremor involves movement at specific joints in specific directions.
- 3. Intention tremor of the arms is seen on finger-nose-finger maneuver in approximately 50% of cases.
- 4. Rest tremor occurs in as many as 20% of cases as a late feature.
- 5. Arm tremor precedes cranial tremor, for which there is a female preponderance.
- 6. The prevalence of neck tremor is greater than that of jaw tremor, which is greater than that of tongue/cheek/forehead tremor.
- 7. There is a tendency for tremor severity to increase over time. (Louis, 2014)

Although the pathophysiology of ET is only partially understood; the notion that ET may be neurodegenerative is one that has been proposed recently (Louis, 2014). An emerging topic is related to how substantial is the role played by the cerebellum in the pathophysiology of underlying movement control pathways in ET. Cerebellar-like symptoms, with abnormalities in tandem gait and balance, have been repeatedly described in ET patients. Intention (i.e., "cerebellar") tremor of the arms occurs in approximately one-half of ET patients, and there are a variety of other motor abnormalities that point to what is likely to be a more pervasive underlying abnormality of cerebellar function in ET. As for the thalamus, it is considered to be a cerebellar outflow region and a final common nodal point for diatheses-affected pathways that emerge from more proximal points within the cerebellar system in ET (Louis, 2014).

Drugs thought to improve tremor include beta-adrenergic antagonists, primidone, topiramate, ethanol, and benzodiazepines (Troiano et al, 2004). Less consistent efficacy is reported with many other medications, usually anti-epileptic drugs (AEDs) (Ondo et al., 2016).

Epilepsy

Epilepsy is a common condition. According to the World Health Organization, epilepsy accounts for 1% of the global burden of disease, more than breast cancer in women and almost as much as lung cancer in men (Murray et al., 2012). It is estimated that as many as sixty-five million people around the world have epilepsy (Sirven and Shafer, 2014).

The International League Against Epilepsy (ILAE) in 2014 recommended broadening the definition of epilepsy to include several different etiology groups, as well as epileptic encephalopathies. Therefore, epilepsy is a disease of the brain defined by any of the following conditions: 1) at least two unprovoked (or reflex) seizures occurring more than 24 hours apart from each other; 2) one unprovoked (or reflex) seizure and a probability of further seizures similar to the expected general recurrence risk after two unprovoked seizures (at least 60%), occurring over the next 10 years (Fisher et al., 2014).

In April 2017 the ILAE presented a revised operational classification of seizure types. Changes include the following:

- 1. "Partial" becomes "focal".
- 2. Awareness is used as a classifier of focal seizures.
- 3. The terms dyscognitive, simple partial, complex partial, psychic, and secondarily generalized are eliminated.
- 4. New focal seizure types include automatisms, behavioral arrest, hyperkinetic seizures, autonomic, cognitive, and emotional seizures.
- 5. Atonic, clonic, epileptic spasms, myoclonic, and tonic seizures can be of either focal or generalized onset.
- 6. Focal to bilateral tonic–clonic seizure replaces the term secondarily generalized seizure.
- New generalized seizure types are absence with eyelid myoclonia, myoclonic absence, myoclonicatonic, myoclonic-tonic-clonic seizures.
- 8. Seizures of unknown onset may have features that can still be classified. The new classification does not represent a fundamental change, but allows greater flexibility and transparency in naming seizure types (Fisher et al., 2017).

Mesial Temporal Sclerosis

Studies have shown that in some patients, the development of epilepsy does not stop at the time of diagnosis and that recurring seizures may contribute to the progression of the disorder, so that to that effect, progressive reduction of hippocampal volumes documented over time on brain MRI, correlates with lower neuron cell counts (Bronen et al., 1991; van Paesschen et al., 1997; Devinski et al., 2013).

Temporal lobe epilepsy (TLE) is the most common form of focal (partial) or location-related epilepsy - it accounts for approximately 60% of all people living with epilepsy (Holmes et al., 2017). There are two types of TLE, one which involves the medial or internal structures of the temporal lobe; while the second, called neocortical temporal lobe epilepsy, involves the outer portion of the temporal lobe. The most common version of these two types is mesial temporal lobe epilepsy (MTLE) (Holmes et al., 2017).

Many patients with MTLE develop pharmacoresistance to antiepileptic drugs, and the most common pathological finding in patients with drug-resistant MTLE is hippocampal sclerosis (HS). HS is a unique pathologic condition characterized by specific neuropathologic features (Berg et al., 2010).

An ILAE task force reviewed previous classification schemes and proposed a system based on semiquantitative hippocampal cell loss patterns that can be applied to any histopathology laboratory and classified three types in anatomically well-preserved hippocampal specimens (Blümcke et al., 2013).

PATIENT SELECTION FOR SURGERY

Surgery for Patients With Movement Disorders

Parkinson's Disease

Traditionally, surgery has been reserved for patients with advanced PD, when medications no longer deliver the desired effect or induce extremely debilitating side effects. A multi-center study evaluated the effect

of surgery in patients less than 60 years of age, Hoehn and Yahr score below 3 and with shorter duration of motor fluctuations (Schüpbach et al., 2013). The primary outcome measure considered was quality of life of the patients and the results suggest that surgery should be considered in this specific population, with very favorable results, although there are still insufficient data to consider that surgical procedures may have some neuroprotective effects or even slow the rate of disease progression (Benabid, 2007).

Surgery should not be offered for patients with short disease time, thus avoiding surgery neither in patients who may eventually develop atypical parkinsonism and not have idiopathic PD, nor for patients with very advanced age and disease, as in these populations surgery will probably have no long-term improvement in quality of life, but can also add unnecessary risks to the patient. More specific criteria such as disease length, minimum or maximum age, and clinical staging of the disease based on appropriate rating scales are still controversial.

A set of very specific criteria adequately selects the possible candidate for surgery for Parkinson's disease, thus providing a better therapeutic result, while also decreasing the incidence of possible complications.

- 1. Certainty about the diagnosis of idiopathic PD despite modern and innovative technologies, both in the field of radiology with high-resolution magnetic resonance imaging (Lotfipour et al., 2012), ultrasonography (Bor-Seng-Shu et al., 2010), PET-scan (Stoessl, 2014) and available biomarkers (Sharma et al., 2013), the diagnosis of PD is still based on clinical criteria and observation of the patient's evolution as the disease progresses.
- 2. Unquestionable response to levodopa therapy, at least in the early years and early stage of the disease. Another important thing to consider is the asymmetric onset with progressive evolution of the symptoms, which is suggestive of idiopathic PD. (Brooks DJ).
- 3. Improvement in motor scores (sub-item 3) of the Unified Parkinson Disease Rating Scale (UPDRS) (Goetz et al., 2008) of at least 30% with the levodopa challenge test comparing the "on" and "off" phases. An exception to this criterion, are patients who present totally incapacitating tremor, which, by itself, already justifies the indication of the procedure (Munhoz et al., 2014).
- 4. A score equal to or less than 4 on Hoehn and Yahr Scale (Hoehn and Yahr, 1967).
- 5. Identification of motor symptoms that are responsive to levodopa and by consequence, will improve with surgical treatment (Munhoz et al., 2014).
- 6. Absence of a structural lesion in preoperative neuroimaging exams that may make it impossible to structurally identifies or physiologically locates the therapeutic target. Patients who have undergone previous ablative or neuromodulatory neurosurgical procedures can be reoperated, provided they are correctly evaluated (Ellis et al., 2008)
- 7. In cases of previous psychiatric history, the patient should be evaluated by a qualified professional, who will determine if the patient is able to understand and mainly collaborate with the surgical procedure (Voon et al., 2006).
- 8. In cases where there is suspicion of, or caregivers report symptoms of cognitive impairment, the patient should undergo a careful preoperative neuropsychological evaluation (Voon et al., 2006)...

Hyperkinetic Movement Disorders

Idiopathic dystonia – both generalized and segmental – is the best indication for deep brain stimulation (DBS), while DBS in for either focal or secondary dystonia is not sufficiently supported by clinical

evidence (Moro et al., 2013). Surgery is indicated when medical therapy is no longer able to provide sufficient improvement of disability and quality of life. Younger patients with idiopathic dystonia and shorter duration of disease seem to benefit the most from DBS. The mechanisms by which DBS leads to symptomatic improvement in dystonia are far from being clear and well understood at present, which might at least partially explain the variability and unpredictability of clinical success of DBS (Mehdorn, 2016).

Essential tremor is the most common movement disorder. The severity of tremor is measured objectively using scales such as the Fahn–Tolosa–Marin Tremor Rating Scale or the Essential Tremor Rating Scale. Having the patient draw spirals or write his/her name is a rapid way of assessing tremor during the awake period of surgery, or on follow-up during routine DBS programming in the office (Larson, 2014).

The suitability of DBS in TS patients depends on many factors, yet there is much variability regarding specific inclusion and exclusion criteria across studies. Typically, core criteria focus on accurate diagnosis, high tic severity (typically a Yale Global Tic Severity Scale [YGTSS] score >35/50) and resistance to at least three different pharmacological agents (Temel and Visser-Vandewalle, 2004). Although psychiatric co-morbidity is a common component of TS, motor and vocal tics should be the main source of disability prompting surgical evaluation. Exclusion criteria typically include major poorly-controlled psychiatric disorders, pregnancy, current substance abuse or dependence, severe cognitive impairment and structural brain abnormalities on MRI (Akbarian-Tefaghi et al., 2016; Temel and Visser-Vandewalle, 2004).

Choice of the Therapeutic Target

Idiopathic Parkinson's Disease

With increased knowledge of PD it is clear that there is no more room for just one single surgical target to treat all symptoms of this multi-faceted disease. (Okun and Foote 2010). The adequate choice of the surgical target must be made individually, depending on the idiosyncrasies of each case. The targets that present the highest degree of evidence and efficacy in the treatment of PD are the GPi and the STN. Stimulating the STN would lead to a higher rate of improvement in the cardinal symptoms of the disease, reducing the dose of dopaminergic equivalents in the immediate postoperative period. Nevertheless, if this withdrawal is made somewhat abruptly, severe bouts of depression may occur, even leading to higher risk of suicide (Voon et al., 2008). There was also an increase in battery life of the neurostimulator in cases of STN, considering that the parameters used were much lower, than those of the GPi. The great advantage of targeting the GPi was restricted to those patients who had extremely pronounced dyskinesia, or previous cognitive deficits, as the use of the STN in these latter cases could lead to a marked worsening of some cognitive functions, more specifically those related to verbal fluency (Follett et al., 2010).

A recent meta-analysis showed that both targets present practically similar results when comparing pre and postoperative motor scores of the sub-item III of the UPDRS scale, corroborating the findings of other groups (Anderson et al., 2005; Nakamura et al., 2007; Weaver et al., 2012), both in bilateral and unilateral procedures. The concept that the GPi is safer from a cognitive point of view has been contested in a recent study comparing STN stimulation with GPi in patients with advanced PD (Odekerken et al., 2013), although the results presented were challenged because of low statistical power to avoid a type II error (Montgomery, 2013), showing how controversial this subject still is. The vast majority of centers that perform DBS surgery use the STN as a target, but this choice is based primarily on open clinical trials, personal conviction, as well as personal experience (Laitinen et al., 1992).

While surgical procedures performed on the STN and the GPi have shown consistent and effective results in the treatment of the cardinal symptoms of PD, their effect on axial symptoms, such as gait and balance, is at best irregular, if not disappointing (St George et al., 2010). The identification of numerous connections between the peduncle-pontine nucleus (PPN) and structures as distant as the cerebral cortex and the spinal cord, associated with the recognition of its extremely important role in gait control, rose the interest for this new target as the ideal one for treatment of axial disorders. Despite the lack of controlled and blinded trials, series with good results on a greater or lesser scale are found in the literature (Moro et al., 2010-a; Thevathasan et al., 2010). Additionally, the isolated stimulation of the PPN is arguably a limiting factor in the improvement of these patients, and some authors proposed that the combined and concomitant stimulation of both the PPN and the STN (Stefani et al., 2009; Khan et al., 2010) hoping that the synergistic and adjuvant effects between the two targets could concurrently improve both appendicular and axial symptoms of the disease. Despite these results, the isolated stimulation of the PPN is still considered experimental in many centers, due to the lack of well-designed trials demonstrating its real efficacy. The main side effects of PPN stimulation are dysesthesias, which usually occur by spreading of the electrical current, with undesired stimulation of structures such as the medial lemniscus and the spinothalamic tract.

The *nucleus ventralis intermedius* (VIM) of the thalamus was for many years the target of choice in the surgical treatment of PD, especially in those patients with incapacitating tremor. Considering that improvement of other cardinal symptoms of the disease, with the exception of rigidity, is poor in those patients who had undergone VIM procedures, the VIM currently plays an extremely limited role in the neurosurgical arsenal, restricting itself only to those patients with unilateral PD of extremely slow evolution and with otherwise intractable tremor. In patients with tremor as the main manifestation, but accompanied by other debilitating symptoms, such as marked bradykinesia and rigidity, the STN is the best target (Hariz et al., 2008).

An area that encompasses the region of the *zona incerta* (Zi) and the pre-lemniscal radiation (Raprl) known as the posterior subthalamic area (PSA) has been used more frequently in the treatment of refractory tremor (Sandvik et al., 2012). The Raprl in patients with PD was used in an attempt to alleviate the tremor and rigidity of patients with advanced disease, with improvement maintained even after 2 years (Velasco et al., 2001). In the same trial, the authors commented that Raprl can be approached by both the neuromodulatory and ablative methods, but that this would have the disadvantage of causing worsening of the patient's bradykinesia. Six years after this publication, where they initially emphasized only improved tremor and rigidity, the same group reported improvement in all cardinal symptoms of the disease (Carrillo-Ruiz et al., 2008). As there was no significant change in stimulation parameters or electrode design, this phenomenon is apparently related to the positioning of the most proximal poles in a more dorsal or dorsal-medial location in relation to the STN, which would include the more caudal portion of Zi.

Other targets like the motor cortex (Arle et al., 2008), the centromedian / parafascicular / habenullae complex of the thalamus (Benabid, 2009; Stefani et al., 2009) and the posterior aspect of the spinal cord (Thevathasan et al., 2010) were anecdotally tried, with a small number of cases and inconsistent results. One clinical trial showed a significant improvement in gait scores and postural stability in 15 patients with PD who underwent epidural spinal cord stimulation (Agari and Date, 2012). However, the main indication was chronic pain in the lumbar region with irradiation to the lower limbs.

Hyperkinetic Movement Disorders

Despite the fact that the GPi had been considered the optimal target for DBS in dystonic patients, the optimal target needs to be considered individually in every single patient depending on the most predominant symptom, and sometimes more than one surgical target may be required to optimize post-surgical results. STN DBS might be an alternative to GPi DBS, and VIM DBS in severe refractory dystonic tremor has also been suggested (Mehdorn, 2016).

Historically, the most commonly used target for DBS in ET is the VIM. Several studies report significant improvement with both unilateral and bilateral VIM stimulation, with long-term follow-up showing reduction in tremor severity rates ranging from 40% to 80% with corresponding improvement in quality of life (Huss et al., 2015). Other targets have emerged as an alternative to the VIM for patients with ET. Currently, the most widely studied area is the one inferior to the thalamus, and posterior and superior to the STN. Several other targets have been stimulated by different centers in this region, including the Zi and the prelemniscal radiations, although it is difficult to determine exactly where the mechanism of action is taking place, as is the case with most DBS targets. A broader anatomic term that incorporates both of these targets is the posterior subthalamic area (PSA). This is a particularly interesting region, not only because it appears to provide tremor reduction rates comparable to those of the VIM, but it may also have a lower adverse effect profile, better efficacy with proximal and intention tremor, and may avoid the development of "tolerance" seen with VIM stimulation over time (Larson, 2014).

In general terms, appendicular tremor responds better than voice tremor or midline tremor, the latter of which usually requires bilateral surgery (Obwegeser et al., 2000). We usually indicate unilateral surgery, aimed at improving the tremor on the more symptomatic side, or on the dominant side, depending on the patient's preference. Bilateral surgery is performed as a staged procedure in highly selected cases of severe bilateral, head or voice tremor. Although DBS is a non-ablative surgery, bilateral surgery, especially if simultaneous, has a specific risk of causing speech disorders and/or ataxia. Bothering induced "paresthesia" is also a relatively common side effect, due to spreading of the current in a posterior direction to the ventro caudalis (VC) nucleus of the thalamus. Many of these side effects can be bypassed with adjustments in generator programming.

Tractography (DTI) has been established as a powerful tool for understanding the functioning of the central nervous system, as it is virtually the only non-invasive method of visualizing structural connectivity in the brain (Calabrese, 2016). With the development of longer electrodes, with a greater number of contacts and the incorporation of tractography, it became possible to identify the entire projection of the dentate-rubro thalamic tract all the way up to VIM. The adequate placement of a longer electrode in a trajectory, which includes in its cranial portion the VIM and its more caudal portion the posterior subthalamic area, allowing the use of two distinct areas with neurophysiological characteristics (the posterior subthalamic area is composed in its majority by axonal fibers, whereas the VIM is made up mostly by neuron cell bodies). (Coenen et al., 2014; Tasker and Kiss, 1995). In cases of refractory tremor, the use of concomitant neuromodulation of these two distinct anatomical and neurophysiological targets has been proven to be effective in the control of the drug-resistant tremor, and they can be stimulated either isolated or in combination. Also, as the stimulation of white fibers usually requires lower voltages, the feasibility of saving the battery of the generators is highly likely to be seen over time (Fenoy and Schiess, 2017).

Currently, nine targets have been used for TS, including the thalamic-centromedian-parafascicular complex (CMPf), the cross point of the centromedian nucleus-substantia periventricularis-nucleus ventro-oralis nucleus (CM-Spv-Voi), the target of the nucleus ventro-oralis and posterior-ventro oralis anterior-

Voi complex (Vop-Voa-Voi), the GPi (anteromedial and posteroventral regions), the nucleus accumbens (NA), the anterior limb of the internal capsule (ALIC), the STN and the globus pallidus externus (GPe). The rationale behind each of these target choices has varied depending on whether tics are considered a movement disorder in which case sensorimotor areas such as the posteroventral pallidum have been stimulated, or if they are considered to be the result of either a compulsion or a failure of inhibition, wherein associative/limbic areas have been targeted. The thalamus and GPi have been the most widely stimulated with a combination of targets used in some studies (Akbarian-Tefaghi et al., 2016).

Surgery for Patients with Epilepsy

Drug resistant epilepsy may be defined as failure of adequate trials of two appropriately chosen antiepileptic drugs (AED) schedules, used in appropriate therapeutic dosages in order to achieve sustained seizure freedom, and well tolerated (with no disabling side-effects), whether as monotherapies or in combination (Kwan et al., 2010).

For patients who are compromised by such seizures, referral to an epilepsy surgery center should be strongly considered (Engel et al., 2003). Surgery is a well-established therapeutic option for people with epilepsy whose seizures are not controlled by anti-epilepsy drugs.

NEUROSURGERY

Movement Disorders

Laitinen et al. (1992) published a series of 38 patients with advanced PD submitted to stereotactic posteroventral pallidotomy with excellent results in improving all cardinal symptoms of the disease, but mainly in the control of dyskinesias caused by chronic levodopa therapy. The results of this paper, combined with the development of more accurate stereotactic systems, the improvement of neuronal activity recording systems and intraoperative stimulation, new modalities of neuroradiology and, mainly, a better understanding of the pathophysiology of the base nuclei through of experimental studies on primates (Alexander et al., 1990; DeLong, 1990), led to a resurgence of stereotactic surgery for PD.

Further studies demonstrated the efficacy of ablative procedures either at the GPi (Lang et al., 1997; Vitek et al., 1997), or at the STN (Alvarez et al., 2001; Patel et al., 2003). These ablative procedures proved to be safe and efficient, and the excellent results presented by the Grenoble group in 1993 with chronic electrical stimulation of the STN (Pollak et al., 1993), led to establish this procedure as the gold standard for the surgical treatment of PD, despite preliminary good results with GPi stimulation (Anderson et al., 2005). The efficacy of these procedures in the treatment of PD was verified up to 10 years after surgery (Moro et al., 2010-b).

Deep Brain Stimulation

Technical Aspects of the Surgical Procedure

Two stereotactic techniques are available for performing the surgical procedure: "frame" or "frameless" (Machado et al., 2006). The first is the more classical technique, while the second one is more recent

and has shown promising results in the field of stereotactic procedures, especially regarding the patient's comfort during the procedure itself. Although some authors still infer that the use of the stereotactic frame seems to offer a greater accuracy in localizing and determining the target (Bjartmarz and Rehncrona, 1990), the development of the frameless technique and the training of the professionals who execute it, with their respective learning curves, may reduce or even nullify this difference in the future (Fukaya et al., 2010; Kelman et al., 2010; Tai et al., 2010).

The stereotactic frame is placed immediately before the surgery and following the frame placement, the patient is referred for preoperative imaging, which may consist of computed tomography (CT), MRI, ventriculography or a combination of these. The authors perform the acquisition of stereotactic images with a CT device, with volumetric acquisition and 1.25 mm thick slices, due to lower image distortion than MRI (Sumanaweera et al., 1995). The stereotactic study is then fused with the preoperative MRI, using designated softwares. MRI images are also obtained volumetrically, in the axial planes, with contrast-enhanced T1-weighted, T2-weighted and Inversion Recovery (IR) sequences acquisitions (Figures 1 and 2). The last two are used for delimitation by direct visualization of the targets and adjacent structures, which is a key aspect of the procedure, providing meaningful information that improves accuracy, while avoiding stimulation induced side effects (Machado et al., 2006). Contrast-enhanced T1 acquisition allows visualization of vessels from both the surface and depth of the brain, which helps in determining the safest path through the brain to the therapeutic target, reducing the risk of intraoperative bleeding and avoiding eloquent brain areas (Starr, 2002).

Targets

The most frequently used targets in the surgical treatment of Parkinson's disease are the subthalamic nucleus (STN), the globus pallidus internus (GPi), and the ventro-intermediate nucleus of the thalamus (VIM), with the best clinical results being obtained when the electrode or lesion are located in the sensory-motor portion of the corresponding target. In the case of the STN, the target must be located in its dorsolateral region (Herzog et al., 2004), whereas in the GPi, in its most lateral postero-ventral aspect (Rezai et al., 2008).

Direct Method of Target Determination

This method is based on direct visualization of the nucleus and its adjacent structures in order to precisely locate the ideal target. The boarders of the STN and adjacent white matter are well delimited in T2-weighted acquisitions (Figure 1). The red nucleus, also easily identifiable in this sequence, is posteromedially located to the STN, and its anterior border can be used as the anterior limit of the anteroposterior coordinate of this target (Machado et al., 2006).

The GPi and optic tract can be visualized in IR acquisitions - the optic tract is easily recognized, particularly in coronal sections in the intersection plane of the mamillary bodies, its dorsal border being adopted as the lateral and inferior limits of the GPi (Figure 2) (Starr, 2002). This coordinate serves, in fact, to facilitate intraoperative stimulation (visual response) and is not necessarily the ultimate target. Often the ablative therapeutic lesion or implant is located discretely in a more lateral position, to avoid the internal capsule, or superiorly depending on intraoperative stimulation findings.

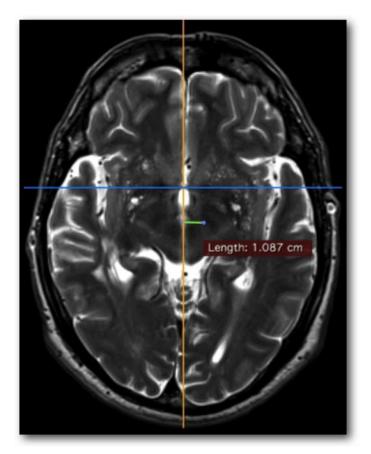


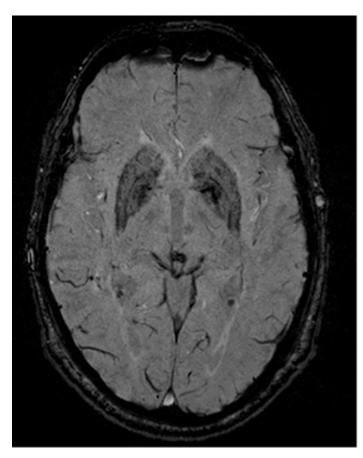
Figure 1. Direct method of target determination - STN. T2-weighted brain MRI, axial plane, with STN visualization at 10.87 mm from the midline.

With the image restrictions still found with conventional MRI machines used for clinical purposes, either with 1.5 or 3 T fields, the VIM is still not precisely delimited, and its identification is performed only by using an indirect method, particularly as visualization of the internal capsule in MRI scans may help in determining the lateral coordinate of the VIM. This limitation may be overcome using unique and specific sequences with the 1.5 T MRI (Vassal et al., 2012) or with future widespread availability of new 7T-MRI (Lenglet et al., 2012).

Indirect Method of Target Determination

This method uses predetermined coordinates, calculated by obtaining the distances between the targets to the mid-commissural point (MCP). The main limitation of the method is the considerable variation between the different stereotactic atlases, developed through a reduced number of brain samples (Rezai et al., 2008). The delimitation of the ideal coordinates is often elaborated from the average of the coordinates with the best clinical response, based on the experience of different centers.

Figure 2. Direct method of target determination – Gpi. IR sequence brain MRI, axial plane, with detailed visualization of both the GPi and the pallido-capsular transition.



Deep Brain Stimulation Systems Implantation

Before the insertion of the microelectrodes, the patient is examined by the neurologist to determine the baseline values of the UPDRS, rating the cardinal symptoms, namely rigidity, bradykinesia and tremor. The microelectrode is then introduced at 10 mm above the target and recording starts, with progression performed every 1 mm, for further microstimulation. The objective is to obtain a trajectory with adequate nuclear extension (STN, GPi or VIM), with adequate recording of sensory-motor driving (cellular activation with passive limb movement) and determination of the borders of the surgical target by the identification of adjacent structures.

Following microregistration and microstimulation, the electrode is positioned at the ventral edge of the ideal target location, defined by the combination of the anatomical target and the refinement of the microrecording. The procedure is monitored by fluoroscopy in order to confirm the positioning and to avoid movement or migration of the electrode during this whole phase. (Machado et al., 2006)

During stimulation of STN, it is possible to observe a reduction in rigidity, bradykinesia and tremor, in addition to defining the thresholds of adverse effects such as contractions of the limbs and/or face (stimulation of the cortico-spinal tract, situated anteriorly), paresthesias (postero-medialy) and conju-

gated deviation of the eyes (due to unwanted stimulation of the third nerve nucleus, situated inferiorly). For practical purposes, improvement of rigidity of the contralateral wrist (the Pollak-Limousin sign) with the lowest possible current without side effects, is the final goal of the macro-stimulation (Benabid et al., 2000). In some instances, contralateral dyskinesias, especially in the foot, can be observed, and they usually correlate with good surgical outcome. In procedures performed in the GPI, the goal is to improve contralateral rigidity and bradykinesia. However, in this case the latency between the start of stimulation train and the evidence of intraoperative clinical improvement is much higher than that observed with stimulation of the STN. In a considerable number of patients improvement of symptoms is often not observed intraoperatively and the role of intraoperative macrostimulation consists essentially in characterizing side effects resulting from stimulation of adjacent areas, such as stimulation of the cortico-spinal tract (postero-medially) and the optic tract (located below) (Kumar et al., 1998). When the surgical target is the VIM, intraoperative stimulation aims at obtaining total tremor control with the lowest possible voltage, while also avoiding side effects, notably paraesthesia (which denotes a very posterior position of the tip of the lead, close to the ventro-caudal nucleus of the thalamus or of the posterior lemniscus), dysarthria and compromise of contralateral movement (demonstrating a lateral position, very close to the internal capsule) (Kiss et al., 2003). As a safety parameter, a threshold of 4 volts for inducing transient side effects is considered adequate for the implantation of the electrode in the STN, whereas for the Gpi, which normally requires higher stimulation values for a better clinical result, a threshold of 1 to 2 volts above the values with clinical improvement is considered satisfactory.

Neuroablative Procedures: Pallidotomy and Thalamotomy

Thalamotomy (Fox et al., 1991; Linhares and Tasker, 2000) and pallidotomy (Iacono et al., 1995; Lang et al., 1997) are well established procedures with consistent results, but the potential for adjustability and reversibility of the neuromodulatory procedure led to a progressive replacement of these procedures by electrical stimulation of deep brain structures. In spite of these advantages, the neuromodulatory procedure is by no means free of limitations. Implanted devices are expensive, create greater interdependence between the patient and the medical center and need to be replaced at fixed intervals, even after the development of rechargeable pulse generators. The main disadvantages of the ablative method are the potential of causing an injury that is unduly localized and also the limitation in performing bilateral procedures. Nonetheless, ablative procedures may still play a role as a therapeutic surgical option in PD, especially in cases where the patient has difficulty attending routine evaluations, as is normally required in patients who have been implanted with neuromodulation devices or in patients with a recurrent history of infections (Pérez-Suarez et al., 2017).

The use of MRI-guided high intensity ultrasound devices has been the subject of recent research for the treatment of some neurological diseases. It consists in performing encephalic therapeutic lesions with real-time intraoperative monitoring by MRI-guided thermometry, causing temporary subclinical lesions, which allow us to evaluate both potential therapeutic and collateral effects prior to performing the definitive lesion, therefore increasing the safety profile of a procedure that is eminently ablative. Although most of the patients with movement disorders treated by this new technology were patients with essential tremor (Elias et al., 2013; Lipsman et al., 2013), it shows great potential for use in other movement disorders and even in psychiatric disorders, mostly due to its safety profile.

Radiosurgical ablative procedures, such as those using devices as the Gamma Knife[®], still do not have a well-defined safety profile for their therapeutic use in procedures in either the GPi or the STN (Bronstein et al., 2013). In some particular situations, the procedure may be performed in patients who require an unilateral thalamic procedure and who are at high risk of developing postoperative complication with the implant of an electrical stimulation device.

Epilepsy

The different surgical approaches used for mesiotemporal epilepsy are standard anterior temporal lobectomy, anterior temporal (also known as key-hole resection), extended lesionectomy, and transsylvian and subtemporal selective amygdalohippocampectomy. Regardless of the selected surgical technique, each and all of them resulted in similar epileptological and neuropsychological results, as well as complication rates. The surgical approach for an individual patient should be tailored according to not only the specific localization of the epileptogenic area, but also based on the experience of the surgeon (Engel et al., 2003; Lopes et al. 2017).

MANAGING PATIENTS TREATED WITH SURGICAL PROCEDURES

Movement Disorders

Postoperative MRI to locate the electrode contacts and/or lesions is obtained only 1 month after the procedure so that any pneumatocele or hemorrhage is absorbed, reducing the risk of brain shift (Figure 3).

Postoperative T2-weighted brain MRI, axial plane, showing the electrode suitably positioned in the left STN topography.

Epilepsy

Approximately two-thirds of patients become seizure free after anterior temporal lobectomy, except for simple partial seizures, whose effects on the patient's quality of life are so mild, that their persistence is acceptable, with no further clinical compromise. This outcome was found in a large number of Class IV series, and was confirmed in a randomized, controlled trial of surgery versus antiepileptic drug therapy. However, ten to 15% of refractory patients remain unimproved after surgery (Engel et al., 2003; Lopes et al., 2017).

For those patients who persist with seizures after resective epilepsy surgery, further available surgical options have insufficient evidence to make firm conclusive statements on their efficacy and safety. These new techniques are all functional, stimulation-based techniques, such as hippocampal deep brain stimulation (DBS), centromedian thalamic DBS, nucleus accumbens DBS and cerebellar stimulation. There is a need for more, larger and well-designed randomized controlled trials to validate and optimize the efficacy and safety of any of these invasive intracranial neurostimulation treatments (Sprengers et al., 2017).

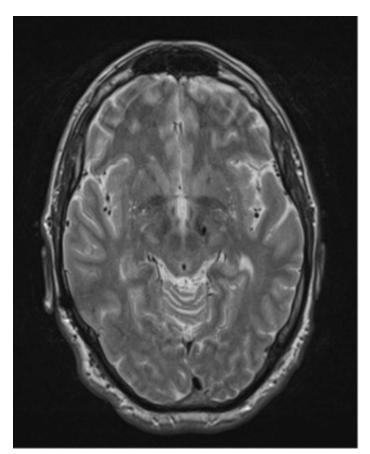


Figure 3. Postoperative MRI to locate the electrode contacts.

FUTURE RESEARCH DIRECTIONS

In studies conducted on non-human primates, researchers have identified that the ideal activity would be related to monitoring a neuron in the motor cortex (Rosin et al., 2011), as this cortical neuron firing would then trigger a reflex stimulus in the GPi of these animals, and the final result seems to be much more effective when compared to the use of continuous stimulation in this model. This is particularly difficult in clinical practice as the stimulation lead is already located in given neuron of a certain structure of the basal ganglia, so that the recording of the neuronal activity that would trigger the stimulation would start in the very structure that was being stimulated. The neurophysiological pattern of beta-oscillations (ranging from 13-30 Hz) has a high correlation with the symptoms of PD and these oscillations are restricted to the territory corresponding to the dorsolateral portion of the STN.

In a recently published clinical trial, Little et al. (2013) evaluated patients who underwent implantation of electrical stimulation electrodes in the STN and who were left with their electrodes externalized for detailed neurophysiological analysis. All patients had the expected improvement in motor symptoms. However, in situations where stimulation was triggered by beta band oscillations from the subthalamic nucleus, stimulation was not only statistically more effective, but there was also a documented reduction in battery consumption. These systems have been known in the field of epilepsy for some time. Alternatively, in recent years there has been a growing interest in the development and application of techniques that, through the infusion of trophic factors, genetic and cellular therapies seek to modify the dysfunctional neuronal circuitry in patients with PD. Several attempts were performed using different techniques such as the use of neurotrophic factors such as neurturin (Marks et al., 2010), enzymes such as glutamic acid decarboxylase (GAD) (LeWitt et al., 2011) and dopaminergic cell transplantation (Freed et al., 2001; Olanow et al., 2003). These studies failed to demonstrate the expected effect on the evaluation of the primary endpoint at a given planned time when compared to the control groups. However, after some time there was some degree of improvement, which came with associated side effects, such as in cases of dopaminergic cell transplantation that led to worsening of previously existing dyskinesia. If on the one hand there is a strong appeal to treat degenerative diseases such as PD by using strategies that try to correct their basic biochemical changes, on the other hand we are faced with the challenge of demonstrating safety and efficacy comparable to electrical stimulation of brain structures, which currently stands as the surgical method of choice.

CONCLUSION

Despite all the knowledge about the pathophysiology of the basal ganglia, many patients who are submitted to deep brain stimulation do not have adequate therapeutic results. It may be possible to identify these intriguing situations even in cases of well-selected patients, with leads precisely located in the target structure and with optimized stimulation parameters. Neurodegenerative symptoms fluctuate a lot during the day and our therapeutic stimulation regime is fixed, creating an imbalance between what the organism needs and what the neuromodulator device really offers. This imbalance would be responsible for less than expected results and for possible premature drainage of batteries. As for other movement disorders, such as essential tremor and dystonia, although there are still no devices that are fully internal with such programmed pacing ability as those for patients with PD, the results of DBS are promising. The key point is to identify some sign of brain neuronal activity that cannot only trigger an immediate and coherent change in the parameters of stimulation, but also accurately translate the driving symptomatology at that specific moment.

REFERENCES

Agari, T., & Date, I. (2012). Spinal cord stimulation for the treatment of abnormal posture and gait disorder in patients with Parkinson's disease. *Neurologia Medico-Chirurgica*, 52(7), 470–474. doi:10.2176/ nmc.52.470 PMID:22850494

Akbarian-Tefaghi, L., Zrinzoand, L., & Foltynie, T. (2016, September). The Use of Deep Brain Stimulation in Tourette Syndrome. *Brain Sciences*, 6(3), 35. doi:10.3390/brainsci6030035 PMID:27548235

Albanese, A., Bhatia, K., Bressman, S. B., Delong, M. R., Fahn, S., Fung, V. S., ... Teller, J. K. (2013, June). Phenomenology and classification of dystonia: A consensus update. *Movement Disorders*, 28(7), 863–873. doi:10.1002/mds.25475 PMID:23649720

Alexander, G. E., Crutcher, M. D., & DeLong, M. R. (1990). Basal ganglia-thalamocortical circuits: Parallel substrates for motor, oculomotor, "prefrontal" and "limbic" functions. *Progress in Brain Research*, *85*, 119–146. doi:10.1016/S0079-6123(08)62678-3 PMID:2094891

Alvarez, L., Macias, R., Guridi, J., Lopez, G., Alvarez, E., Maragoto, C., ... Obeso, J. A. (2001, January). Dorsal subthalamotomy for Parkinson's disease. *Movement Disorders*, *16*(1), 72–78. doi: PMID:11215596

American Psychiatric Association. (2013). Diagnostic and Statistical Manual of Mental Disorders (5th ed.). Washington, DC: American Psychiatric Association Press.

Anderson, V. C., Burchiel, K. J., Hogarth, P., Favre, J., & Hammerstad, J. P. (2005, April). Pallidal vs subthalamic nucleus deep brain stimulation in Parkinson disease. *Archives of Neurology*, *62*(4), 554–560. doi:10.1001/archneur.62.4.554 PMID:15824252

Arle, J. E., Apetauerova, D., Zani, J., Deletis, D. V., Penney, D. L., Hoit, D., ... Shils, J. L. (2008, July). Motor cortex stimulation in patients with Parkinson disease: 12-month follow-up in 4 patients. *Journal of Neurosurgery*, *109*(1), 133–139. doi:10.3171/JNS/2008/109/7/0133 PMID:18590444

Benabid,, A. L., Krack, P. P., Benazzouz, A., Limousin, P., Koudsie, A., & Pollak, P. (2000). Deep brain stimulation of the subthalamic nucleus for Parkinson's disease: Methodologic aspects and clinical criteria. *Neurology*, *55*(12Suppl 6), S40–S44. PMID:11188974

Benabid, A. L. (2007, November). What the future holds for deep brain stimulation. *Expert Review of Medical Devices*, 4(6), 895–903. doi:10.1586/17434440.4.6.895 PMID:18035954

Benabid, A. L. (2009). Targeting the caudal intralaminar nuclei for functional neurosurgery of movement disorders. *Brain Research Bulletin*, 78(2-3), 109–112. doi:10.1016/j.brainresbull.2008.08.020 PMID:18812212

Berg, A. T., Berkovic, S. F., Brodie, M. J., Buchhalter, J., Cross, J. H., van Emde Boas, W., ... Scheffer, I. E. (2010). Revised terminology and concepts for organization of seizures and epilepsies: Report of the ILAE Commission on Classification and Terminology, 2005–2009. *Epilepsia*, *51*(4), 676–685. doi:10.1111/j.1528-1167.2010.02522.x PMID:20196795

Bjartmarz, H., & Rehncrona, S. (2007). Comparison of accuracy and precision between frame-based and frameless stereotactic navigation for deep brain stimulation electrode implantation. *Stereotactic and Functional Neurosurgery*, *85*(5), 235–242. doi:10.1159/000103262 PMID:17534136

Blümcke, I., Thom, M., Aronica, E., Armstrong, D. D., Bartolomei, F., Bernasconi, A., ... Spreafico, R. (2013). International consensus classification of hippocampal sclerosis in temporal lobe epilepsy: A Task Force report from the ILAE Commission on Diagnostic Methods. *Epilepsia*, *54*(7), 1315–1329. doi:10.1111/epi.12220 PMID:23692496

Bor-Seng-Shu, E., Fonoff, E. T., Barbosa, E. R., & Teixeira, M. J. (2010, December). Substantia nigra hyperechogenicity in Parkinson's disease. *Acta Neurochirurgica*, *152*(12), 2085–2087. doi:10.100700701-010-0736-0 PMID:20623147

Bronen, R. A., Cheung, C., & Charles, J. T. (1991). Imaging findings in hippocampal sclerosis: Correlation whith Patology. *AJNR. American Journal of Neuroradiology*, *12*, 933–940. PMID:1950925

Bronstein, J. M., Tagliati, M., Alterman, R. L., Lozano, A. M., Volkmann, J., Stefani, A., ... DeLong, M. R. (2011, February). Deep brain stimulation for Parkinson disease: An expert consensus and review of key issues. *Archives of Neurology*, *68*(2), 165. doi:10.1001/archneurol.2010.260 PMID:20937936

Brooks, D. J. (2002). Diagnosis and management of atypical parkinsonian symptoms. *Journal of Neurology, Neurosurgery, and Psychiatry*, 72, i10–i16. PMID:11870198

Calabrese, E. (2016). Diffusion Tractography in Deep Brain Stimulation Surgery: A Review. *Frontiers in Neuroanatomy*, *10*, 45. doi:10.3389/fnana.2016.00045 PMID:27199677

Callesen, M. B., Hansen, K. V., Gjedde, A., Linnet, J., & Moller, A. (2013). Dopaminergic and clinical correlates of pathological gambling in Parkinson's disease: A case report. *Frontiers in Behavioral Neuroscience*, (7): 1–8. PMID:23908610

Camargo, C. H., Camargos, S. T., Cardoso, F. E., & Teive, H. A. (2015). The genetics of the dystonias – a review based on the new classification of the dystonias. *Arquivos de Neuro-Psiquiatria*, 73(4), 350–358. doi:10.1590/0004-282X20150030 PMID:25992527

Carrillo-Ruiz, J. D., Velasco, F., Jimenez, F., Castro, G., Velasco, A. L., Hernandez, J. A., ... Velasco, M. (2008). Feb). Bilateral electrical stimulation of prelemniscal radiations in the treatment of advanced Parkinson's disease. *Neurosurgery*, *62*(2), 347–357. doi:10.1227/01.neu.0000316001.03765.e8 PMID:18382312

Coenen, V. A., Mädler, B., Schiffbauer, H., Urbach, H., & Allert, N. (2011, April). Individual fiber anatomy of the subthalamic region revealed with diffusion tensor imaging: A concept to identify the deep brain stimulation target for tremor suppression. *Neurosurgery*, *68*(4), 1069–1075. doi:10.1227/ NEU.0b013e31820a1a20 PMID:21242831

Coral, P., Teive, H. A., & Werneck, L. C. (2000). Hemiballism: Report of eight cases. *Arquivos de Neuro-Psiquiatria*, 58(3A), 698–703. doi:10.1590/S0004-282X2000000400016 PMID:10973112

DeLong, M. R. (1990, July). Primate models of movement disorders of basal ganglia origin. *Trends in Neurosciences*, *13*(7), 281–285. doi:10.1016/0166-2236(90)90110-V PMID:1695404

Devinsky, O., Vezzani, A., Najjar, S., De Lanerolle, N. C., & Rogawski, M. A. (2013). Glia and epilepsy: Excitability and inflammation. *Trends in Neurosciences*, *36*(3), 174–184. doi:10.1016/j.tins.2012.11.008 PMID:23298414

Dewey, R. B., & Jankovic, J. (1989). Hemiballism-hemichorea: Clinical and pharmacologic findings in 21 patients. *Archives of Neurology*, *46*(8), 862–867. doi:10.1001/archneur.1989.00520440044020 PMID:2757526

Du, J. J., & Chen, S. D. (2017). Current Nondopaminergic Therapeutic Options for Motor Symptoms of Parkinson's Disease. *Chinese Medical Journal*, *130*(15), 1856–1866. doi:10.4103/0366-6999.211555 PMID:28748860

Elias, W. J., Huss, D., Voss, T., Loomba, J., Khaled, M., Zadicario, E., ... Wintermark, M. (2013). A pilot study of focused ultrasound thalamotomy for essential tremor. *The New England Journal of Medicine*, *369*(7), 640–648. doi:10.1056/NEJMoa1300962 PMID:23944301

Ellis, T. M., Foote, K. D., Fernandez, H. H., Sudhyadhom, A., Rodriguez, R. L., Zeilman, P., ... Okun, M. S. (2008). Reoperation for suboptimal outcomes after deep brain stimulation surgery. *Neurosurgery*, *63*(4), 754–760. doi:10.1227/01.NEU.0000325492.58799.35 PMID:18981887

Engel, J. Jr, Wiebe, S., French, J., Sperling, M., Williamson, P., Spencer, D., ... Enos, B. (2003). Practice parameter: Temporal lobe and localized neocortical resections for epilepsy. *Neurology*, *60*(4), 538–547. doi:10.1212/01.WNL.0000055086.35806.2D PMID:12601090

Fenoy, A. J., & Schiess, M. C. (2017, July). Deep Brain Stimulation of the Dentato-Rubro-Thalamic Tract: Outcomes of Direct Targeting for Tremor. *Neuromodulation*, 20(5), 429–436. doi:10.1111/ner.12585 PMID:28256785

Fisher, R. S., Acevedo, C., Arzimanoglou, A., Bogacz, A., Cross, J. H., Elger, C. E., ... Wiebe, S. (2014). A practical clinical definition of epilepsy. *Epilepsia*, 55(4), 475–482. doi:10.1111/epi.12550 PMID:24730690

Fisher, R. S., Cross, J. H., French, J. A., Higurashi, N., Hirsch, E., Jansen, F. E., ... Zuberi, S. M. (2017). Operational classification of seizure types by the International League Against Epilepsy: Position Paper of the ILAE Commission for Classification and Terminology. *Epilepsia*, *58*(4), 522–530. doi:10.1111/epi.13670 PMID:28276060

Follett, K. A., Weaver, F. M., Stern, M., Hur, K., Harris, C. L., Luo, P., ... Reda, D. J. (2010). Pallidal versus subthalamic deep-brain stimulation for Parkinson's disease. *The New England Journal of Medicine*, *362*(22), 2077–2091. doi:10.1056/NEJMoa0907083 PMID:20519680

Fox, M. W., Ahlskog, J. E., & Kelly, P. J. (1991). Stereotactic ventrolateralis thalamotomy for medically refractory tremor in post-levodopa era Parkinson's disease patients. *Journal of Neurosurgery*, *75*(5), 723–730. doi:10.3171/jns.1991.75.5.0723 PMID:1919694

Freed, C. R., Greene, P. E., Breeze, R. E., Tsai, W. Y., DuMouchel, W., Kao, R., ... Fahn, S. (2001). Transplantation of embryonic dopamine neurons for severe Parkinson's disease. *The New England Journal of Medicine*, *344*(10), 710–719. doi:10.1056/NEJM200103083441002 PMID:11236774

Frucht, S. J. (2013). The definition of dystonia: Current concepts and controversies. *Movement Disorders*, 28(7), 884–888. doi:10.1002/mds.25529 PMID:23893444

Fukaya, C., Sumi, K., Otaka, T., Obuchi, T., Kano, T., Kobayashi, K., ... Katayama, Y. (2010). Nexframe frameless stereotaxy with multitract microrecording: Accuracy evaluated by frame-based stereotactic X-ray. *Stereotactic and Functional Neurosurgery*, 88(3), 163–168. doi:10.1159/000313868 PMID:20431327

Germiniani, F. M., Miranda, A. P., Ferenczy, P., Munho, R. P., & Teive, H. A. (2012, July). Tourette's syndrome: From demonic possession and psychoanalysis to the discovery of gene. *Arquivos de Neuro-Psiquiatria*, 70(7), 547–549. doi:10.1590/S0004-282X2012000700014 PMID:22836463

Goetz, C. G., Tilley, B. C., Shaftman, S. R., Stebbins, G. T., Fahn, S., Martinez-Martin, P., ... LaPelle, N. (2008). Movement Disorder Society-Sponsored Revision of the Unified Parkinson's Disease Rating Scale (MDS-UPDRS): Scale presentation and clinimetric testing results. *Movement Disorders*, 23(15), 2129–2170. doi:10.1002/mds.22340 PMID:19025984

Hariz, M. I., Krack, P., Alesch, F., Augustinsson, L. E., Bosch, A., Ekberg, R., ... Benabid, A.-L. (2008, June). Multicentre European study of thalamic stimulation for parkinsonian tremor: A 6 year follow-up. *Journal of Neurology, Neurosurgery, and Psychiatry*, 79(6), 694–699. doi:10.1136/jnnp.2007.118653 PMID:17898034

Herzog, J., Fietzek, U., Hamel, W., Morsnowski, A., Steigerwald, F., Schrader, B., ... Volkmann, J. (2004). Most effective stimulation site in subthalamic deep brain stimulation for Parkinson's disease. *Movement Disorders*, *19*(9), 1050–1054. doi:10.1002/mds.20056 PMID:15372594

Hoehn, M. M., & Yahr, M. D. (1967, May). Parkinsonism: Onset, progression and mortality. *Neurology*, *17*(5), 427–442. doi:10.1212/WNL.17.5.427 PMID:6067254

Holmes, G., Sirven, J., & Fisher, R. S. (2014). *Temporal Lobe Epilepsy*. Epilepsy Foundation and Epilepsy Therapy Project. Available at: http://www.epilepsy.com/learn/types-epilepsy-syndromes/ temporal-lobe-epilepsy

Huss, D. S., Dallapiazza, R. F., Shah, B. B., Harrison, M. B., Diamond, J., & Elias, W. J. (2015, December). Functional assessment and quality of life in essential tremor with bilateral or unilateral DBS and focused ultrasound thalamotomy. *Movement Disorders*, *30*(14), 1937–1943. doi:10.1002/mds.26455 PMID:26769606

Iacono, R. P., Shima, F., Lonser, R. R., Kuniyoshi, S., Maeda, G., & Yamada, S. (1995, June). The results, indications, and physiology of posteroventral pallidotomy for patients with Parkinson's disease. *Neurosurgery*, *36*(6), 1118–1125. doi:10.1227/00006123-199506000-00008 PMID:7643990

Jabbari, B. (2016). History of Botulinum Toxin Treatment in Movement Disorders. *Tremor and Other Hyperkinetic Movements (New York, N.Y.)*, 6, 394. PMID:27917308

Kelman, C., Ramakrishnan, V., Davies, A., & Holloway, K. (2010). Analysis of stereotactic accuracy of the cosman-robert-wells frame and nexframe frameless systems in deep brain stimulation surgery. *Stereotactic and Functional Neurosurgery*, 88(5), 288–295. doi:10.1159/000316761 PMID:20588080

Khan, S., Gill, S. S., Mooney, L., White, P., Whone, A., Brooks, D. J., & Pavese, N. (2010). Combined pedunculopontine-subthalamic stimulation in Parkinson disease. *Neurology*, 78(14), 1090–1095. doi:10.1212/WNL.0b013e31824e8e96 PMID:22402859

Kiss, Z. H., Wilkinson, M., Krcek, J., Suchowersky, O., Hu, B., Murphy, W. F., ... Tasker, R. R. (2003, October). Is the target for thalamic deep brain stimulation the same as for thalamotomy? *Movement Disorders*, *18*(10), 1169–1175. doi:10.1002/mds.10524 PMID:14534922

Kumar, R., Lozano, A. M., Montgomery, E., & Lang, A. E. (1998). Pallidotomy and deep brain stimulation of the pallidum and subthalamic nucleus in advanced Parkinson's disease. *Movement Disorders*, *13*(Suppl 1), 73–82. PMID:9613722

Kwan, P., Arzimanoglou, A., Berg, A. T., Brodie, M. J., Allen Hauser, W., Mathern, G., ... French, J. (2010). Definition of drug resistant epilepsy: Consensus proposal by the ad hoc Task Force of the ILAE Commission on Therapeutic Strategies. *Epilepsia*, *51*(6), 1069–1077. doi:10.1111/j.1528-1167.2009.02397.x PMID:19889013

Laitinen, L. V., Bergenheim, A. T., & Hariz, M. I. (1992, January). Leksell's posteroventral pallidotomy in the treatment of Parkinson's disease. *Journal of Neurosurgery*, *76*(1), 53–61. doi:10.3171/ jns.1992.76.1.0053 PMID:1727169

Lang, A. E., Duff, J., Saint-Cyr, J. A., Trepanier, L., Gross, R. E., Lombardi, W., ... Lozano, A. M. (1999, September). Posteroventral medial pallidotomy in Parkinson's disease. *Journal of Neurology*, 246(S2Suppl 2), II28–II41. doi:10.1007/BF03161079 PMID:10526000

Lang, A. E., Lozano, A. M., Montgomery, E., Duff, J., Tasker, R., & Hutchinson, W. (1997). Posteroventral medial pallidotomy in advanced Parkinson's disease. *The New England Journal of Medicine*, *337*(15), 1036–1042. doi:10.1056/NEJM199710093371503 PMID:9321531

Larson, P. S. (2014). Deep brain stimulation for movement disorders. *Neurotherapeutics; the Journal of the American Society for Experimental NeuroTherapeutics*, *11*(3), 465–474. doi:10.100713311-014-0274-1 PMID:24833244

Lees, A. J., Hardy, J., & Revesz, T. (2009). Parkinson's disease. *Lancet*, 373(9680), 2055–2066. doi:10.1016/S0140-6736(09)60492-X PMID:19524782

Lenglet, C., Abosch, A., Yacoub, E., De Martino, F., Sapiro, G., & Harel, N. (2012). Comprehensive in vivo mapping of the human basal ganglia and thalamic connectome in individuals using 7T MRI. *PLoS One*, *7*(1), e29153. doi:10.1371/journal.pone.0029153 PMID:22235267

LeWitt, P. A., Rezai, A. R., Leehey, M. A., Ojemann, S. G., Flaherty, A. W., Eskandar, E. N., ... Feigin, A. (2011, April). AAV2-GAD gene therapy for advanced Parkinson's disease: A double-blind, sham-surgery controlled, randomised trial. *Lancet Neurology*, *10*(4), 309–319. doi:10.1016/S1474-4422(11)70039-4 PMID:21419704

Linhares, M. N., & Tasker, R. R. (2000, February). Microelectrode-guided thalamotomy for Parkinson's disease. *Neurosurgery*, 46(2), 390–395. doi:10.1097/00006123-200002000-00024 PMID:10690728

Lipsman, N., Schwartz, M. L., Huang, Y., Lee, L., Sankar, T., Chapman, M., ... Lozano, A. M. (2013, May). MR-guided focused ultrasound thalamotomy for essential tremor: A proof-of-concept study. *Lancet Neurology*, *12*(5), 462–468. doi:10.1016/S1474-4422(13)70048-6 PMID:23523144

Little, S., Pogosyan, A., Neal, S., Zavala, B., Zrinzo, L., Hariz, M., ... Brown, P. (2013, September). Adaptive deep brain stimulation in advanced Parkinson disease. *Annals of Neurology*, 74(3), 449–457. doi:10.1002/ana.23951 PMID:23852650

Lopes, M. A., Richardson, M. P., Abela, E., Rummel, C., Schindler, K., Goodfellow, M., & Terry, J. R. (2017). An optimal strategy for epilepsy surgery: Disruption of the rich-club? *PLoS Computational Biology*, *13*(8), e1005637. doi:10.1371/journal.pcbi.1005637 PMID:28817568

Lotfipour, A. K., Wharton, S., Schwarz, S. T., Gontu, V., Schafer, A., Peters, A. M., ... Bajaj, N. P. S. (2012, January). High resolution magnetic susceptibility mapping of the substantia nigra in Parkinson's disease. *Journal of Magnetic Resonance Imaging*, *35*(1), 48–55. doi:10.1002/jmri.22752 PMID:21987471

Louis, E. D. (2014). Essential Tremor: From Bedside to Bench and Back to Bedside. *Current Opinion in Neurology*, 27(4), 461–467. doi:10.1097/WCO.00000000000115 PMID:24950011

Machado, A., Rezai, A. R., Kopell, B. H., Gross, R. E., Sharan, A. D., & Benabid, A. L. (2006). Deep brain stimulation for Parkinson's disease: Surgical technique and perioperative management. *Movement Disorders*, *21*(S14Suppl 14), S247–S258. doi:10.1002/mds.20959 PMID:16810722

Marks, W. J. Jr, Bartus, R. T., Siffert, J., Davis, C. S., Lozano, A., Boulis, N., ... Olanow, C. W. (2010, December). Gene delivery of AAV2-neurturin for Parkinson's disease: A double-blind, randomised, controlled trial. *Lancet Neurology*, *9*(12), 1164–1172. doi:10.1016/S1474-4422(10)70254-4 PMID:20970382

Mehdorn, H. M. (2016, June). Deep brain stimulation for dystonia: Review of the literature. *Journal of Neurosurgical Sciences*, 60(2), 199–210. PMID:26977634

Montgomery, E. B. Jr. (2013, April). Subthalamic versus globus pallidus deep brain stimulation. *Lancet Neurology*, *12*(4), 329. doi:10.1016/S1474-4422(13)70045-0 PMID:23518323

Moro, E., Gross, R. E., & Krauss, J. K. (2013, June). What's new in surgical treatment for dystonia? *Movement Disorders*, 28(7), 1013–1020. doi:10.1002/mds.25550 PMID:23893457

Moro, E., Hamani, C., Poon, Y. Y., Al-Khairallah, T., Dostrovsky, J. O., Hutchison, W. D., & Lozano, A. M. (2010, January 01). (2010-a). Unilateral pedunculopontine stimulation improves falls in Parkinson's disease. *Brain*, *133*(Pt 1), 215–224. doi:10.1093/brain/awp261 PMID:19846583

Moro, E., Lozano, A. M., Pollak, P., Agid, Y., Rehncrona, S., Volkmann, J., ... Lang, A. E. (2010, April 15). (2010-b). Long-term results of a multicenter study on subthalamic and pallidal stimulation in Parkinson's disease. *Movement Disorders*, *25*(5), 578–586. doi:10.1002/mds.22735 PMID:20213817

Munhoz, R. P., Cerasa, A., & Okun, M. S. (2014). Surgical treatment of dyskinesia in Parkinson's disease. *Frontiers in Neurology*, *5*, 65. doi:10.3389/fneur.2014.00065 PMID:24808889

Murray, C. J. L., Vos, T., Lozano, R., Naghavi, M., Flaxman, A. D., Michaud, C., ... Lopez, A. D. (2012). Disability adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: A systematic analysis for the Global Burden of Disease Study 2010. *Lancet*, *380*(9859), 2197–2223. doi:10.1016/S0140-6736(12)61689-4 PMID:23245608

Nakamura, K., Christine, C. W., Starr, P. A., & Marks, W. J. Jr. (2007). Effects of unilateral subthalamic and pallidal deep brain stimulation on fine motor functions in Parkinson's disease. *Movement Disorders*, 22(5), 619–626. doi:10.1002/mds.21300 PMID:17230483

Obwegeser, A. A., Uitti, R. J., Turk, M. F., Strongosky, A. J., & Wharen, R. E. (2000). Thalamic stimulation for the treatment of midline tremors in essential tremor patients. *Neurology*, *54*(12), 2342–2344. doi:10.1212/WNL.54.12.2342 PMID:10881269

Odekerken, V. J., van Laar, T., Staal, M. J., Mosch, A., Hoffmann, C. F., Nijssen, P. C., ... de Bie, R. M. A. (2013, January). Subthalamic nucleus versus globus pallidus bilateral deep brain stimulation for advanced Parkinson's disease (NSTAPS study): A randomised controlled trial. *Lancet Neurology*, *12*(1), 37–44. doi:10.1016/S1474-4422(12)70264-8 PMID:23168021

Okun, M. S., & Foote, K. D. (2010, December). Parkinson's disease DBS: What, when, who and why? The time has come to tailor DBS targets. *Expert Review of Neurotherapeutics*, *10*(12), 1847–1857. doi:10.1586/ern.10.156 PMID:21384698

Olanow, C. W., Goetz, C. G., Kordower, J. H., Stoessl, A. J., Sossi, V., Brin, M. F., ... Freeman, T. B. (2003, September). A double-blind controlled trial of bilateral fetal nigral transplantation in Parkinson's disease. *Annals of Neurology*, *54*(3), 403–414. doi:10.1002/ana.10720 PMID:12953276

Ondo, W. (2016). Essential Tremor: What We Can Learn from Current Pharmacotherapy. *Tremor and Other Hyperkinetic Movements (New York, N.Y.)*, 6, 356. PMID:26989572

Patel, N. K., Heywood, P., O'Sullivan, K., McCarter, R., Love, S., & Gill, S. S. (2003, May). Unilateral subthalamotomy in the treatment of Parkinson's disease. *Brain*, *126*(Pt 5), 1136–1145. doi:10.1093/brain/awg111 PMID:12690053

Pérez-Suárez, J., Torres Díaz, C.V., López Manzanares, L., Navas García, M., Pastor, J., & Barrio Fernández, P., & de Sola, R. (2017). Radiofrequency Lesions through Deep Brain Stimulation Electrodes in Movement Disorders: Case Report and Review of the Literature. *Stereotactic and Functional Neurosurgery*, *95*(3), 137–141. doi:10.1159/000454891 PMID:28433987

Poewe, W. (2008). Non-motor symptoms in Parkinson's disease. *European Journal of Neurology*, *15*(s1), 14–20. doi:10.1111/j.1468-1331.2008.02056.x PMID:18353132

Pollak, P., Benabid, A. L., Gross, C., Gao, D. M., Laurent, A., Benazzouz, A., & (1993). Effects of the stimulation of the subthalamic nucleus in Parkinson disease. *Revue Neurologique*, *149*(3), 175–176. PMID:8235208

Postuma, R. B., Berg, D., Stern, M., Poewe, W., Olanow, C. W., Oertel, W., ... Deuschl, G. (2015). MDS clinical diagnostic criteria for Parkinson's disease. *Movement Disorders*, *30*(12), 1591–1601. doi:10.1002/mds.26424 PMID:26474316

Rezai, A. R., Machado, A. G., Deogaonkar, M., Azmi, H., Kubu, C., & Boulis, N. M. (2008, February). Surgery for movement disorders. *Neurosurgery*, *62*(Suppl 2), 809–838. doi:10.1227/01. neu.0000316285.52865.53 PMID:18596424

Rosin, B., Slovik, M., Mitelman, R., Rivlin-Etzion, M., Haber, S. N., Israel, Z., ... Bergman, H. (2011, October). Closed-loop deep brain stimulation is superior in ameliorating parkinsonism. *Neuron*, 72(2), 370–384. doi:10.1016/j.neuron.2011.08.023 PMID:22017994

Salat, D., Noyce, A. J., Schrag, A., & Tolosa, E. (2016). Challenges of modifying disease progression in prediagnostic Parkinson's disease. *Lancet Neurology*, 1–12. PMID:26993435

Sandvik, U., Koskinen, L. O., Lundquist, A., & Blomstedt, P. (2012, April). Thalamic and subthalamic deep brain stimulation for essential tremor: Where is the optimal target? *Neurosurgery*, *70*(4), 840–845. doi:10.1227/NEU.0b013e318236a809 PMID:22426044

Schüpbach, W. M., Rau, J., Knudsen, K., Volkmann, J., Krack, P., Timmermann, L., & ... (2013). Neurostimulation for Parkinson's disease with early motor complications. *The New England Journal of Medicine*, *368*(7), 610–622. doi:10.1056/NEJMoa1205158 PMID:23406026

Seppi, K., Weintraub, D., Coelho, M., Perez-Lloret, S., Fox, S. H., Katzenschlager, R., ... Sampaio, C. (2011). The Movement Disorder Society Evidence-Based Medicine Review Update: Treatments for the non-Motor symptoms of Parkinson's disease. *Movement Disorders*, *26*(3), 42–80. doi:10.1002/mds.23884 PMID:22021174

Shannon, K. M. (1998). Ballism. In J. Jankovic & E. Tolosa (Eds.), *Parkinson's disease and movement disorders*. *3* (pp. 365–375). Baltimore, MD: Williams and Wilkins.

Sharma, S., Moon, C. S., Khogali, A., Haidous, A., Chabenne, A., Ojo, C., ... Ebadi, M. (2013, September). Biomarkers in Parkinson's disease (recent update). *Neurochemistry International*, *63*(3), 201–229. doi:10.1016/j.neuint.2013.06.005 PMID:23791710

Sirven, J. I., & Shafer, P. O. (2014). *What is Epilepsy?* Epilepsy Foundation and Epilepsy Therapy Project. Available at: http://www.epilepsy.com/learn/about-epilepsy-basics/what-epilepsy

Sprengers, M., Vonck, K., Carrette, E., & Boon, P. (2017). Deep brain and cortical stimulation for epilepsy. *Cochrane Database of Systematic Reviews*, 7. PMID:28718878

St George, R. J., Nutt, J. G., Burchiel, K. J., & Horak, F. B. (2010, October). A meta-regression of the long-term effects of deep brain stimulation on balance and gait in PD. *Neurology*, 75(14), 1292–1299. doi:10.1212/WNL.0b013e3181f61329 PMID:20921515

Starr, P. A. (2002). Placement of deep brain stimulators into the subthalamic nucleus or Globus pallidus internus: Technical approach. *Stereotactic and Functional Neurosurgery*, 79(3-4), 118–145. doi:10.1159/000070828 PMID:12890973

Stefani, A., Peppe, A., Pierantozzi, M., Galati, S., Moschella, V., Stanzione, P., & Mazzone, P. (2009). Multi-target strategy for Parkinsonian patients: The role of deep brain stimulation in the centromedian-parafascicularis complex. *Brain Research Bulletin*, 78(2-3), 113–118. doi:10.1016/j.brainresbull.2008.08.007 PMID:18812214

Stoessl, A. J. (2014, January). Developments in neuroimaging: Positron emission tomography. *Parkinsonism & Related Disorders*, 20(Suppl 1), S180–S183. doi:10.1016/S1353-8020(13)70042-7 PMID:24262176

Strauss, I., Kalia, S. K., & Lozano, A. M. (2014, January). Where are we with surgical therapies for Parkinson's disease? *Parkinsonism & Related Disorders*, 20(Suppl 1), S187–S191. doi:10.1016/S1353-8020(13)70044-0 PMID:24262178

Sumanaweera, T. S., Glover, G. H., Hemler, P. F., van den Elsen, P. A., Martin, D., Adler, J. R., & Napel, S. (1995, July). MR geometric distortion correction for improved frame-based stereotaxic target localization accuracy. *Magnetic Resonance in Medicine*, *34*(1), 106–113. doi:10.1002/mrm.1910340116 PMID:7674887

Tai, C. H., Wu, R. M., Lin, C. H., Pan, M. K., Chen, Y. F., Liu, H. M., ... Tseng, S.-H. (2010, November). Deep brain stimulation therapy for Parkinson's disease using frameless stereotaxy: Comparison with frame-based surgery. *European Journal of Neurology*, *17*(11), 1377–1385. doi:10.1111/j.1468-1331.2010.03035.x PMID:20443976

Tasker, R. R., & Kiss, Z. H. (1995, January). The role of the thalamus in functional neurosurgery. *Neurosurgery Clinics of North America*, 6(1), 73–104. PMID:7696876

Teive, H. A., Germiniani, F. M., Della Coletta, M. V., & Werneck, L. C. (2001, September). (2001-a). Tics and Tourette syndrome: Clinical evaluation of 44 cases. *Arquivos de Neuro-Psiquiatria*, *59*(3B), 725–728. doi:10.1590/S0004-282X2001000500014 PMID:11593273

Teive, H. A., Zavala, J. A., Iwamoto, F. M., Sá, D., Carraro, H. Jr, & Werneck, L. C. (2001, September). (2001 –b). As contribuições de Charcot e de Marsden para o desenvolvimento dos distúrbios do movimento nos séculos XIX e XX. *Arquivos de Neuro-Psiquiatria*, *59*(3-A), 633–636. doi:10.1590/S0004-282X2001000400031 PMID:11588652

Teixeira, M. J., & Fonoff, E. T. (2004). Tratamento cirúrgico da Doença de Parkinson. *Rev Med (São Paulo)*, 83(1-2), 1–16.

Temel, Y., Ackermans, L., Celik, H., Spincemaille, G. H., van der Linden, C., Walenkamp, G. H., & (2004). Management of hardware infections following deep brain stimulation. *Acta Neurochirurgica*, *146*(4), 355–361. doi:10.100700701-004-0219-2 PMID:15057529

Termsarasab, P., Thammongkolchai, T., & Frucht, S. J. (2016). Medical treatment of dystonia. *Journal of Clinical Movement Disorders*, *3*(1), 19. doi:10.118640734-016-0047-6 PMID:28031858

Thenganatt, M. A., & Jankovic, J. (2016). Recent Advances in Understanding and Managing Tourette Syndrome. *F1000 Research*, *5*, F1000. PMID:26918185

Thevathasan, W., Mazzone, P., Jha, A., Djamshidian, A., Dileone, M., Di Lazzaro, V., & Brown, P. (2010). Spinal cord stimulation failed to relieve akinesia or restore locomotion in Parkinson disease. *Neurology*, 74(16), 1325–1327. doi:10.1212/WNL.0b013e3181d9ed58 PMID:20404313

Troiano, A. R., Teive, H. A., Fabiani, G. B., Zavala, J. A., Sã, D. S., Ferminiani, F. M., ... Werneck, L. C. (2004). Clinical response to long action propranolol in 40 patients diagnosed with essential tremor with no previous treatment: An open, non-controlled study. *Arquivos de Neuro-Psiquiatria*, 62(1), 86–90. doi:10.1590/S0004-282X2004000100015 PMID:15122439

Van Paesschen, W., Revesz, T., Duncan, J. S., King, M. D., & Connelly, A. (1997). Quantitative neuropathology and quantitative magnetic resonance imaging for the hippocampus in temporal lobe epilepsy. *Annals of Neurology*, *42*(5), 756–766. doi:10.1002/ana.410420512 PMID:9392575

Vassal, F., Coste, J., Derost, P., Mendes, V., Gabrillargues, J., Nuti, C., ... Lemaire, J.-J. (2012). Direct stereotactic targeting of the ventrointermediate nucleus of the thalamus based on anatomic 1.5-T MRI mapping with a white matter attenuated inversion recovery (WAIR) sequence. *Brain Stimulation*, *5*(4), 625–633. doi:10.1016/j.brs.2011.10.007 PMID:22405744

Velasco, F., Jimenez, F., Perez, M. L., Carrillo-Ruiz, J. D., Velasco, A. L., Ceballos, J., & ... (2001, August). Electrical stimulation of the prelemniscal radiation in the treatment of Parkinson's disease: An old target revised with new techniques. *Neurosurgery*, *49*(2), 293–306. PMID:11504105

Vidakovic, A., Dragasevic, N., & Kostic, V. S. (1994). Hemiballism: Report of 25 cases. *Journal of Neurology, Neurosurgery, and Psychiatry*, 57(8), 945–949. doi:10.1136/jnnp.57.8.945 PMID:7914529

Vitek, J. L., Bakay, R. A., & DeLong, M. R. (1997). Microelectrode-guided pallidotomy for medically intractable Parkinson's disease. *Advances in Neurology*, *74*, 183–198. PMID:9348414

Voon, V., Krack, P., Lang, A. E., Lozano, A. M., Dujardin, K., Schupbach, M., ... Moro, E. (2008). A multicentre study on suicide outcomes following subthalamic stimulation for Parkinson's disease. *Brain*, *131*(Pt 10), 2720–2728. doi:10.1093/brain/awn214 PMID:18941146

Voon, V., Kubu, C., Krack, P., Houeto, J. L., & Tröster, A. I. (2006). Deep brain stimulation: Neuropsychological and neuropsychiatric issues. *Movement Disorders*, *21*(S14Suppl 14), S305–S327. doi:10.1002/ mds.20963 PMID:16810676

Weisman, H., Qureshi, I. A., Leckman, J. F., Scahill, L., & Bloch, M. H. (2013). Systematic Review: Pharmacological Treatment of Tic Disorders – Efficacy of Antipsychotic and Alpha-2 Adrenergic Agonist Agents. *Neuroscience and Biobehavioral Reviews*, *37*(6), 1162–1171. doi:10.1016/j.neubio-rev.2012.09.008 PMID:23099282

Yoo, H. K., Joung, Y. S., Lee, J. S., Song, D. H., Lee, Y. S., Kim, J.-W., ... Cho, S. C. (2013). A multicenter, randomized, double-blind, placebo-controlled study of aripiprazole in children and adolescents with Tourette's disorder. *The Journal of Clinical Psychiatry*, 74(8), e772–e780. doi:10.4088/JCP.12m08189 PMID:24021518

Compilation of References

Aarsland, D., Andersen, K., Larsen, J. P., Lolk, A., & Kragh-Sorensen, P. (2003). Prevalence and characteristics of dementia in Parkinson disease: An 8-year prospective study. *Archives of Neurology*, *60*(3), 387–392. doi:10.1001/archneur.60.3.387 PMID:12633150

Aarsland, D., Ballard, C. G., & Halliday, G. (2004). Are Parkinson's disease with dementia and dementia with Lewy bodies the same entity? *Journal of Geriatric Psychiatry and Neurology*, *17*(3), 137–145. doi:10.1177/0891988704267470 PMID:15312277

Aarsland, D., Brønnick, K., Ehrt, U., De Deyn, P. P., Tekin, S., Emre, M., & Cummings, J. L. (2007). Neuropsychiatric symptoms in patients with Parkinson's disease and dementia: Frequency, profile and associated care giver stress. *Journal of Neurology, Neurosurgery, and Psychiatry*, 78(1), 36–42. doi:10.1136/jnnp.2005.083113 PMID:16820421

Aarsland, D., Bronnick, K., Williams-Gray, C., Weintraub, D., Marder, K., Kulisevsky, J., ... Emre, M. (2010). Mild cognitive impairment in Parkinson disease: A multicenter pooled analysis. *Neurology*, 75(12), 1062–1069. doi:10.1212/WNL.0b013e3181f39d0e PMID:20855849

Aarsland, D., Kvaløy, J. T., Andersen, K., Larsen, J. P., Tang, M. X., Lolk, A., ... Marder, K. (2007). The effect of age of onset of PD on risk of dementia. *Journal of Neurology*, 254(1), 38–45. doi:10.100700415-006-0234-8 PMID:17508138

Aarsland, D., Marsh, L., & Schrag, A. (2009). Neuropsychiatric symptoms in Parkinson's disease. *Movement Disorders*, 24(15), 2175–2186. doi:10.1002/mds.22589 PMID:19768724

Aarsland, D., Mosimann, U. P., & McKeith, I. G. (2004). Role of cholinesterase inhibitors in Parkinson's disease and dementia with Lewy bodies. *Journal of Geriatric Psychiatry and Neurology*, *17*(3), 164–171. doi:10.1177/0891988704267463 PMID:15312280

Aarsland, D., Perry, R., Larsen, J. P., McKeith, I. G., O'Brien, J. T., Perry, E. K., ... Ballard, C. G. (2005b). Neuroleptic sensitivity in Parkinson's disease and parkinsonian dementias. *The Journal of Clinical Psychiatry*, *66*(05), 633–637. doi:10.4088/JCP.v66n0514 PMID:15889951

Aarsland, D., Zaccai, J., & Brayne, C. (2005a). A systematic review of prevalence studies of dementia in Parkinson's disease. *Movement Disorders*, 20(10), 1255–1263. doi:10.1002/mds.20527 PMID:16041803

Abajirao, S. A., Nair, M., & Kambale, H. (2013). Subacute sclerosing panencephalitis: A clinical appraisal. *Annals of Indian Academy of Neurology*, *16*(4), 631. doi:10.4103/0972-2327.120497 PMID:24339595

Abeliovich, A., & Flint Beal, M. (2006). Parkinsonism genes: Culprits and clues. *Journal of Neurochemistry*, 99(4), 1062–1072. doi:10.1111/j.1471-4159.2006.04102.x PMID:16836655

Aberg, L. E., Backman, M., Kirveskari, E., & Santavuori, P. (2000). Epilepsy and Antiepileptic Drug Therapy in Juvenile Neuronal Ceroid Lipofuscinosis. *Epilepsia*, *41*(10), 1296–1302. doi:10.1111/j.1528-1157.2000.tb04608.x PMID:11051125

Compilation of References

Abhinav, K., Stanton, B., Johnston, C., Hardstaff, J., Orrell, R. W., Howard, R., ... Shaw, C. E. (2007). Amyotrophic lateral sclerosis in South-East England: A population-based study. The South-East England register for amyotrophic lateral sclerosis (SEALS Registry). *Neuroepidemiology*, *29*(1-2), 44–48. doi:10.1159/000108917 PMID:17898523

Abou-Sleiman, P. M., Muqit, M. M. K., & Wood, N. W. (2006). Expanding insights of mitochondrial dysfunction in Parkinson's disease. *Nature Reviews. Neuroscience*, 7(3), 207–219. doi:10.1038/nrn1868 PMID:16495942

Acevedo-Morantes, C. Y., & Wille, H. (2014). The Structure of Human Prions: From Biology to Structural Models— Considerations and Pitfalls. *Viruses*, *6*(10), 3875–3892. doi:10.3390/v6103875 PMID:25333467

Acevedo-Torres, K., Berríos, L., Rosario, N., Dufault, V., Skatchkov, S., Eaton, M. J., & Ayala-Torres, S. (2009). Mitochondrial DNA damage is a hallmark of chemically induced and the R6/2 transgenic model of Huntington's disease. *DNA Repair*, 8(1), 126–136. doi:10.1016/j.dnarep.2008.09.004 PMID:18935984

Acquatella-Tran, V. B. I., Imberdis, T., & Perrier, V. (2013). From prion diseases to prion-like propagation mechanisms of neurodegenerative diseases. *International Journal of Cell Biology*, 975832. PMID:24222767

Acsadi, G., Lee, I., Li, X., Khaidakov, M., Pecinova, A., Parker, G. C., & Hüttemann, M. (2009). Mitochondrial dysfunction in a neural cell model of spinal muscular atrophy. *Journal of Neuroscience Research*, 87(12), 2748–2756. doi:10.1002/jnr.22106 PMID:19437551

Acton, Q. A. (2012). Amyotrophic Lateral Sclerosis: New insights for the healthcare professional. Scholarly Editions.

Adams, H. P. Jr, Bendixen, B. H., Kappelle, L. J., Biller, J., Love, B. B., Gordon, D. L., & Marsh, E. E. III. (1993). Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. TOAST. Trial of Org 10172 in Acute Stroke Treatment. *Stroke*, 24(1), 35–41. doi:10.1161/01.STR.24.1.35 PMID:7678184

Adams, J. M., & Cory, S. (1998). The Bcl-2 protein family: Arbiters of cell survival. *Science*, *281*(5381), 1322–1326. doi:10.1126cience.281.5381.1322 PMID:9735050

Adams, R. D., van Bogaert, L., & Vander Eecken, H. (1964). Striato-nigral degeneration. *Journal of Neuropathology* and *Experimental Neurology*, 23(4), 584–608. PMID:14219099

Adler, C. H., Connor, D. J., Hentz, J. G., Sabbagh, M. N., Caviness, J. N., Shill, H. A., ... Beach, T. G. (2010). Incidental Lewy body disease: Clinical comparison to a control cohort. *Movement Disorders*, 25(5), 642–646. doi:10.1002/ mds.22971 PMID:20175211

Adolphs, R., Gosselin, F., Buchanan, T. W., Tranel, D., Schyns, P., & Damasio, A. R. (2005). A mechanism for impaired fear recognition after amygdala damage. *Nature*, *433*(7021), 68–72. doi:10.1038/nature03086 PMID:15635411

Aflaki, E., Westbroek, W., & Sidransky, E. (2017). The Complicated Relationship between Gaucher Disease and Parkinsonism: Insights from a Rare Disease. *Neuron*, *93*(4), 737–746. doi:10.1016/j.neuron.2017.01.018 PMID:28231462

Agari, T., & Date, I. (2012). Spinal cord stimulation for the treatment of abnormal posture and gait disorder in patients with Parkinson's disease. *Neurologia Medico-Chirurgica*, 52(7), 470–474. doi:10.2176/nmc.52.470 PMID:22850494

Agrawal, M., & Biswas, A. (2015). Molecular diagnostics of neurodegenerative disorders. *Frontiers in Molecular Biosciences*, 2. PMID:26442283

Aguirre, T., van Den Bosch, L., Goetschalckx, K., Tilkin, P., Mathijis, G., Cassiman, J.-J., & Robberecht, W. (1998). Increased sensitivity of fibroblasts from amyotrophic lateral sclerosis patients to oxidative stress. *Annals of Neurology*, *43*(4), 452–457. doi:10.1002/ana.410430407 PMID:9546325

Aguzzi, A., Sigurdson, C., & Heikenwaelder, M. (2008). Molecular mechanisms of prion pathogenesis. *Annual Review of Pathology: Mechanisms of Disease*, 3(1), 11–40. doi:10.1146/annurev.pathmechdis.3.121806.154326 PMID:18233951

Aguzzi, A., & Zhu, C. (2012). Five Questions on Prion Diseases. *PLoS Pathogens*, 8(5). doi:10.1371/journal.ppat.1002651 PMID:22570608

Akbarian-Tefaghi, L., Zrinzoand, L., & Foltynie, T. (2016, September). The Use of Deep Brain Stimulation in Tourette Syndrome. *Brain Sciences*, *6*(3), 35. doi:10.3390/brainsci6030035 PMID:27548235

Alami, N. H., Smith, R. B., Carrasco, M. A., Williams, L. A., Winborn, C. S., Han, S. S., ... Taylor, J. P. (2014). Axonal transport of TDP-43 mRNA granules is impaired by ALS-causing mutations. *Neuron*, *81*(3), 536–543. doi:10.1016/j. neuron.2013.12.018 PMID:24507191

Alam, Z. I., Daniel, S. E., Lees, A. J., Marsden, D. C., Jenner, P., & Halliwell, B. (1997). A generalised increase in protein carbonyls in the brain in Parkinson's but not incidental Lewy body disease. *Journal of Neurochemistry*, 69(3), 1326–1329. doi:10.1046/j.1471-4159.1997.69031326.x PMID:9282961

Alam, Z. I., Jenner, A., Daniel, S. E., Lees, A. J., Cairns, N., Marsden, C. D., ... Halliwell, B. (1997). Oxidative DNA damage in the parkinsonian brain: An apparent selective increase in 8-hydroxyguanine levels in substantia nigra. *Journal of Neurochemistry*, *69*(3), 1196–1203. doi:10.1046/j.1471-4159.1997.69031196.x PMID:9282943

Alavi Naini, S. M., & Soussi-yanicostas, N. (2015). Tau hyperphosphorylation and oxidative stress, a critical vicious circle in neurodegenerative tauopathies? *Oxidative Medicine and Cellular Longevity*, 151979. PMID:26576216

Albanese, A., Bhatia, K., Bressman, S. B., Delong, M. R., Fahn, S., Fung, V. S., ... Teller, J. K. (2013, June). Phenomenology and classification of dystonia: A consensus update. *Movement Disorders*, 28(7), 863–873. doi:10.1002/mds.25475 PMID:23649720

Al-Essa, M. A., Bakheet, S. M., Patay, Z. J., Powe, J. E., & Ozand, P. T. (2000). Clinical and cerebral fdg pet scan in a patient with krabbe's disease. *Pediatric Neurology*, 22(1), 44–47. doi:10.1016/S0887-8994(99)00107-1 PMID:10669205

Alexander, G. E. (2004). Biology of Parkinson's disease: Pathogenesis and pathophysiology of a multisystem neurodegenerative disorder. *Dialogues in Clinical Neuroscience*, 6(3), 259. PMID:22033559

Alexander, G. E., Crutcher, M. D., & DeLong, M. R. (1990). Basal ganglia-thalamocortical circuits: Parallel substrates for motor, oculomotor, "prefrontal" and "limbic" functions. *Progress in Brain Research*, *85*, 119–146. doi:10.1016/S0079-6123(08)62678-3 PMID:2094891

Alexander, G. E., DeLong, M. R., & Strick, P. L. (1986). Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annual Review of Neuroscience*, *9*(1), 357–381. doi:10.1146/annurev.ne.09.030186.002041 PMID:3085570

Alexander, W. S. (1949). Progressive Fibrinoid Degeneration of Fibrillary Astrocytes Associated with Mental Retardation in a Hydrocephalic Infant. *Brain*, 72(3), 373–381. doi:10.1093/brain/72.3.373 PMID:15409268

Alexeyev, M. F., LeDOUX, S. P., & Wilson, G. L. (2004). Mitochondrial DNA and aging. *Clinical Science*, *107*(4), 355–364. doi:10.1042/CS20040148 PMID:15279618

Algahtani, H. A., & Aldarmahi, A. A. (2014). Cerebral venous sinus thrombosis. *Neurosciences (Riyadh)*, 19(1), 11–16. PMID:24419443

Alhola, P., & Polo-Kantola, P. (2007). Sleep deprivation: Impact on cognitive performance. *Neuropsychiatric Disease* and *Treatment*, *3*(5), 553. PMID:19300585

Aliev, G., Ashraf, G. M., Horecký, J., Vancova, O., Gvozdjakova, A., & Kucharská, J. ... Bachurin, S. (2011). Potential Preventive Effects of Coenzyme Q and Creatine Supplementation on Brain Energy Metabolism in Rats Exposed to Chronic Cerebral Hypoperfusion. In Systems Biology of Free Radicals and Antioxidants. Berlin: Springer.

Allan, L. M., Rowan, E. N., Firbank, M. J., Thomas, A. J., Parry, S. W., Polvikoski, T. M., ... Kalaria, R. N. (2011). Long term incidence of dementia, predictors of mortality and pathological diagnosis in older stroke survivors. *Brain*, *134*(12), 3716–3727. doi:10.1093/brain/awr273 PMID:22171356

Allen, J. A., & Coombs, M. M. (1980). Covalent binding of polycyclic aromatic compounds to mitochondrial and nuclear DNA. *Nature*, 287(5779), 244–245. doi:10.1038/287244a0 PMID:7432460

Allen, N., Robinson, A. C., Snowden, J., Davidson, Y. S., & Mann, D. M. A. (2014). Patterns of cerebral amyloid angiopathy define histopathological phenotypes in Alzheimer's disease. *Neuropathology and Applied Neurobiology*, *40*(2), 136–148. doi:10.1111/nan.12070 PMID:23808763

Alonso, A., Logroscino, G., Jick, S. S., & Hernan, M. A. (2010). Association of smoking with amyotrophic lateral sclerosis risk and survival in men and women: A prospective study. *BMC Neurology*, *10*(1), 6. doi:10.1186/1471-2377-10-6 PMID:20074360

Alvarez, L., Macias, R., Guridi, J., Lopez, G., Alvarez, E., Maragoto, C., ... Obeso, J. A. (2001, January). Dorsal subthalamotomy for Parkinson's disease. *Movement Disorders*, *16*(1), 72–78. doi: PMID:11215596

Alzheimer's Association. (2014). 2014 Alzheimer's disease facts and figures. *Alzheimer's & Dementia*, *10*(2), e47–e92. doi:10.1016/j.jalz.2014.02.001 PMID:24818261

Alzheimer's Association. (2017a). *Stages of Alzheimer's and Symptoms* | *Alzheimer's Association*. Retrieved March 6, 2017, from http://www.alz.org/alzheimers_disease_stages_of_alzheimers.asp

Alzheimer's Association. (2017b). *Types of Dementia*. Retrieved April 1, 2017, from http://www.alz.org/dementia/types-of-dementia.asp

Alzheimer's Association. (2017c). *What is Dementia?* Retrieved April 1, 2017, from http://www.alz.org/what-is-dementia. asp

American Psychiatric Association. (2013). Diagnostic and Statistical Manual of Mental Disorders (5th ed.). Washington, DC: American Psychiatric Association Press.

Ames, B. N. (1989). Endogenous oxidative DNA damage, aging, and cancer. *Free Radical Research Communications*, 7(3-6), 121–128. doi:10.3109/10715768909087933 PMID:2684796

Ameur, A., Zaghlool, A., Halvardson, J., Wetterbom, A., Gyllensten, U., Cavelier, L., & Feuk, L. (2011). Total RNA sequencing reveals nascent transcription and widespread co-transcriptional splicing in the human brain. *Nature Structural & Molecular Biology*, *18*(12), 1435–1440. doi:10.1038/nsmb.2143 PMID:22056773

Amgen. (2017). Pipeline. Retrieved March 7, 2017, from http://www.amgenpipeline.com/pipeline/

Anand, A., Khurana, P., Chawla, J., Sharma, N., & Khurana, N. (2017). Emerging treatments for the behavioral and psychological symptoms of dementia. *CNS Spectrums*, 1–9. doi:10.1017/S1092852917000530 PMID:28911339

Anand, A., Patience, A. A., Sharma, N., & Khurana, N. (2017). The present and future of pharmacotherapy of Alzheimer's disease: A comprehensive review. *European Journal of Pharmacology*, *815*, 364–375. doi:10.1016/j.ejphar.2017.09.043 PMID:28978455

Andersen, P. M., Borasio, G. D., Dengler, R., Hardiman, O., Kollewe, K., Leigh, P. N., ... Tomik, B. (2005). EFNS task force on management of amyotrophic lateral sclerosis: Guidelines for diagnosing and clinical care of patients and relatives. *European Journal of Neurology*, *12*(12), 921–938. doi:10.1111/j.1468-1331.2005.01351.x PMID:16324086

Anderson, J. K. (2004). Oxidative stress in neurodegeneration: Cause or consequence? *Nat Neurosci Rev*, 5(7), S18–S25. doi:10.1038/nrn1434 PMID:15298006

Anderson, R. A., Donnelly, C. A., Ferguson, N. M., Woolhouse, M. E. J., Watt, C. J., Udy, H. J., ... Wells, G. A. H. (1996). Transmission dynamics and epidemiology of BSE in British cattle. *Nature*, *382*(6594), 779–788. doi:10.1038/382779a0 PMID:8752271

Anderson, S., Bankier, A. T., Barrell, B. G., De Bruijn, M. H., Coulson, A. R., Drouin, J., ... Schreier, P. H. (1981). Sequence and organization of the human mitochondrial genome. *Nature*, 290(5806), 457–465. doi:10.1038/290457a0 PMID:7219534

Anderson, V. C., Burchiel, K. J., Hogarth, P., Favre, J., & Hammerstad, J. P. (2005, April). Pallidal vs subthalamic nucleus deep brain stimulation in Parkinson disease. *Archives of Neurology*, *62*(4), 554–560. doi:10.1001/archneur.62.4.554 PMID:15824252

Andreassen, O. A., Dedeoglu, A., Klivenyi, P., Beal, M. F., & Bush, A. I. (2000). N-acetyl-L-cysteine improves survival and preserves motor performance in an animal model of familial amyotrophic lateral sclerosis. *Neuroreport*, *11*(11), 2491–2493. doi:10.1097/00001756-200008030-00029 PMID:10943709

Andres-Mateos, E., Perier, C., Zhang, L., Blanchard-Fillion, B., Greco, T. M., Thomas, B., & Dawson, T. M. (2007). DJ-1 gene deletion reveals that DJ-1 is an atypical peroxiredoxin-like peroxidase. *Proceedings of the National Academy of Sciences of the United States of America*, *104*(37), 14807–14812. doi:10.1073/pnas.0703219104 PMID:17766438

Andres, R. H., Ducray, A. D., Schlattner, U., Wallimann, T., & Widmer, H. R. (2008). Functions and effects of creatine in the central nervous system. *Brain Research Bulletin*, *76*(4), 329–343. doi:10.1016/j.brainresbull.2008.02.035 PMID:18502307

Andriollo-Sanchez, M., Hininger-Favier, I., Meunier, N., Venneria, E., O'Connor, J.M., Maiani, G., Coudray, C., & Roussel, A.M. (2005). Age-related oxidative stress and antioxidant parameters in middle-aged and older European subjects: the ZENITH study. *European Journal of Clinical Nutrition*, *59*(Suppl. 2), S 58–S62.

Ann, R. S., Eric, G. B. E., Jillian, L. M., Heidi, C. M., Jeffrey, G. P., & Glenn, L. M. (2013). Zinc drives a tertiary fold in the prion protein with familial disease mutation sites at the interface. *Structure (London, England)*, *21*(2), 236–246. doi:10.1016/j.str.2012.12.002 PMID:23290724

Antonini, A., Abbruzzese, G., Barone, P., Bonuccelli, U., Lopiano, L., Onofrj, M., ... Quattrone, A. (2008). COMT inhibition with tolcapone in the treatment algorithm of patients with Parkinson's disease (PD): Relevance for motor and non-motor features. *Neuropsychiatric Disease and Treatment*, 4(1), 1–9. doi:10.2147/NDT.S2404 PMID:18728767

Apel, K., & Hirt, H. (2004). Reactive oxygen species: Metabolism, oxidative stress, and signal transduction. *Annual Review of Plant Biology*, 55(1), 373–399. doi:10.1146/annurev.arplant.55.031903.141701 PMID:15377225

Apweiler, R., Bairoch, A., Wu, C. H., Barker, W. C., Boeckmann, B., & Ferro, S. (2004). UniProt: The Universal Protein knowledgebase. *Nucleic Acids Research*, *32*(Database issue), D115–D119. doi:10.1093/nar/gkh131 PMID:14681372

Arber, C. E., Houlden, H., Li, A., & Wray, S. (2016). Review: Insights into molecular mechanism of disease in neurodegeneration with brain iron accumulation: unifying theories. *Neuropathology and Applied Neurobiology*, *42*(3), 200–241. doi:10.1111/nan.12242 PMID:25870938

Arena, G., Gelmetti, V., Torosantucci, L., Vignone, D., Lamorte, G., De Rosa, P., ... Valente, E. M. (2013). PINK1 protects against cell death induced by mitochondrial depolarization, by phosphorylating Bcl-xL and impairing its pro-apoptotic cleavage. *Cell Death and Differentiation*, 20(7), 920–930. doi:10.1038/cdd.2013.19 PMID:23519076

Arendt, T., Bigl, V., Arendt, A., & Tennstedt, A. (1983). Loss of neurons in the nucleus basalis of Meynert in Alzheimer's disease, paralysis agitans and Korsakoff's disease. *Acta Neuropathologica*, *61*(2), 101–108. doi:10.1007/BF00697388 PMID:6637393

Areosa, S., & Grimley, E. (2002). Effect of the treatment of Type II diabetes mellitus on the development of cognitive impairment and dementia. *Cochrane Database of Systematic Reviews*, (4): CD003804–CD003804. PMID:12519608

Argov, Z., & Navon, R. (1984). Clinical and genetic variations in the syndrome of adult GM2 gangliosidosis resulting from hexosaminidase A deficiency. *Annals of Neurology*, *16*(1), 14–20. doi:10.1002/ana.410160105 PMID:6235771

Ariesen, M. J., Claus, S. P., Rinkel, G. J., & Algra, A. (2003). Risk factors for intracerebral hemorrhage in the general population: A systematic review. *Stroke*, *34*(8), 2060–2065. doi:10.1161/01.STR.0000080678.09344.8D PMID:12843354

Ariga, H., Takahashi-Niki, K., Kato, I., Maita, H., Niki, T., & Iguchi-Ariga, S. M. M. (2013). Neuroprotective Function of DJ-1 in Parkinson's Disease. *Oxidative Medicine and Cellular Longevity*, 2013, 683920. doi:10.1155/2013/683920 PMID:23766857

Arispe, N., Rojas, E., & Pollard, H. B. (1993). Alzheimer disease amyloid beta protein forms calcium channels in bilayer membranes: Blockade by tromethamine and aluminum. *Proceedings of the National Academy of Sciences of the United States of America*, *90*(2), 567–571. doi:10.1073/pnas.90.2.567 PMID:8380642

Arle, J. E., Apetauerova, D., Zani, J., Deletis, D. V., Penney, D. L., Hoit, D., ... Shils, J. L. (2008, July). Motor cortex stimulation in patients with Parkinson disease: 12-month follow-up in 4 patients. *Journal of Neurosurgery*, *109*(1), 133–139. doi:10.3171/JNS/2008/109/7/0133 PMID:18590444

Arlt, S., Beisiegel, U., & Kontush, A. (2002). Lipid peroxidation in neurodegeneration: New insights into Alzheimer's disease. *Current Opinion in Lipidology*, *13*(3), 289–294. doi:10.1097/00041433-200206000-00009 PMID:12045399

Armon, C., & Lorenzo, N. (2017). *Amyotrophic lateral sclerosis. Practical essentials*. Available at http://emedicine. medscape.com/article/1170097-overview

Armon, C. (2003). An evidence-based medicine approach to the evaluation of the role of exogenous risk factors in sporadic amyotrophic lateral sclerosis. *Neuroepidemiology*, 22(4), 217–228. doi:10.1159/000070562 PMID:12792141

Armon, C. (2009). Smoking may be considered an established risk factor for sporadic ALS. *Neurology*, 73(20), 1693–1698. doi:10.1212/WNL.0b013e3181c1df48 PMID:19917993

Armstrong, M. J. (2014). Diagnosis and treatment of corticobasal degeneration. *Current Treatment Options in Neurology*, *16*(3), 282. doi:10.100711940-013-0282-1 PMID:24469408

Armstrong, M. J., Litvan, I., Lang, A. E., Bak, T. H., Bhatia, K. P., Borroni, B., ... Weiner, W. J. (2013). Criteria for the diagnosis of corticobasal degeneration. *Neurology*, *80*(5), 496–503. doi:10.1212/WNL.0b013e31827f0fd1PMID:23359374

Armstrong, R. A., Cairns, N. J., & Lantos, P. L. (2001). What does the study of the spatial patterns of pathological lesions tell us about the pathogenesis of neurodegenerative disorders? *Neuropathology*, 21(1), 1–12. doi:10.1046/j.1440-1789.2001.00373.x PMID:11304036

Arnold, P. D., Siegel-Bartelt, J., Cytrynbaum, C., Teshima, I., & Schachar, R. (2001). Velo-cardio-facial syndrome: Implications of microdeletion 22q11 for schizophrenia and mood disorders. *American Journal of Medical Genetics*, *105*(4), 354–362. doi:10.1002/ajmg.1359 PMID:11378850 Arnold, W. D., & Burghes, A. H. (2013). Spinal muscular atrophy: Development and implementation of potential treatments. *Annals of Neurology*, 74(3), 348–362. doi:10.1002/ana.23995 PMID:23939659

Arnold, W. D., Kassar, D., & Kissel, J. T. (2015). Spinal muscular atrophy: Diagnosis and management in a new therapeutic era. *Muscle & Nerve*, 51(2), 157–167. doi:10.1002/mus.24497 PMID:25346245

Aronis, A., Melendez, J. A., Golan, O., Shilo, S., Dicter, N., & Tirosh, O. (2003). Potentiation of Fas-mediated apoptosis by attenuated production of mitochondria-derived reactive oxygen species. *Cell Death and Differentiation*, *10*(3), 335–344. doi:10.1038j.cdd.4401150 PMID:12700633

Artero, S., Tiemeier, H., Prins, N., Sabatier, R., Breteler, M., & Ritchie, K. (2004). Neuroanatomical localisation and clinical correlates of white matter lesions in the elderly. *Journal of Neurology, Neurosurgery, and Psychiatry*, 75(9), 1304–1308. doi:10.1136/jnnp.2003.023713 PMID:15314121

Arvanitakis, Z., Leurgans, S. E., Barnes, L. L., Bennett, D. A., & Schneider, J. A. (2014). Microinfarct pathology, dementia, and cognitive systems. *Stroke*, 42(3), 722–727. doi:10.1161/STROKEAHA.110.595082 PMID:21212395

Ascherio, A., Chen, H., Weisskopf, M. G., O'Reilly, E., McCullough, M. L., Calle, E. E., ... Thun, M. J. (2006). Pesticide exposure and risk for Parkinson's disease. *Annals of Neurology*, 60(2), 197–203. doi:10.1002/ana.20904 PMID:16802290

Ashaq, I., Shajrul, A., Akbar, M., & Rashid, F. (2015). Protein Misfolding Diseases: In Perspective of Gain and Loss of Function. In Proteostasis and Chaperone Surveillance (pp. 105-118). Springer India.

Ash, P. E. A., Bieniek, K. F., Gendron, T. F., Caulfield, T., Lin, W. L., Dejesus-Hernandez, M., ... Joseph, W. (2013). Unconventional translation of C9ORF72 GGGGCC expansion generates insoluble polypeptides specific to c9FTD/ALS. *Neuron*, 77(4), 639–646. doi:10.1016/j.neuron.2013.02.004 PMID:23415312

Ashraf, W., Pfeiffer, R. F., Park, F., Lof, J., & Quigley, E. M. (1997). Constipation in Parkinson's disease: Objective assessment and response to psyllium. *Movement Disorders*, *12*(6), 946–951. doi:10.1002/mds.870120617 PMID:9399219

Atlante, A., Calissano, P., Bobba, A., Giannattasio, S., Marra, E., & Passarella, S. (2001). Glutamate neurotoxicity, oxidative stress and mitochondria. *Federation of European Biochemical Societies (FEBS)*. *Letters*, 497(1), 1–5.

Averill, A. J., Kasarskis, E. J., & Segerstrom, S. C. (2007). Psychological health in patients with amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases*, 8(4), 243–254. doi:10.1080/17482960701374643 PMID:17653923

Avila, M. F., Torrente, D., Cabezas, R., Morales, L., García-Segura, L. M., Ganzalez, J., & George, E. B. (2014). Structural insights from GRP78-NF-kappaB binding interactions: A computational approach to understand a possible neuroprotective pathway in brain injuries. *Journal of Theoretical Biology*, *345*, 43–51. doi:10.1016/j.jtbi.2013.12.010 PMID:24361327

Ayano, G. (2016). Parkinson's Disease: A Concise Overview of Etiology, Epidemiology, Diagnosis, Comorbidity and Management. *Journal of Neurological Disorders*, 4(6), 1–6. doi:10.4172/2329-6895.1000298

Azuaje, F. J. (2014). Selecting biologically informative genes in co-expression networks with a centrality score. *Biology Direct*, *9*(1), 12. doi:10.1186/1745-6150-9-12 PMID:24947308

Babi, M. A., Kraft, B. D., Sengupta, S., Peterson, H., Orgel, R., Wegermann, Z., & Luedke, M. W. (2016). Related or not? Development of spontaneous Creutzfeldt–Jakob disease in a patient with chronic, well-controlled HIV: A case report and review of the literature. *SAGE Open Medical Case Reports*, *4*.

Backman, M. L., Santavuori, P. R., Aberg, L. E., & Aronen, E. T. (2005). Psychiatric symptoms of children and adolescents with juvenile neuronal ceroid lipofuscinosis. *Journal of Intellectual Disability Research*, 49(1), 25–32. doi:10.1111/j.1365-2788.2005.00659.x PMID:15634309

Bailey, R. (2017). *Divisions of the Brain. Retrieved from Divisions of the Brain - Forebrain, Midbrain, Hindbrain.* Retrieved from https://www.thoughtco.com/divisions-of-the-brain-4032899

Balaban, R. S., Nemoto, S., & Finkel, T. (2005). Mitochondria, oxidants, and aging. *Cell*, *120*(4), 483–495. doi:10.1016/j. cell.2005.02.001 PMID:15734681

Ballard, C. G., Aarsland, D., McKeith, I., O'brien, J., Gray, A., Cormack, F., ... Brown, R. (2002). Fluctuations in attention PD dementia vs DLB with parkinsonism. *Neurology*, *59*(11), 1714–1720. doi:10.1212/01.WNL.0000036908.39696. FD PMID:12473758

Baloyannis, S. J. (2006). Mitochondrial alterations in Alzheimer's disease. *Journal of Alzheimer's Disease*, 9(2), 119–126. doi:10.3233/JAD-2006-9204 PMID:16873959

Baloyannis, S. J. (2014). Golgi apparatus and protein trafficking in Alzheimer's disease. *Journal of Alzheimer's Disease*, 42(3), 153–162. PMID:24946873

Bandopadhyay, R., & de Belleroche, J. (2010). Pathogenesis of Parkinson's disease: Emerging role of molecular chaperones. *Trends in Molecular Medicine*, *16*(1), 27–36. doi:10.1016/j.molmed.2009.11.004 PMID:20036196

Bansal, A., & Srivastava, P. A. (2018). Transcriptomics to Metabolomics: A Network Perspective for Big Data. IGI Global.

Bansal, A., & Ramana, J. (2015). TCGDB: A Compendium of Molecular Signatures of Thyroid Cancer and Disorders. *Journal of Cancer Science & Therapy*, 7(7). Retrieved from https://www.omicsonline.org/open-access/tcgdb-a-compendium-of-molecular-signatures-of-thyroid-cancer-and-disorders-1948-5956-1000350.php?aid=57693

Bansal, A., Singh, T. R., & Chauhan, R. S. (2017). A novel miRNA analysis framework to analyze differential biological networks. *Scientific Reports*, 7(1), 14604. doi:10.103841598-017-14973-x PMID:29097749

Bansal, R., & Singh, R. (2017). Exploring the potential of natural and synthetic neuroprotective steroids against neurodegenerative disorders: A literature review. *Medicinal Research Reviews*, 1098–1128. PMID:28697282

Barabási, A.-L., Gulbahce, N., & Loscalzo, J. (2011). Network medicine: A network-based approach to human disease. *Nature Reviews. Genetics*, *12*(1), 56–68. doi:10.1038/nrg2918 PMID:21164525

Baradaran, N., Tan, S. N., Liu, A., Ashoori, A., Palmer, S. J., Wang, Z. J., ... McKeown, M. J. (2013). Parkinson's Disease Rigidity: Relation to Brain Connectivity and Motor Performance. *Frontiers in Neurology*, *4*, 67. doi:10.3389/fneur.2013.00067 PMID:23761780

Baran, E. J. (2000). Metal complexes of carnosine. Biochemistry, 65, 789-797. PMID:10951097

Baranello, R. J., Bharani, K. L., Padmaraju, V., Chopra, N., Lahiri, D. K., Greig, N. H., ... Sambamurti, K. (2015). Amyloid-beta protein clearance and degradation (ABCD) pathways and their role in Alzheimer's disease. *Current Alzheimer Research*, *12*(1), 32–46. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/25523424

Barker, W. W., Luis, C. A., Kashuba, A., Luis, M., Harwood, D. G., Loewenstein, D., ... Sevush, S. (2002). Relative frequencies of Alzheimer disease, Lewy body, vascular and frontotemporal dementia, and hippocampal sclerosis in the State of Florida Brain Bank. *Alzheimer Disease and Associated Disorders*, *16*(4), 203–212. doi:10.1097/00002093-200210000-00001 PMID:12468894

Baron-Cohen, S., Lombardo, M. V., Auyeung, B., Ashwin, E., Chakrabarti, B., & Knickmeyer, R. (2011). Why are autism spectrum conditions more prevalent in males? *PLoS Biology*, *9*(6), 1001081. doi:10.1371/journal.pbio.1001081 PMID:21695109

Barreto, G. E., Ganzalez, J., Torres, Y., & Morales, L. (2011). Astrocytic-neuronal crosstalk: Implications for neuroprotection from brain injury. *Neuroscience Research*, 71(2), 107–113. doi:10.1016/j.neures.2011.06.004 PMID:21693140

Barreto, G. E., Santos-Galindo, M., & Garcia-Segura, L. M. (2014). Selective estrogen receptor modulators regulate reactive microglia after penetrating brain injury. *Frontiers in Aging Neuroscience*, *6*, 132. doi:10.3389/fnagi.2014.00132 PMID:24999330

Barten, D. M., Meredith, J. E. Jr, Zaczek, R., Houston, J. G., & Albright, C. F. (2006). Gamma-secretase inhibitors for Alzheimer's disease: Balancing efficacy and toxicity. *Drugs in R&D.*, 7(2), 87–97. doi:10.2165/00126839-200607020-00003 PMID:16542055

Bartus, R. T., Dean, R., Beer, B., & Lippa, A. (1982). The cholinergic hypothesis of geriatric memory dysfunction. *Science*, 217(4558), 408–414. doi:10.1126cience.7046051 PMID:7046051

Baslow, M. H., & Guilfoyle, D. N. (2013). Canavan disease, a rare early-onset human spongiform leukodystrophy: Insights into its genesis and possible clinical interventions. *Biochimie*, 95(4), 946–956. doi:10.1016/j.biochi.2012.10.023 PMID:23151389

Bassett, A. S., & Chow, E. W. (1999). 22q11 deletion syndrome: A genetic subtype of schizophrenia. *Biological Psychiatry*, *46*(7), 882–891. doi:10.1016/S0006-3223(99)00114-6 PMID:10509171

Basso, M., Massignan, T., Samengo, G., Cheroni, C., De Biasi, S., Salmona, M., ... Bonetto, V. (2006). Insoluble mutant SOD1 is partly oligoubiquitinated in amyotrophic lateral sclerosis mice. *The Journal of Biological Chemistry*, 281(44), 33325–33335. doi:10.1074/jbc.M603489200 PMID:16943203

Baughman, J. M., Perocchi, F., Girgis, H. S., Plovanich, M., Belcher-Timme, C. A., Sancak, Y., ... Mootha, V. K. (2011). Integrative genomics identifies MCU as an essential component of the mitochondrial calcium uniporter. *Nature*, 476(7360), 341–345. doi:10.1038/nature10234 PMID:21685886

Bäumer, D., Talbot, K., & Turner, M. R. (2014). Advances in motor neurone disease. *Journal of the Royal Society of Medicine*, *107*(1), 14–21. doi:10.1177/0141076813511451 PMID:24399773

Bayram, B., Esatbeyoglu, T., Schulze, N., Ozcelik, B., Frank, J., & Rimbach, G. (2012). Comprehensive analysis of polyphenols in 55 extra virgin olive oils by HPLC-ECD and their correlation with antioxidant activities. *Plant Foods for Human Nutrition (Dordrecht, Netherlands)*, 67(4), 326–336. doi:10.100711130-012-0315-z PMID:23070730

Beal, M. F. (2005). Mitochondria take center stage in aging and neurodegeneration. *Annals of Neurology*, 58(4), 495–505. doi:10.1002/ana.20624 PMID:16178023

Becker, C., Jick, S. S., & Meier, C. R. (2008). Use of antihypertensives and the risk of Parkinson disease. *Neurology*, 70(16 Part 2), 1438–1444. doi:10.1212/01.wnl.0000303818.38960.44 PMID:18256367

Beckman, K. B., & Ames, B. N. (1998). The free radical theory of aging matures. *Physiological Reviews*, 78(2), 547–581. doi:10.1152/physrev.1998.78.2.547 PMID:9562038

Bedford, L., Hay, D., Devoy, A., Paine, S., Powe, D. G., Seth, R., ... Mee, M. (2008). Depletion of 26S proteasomes in mouse brain neurons causes neurodegeneration and Lewy-like inclusions resembling human pale bodies. *The Journal of Neuroscience*, 28(33), 8189–8198. doi:10.1523/JNEUROSCI.2218-08.2008 PMID:18701681

Beeldman, E., Raaphorst, J., Twennaar, M. K., de Visser, M., Schmand, B. A., & de Haan, R. J. (2016). The cognitive profile of ALS: A systematic review and metaanalysis update. *Journal of Neurology, Neurosurgery, and Psychiatry*, 87(6), 611–619. doi:10.1136/jnnp-2015-310734 PMID:26283685

Beghi, E. (2013). Are professional soccer players at higher risk for ALS? *Amyotrophic Lateral Sclerosis & Frontotemporal Degeneration*, 14(7-8), 501–506. doi:10.3109/21678421.2013.809764 PMID:23859483

Beghi, E., Balzarini, C., Bogliun, G., Logroscino, G., Manfredi, L., Mazzini, L., ... Vitelli, E. (2002). Reliability of the El Escorial diagnostic criteria for amyotrophic lateral sclerosis. *Neuroepidemiology*, *21*(6), 265–270. doi:10.1159/000065524 PMID:12411728

Beghi, E., Millul, A., Micheli, A., Vitelli, E., & Logroscino, G. (2007). Incidence of ALS in Lombardy, Italy. *Neurology*, 68(2), 141–145. doi:10.1212/01.wnl.0000250339.14392.bb PMID:17210896

Beilina, A., Rudenko, I. N., Kaganovich, A., Civiero, L., Chau, H., Kalia, S. K., ... Cookson, M. R. (2014). Unbiased screen for interactors of leucine-rich repeat kinase 2 supports a common pathway for sporadic and familial Parkinson disease. *Proceedings of the National Academy of Sciences of the United States of America*, *111*(7), 2626–2631. doi:10.1073/ pnas.1318306111 PMID:24510904

Beiske, A. G., Loge, J. H., Rønningen, A., & Svensson, E. (2009). Pain in Parkinson's disease: Prevalence and characteristics. Pain. *International Association for the Study of Pain*, *141*(1), 173–177. PMID:19100686

Bekris, L. M., Mata, I. F., & Zabetian, C. P. (2010). The genetics of Parkinson disease. *Journal of Geriatric Psychiatry* and Neurology, 23(4), 228–242. doi:10.1177/0891988710383572 PMID:20938043

Bekris, L. M., Yu, C. E., Bird, T. D., & Tsuang, D. W. (2010). Review article: Genetics of Alzheimer disease. *Journal of Geriatric Psychiatry and Neurology*, 23(4), 213–227. doi:10.1177/0891988710383571 PMID:21045163

Belay, E. D. (2004). Prions and Prion Diseases: Current Perspectives. *Emerging Infectious Diseases*, *10*(12), 2265–2266. doi:10.3201/eid1012.3040847

Belkacemi, A., & Ramassamy, C. (2012). Time sequence of oxidative stress in the brain from transgenic mouse models of Alzheimer's disease related to the amyloid-β cascade. *Free Radical Biology & Medicine*, *52*(3), 593–600. doi:10.1016/j. freeradbiomed.2011.11.020 PMID:22172527

Benabid,, A. L., Krack, P. P., Benazzouz, A., Limousin, P., Koudsie, A., & Pollak, P. (2000). Deep brain stimulation of the subthalamic nucleus for Parkinson's disease: Methodologic aspects and clinical criteria. *Neurology*, *55*(12Suppl 6), S40–S44. PMID:11188974

Benabid, A. L. (2007, November). What the future holds for deep brain stimulation. *Expert Review of Medical Devices*, 4(6), 895–903. doi:10.1586/17434440.4.6.895 PMID:18035954

Benabid, A. L. (2009). Targeting the caudal intralaminar nuclei for functional neurosurgery of movement disorders. *Brain Research Bulletin*, 78(2-3), 109–112. doi:10.1016/j.brainresbull.2008.08.020 PMID:18812212

Benajiba, L., Le Ber, I., Camuzat, A., Lacoste, M., Thomas-Anterion, C., Couratier, P., ... Brice, A. (2009). TARDBP mutations in motoneuron disease with frontotemporal lobar degeneration. *Annals of Neurology*, *65*(4), 470–473. doi:10.1002/ana.21612 PMID:19350673

Benchoua, A., Trioulier, Y., Zala, D., Gaillard, M.-C., Lefort, N., Dufour, N., ... Hantraye, P. (2006). Involvement of mitochondrial complex II defects in neuronal death produced by N-terminus fragment of mutated huntingtin. *Molecular Biology of the Cell*, *17*(4), 1652–1663. doi:10.1091/mbc.E05-07-0607 PMID:16452635

Bender, A., Krishnan, K. J., Morris, C. M., Taylor, G. A., Reeve, A. K., Perry, R. H., ... Klopstock, T. (2006). High levels of mitochondrial DNA deletions in substantia nigra neurons in aging and Parkinson disease. *Nature Genetics*, *38*(5), 515–517. doi:10.1038/ng1769 PMID:16604074

Bendor, J. T., Logan, T. P., & Edwards, R. H. (2013). The function of alpha-synuclein. *Neuron*, 79(6), 1044–1066. doi:10.1016/j.neuron.2013.09.004 PMID:24050397

Bennett, B. D., Denis, P., Haniu, M., Teplow, D. B., Kahn, S., Louis, J.-C., ... Vassar, R. (2000). A Furin-like Convertase Mediates Propeptide Cleavage of BACE, the Alzheimer's β-Secretase. *The Journal of Biological Chemistry*, 275(48), 37712–37717. doi:10.1074/jbc.M005339200 PMID:10956649

Bennett, D. A., Wilson, R. S., Gilley, D. W., & Fox, J. H. (1990). Clinical diagnosis of Binswanger's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 53(11), 961–965. doi:10.1136/jnnp.53.11.961 PMID:2283526

Bennett, S., Grant, M. M., & Aldred, S. (2009). Oxidative stress in vascular dementia and Alzheimer's disease: A common pathology. *Journal of Alzheimer's Disease*, 17(2), 245–257. doi:10.3233/JAD-2009-1041 PMID:19221412

Bensimon, G., Lacomblez, L., & Meininger, V. (1994). A controlled trial of riluzole in amyotrophic lateral sclerosis: ALS/ Riluzole Study Group. *The New England Journal of Medicine*, *330*(9), 585–591. doi:10.1056/NEJM199403033300901 PMID:8302340

Bentmann, E., Neumann, M., Tahirovic, S., Rodde, R., Dormann, D., & Haass, C. (2012). Requirements for stress granule recruitment of fused in sarcoma (FUS) and TAR DNA-binding protein of 43 kDa (TDP-43). *The Journal of Biological Chemistry*, 287(27), 23079–23094. doi:10.1074/jbc.M111.328757 PMID:22563080

Bento, C. F., Ashkenazi, A., Jimenez-Sanchez, M., & Rubinsztein, D. C. (2016). The Parkinson's disease-associated genes *ATP13A2* and *SYT11* regulate autophagy via a common pathway. *Nature Communications*, 7, 11803. doi:10.1038/ ncomms11803 PMID:27278822

Berg, A. T., Berkovic, S. F., Brodie, M. J., Buchhalter, J., Cross, J. H., van Emde Boas, W., ... Scheffer, I. E. (2010). Revised terminology and concepts for organization of seizures and epilepsies: Report of the ILAE Commission on Classification and Terminology, 2005–2009. *Epilepsia*, *51*(4), 676–685. doi:10.1111/j.1528-1167.2010.02522.x PMID:20196795

Berg, D., Postuma, R. B., Bloem, B., Chan, P., Dubois, B., Gasser, T., ... Deuschl, G. (2014). Time to Redefine PD? Introductory Statement of the MDS Task Force on the Definition of Parkinson's Disease. *Movement Disorders*, 29(4), 454–462. doi:10.1002/mds.25844 PMID:24619848

Bergersen, L. H. (2015). Lactate Transport and Signaling in the Brain: Potential Therapeutic Targets and Roles in Body—Brain Interaction. *Journal of Cerebral Blood Flow and Metabolism*, 35(2), 176–185. doi:10.1038/jcbfm.2014.206 PMID:25425080

Berger, Z., Smith, K. A., & LaVoie, M. J. (2010). Membrane localization of LRRK2 is associated with increased formation of the highly active LRRK2 dimer and changes in its phosphorylation. *Biochemistry*, 49(26), 5511–5523. doi:10.1021/bi100157u PMID:20515039

Berk, C., & Sabbagh, M. N. (2013). Successes and failures for drugs in late-stage development for Alzheimer's disease. *Drugs & Aging*, *30*(10), 783–792. doi:10.100740266-013-0108-6 PMID:23943247

Bernadeta, S. (2013). Zinc homeostasis and neurodegenerative disorders. *Frontiers in Aging Neuroscience*, 5, 33. PMID:23882214

Berridge, M. J. (2010). Calcium hypothesis of Alzheimer's disease. *Pflügers Archiv*, 459(3), 441–449. doi:10.100700424-009-0736-1 PMID:19795132

Bersano, A., Bedini, G., Oskam, J., Mariotti, C., Taroni, F., Baratta, S., & Parati, E. A. (2017). CADASIL: Treatment and Management Options. *Current Treatment Options in Neurology*, 19(9), 31. doi:10.100711940-017-0468-zPMID:28741120

Berument, S. K., Rutter, M., Lord, C., Pickles, A., & Bailey, A. (1999). Autism screening questionnaire: Diagnostic validity. *The British Journal of Psychiatry*, *175*(05), 444–451. doi:10.1192/bjp.175.5.444 PMID:10789276

Betarbet, R., Sherer, T. B., MacKenzie, G., Garcia-Osuna, M., Panov, A. V., & Greenamyre, J. T. (2000). Chronic systemic pesticide exposure reproduces features of Parkinson's disease. *Nature Neuroscience*, *3*(12), 1301–1306. doi:10.1038/81834 PMID:11100151

Beurel, E., Grieco, S. F., & Jope, R. S. (2015). Glycogen synthase kinase-3 (GSK3): Regulation, actions, and diseases. *Pharmacology & Therapeutics*, 0, 114–131. doi:10.1016/j.pharmthera.2014.11.016 PMID:25435019

Beyer, M. K., Janvin, C. C., Larsen, J. P., & Aarsland, D. (2007). A magnetic resonance imaging study of patients with Parkinson's disease with mild cognitive impairment and dementia using voxel-based morphometry. *Journal of Neurology, Neurosurgery, and Psychiatry*, 78(3), 254–259. doi:10.1136/jnnp.2006.093849 PMID:17028119

Bhat, A. H., Dar, K. B., Anees, S., Zargar, M. A., Masood, A., Sofi, M. A., & Ganie, S. A. (2015). Oxidative stress, mitochondrial dysfunction and neurodegenerative diseases; a mechanistic insight. *Biomedicine and Pharmacotherapy*, 74, 101–110. doi:10.1016/j.biopha.2015.07.025 PMID:26349970

Biasini, E., Turnbaugh, J. A., Unterberger, U., & Harris, D. A. (2012). Prion protein at the crossroads of physiology and disease. *Trends in Neurosciences*, *35*(2), 92–103. doi:10.1016/j.tins.2011.10.002 PMID:22137337

Bickford, P. C., Flowers, A., & Grimmig, B. (2017). Aging leads to altered microglial function that reduces brain resiliency increasing vulnerability to neurodegenerative diseases. *Experimental Gerontology*, *94*, 4–8. doi:10.1016/j. exger.2017.01.027 PMID:28163132

Bidichandani, S. I., & Delatycki, M. B. (2017). Friedreich ataxia. Academic Press.

Bilheude, J., Uro-coste, E., Basset-leobon, C., Perret-liaudet, A., Ironside, J. W., Peoch, K., & Andre, O. (2008). Beyond PrP res Type 1 / Type 2 Dichotomy in Creutzfeldt- Jakob Disease. *PLoS Pathogens*, 4(3), 1–9. PMID:18383623

Billette de Villemeur, T., Deslys, J. P., Pradel, A., Soubrié, C., Alpérovitch, A., Tardieu, M., & Agid, Y. (1996). Creutzfeldt-Jakob disease from contaminated growth hormone extracts in France. *Neurology*, *47*(3), 690–695. doi:10.1212/ WNL.47.3.690 PMID:8797466

Bilsland, L. G., Dick, J. R., Pryce, G., Petrosino, S., Di Marzo, V., Baker, D., & Greensmith, L. (2006). Increasing cannabinoid levels by pharmacological and genetic manipulation delay disease progression in SOD1 mice. *The FASEB Journal*, *20*(7), 1003–1005. doi:10.1096/fj.05-4743fje PMID:16571781

Binder, L. I., Frankfurter, A., & Rebhun, L. I. (1985). The distribution of tau in the mammalian central nervous system. *The Journal of Cell Biology*, *101*(4), 1371–1378. doi:10.1083/jcb.101.4.1371 PMID:3930508

Biogen. (2017). *Research Pipeline*. Retrieved March 7, 2017, from https://www.biogen.com/en_us/research-pipeline/biogen-pipeline.html

Birben, E., Sahiner, U., Sackesen, C., Erzurum, S., & Kalayci, O. (2012). Oxidative stress and antioxidant defense. *The World Allergy Organization Journal*, *5*(1), 9–19. doi:10.1097/WOX.0b013e3182439613 PMID:23268465

Bishop, N. A., Lu, T., & Yankner, B. A. (2010). Neural mechanisms of ageing and cognitive decline. *Nature*, 464(7288), 529–535. doi:10.1038/nature08983 PMID:20336135

Bishop, S. J. (2007). Neurocognitive mechanisms of anxiety: An integrative account. *Trends in Cognitive Sciences*, *11*(7), 307–316. doi:10.1016/j.tics.2007.05.008 PMID:17553730

Biskup, S., & West, A. B. (2009). Zeroing in on LRRK2-linked pathogenic mechanisms in Parkinson's disease. *Biochimica et Biophysica Acta (BBA)-*. *Molecular Basis of Disease*, 1792(7), 625–633. doi:10.1016/j.bbadis.2008.09.015 PMID:18973807

Bjartmarz, H., & Rehncrona, S. (2007). Comparison of accuracy and precision between frame-based and frameless stereotactic navigation for deep brain stimulation electrode implantation. *Stereotactic and Functional Neurosurgery*, 85(5), 235–242. doi:10.1159/000103262 PMID:17534136

Björkblom, B., Adilbayeva, A., Maple-Grødem, J., Piston, D., Ökvist, M., Xu, X. M., ... Møller, S. G. (2013). Parkinson Disease Protein DJ-1 Binds Metals and Protects against Metal-induced Cytotoxicity. *The Journal of Biological Chemistry*, 288(31), 22809–22820. doi:10.1074/jbc.M113.482091 PMID:23792957

Blalock, E. M., Buechel, H. M., Popovic, J., Geddes, J. W., & Landfield, P. W. (2011). Microarray analyses of lasercaptured hippocampus reveal distinct gray and white matter signatures associated with incipient Alzheimer's disease. *Journal of Chemical Neuroanatomy*, 42(2), 118–126. doi:10.1016/j.jchemneu.2011.06.007 PMID:21756998

Bloom, G. S. (2014). Amyloid-β and Tau. JAMA Neurology, 71(4), 505. doi:10.1001/jamaneurol.2013.5847 PMID:24493463

Blümcke, I., Thom, M., Aronica, E., Armstrong, D. D., Bartolomei, F., Bernasconi, A., ... Spreafico, R. (2013). International consensus classification of hippocampal sclerosis in temporal lobe epilepsy: A Task Force report from the ILAE Commission on Diagnostic Methods. *Epilepsia*, *54*(7), 1315–1329. doi:10.1111/epi.12220 PMID:23692496

Bøe, R., Gjertsen, B. T., Vintermyr, O. K., Houge, G., Lanotte, M., & Døskeland, S. O. (1991). The protein phosphatase inhibitor okadaic acid induces morphological changes typical of apoptosis in mammalian cells. *Experimental Cell Research*, *195*(1), 237–246. doi:10.1016/0014-4827(91)90523-W PMID:1647324

Boeve, B. F., Silber, M. H., Ferman, T. J., Lin, S. C., Benarroch, E. E., Schmeichel, A. M., ... Dickson, D. W. (2013). Clinicopathologic correlations in 172 cases of rapid eye movement sleep behavior disorder with or without a coexisting neurologic disorder. *Sleep Medicine*, *14*(8), 754–762. doi:10.1016/j.sleep.2012.10.015 PMID:23474058

Bohnen, N. I., & Albin, R. L. (2011). The Cholinergic System and Parkinson Disease. *Behavioural Brain Research*, 221(2), 564–573. doi:10.1016/j.bbr.2009.12.048 PMID:20060022

Bohnen, N. I., Müller, M. L. T. M., Koeppe, R. A., Studenski, S. A., Kilbourn, M. A., Frey, K. A., & Albin, R. L. (2009). History of falls in Parkinson disease is associated with reduced cholinergic activity. *Neurology*, *73*(20), 1670–1676. doi:10.1212/WNL.0b013e3181c1ded6 PMID:19917989

Bohnen, N. I., Müller, M. L. T. M., Kotagal, V., Koeppe, R. A., Kilbourn, M. R., Gilman, S., ... Frey, K. A. (2012). Heterogeneity of cholinergic denervation in Parkinson's disease without dementia. *Journal of Cerebral Blood Flow and Metabolism*, *32*(8), 1609–1617. doi:10.1038/jcbfm.2012.60 PMID:22569194

Bolisetty, S., & Jaimes, E. A. (2013). Mitochondria and reactive oxygen species: Physiology and pathophysiology. *International Journal of Molecular Sciences*, *14*(3), 6306–6344. doi:10.3390/ijms14036306 PMID:23528859

Bolton, P., Macdonald, H., Pickles, A., Rios, P., Goode, S., Crowson, M. A., ... Rutter, M. (1994). Case-control family history study of autism. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, *35*(5), 877–900. doi:10.1111/j.1469-7610.1994.tb02300.x PMID:7962246

Bonda, D. J., Manjila, S., Mehndiratta, P., Khan, F., Miller, B. R., Onwuzulike, K., & Cali, I. (2016). Human prion diseases: Surgical lessons learned from iatrogenic prion transmission. *Neurosurgical Focus*, *41*(1), E10. doi:10.3171/2016.5.FOCUS15126 PMID:27364252

Bonifati, V., Rizzu, P., Van Baren, M. J., Schaap, O., Breedveld, G. J., Krieger, E., & van Dongen, J. W. (2003). Mutations in the DJ-1 gene associated with autosomal recessive early-onset Parkinsonism. *Science*, 299(5604), 256–259. doi:10.1126cience.1077209 PMID:12446870

Bonuccelli, U., Del Dotto, P., & Rascol, O. (2009). Role of dopamine receptor agonists in the treatment of early Parkinson's disease. *Parkinsonism & Related Disorders*, *15*, S44–S53. doi:10.1016/S1353-8020(09)70835-1 PMID:20123557

Boot, B. P., Orr, C. F., Ahlskog, J. E., Ferman, T. J., Roberts, R., Pankratz, V. S., ... Boeve, B. F. (2013). Risk factors for dementia with Lewy bodies: A case-control study. *Neurology*, *81*(9), 833–840. doi:10.1212/WNL.0b013e3182a2cbd1 PMID:23892702

Borchelt, D. R., Lee, M. K., Slunt, H. S., Guarnieri, M., Xu, Z. S., Wong, P. C., Brown, R. H., Jr., & Cleveland, D. W. (1994). Superoxide dismutase 1 with mutations linked to familial amyotrophic lateral sclerosis possesses significant activity. *Proceedings of the National Academy of Sciences*, *91*, 8292–8296.

Borchelt, D. R., Guarnieri, M., Wong, P. C., Lee, M. K., Slunt, H. S., Xu, Z. S., ... Cleveland, D. W. (1995). Superoxide dismutase 1 subunits with mutations linked to familial amyotrophic lateral sclerosis do not affect wild-type subunit function. *The Journal of Biological Chemistry*, 270(7), 3234–3238. doi:10.1074/jbc.270.7.3234 PMID:7852409

Bordet, T., Buisson, B., Michaud, M., Drouot, C., Galea, P., Delaage, P., ... Lacapere, J. J. (2007). Identification and characterization of cholest-4-en-3-one, oxime (TRO19622), a novel drug candidate for amyotrophic lateral sclerosis. *The Journal of Pharmacology and Experimental Therapeutics*, *322*(2), 709–720. doi:10.1124/jpet.107.123000 PMID:17496168

Borovac, J. A. (2016). Side effects of a dopamine agonist therapy for Parkinson's disease: A mini-review of clinical pharmacology. *The Yale Journal of Biology and Medicine*, 89(1), 37–47. PMID:27505015

Borroni, B., Bonvicini, C., Alberici, A., Buratti, E., Agosti, C., Archetti, S., ... Padovani, A. (2009). Mutation within TARDBP leads to frontotemporal dementia without motor neuron disease. *Human Mutation*, *30*(11), E974–E983. doi:10.1002/humu.21100 PMID:19655382

Borroni, B., Premi, E., Formenti, A., Turrone, R., Alberici, A., Cottini, E., ... Padovani, A. (2015). Structural and functional imaging study in dementia with Lewy bodies and Parkinson's disease dementia. *Parkinsonism & Related Disorders*, *21*(9), 1049–1055. doi:10.1016/j.parkreldis.2015.06.013 PMID:26109553

Borsello, T., Clarke, P. G., Hirt, L., Vercelli, A., Repici, M., Schorderet, D. F., ... Bonny, C. (2003). A peptide inhibitor of c-Jun N-terminal kinase protects against excitotoxicity and cerebral ischemia. *Nature Medicine*, *9*(9), 1180–1186. doi:10.1038/nm911 PMID:12937412

Bor-Seng-Shu, E., Fonoff, E. T., Barbosa, E. R., & Teixeira, M. J. (2010, December). Substantia nigra hyperechogenicity in Parkinson's disease. *Acta Neurochirurgica*, *152*(12), 2085–2087. doi:10.100700701-010-0736-0 PMID:20623147

Bosgraaf, L., & Van Haastert, P. J. (2003). Roc, a Ras/GTPase domain in complex proteins. *Biochimica et Biophysica Acta (BBA)-*. *Molecular Cell Research*, *1643*(1), 5–10.

Botella, J. A., Bayersdorfer, F., & Schneuwly, S. (2008). Superoxide dismutase overexpression protects dopaminergic neurons in a Drosophila model of Parkinson's disease. *Neurobiology of Disease*, *30*(1), 65–73. doi:10.1016/j. nbd.2007.11.013 PMID:18243716

Boucard, C. C., Hernowo, A. T., Maguire, R. P., Jansonius, N. M., Roerdink, J. B., Hooymans, J. M., & Cornelissen, F. W. (2009). Changes in cortical grey matter density associated with long-standing retinal visual field defects. *Brain*, *132*(7), 1898–1906. doi:10.1093/brain/awp119 PMID:19467992

Bouteille, M., Fontaine, C., & Vedrenne, C. L. (1965). Sur uncas d'encephalite subaiguea inclusions. Etude anatomoclinique et ultra structurale. *Revue Neurologique*, *113*, 454–458.

Boveris, A., & Navarro, A. (2008). Brain mitochondrial dysfunction in aging. *IUBMB Life*, 60(5), 308–314. doi:10.1002/ iub.46 PMID:18421773

Bowler, J., & Hachinsky, V. (1996). History of the concept of vascular dementia: two opposing views on current definitions and criteria for vascular dementia. *Vascular dementia: current concepts*, 1-24.

Bowling, A. C., Schulz, J. B., Brown, R. H. Jr, & Beal, M. F. (1993). Superoxide dismutase activity, oxidative damage, and mitochondrial energy metabolism in familial and sporadic amyotrophic lateral sclerosis. *Journal of Neurochemistry*, *61*(6), 2322–2325. doi:10.1111/j.1471-4159.1993.tb07478.x PMID:8245985

Boxer, A. L., Mackenzie, I. R., Boeve, B. F., Baker, M., Seeley, W. W., Crook, R., ... Rademakers, R. (2011). Clinical, neuroimaging and neuropathological features of a new chromosome 9p-linked FTD-ALS family. *Journal of Neurology, Neurosurgery, and Psychiatry*, 82(2), 196–203. doi:10.1136/jnnp.2009.204081 PMID:20562461

Boyle, P. A., Yu, L., Nag, S., Leurgans, S., Wilson, R. S., Bennett, D. A., & Schneider, J. A. (2015). Cerebral amyloid angiopathy and cognitive outcomes in community-based older persons. *Neurology*, 85(22), 1930–1936. doi:10.1212/WNL.000000000002175 PMID:26537052

Braak, H., & Braak, E. (1991). Neuropathological stageing of Alzheimer-related changes. *Acta Neuropathologica*, 82(4), 239–259. doi:10.1007/BF00308809 PMID:1759558

Braak, H., & Braak, E. (2000). Pathoanatomy of Parkinson's disease. *Journal of Neurology*, 247(S2), II3–II10. doi:10.1007/PL00007758 PMID:10991663

Braak, H., Del Tredici, K., Bratzke, H., Hamm-Clement, J., Sandmann-Keil, D., & Rüb, U. (2002). Staging of the intracerebral inclusion body pathology associated with idiopathic Parkinson's disease (preclinical and clinical stages). *Journal of Neurology*, *249*(0), III-1. doi:10.100700415-002-1301-4 PMID:12528692

Braak, H., Del Tredici, K., Rüb, U., de Vos, R. A., Steur, E. N. J., & Braak, E. (2003). Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiology of Aging*, 24(2), 197–211. doi:10.1016/S0197-4580(02)00065-9 PMID:12498954

Braak, H., Rüb, U., Gai, W. P., & Del Tredici, K. (2003). Idiopathic Parkinson's disease: Possible routes by which vulnerable neuronal types may be subject to neuroinvasion by an unknown pathogen. *Journal of Neural Transmission (Vienna, Austria)*, *110*(5), 517–536. doi:10.100700702-002-0808-2 PMID:12721813

Braak, H., Rüb, U., Steur, E. J., Del Tredici, K., & De Vos, R. A. I. (2005). Cognitive status correlates with neuropathologic stage in Parkinson disease. *Neurology*, 64(8), 1404–1410. doi:10.1212/01.WNL.0000158422.41380.82 PMID:15851731

Brady, R. O., Kanfer, J., & Shapiro, D. (1965). The metabolism of glucocerebrosides.I. Purification and properties of a glucocerebroside-cleaving enzyme from spleen tissue. *The Journal of Biological Chemistry*, 240, 39–43. PMID:14253443

Brandel, J. P., Salomon, D., Capek, I., Vaillant, V., & Alpérovitch, A. (2009). Epidemiological surveillance of Creutzfeldt-Jakob in France. *Revue Neurologique*, *165*(8-9), 684–693. doi:10.1016/j.neurol.2009.04.006 PMID:19467685

Brandt, T. (Ed.). (2003). Neurological disorders: course and treatment. Gulf Professional Publishing.

Bratic, A., & Larsson, N. G. (2013). The role of mitochondria in aging. *The Journal of Clinical Investigation*, *123*(3), 951–957. doi:10.1172/JCI64125 PMID:23454757

Brazier, M. W., Doctrow, S. R., Masters, C. L., & Collins, S. J. (2008). A manganese-superoxide dismutase/catalase mimetic extends survival in a mouse model of human prion disease. *Free Radical Biology & Medicine*, 45(2), 184–192. doi:10.1016/j.freeradbiomed.2008.04.006 PMID:18455516

Bredesen, D. E., Rao, R. V., & Mehlen, P. (2006). Cell death in the nervous system. *Nature*, 443(7113), 796–802. doi:10.1038/nature05293 PMID:17051206

Brenner, M., Goldman, J. E., Quinlan, R. A., & Messing, A. (2009). Alexander Disease: A Genetic Disorder of Astrocytes. In P. Haydon & V. Parpura (Eds.), *Astrocytes in (Patho)Physiology of the Nervous System* (pp. 591–648). Boston, MA: Springer. doi:10.1007/978-0-387-79492-1_24

Brettschneider, J., Del Tredici, K., & Lee, V. M. (2015). Spreading of pathology in neurodegenerative diseases: A focus on human studies. *Nature Reviews. Neuroscience*, *16*(2), 109–120. doi:10.1038/nrn3887 PMID:25588378

Brettschneider, J., Libon, D. J., Toledo, J. B., Xie, S. X., McCluskey, L., Elman, L., ... Trojanowski, J. Q. (2012). Microglial activation and TDP-43 pathology correlate with executive dysfunction in amyotrophic lateral sclerosis. *Acta Neuropathologica*, *123*(3), 395–407. doi:10.100700401-011-0932-x PMID:22210083

Brewer, G. J., Kanzer, S. H., Zimmerman, E. A., Molho, E. S., Celmins, D. F., Heckman, S. M., & Dick, R. (2010). Subclinical zinc deficiency in Alzheimer's disease and parkinson's disease. *American Journal of Alzheimer's Disease and Other Dementias*, 2(7), 572–575. doi:10.1177/1533317510382283 PMID:20841345

Breydo, L., Wu, J. W., & Uversky, V. N. (2012). α-Synuclein misfolding and Parkinson's disease. *Biochimica et Biophysica* Acta (BBA)-. Molecular Basis of Disease, 1822(2), 261–285. doi:10.1016/j.bbadis.2011.10.002

Brieger, K., Schiavone, S., Miller, F. J. Jr, & Krause, K. H. (2012). Reactive oxygen species: From health to disease. *Swiss Medical Weekly*, *142*, w13659. PMID:22903797

Brissaud, É. (1895). Leçons sur les maladies nerveuses: Salpêtrière, 1893-1894 (Vol. 1). G. Masson.

Brocks, D. R. (1999). Anticholinergic drugs used in Parkinson's disease: An overlooked class of drugs from a pharmacokinetic perspective. *Journal of Pharmacy & Pharmaceutical Sciences*, 2(2), 39–46. PMID:10952768

Bronen, R. A., Cheung, C., & Charles, J. T. (1991). Imaging findings in hippocampal sclerosis: Correlation whith Patology. *AJNR. American Journal of Neuroradiology*, *12*, 933–940. PMID:1950925

Bronisz, A., Carey, H. A., Godlewski, J., Sif, S., Ostrowski, M. C., & Sharma, S. M. (2014). The multifunctional protein Fused in Sarcoma (FUS) is a coactivator of Microphthalmia-associated Transcription Factor (MITF). *The Journal of Biological Chemistry*, 289(1), 326–334. doi:10.1074/jbc.M113.493874 PMID:24257758

Bronstein, J. M., Tagliati, M., Alterman, R. L., Lozano, A. M., Volkmann, J., Stefani, A., ... DeLong, M. R. (2011, February). Deep brain stimulation for Parkinson disease: An expert consensus and review of key issues. *Archives of Neurology*, 68(2), 165. doi:10.1001/archneurol.2010.260 PMID:20937936

Brookmeyer, R., Gray, S., & Kawas, C. (1998). Projections of Alzheimer's disease in the United States and the public health impact of delaying disease onset. *American Journal of Public Health*, 88(9), 1337–1342. doi:10.2105/AJPH.88.9.1337 PMID:9736873

Brooks, B. R., Miller, R. G., Swash, M., & Munsat, T. L. (2000). El Escorial revisited: Revised criteria for the diagnosis of amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, *41*(5), 293–299. doi:10.1080/146608200300079536 PMID:11464847

Brooks, D. J. (2002). Diagnosis and management of atypical parkinsonian symptoms. *Journal of Neurology, Neurosurgery, and Psychiatry*, 72, i10–i16. PMID:11870198

Brouillet, E., Hantraye, P., Ferrante, R. J., Dolan, R., Leroy-Willig, A., Kowall, N. W., & Beal, M. F. (1995). Chronic mitochondrial energy impairment produces selective striatal degeneration and abnormal choreiform movements in primates. *Proceedings of the National Academy of Sciences of the United States of America*, 92(15), 7105–7109. doi:10.1073/ pnas.92.15.7105 PMID:7624378

Brown, C. E., & Antholine, W. E. (1979). Chelation chemistry of carnosine. Evidence that mixed complexes may occur in vivo. *Journal of Physical Chemistry*, *83*(26), 3314–3319. doi:10.1021/j100489a002

Brown, D. R. (2005). ORGINAL ARTICLE Neurodegeneration and oxidative stress: Prion disease results from loss of antioxidant defence. *Folia Neuropathologica*, 43(4), 229–243. PMID:16416388

Brown, D. R., Boon-Seng, W., Hafiz, F., & Clive, C. (1999). Normal prion protein has an activity like that of superoxide dismutase. *The Biochemical Journal*, *344*(1), 1–5. doi:10.1042/bj3440001 PMID:10548526

Brown, D. R., Schmidt, B., & Kretzschmar, H. A. (1997). Effects of oxidative stress on prion protein expression in PC12 cells. *International Journal of Developmental Neuroscience*, *15*(8), 961–972. doi:10.1016/S0736-5748(97)00042-7 PMID:9641527

Brown, D. R., Schmidt, B., & Kretzschmar, H. A. (1998). A prion protein fragment primes type 1 astrocytes to proliferation signals from microglia. *Neurobiology of Disease*, 4(6), 410–422. doi:10.1006/nbdi.1998.0169 PMID:9666480

Brown, D. R., Schulz-Schaeffer, W. J., Schmidt, B., & Kretzschmar, H. A. (1997). Prion Protein-Deficient Cells Show Altered Response to Oxidative Stress Due to Decreased SOD-1 Activity. *Experimental Neurology*, *146*(1), 104–112. doi:10.1006/exnr.1997.6505 PMID:9225743

Browne, S. E., Bowling, A. C., Baik, M. J., Gurney, M., Brown, R. H. Jr, & Beal, M. F. (1998). Metabolic dysfunction in familial, but not sporadic, amyotrophic lateral sclerosis. *Journal of Neurochemistry*, *71*(1), 281–287. doi:10.1046/j.1471-4159.1998.71010281.x PMID:9648876

Brown, P., Rodgers-Johnson, P., Cathala, F., Gibbs, C. J., & Gajdusek, D. C. (1984). Creutzfeldt-Jakob disease of long duration: Clinicopathological characteristics, transmissibility, and differential diagnosis. *Annals of Neurology*, *16*(3), 295–304. doi:10.1002/ana.410160305 PMID:6385823

Brown, R. C., Lockwood, A. H., & Sonawane, B. R. (2005). Neurodegenerative diseases: An overview of environmental risk factors. *Environmental Health Perspectives*, *113*(9), 1250–1256. doi:10.1289/ehp.7567 PMID:16140637

Bruijn, L. I., Becher, M. W., Lee, M. K., Anderson, K. L., Jenkins, N. A., Copeland, N. G., ... Cleveland, D. W. (1997). ALS-linked SOD1 mutant G85R mediates damage to astrocytes and promotes rapidly progressive disease with SOD1-containing inclusions. *Neuron*, *18*(2), 327–338. doi:10.1016/S0896-6273(00)80272-X PMID:9052802

Bruijn, L. I., Houseweart, M. K., Kato, S., Anderson, K. L., Anderson, S. D., & Ohama, E. (1851–1854). ... Cleveland, D. W. (1998). Aggregation and motor neuron toxicity of an ALS-linked SOD1 mutant independent from wild-type SOD1. *Science*, 281.

Brundin, P., Ma, J., & Kordower, J. H. (2016, August). How strong is the evidence that Parkinson's disease is a prion disorder? *Current Opinion in Neurology*, 29(4), 459–466. doi:10.1097/WCO.00000000000349 PMID:27257944

Bryson, S. E., Rogers, S. J., & Fombonne, E. (2003). Autism spectrum disorders: Early detection, intervention, education, and psychopharmacological management. *Canadian Journal of Psychiatry*, *48*(8), 506–516. doi:10.1177/070674370304800802 PMID:14574826

Buday, E. (1995). The effects of signed and spoken words taught with music on sign and speech imitation by children with autism. *Journal of Music Therapy*, *32*(3), 189–202. doi:10.1093/jmt/32.3.189

Budge, M., Johnston, C., Hogervorst, E., De Jager, C., Milwain, E., Iversen, S., ... Smith, A. (2000). Plasma total homocysteine and cognitive performance in a volunteer elderly population. *Annals of the New York Academy of Sciences*, 903(1), 407–410. doi:10.1111/j.1749-6632.2000.tb06392.x PMID:10818531

Buechel, H. M., Popovic, J., Searcy, J. L., Porter, N. M., Thibault, O., & Blalock, E. M. (2011). Deep Sleep and Parietal Cortex Gene Expression Changes Are Related to Cognitive Deficits with Age. *PLoS One*, *6*(4), e18387. doi:10.1371/ journal.pone.0018387 PMID:21483696

Bunton-Stasyshyn, R. K., Saccon, R. A., Fratta, P., & Fisher, E. M. (2015). SOD1 Function and Its Implications for Amyotrophic Lateral Sclerosis Pathology: New and Renascent Themes. *The Neuroscientist*, 21(5), 519–529. doi:10.1177/1073858414561795 PMID:25492944

Burd, L., Severud, R., Kerbeshian, J., & Klug, M. (1999). Prenatal and perinatal risk factors for autism. *Journal of Perinatal Medicine*, 27, 441–450. PMID:10732302

Burnet, F. M. (1981). A possible role of zinc in the pathology of dementia. *Lancet*, *1*(8213), 186–188. doi:10.1016/S0140-6736(81)90062-3 PMID:6162062

Burns, M. P., Zhang, L., Rebeck, G. W., Querfurth, H. W., & Moussa, C. E. H. (2009). Parkin promotes intracellular Aβ 1–42 clearance. *Human Molecular Genetics*, *18*(17), 3206–3216. doi:10.1093/hmg/ddp258 PMID:19483198

Burton, E. J., McKeith, I. G., Burn, D. J., Williams, E. D., & O'Brien, J. T. (2004). Cerebral atrophy in Parkinson's disease with and without dementia: A comparison with Alzheimer's disease, dementia with Lewy bodies and controls. *Brain*, *127*(4), 791–800. doi:10.1093/brain/awh088 PMID:14749292

Burxer, G. V. (1974). Stress in farm animals. Veterinaria, 8, 92-94.

Busciglio, J., Pelsman, A., Wong, C., Pigino, G., Yuan, M., Mori, H., & Yankner, B. A. (2002). Altered metabolism of the amyloid β precursor protein is associated with mitochondrial dysfunction in Down's syndrome. *Neuron*, *33*(5), 677–688. doi:10.1016/S0896-6273(02)00604-9 PMID:11879646

Butterfield, D. A., & Boyd-Kimball, D. (2004). Amyloid β-peptide (1-42) contributes to the oxidative stress and neurodegeneration found in Alzheimer disease Brain. *Brain Pathology (Zurich, Switzerland)*, *14*(4), 426–432. doi:10.1111/j.1750-3639.2004.tb00087.x PMID:15605990

Butterfield, D. A., Castegna, A., Lauderback, C. M., & Drake, J. (2002). Evidence that amyloid β -peptide induced lipid peroxidation and its sequelae in Alzheimer's disease brain contribute to neuronal death. *Neurobiology of Aging*, 23(5), 655–664. doi:10.1016/S0197-4580(01)00340-2 PMID:12392766

Butterfield, D. A., Di Domenico, F., Swomley, A. M., Head, E., & Perluigi, M. (2014). Redox proteomics analysis to decipher the neurobiology of Alzheimer-like neurodegeneration: Overlaps in Down's syndrome and Alzheimer's disease brain. *The Biochemical Journal*, *463*(2), 177–189. doi:10.1042/BJ20140772 PMID:25242166

Butterfield, D. A., Drake, J., Pocernich, C., & Castegna, A. (2001). Evidence of oxidative damage in Alzheimer's disease brain: Central role for amyloid β -peptide. *Trends in Molecular Medicine*, 7(12), 548–554. doi:10.1016/S1471-4914(01)02173-6 PMID:11733217

Butterfield, D. A., Swomley, A. M., & Sultana, S. (2013). Amyloid β-peptide (1–42)-induced oxidative stress in Alzheimer disease: Importance in disease pathogenesis and progression. *Antioxidants & Redox Signalling*, *19*(8), 823–835. doi:10.1089/ars.2012.5027 PMID:23249141 Butterfield, D., Castegna, A., Pocernich, C., Drake, J., Scapagnini, G., & Calabrese, V. (2002). Nutritional approaches to combat oxidative stress in Alzheimer's disease. *The Journal of Nutritional Biochemistry*, *13*(8), 444–461. doi:10.1016/S0955-2863(02)00205-X PMID:12165357

Cabeza, R., Nyberg, L., & Park, D. C. (2016). *Cognitive neuroscience of aging: Linking cognitive and cerebral aging*. Oxford University Press. doi:10.1093/acprof:oso/9780199372935.001.0001

Cabezas, R., Ávila, M., Gonzalez, J., El-Bachá, R. S., Báez, E., García-Segura, L. M., ... Barreto, G. E. (2014). Astrocytic modulation of blood brain barrier: Perspectives on Parkinson's disease. *Frontiers in Cellular Neuroscience*, *8*, 211. doi:10.3389/fncel.2014.00211 PMID:25136294

Cabezas, R., El-Bachá, R. S., González, J., & Barreto, G. E. (2012). Mitochondrial functions in astrocytes: Neuroprotective implications from oxidative damage by rotenone. *Neuroscience Research*, 74(2), 80–90. doi:10.1016/j.neures.2012.07.008 PMID:22902554

Cadet, J. L. (1988). Free radical mechanisms in the central nervous system: An overview. *The International Journal of Neuroscience*, 40(1-2), 13–18. doi:10.3109/00207458808985722 PMID:2840405

Cai, Q., & Tammineni, P. (2017). Mitochondrial aspects of synaptic dysfunction in Alzheimer's disease. *Journal of Alzheimer's Disease*, 57(4), 1087–1103. doi:10.3233/JAD-160726 PMID:27767992

Caixeta, L., Costa, J. N., Vilela, A. C., & Nóbrega, M. D. (2014). The development of the dementia concept in 19th century. *Arquivos de Neuro-Psiquiatria*, 72(7), 564–567. doi:10.1590/0004-282X20140069 PMID:25054992

Cakir, E., Usul, H., Peksoylu, B., Sayin, O. C., Alver, A., Topbas, M., ... Kuzeyli, K. (2005). Effects of citicoline on experimental spinal cord injury. *Journal of Clinical Neuroscience*, *12*(8), 923–926. doi:10.1016/j.jocn.2005.03.013 PMID:16257217

Calabrese, E. (2016). Diffusion Tractography in Deep Brain Stimulation Surgery: A Review. *Frontiers in Neuroanatomy*, *10*, 45. doi:10.3389/fnana.2016.00045 PMID:27199677

Calì, T., Ottolini, D., Negro, A., & Brini, M. (2012). α-Synuclein controls mitochondrial calcium homeostasis by enhancing endoplasmic reticulum-mitochondria interactions. *The Journal of Biological Chemistry*, 287(22), 17914–17929. doi:10.1074/jbc.M111.302794 PMID:22453917

Calkins, M. J., Manczak, M., Mao, P., Shirendeb, U., & Reddy, P. H. (2011). Impaired mitochondrial biogenesis, defective axonal transport of mitochondria, abnormal mitochondrial dynamics and synaptic degeneration in a mouse model of Alzheimer's disease. *Human Molecular Genetics*, 20(23), 4515–4529. doi:10.1093/hmg/ddr381 PMID:21873260

Callesen, M. B., Hansen, K. V., Gjedde, A., Linnet, J., & Moller, A. (2013). Dopaminergic and clinical correlates of pathological gambling in Parkinson's disease: A case report. *Frontiers in Behavioral Neuroscience*, 7, 1–8. doi:10.3389/fnbeh.2013.00095 PMID:23908610

Camara, A. K. S., Lesnefsky, E. J., & Stowe, D. F. (2010). Potential therapeutic benefits of strategies directed to mitochondria. *Antioxidants & Redox Signalling*, *13*(3), 279–347. doi:10.1089/ars.2009.2788 PMID:20001744

Camargo, C. H., Camargos, S. T., Cardoso, F. E., & Teive, H. A. (2015). The genetics of the dystonias – a review based on the new classification of the dystonias. *Arquivos de Neuro-Psiquiatria*, 73(4), 350–358. doi:10.1590/0004-282X20150030 PMID:25992527

Camicioli, R., Moore, M. M., Kinney, A., Corbridge, E., Glassberg, K., & Kaye, J. A. (2003). Parkinson's disease is associated with hippocampal atrophy. *Movement Disorders*, 18(7), 784–790. doi:10.1002/mds.10444 PMID:12815657

Campbell, B. C., McLean, C. A., Culvenor, J. G., Gai, W. P., Blumbergs, P. C., Jäkälä, P., ... Li, Q.-X. (2001). The solubility of α-synuclein in multiple system atrophy differs from that of dementia with Lewy bodies and Parkinson's disease. *Journal of Neurochemistry*, *76*(1), 87–96. doi:10.1046/j.1471-4159.2001.00021.x PMID:11145981

Campbell, H., Andrews, N., Brown, K. E., & Miller, E. (2007, December). Review of the effect of measles vaccination on the epidemiology of SSPE. *International Journal of Epidemiology*, *36*(6), 1334–1348. doi:10.1093/ije/dym207 PMID:18037676

Campêlo, C. L., Cagni, F. C., de Siqueira Figueredo, D., Oliveira Jr, L. G., Silva-Neto, A. B., Macêdo, P. T., ... de Oliveira Godeiro Jr, C. (2017). Variants in SNCA Gene Are Associated with Parkinson's Disease Risk and Cognitive Symptoms in a Brazilian Sample. *Frontiers in Aging Neuroscience*, 9. PMID:28676755

Canavan, M. M. (1931). Schilder's encephalitis periaxialis diffusa - Report or a case in a child aged sixteen and onehalf months. *Archives of Neurology and Psychiatry*, 25(2), 299–308. doi:10.1001/archneurpsyc.1931.02230020085005

Candy, J., Perry, R. H., Perry, E. K., Irving, D., Blessed, G., Fairbairn, A. F., & Tomlinson, B. E. (1983). Pathological changes in the nucleus of Meynert in Alzheimer's and Parkinson's diseases. *Journal of the Neurological Sciences*, *59*(2), 277–289. doi:10.1016/0022-510X(83)90045-X PMID:6854353

Canello, T., Engelstein, R., Moshel, O., Xanthopoulos, K., Juanes, M. E., Langeveld, J., ... Gabizon, R. (2008). Methionine sulfoxides on PrPSc: A prion-specific covalent signature. *Biochemistry*, 47(34), 8866–8873. doi:10.1021/bi800801f PMID:18680312

Caporaso, G. L., Gandy, S. E., Buxbaum, J. D., & Greengard, P. (1992). Chloroquine inhibits intracellular degradation but not secretion of Alzheimer beta/A4 amyloid precursor protein. *Proceedings of the National Academy of Sciences of the United States of America*, 89(6), 2252–2256. doi:10.1073/pnas.89.6.2252 PMID:1549591

Carboni, E., & Lingor, P. (2015). Insights on the interaction of alpha-synuclein and metals in the pathophysiology of Parkinson's disease. *Metallomics*, 7(3), 395–404. doi:10.1039/C4MT00339J PMID:25648629

Carrell, R. W., & Lomas, D. A. (1997). Conformational disease. *Lancet*, 350(9071), 134–138. doi:10.1016/S0140-6736(97)02073-4 PMID:9228977

Carrillo-Ruiz, J. D., Velasco, F., Jimenez, F., Castro, G., Velasco, A. L., Hernandez, J. A., ... Velasco, M. (2008). Feb). Bilateral electrical stimulation of prelemniscal radiations in the treatment of advanced Parkinson's disease. *Neurosurgery*, 62(2), 347–357. doi:10.1227/01.neu.0000316001.03765.e8 PMID:18382312

Carrì, M. T., Valle, C., Bozzo, F., & Cozzolino, M. (2015). Oxidative stress and mitochondrial damage: Importance in non-SOD1 ALS. *Frontiers in Cellular Neuroscience*, *9*, 4. PMID:25741238

Carson, M. J., Bilousova, T. V., Puntambekar, S. S., Melchior, B., Doose, J. M., & Ethell, I. M. (2007). A rose by any other name: the potential consequences of microglial heterogeneity during CNS health and disease. *Neurotherapeutics; the Journal of the American Society for Experimental NeuroTherapeutics*, *4*(4), 571–579. doi:10.1016/j.nurt.2007.07.002 PMID:17920538

Cartier, N., Hacein-Bey-Abina, S., Bartholomae, C. C., Veres, G., Schmidt, M., Kutschera, I., ... Aubourg, P. (2009). Hematopoietic Stem Cell Gene Therapy with a Lentiviral Vector in X-Linked Adrenoleukodystrophy. *Science*, *326*(5954), 818–823. doi:10.1126cience.1171242 PMID:19892975

Carvalho, A. N., Lim, J. L., Nijland, P. G., Witte, M. E., & Van Horssen, J. (2014). Glutathione in multiple sclerosis: More than just an antioxidant? *Multiple Sclerosis Journal*, 20(11), 1425–1431. doi:10.1177/1352458514533400 PMID:24842957

Casanova, M. F. (2007). The neuropathology of autism. *Brain Pathology (Zurich, Switzerland)*, 417(4), 422–433. doi:10.1111/j.1750-3639.2007.00100.x PMID:17919128

Casley, C. S., Canevari, L., Land, J. M., Clark, J. B., & Sharpe, M. A. (2002). β-Amyloid inhibits integrated mitochondrial respiration and key enzyme activities. *Journal of Neurochemistry*, 80(1), 91–100. doi:10.1046/j.0022-3042.2001.00681.x PMID:11796747

Cassano, T., Pace, L., Bedse, G., Lavecchia, A. M., De Marco, F., Gaetani, S., & Gaetano, S. (2016). Glutamate and mitochondria: Two prominent players in the oxidative stress-induced neurodegeneration. *Current Alzheimer Research*, *13*(2), 185–197. doi:10.2174/1567205013666151218132725 PMID:26679860

Cathey, S. S., Leroy, J. G., Wood, T., Eaves, K., Simensen, R. J., Kudo, M., ... Friez, M. J. (2010). Phenotype and genotype in mucolipidoses II and III alpha/beta: A study of 61 probands. *Journal of Medical Genetics*, 47(1), 38–48. doi:10.1136/jmg.2009.067736 PMID:19617216

Catterall, W. A. (2011). Voltage-Gated Calcium Channels. *Cold Spring Harbor Perspectives in Biology*, *3*(8), a003947. doi:10.1101/cshperspect.a003947 PMID:21746798

Caudle, W. M., Colebrooke, R. E., Emson, P. C., & Miller, G. W. (2008). Altered vesicular dopamine storage in Parkinson's disease: A premature demise. *Trends in Neurosciences*, *31*(6), 303–308. doi:10.1016/j.tins.2008.02.010 PMID:18471904

Caughey, B., & Baron, G. S. (2006). Prions and their partners in crime. *Nature*, 443(7113), 803–810. doi:10.1038/ nature05294 PMID:17051207

Centers for Disease Control and Prevention (CDC). (2015). Creutzfeldt-Jakob Disease Surveillance and Diagnosis. *MMWR. Morbidity and Mortality Weekly Report*, *45*(31), 665.

Centers for Disease Control and Prevention. (1996). World Health Organization consultation on public health issues related to bovine spongiform encephalopathy and the emergence of a new variant of Creutzfeldt-Jakob disease. *MMWR*. *Morbidity and Mortality Weekly Report*, 45(14), 295. PMID:8598828

Centers for Disease Control and Prevention. (2017). *Leading Causes of Death*. Retrieved April 4, 2017, from https://www.cdc.gov/nchs/fastats/leading-causes-of-death.htm

Chabriat, H., Hervé, D., Duering, M., Godin, O., Jouvent, E., Opherk, C., ... Dichgans, M. (2016). Predictors of clinical worsening in cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy: Prospective cohort study. *Stroke*, *47*(1), 4–11. doi:10.1161/STROKEAHA.115.010696 PMID:26578659

Chakrabarti, S., Munshi, S., Banerjee, K., Thakurta, I. G., Sinha, M., & Bagh, M. B. (2011). Mitochondrial Dysfunction during Brain Aging: Role of Oxidative Stress and Modulation by Antioxidant Supplementation. *Aging and Disease*, 2(3), 242–256. PMID:22396876

Chakraborty, S., Skolnick, B., & Narayan, R. K. (2016). Neuroprotection trials in traumatic brain injury. *Current Neurology and Neuroscience Reports*, *16*(4), 29. doi:10.100711910-016-0625-x PMID:26883431

Chan, D., Fox, N. C., Scahill, R. I., Crum, W. R., Whitwell, J. L., Leschziner, G., ... Rossor, M. N. (2001). Patterns of temporal lobe atrophy in semantic dementia and Alzheimer's disease. *Annals of Neurology*, *49*(4), 433–442. doi:10.1002/ana.92 PMID:11310620

Chandra, S., Gallardo, G., Fernández-Chacón, R., Schlüter, O. M., & Südhof, T. C. (2005). *α-synuclein* cooperates with CSPα in preventing neurodegeneration. *Cell*, *123*(3), 383–396. doi:10.1016/j.cell.2005.09.028 PMID:16269331

Chandrasekaran, K., Hatanpää, K., Brady, D. R., & Rapoport, S. I. (1996). Evidence for physiological down-regulation of brain oxidative phosphorylation in Alzheimer's disease. *Experimental Neurology*, *142*(1), 80–88. doi:10.1006/exnr.1996.0180 PMID:8912900

Chan, P., DeLanney, L. E., Irwin, I., Langston, J. W., & Monte, D. (1991). Rapid ATP Loss Caused by 1-Methyl-4-Phenyl-1, 2, 3, 6-Tetrahydropyridine in Mouse Brain. *Journal of Neurochemistry*, *57*(1), 348–351. doi:10.1111/j.1471-4159.1991. tb02134.x PMID:2051170

Charcot, J. M. (1872). Cinquième Leçon. De la paralysie agitante. *Oeuvres completes, Recueillies et publiées par Bourneville. Bureaux du Progrès Médical, Paris, France, 1*, 155–189.

Charidimou, A., Gang, Q., & Werring, D. J. (2012). Sporadic cerebral amyloid angiopathy revisited: Recent insights into pathophysiology and clinical spectrum. *Journal of Neurology, Neurosurgery, and Psychiatry*, 83(2), 124–137. doi:10.1136/jnnp-2011-301308 PMID:22056963

Chartier-Harlin, M. C., Kachergus, J., Roumier, C., Mouroux, V., Douay, X., Lincoln, S., ... Waucquier, N. (2004). α -synuclein locus duplication as a cause of familial Parkinson's disease. *Lancet*, *364*(9440), 1167–1169. doi:10.1016/S0140-6736(04)17103-1 PMID:15451224

Chatterjee, A., Dasgupta, S., & Sidransky, D. (2011). Mitochondrial Subversion in Cancer. *Cancer Prevention Research* (*Philadelphia*, *Pa.*), *4*(5), 638–654. doi:10.1158/1940-6207.CAPR-10-0326 PMID:21543342

Chauhan, A., Audhya, T., & Chauhan, V. (2012). Brain region-specific glutathione redox imbalance in autism. *Neuro-chemical Research*, *37*(8), 1681–1689. doi:10.100711064-012-0775-4 PMID:22528835

Chauhan, A., Gu, F., Essa, M. M., Wegiel, J., Kaur, K., Brown, W. T., & Chauhan, V. (2011). Brain region-specific deficit in mitochondrial electron transport chain complexes in children with autism. *Journal of Neurochemistry*, *117*(2), 209–220. doi:10.1111/j.1471-4159.2011.07189.x PMID:21250997

Chen, C., & Dong, X. P. (2016). Epidemiological characteristics of human prion diseases. *Infectious Diseases of Poverty*, *5*(1), 47. doi:10.118640249-016-0143-8 PMID:27251305

Cheng, F., Vivacqua, G., & Yu, S. (2011). The role of alpha-synuclein in neurotransmission and synaptic plasticity. *Journal of Chemical Neuroanatomy*, *42*(4), 242–248. doi:10.1016/j.jchemneu.2010.12.001 PMID:21167933

Cheng, K. C., Cahill, D. S., Kasai, H., Nishimura, S., & Loeb, L. A. (1992). 8-Hydroxyguanine, an abundant form of oxidative DNA damage, causes G----T and A----C substitutions. *The Journal of Biological Chemistry*, 267(1), 166–172. PMID:1730583

Chen, H. S., & Lipton, S. A. (2006, June). The chemical biology of clinically tolerated NMDA receptor antagonists. *Journal of Neurochemistry*, 97(6), 1611–1626. doi:10.1111/j.1471-4159.2006.03991.x PMID:16805772

Chen, L., Ding, Y., Cagniard, B., Van Laar, A. D., Mortimer, A., Chi, W., ... Zhuang, X. (2008). Unregulated cytosolic dopamine causes neurodegeneration associated with oxidative stress in mice. *The Journal of Neuroscience*, 28(2), 425–433. doi:10.1523/JNEUROSCI.3602-07.2008 PMID:18184785

Chen, X. F., Zhang, Y. W., Xu, H., & Bu, G. (2013). Transcriptional regulation and its misregulation in Alzheimer's disease. *Molecular Brain*, 6(1), 44. doi:10.1186/1756-6606-6-44 PMID:24144318

Chen, X., & Burgoyne, R. D. (2012). Identification of common genetic modifiers of neurodegenerative diseases from an integrative analysis of diverse genetic screens in model organisms. *BMC Genomics*, *13*(1), 71. doi:10.1186/1471-2164-13-71 PMID:22333271

Chernova, T. A., Wilkinson, K. D., & Chernoff, Y. O. (2014). Physiological and environmental control of yeast prions. *FEMS Microbiology Reviews*, *38*(2), 326–344. doi:10.1111/1574-6976.12053 PMID:24236638

Chez, M. G., Dowling, T., Patel, P. B., Khanna, P., & Kominsky, M. (2007). Elevation of tumor necrosis factor-alpha in cerebrospinal fluid of autistic children. *Pediatric Neurology*, *36*(6), 361–365. doi:10.1016/j.pediatrneurol.2007.01.012 PMID:17560496

Chiba, K., Trevor, A., & Castagnoli, N. Jr. (1984). Metabolism of the neurotoxic tertiary amine, MPTP, by brain monoamine oxidase. *Biochemical and Biophysical Research Communications*, *120*(2), 574–578. doi:10.1016/0006-291X(84)91293-2 PMID:6428396

Chien, P., Weissman, J. S., & DePace, A. H. (2004). Emerging principles of conformation-based prion inheritance. *Annual Review of Biochemistry*, 73(1), 617–656. doi:10.1146/annurev.biochem.72.121801.161837 PMID:15189155

Chio, A., Traynor, B. J., Lombardo, F., Fimognari, M., Calvo, A., Ghiglione, P., ... Restagno, G. (2008). Prevalence of SOD1 mutations in the Italian ALS population. *Neurology*, *70*(7), 533–537. doi:10.1212/01.wnl.0000299187.90432.3f PMID:18268245

Chipuk, J. E., Moldoveanu, T., Llambi, F., Parsons, M. J., & Green, D. R. (2010). The BCL-2 family reunion. *Molecular Cell*, *37*(3), 299–310. doi:10.1016/j.molcel.2010.01.025 PMID:20159550

Choi, B. Y., Jang, B. G., Kim, J. H., Lee, B. E., Sohn, M., Song, H. K., & Suh, S. W. (2012). Prevention of traumatic brain injury-induced neuronal death by inhibition of NADPH oxidase activation. *Brain Research*, *1481*, 49–58. doi:10.1016/j. brainres.2012.08.032 PMID:22975130

Choi, D. W., Yokoyama, M., & Koh, J. (1988). Zinc neurotoxicity in cortical cell culture. *Neuroscience*, *24*(1), 67–79. doi:10.1016/0306-4522(88)90312-0 PMID:3368058

Choonara, Y. E., Pillay, V., Du Toit, L. C., Modi, G., Naidoo, D., Ndesendo, V. M., & Sibambo, S. R. (2009). Trends in the molecular pathogenesis and clinical therapeutics of common neurodegenerative disorders. *International Journal of Molecular Sciences*, *10*(6), 2510–2557. doi:10.3390/ijms10062510 PMID:19582217

Christensen, M. A., Zhou, W., Qing, H., Lehman, A., Philipsen, S., & Song, W. (2004). Transcriptional regulation of BACE1, the beta-amyloid precursor protein beta-secretase, by Sp1. *Molecular and Cellular Biology*, 24(2), 865–874. doi:10.1128/MCB.24.2.865-874.2004 PMID:14701757

Christian, H. (2011). Identifying and validating biomarkers for Alzheimer's disease. *Trends in Biotechnology*, 29(1), 26–32. doi:10.1016/j.tibtech.2010.09.007 PMID:20971518

Chui, H. C., Mortimer, J. A., Slager, U., Zarow, C., Bondareff, W., & Webster, D. D. (1986). Pathologic correlates of dementia in Parkinson's disease. *Archives of Neurology*, *43*(10), 991–995. doi:10.1001/archneur.1986.00520100013007 PMID:3753274

Chui, H. C., & Ramirez-Gomez, L. (2015). Clinical and imaging features of mixed Alzheimer and vascular pathologies. *Alzheimer's Research & Therapy*, 7(1), 21. doi:10.118613195-015-0104-7 PMID:25722748

Chung, Y. C., Bok, E., Huh, S. H., Park, J.-Y., Yoon, S.-H., Kim, S. R., ... Jin, B. K. (2011). Cannabinoid receptor type 1 protects nigrostriatal dopaminergic neurons against MPTP neurotoxicity by inhibiting microglial activation. *Journal of Immunology (Baltimore, Md.: 1950), 187*(12), 6508–6517. doi:10.4049/jimmunol.1102435 PMID:22079984

Church, E. W., Gundersen, A., Glantz, M. J., & Simon, S. D. (2017). Number needed to treat for stroke thrombectomy based on a systematic review and meta-analysis. *Clinical Neurology and Neurosurgery*, *156*, 83–88. doi:10.1016/j. clineuro.2017.03.005 PMID:28359980

Chvapil, M., Ryan, J. N., & Zukoski, C. F. (1972). The effect of zinc and other metals on the stability of lysosomes. *Proceedings of the Society for Experimental Biology and Medicine*, *140*(2), 642–646. doi:10.3181/00379727-140-36521 PMID:5037603

Cicchetti, F., Drouin-Ouellet, J., & Gross, R. E. (2009). Environmental toxins and Parkinson's disease: What have we learned from pesticide-induced animal models? *Trends in Pharmacological Sciences*, *30*(9), 475–483. doi:10.1016/j. tips.2009.06.005 PMID:19729209

Ciechanover, A., & Kwon, Y. T. (2015). Degradation of misfolded proteins in neurodegenerative diseases: Therapeutic targets and strategies. *Experimental & Molecular Medicine*, 47(3), e147. doi:10.1038/emm.2014.117 PMID:25766616

Cipolla, M. J. (2009). The Cerebral Circulation. Morgan & Claypool Life Sciences.

Citron, M. (2002). Alzheimer's disease: Treatments in discovery and development. *Nature Neuroscience*, 5(11s), 1055–1057. doi:10.1038/nn940 PMID:12403985

Citron, M., Teplow, D. B., & Selkoe, D. J. (1995). Generation of amyloid beta protein from its precursor is sequence specific. *Neuron*, *14*(3), 661–670. doi:10.1016/0896-6273(95)90323-2 PMID:7695913

Clark, B. A., Farrant, M., & Cull-Candy, S. G. (1997). A direct comparison of the single-channel properties of synaptic and extrasynaptic NMDA receptors. *The Journal of Neuroscience*, *17*(1), 107–116. PMID:8987740

Clarke, C. E. (2002). Medical management of Parkinson's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 72(suppl 1), i22–i27. doi:10.1136/jnnp.72.suppl_1.i22 PMID:11870200

Clark, L. N., Poorkaj, P., Wszolek, Z., Geschwind, D. H., Nasreddine, Z. S., Miller, B., ... Wilhelmsen, K. C. (1998). Pathogenic implications of mutations in the tau gene in pallido-ponto-nigral degeneration and related neurodegenerative disorders linked to chromosome 17. *Proceedings of the National Academy of Sciences of the United States of America*, 95(22), 13103–13107. doi:10.1073/pnas.95.22.13103 PMID:9789048

Clerc, P., Lipnick, S., & Willett, C. (2016). A look into the future of ALS research. *Drug Discovery Today*, 21(6), 939–949. doi:10.1016/j.drudis.2016.02.002 PMID:26861067

Clinton, L. K., Blurton-Jones, M., Myczek, K., Trojanowski, J. Q., & LaFerla, F. M. (2007). Synergistic Interactions Among Abeta, Tau, and Alpha-synuclein: Acceleration of Neuropathology and Cognitive Decline. *The Journal of Neuroscience*, *30*(21), 7281–7289. doi:10.1523/JNEUROSCI.0490-10.2010 PMID:20505094

Cluskey, S., & Ramsden, D. B. (2001). Mechanisms of neurodegeneration in amyotrophic lateral sclerosis. *Molecular Pathology*, 54(6), 386. PMID:11724913

Coelho, R. C. L. A., Hermsdorff, H. H. M., & Bressan, J. (2013). Anti-inflammatory properties of orange juice: Possible favorable molecular and metabolic effects. *Plant Foods for Human Nutrition (Dordrecht, Netherlands)*, 68(1), 1–10. doi:10.100711130-013-0343-3 PMID:23417730

Coenen, V. A., Mädler, B., Schiffbauer, H., Urbach, H., & Allert, N. (2011, April). Individual fiber anatomy of the subthalamic region revealed with diffusion tensor imaging: A concept to identify the deep brain stimulation target for tremor suppression. *Neurosurgery*, *68*(4), 1069–1075. doi:10.1227/NEU.0b013e31820a1a20 PMID:21242831

Coleman, R. J., Robb, S. A., Lake, B. D., Brett, E. M., & Harding, A. E. (1998). The diverse neurological features of Niemann-Pick disease type C: A report of two cases. *Movement Disorders*, *3*(4), 295–299. doi:10.1002/mds.870030403 PMID:3145417

Cole, S. L., & Vassar, R. (2008). The role of amyloid precursor protein processing by BACE1, the beta-secretase, in Alzheimer disease pathophysiology. *The Journal of Biological Chemistry*, 283(44), 29621–29625. doi:10.1074/jbc. R800015200 PMID:18650431

Collier, T. J., Kanaan, N. M., & Kordower, J. H. (2011). Ageing as a primary risk factor for Parkinson's disease: Evidence from studies of non-human primates. *Nature Reviews. Neuroscience*, *12*(6), 359–366. doi:10.1038/nrn3039 PMID:21587290

Collinge, J., & Clarke, A. R. (2007). A general model of prion strains and their pathogenicity. *Science*, *318*(5852), 930–936. doi:10.1126cience.1138718 PMID:17991853

Colombo, G., Meli, M., Morra, G., Gabizon, R., & Gasset, M. (2009). Methionine sulfoxides on prion protein helix-3 switch on the α -fold destabilization required for conversion. *PLoS One*, 4(1), e4296. doi:10.1371/journal.pone.0004296 PMID:19172188

Concha-Marambio, Pritzkow, Moda, Tagliavini, & Ironside. (2016). New detection method could lead to noninvasive diagnosis of Creutzfeldt-Jakob disease. *Science Translational Medicine*, 8(370), 1–2.

Conn, P. J., Battaglia, G., Marino, M. J., & Nicoletti, F. (2005). Metabotropic glutamate receptors in the basal ganglia motorcircuit. *Nature Reviews. Neuroscience*, *6*(10), 787–798. doi:10.1038/nrn1763 PMID:16276355

Conway, K. A., Rochet, J. C., Bieganski, R. M., & Lansbury, P. T. (2001). Kinetic stabilization of the α-synuclein protofibril by a dopamine-α-synuclein adduct. *Science*, 294(5545), 1346–1349. doi:10.1126cience.1063522 PMID:11701929

Cooke, A., DeVita, C., Gee, J., Alsop, D., Detre, J., Chen, W., & Grossman, M. (2003). Neural basis for sentence comprehension deficits in frontotemporal dementia. *Brain and Language*, 85(2), 211–221. doi:10.1016/S0093-934X(02)00562-X PMID:12735939

Cooke, A., Grossman, M., DeVita, C., Gonzalez-Atavales, J., Moore, P., Chen, W., ... Detre, J. (2006). Large-scale neural network for sentence processing. *Brain and Language*, *96*(1), 14–36. doi:10.1016/j.bandl.2005.07.072 PMID:16168473

Cookson, M. R. (2009). α-Synuclein and neuronal cell death. *Molecular Neurodegeneration*, 4(1), 9. doi:10.1186/1750-1326-4-9 PMID:19193223

Cookson, M. R., & Shaw, P. J. (1999). Oxidative stress and motor neurone disease. *Brain Pathology (Zurich, Switzerland)*, 9(1), 165–186. doi:10.1111/j.1750-3639.1999.tb00217.x PMID:9989458

Cooper, A. J. (2012). The role of glutamine synthetase and glutamate dehydrogenase in cerebral ammonia homeostasis. *Neurochemical Research*, *37*(11), 2439–2455. doi:10.100711064-012-0803-4 PMID:22618691

Cooper, J. M., Wiklander, P. B., Nordin, J. Z., Al-Shawi, R., Wood, M. J., Vithlani, M., ... Alvarez-Erviti, L. (2014). Systemic exosomal siRNA delivery reduced alpha-synuclein aggregates in brains of transgenic mice. *Movement Disorders*, 29(12), 1476–1485. doi:10.1002/mds.25978 PMID:25112864

Cooper, S. A., Murray, K. L., Heath, C. A., Will, R. G., & Knight, R. S. (2006). Sporadic Creutzfeldt–Jakob disease with cerebellar ataxia at onset in the UK. *Journal of Neurology, Neurosurgery, and Psychiatry*, 77(11), 1273–1275. doi:10.1136/jnnp.2006.088930 PMID:16835290

Coral, P., Teive, H. A., & Werneck, L. C. (2000). Hemiballism: Report of eight cases. *Arquivos de Neuro-Psiquiatria*, 58(3A), 698–703. doi:10.1590/S0004-282X2000000400016 PMID:10973112

Cordery, R. J., Hall, M., Cipolotti, L., Davidson, L., Adlard, P., & Rossor, M. N. (2003). Early cognitive decline in Creutzfeldt-Jakob disease associated with human growth hormone treatment. *JOURNAL OF Neurology. Neurosurgery and Psychiatry*, *10*(74), 1412–1416. doi:10.1136/jnnp.74.10.1412

Cordonnier, C., Leys, D., Dumont, F., Deramecourt, V., Bordet, R., Pasquier, F., & Hénon, H. (2010). What are the causes of pre-existing dementia in patients with intracerebral haemorrhages? *Brain*, *133*(11), 3281–3289. doi:10.1093/ brain/awq246 PMID:20852266

Corps, K. N., Roth, T. L., & McGavern, D. B. (2015). Inflammation and neuroprotection in traumatic brain injury. *JAMA Neurology*, 72(3), 355–362. doi:10.1001/jamaneurol.2014.3558 PMID:25599342

Corral-Debrinski, M., Horton, T., Lott, M. T., Shoffner, J. M., Beal, M. F., & Wallace, D. C. (1992). Mitochondrial DNA deletions in human brain: Regional variability and increase with advanced age. *Nature Genetics*, *2*(4), 324–329. doi:10.1038/ng1292-324 PMID:1303288

Corral-Debrinski, M., Horton, T., Lott, M. T., Shoffner, J. M., McKee, A. C., Beal, M. F., ... Wallace, D. C. (1994). Marked changes in mitochondrial DNA deletion levels in Alzheimer brains. *Genomics*, 23(2), 471–476. doi:10.1006/ geno.1994.1525 PMID:7835898

Correia-Melo, C., & Passos, J. F. (2015). Mitochondria: Are they causal players in cellular senescence? *Biochimica et Biophysica Acta*, *1847*(11), 1373–1379. doi:10.1016/j.bbabio.2015.05.017 PMID:26028303

Corti, O., Fournier, M., & Brice, A. (2009). Neurodegeneration in Parkinson's disease: Genetics enlightens physiopathology. In Birth, Life and Death of Dopaminergic Neurons in the Substantia Nigra (pp. 215–221). Springer. doi:10.1007/978-3-211-92660-4_17

Coskun, P. E., Beal, M. F., & Wallace, D. C. (2004). Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proceedings of the National Academy of Sciences of the United States of America*, *101*(29), 10726–10731. doi:10.1073/pnas.0403649101 PMID:15247418

Costello, L. C., Fenselau, C. C., & Franklin, R. B. (2011). Evidence for operation of the direct zinc ligand exchange mechanism for trafficking, transport, and reactivity of zinc in mammalian cells. *Journal of Inorganic Biochemistry*, *105*(5), 589–599. doi:10.1016/j.jinorgbio.2011.02.002 PMID:21440525

Costello, S., Cockburn, M., Bronstein, J., Zhang, X., & Ritz, B. (2009). Parkinson's disease and residential exposure to maneb and paraquat from agricultural applications in the central valley of California. *American Journal of Epidemiology*, *169*(8), 919–926. doi:10.1093/aje/kwp006 PMID:19270050

Cote, A., Chiasson, M., Peralta, M. R. III, Lafortune, K., Pellegrini, L., & Tóth, K. (2005). Cell type-specific action of seizure-induced intracellular zinc accumulation in the rat hippocampus. *The Journal of Physiology*, *566*(3), 821–837. doi:10.1113/jphysiol.2005.089458 PMID:15919712

Council, N. R. (2001). New horizons in health: An integrative approach (Vol. 277). National Academies Press.

Cousins, R. J., Liuzzi, J. P., & Lichten, L. A. (2006). Mammalian zinc transport, trafficking, and signals. *The Journal of Biological Chemistry*, 281(34), 24085–24089. doi:10.1074/jbc.R600011200 PMID:16793761

Cozzolino, M., Ferri, A., & Carri, M. T. (2008). Amyotrophic lateral sclerosis: From current developments in the laboratory to clinical implications. *Antioxidants & Redox Signalling*, *10*(3), 405–443. doi:10.1089/ars.2007.1760 PMID:18370853

Creutzfeldt, H. G. (2002). Clinical diagnosis and differential diagnosis of CJD and vCJD With special emphasis on laboratory tests. *Journal of Neurology, Neurosurgery and Psychiatry*, *1*(3), 88–98.

Crisp, M. J., Beckett, J., Coates, J. R., & Miller, T. M. (2013). Canine degenerative myelopathy: Biochemical characterization of superoxide dismutase 1 in the first naturally occurring non-human amyotrophic lateral sclerosis model. *Experimental Neurology*, 248, 1–9. doi:10.1016/j.expneurol.2013.05.009 PMID:23707216 Cristóvão, A. C., Guhathakurta, S., Bok, E., Je, G., Yoo, S. D., Choi, D.-H., & Kim, Y.-S. (2012). NADPH oxidase 1 mediates α-synucleinopathy in Parkinson's disease. *The Journal of Neuroscience*, *32*(42), 14465–14477. doi:10.1523/JNEUROSCI.2246-12.2012 PMID:23077033

Cronin, S., Hardiman, O., & Traynor, B. J. (2007). Ethnic variation in the incidence of ALS: A systematic review. *Neurology*, *68*(13), 1002–1007. doi:10.1212/01.wnl.0000258551.96893.6f PMID:17389304

Crouch, P. J., Blake, R., Duce, J. A., Ciccotosto, G. D., Li, Q. X., Barnham, K. J., & Masters, C. L. (2005). Copperdependent inhibition of human cytochrome c oxidase by a dimeric conformer of amyloid-β1-42. *The Journal of Neuroscience*, *25*(3), 672–679. doi:10.1523/JNEUROSCI.4276-04.2005 PMID:15659604

Csermely, P., Szamel, M., Resch, K., & Somogyi, J. (1988). Zinc can increase the activity of protein kinase C and contributes to its binding to plasma membranes in T-lymphocytes. *The Journal of Biological Chemistry*, 263(14), 6487–6490. PMID:3258866

Csiszar, A., Ungvari, Z., Koller, A., Edwards, J. G., & Kaley, G. (2004). Proinflammatory phenotype of coronary arteries promotes endothelial apoptosis in aging. *Physiological Genomics*, *17*(1), 21–30. doi:10.1152/physiolgenom-ics.00136.2003 PMID:15020720

Cuajungco, M. P., & Lees, G. J. (1997). Zinc metabolism in the brain: Relevance to human neurodegenerative disorders. *Neurobiology of Disease*, 4(3-4), 137–169. doi:10.1006/nbdi.1997.0163 PMID:9361293

Cudkowicz, M. E., Shefner, J. M., Schoenfeld, D. A., Zhang, H., Andreasson, K. I., Rothstein, J. D., & Drachman, D. B. (2004). Clinical trial of celecoxib in subjects with amyotrophic lateral sclerosis. *Annals of Neurology*, *5*, 25–26.

Cuervo, A. M., & Wong, E. (2013). Chaperone-mediated autophagy: Roles in disease and aging. *Cell Research*, 24(1), 92–104. doi:10.1038/cr.2013.153 PMID:24281265

Cui, H., Kong, Y., & Zhang, H. (2012). Oxidative Stress, Mitochondrial Dysfunction, and Aging. *Journal of Signal Transduction*, 2012, 646354. doi:10.1155/2012/646354 PMID:21977319

Cummings, J. L. (1993). Frontal-subcortical circuits and human behavior. *Archives of Neurology*, *50*(8), 873–880. doi:10.1001/archneur.1993.00540080076020 PMID:8352676

Cummings, J. L., & Duchen, L. W. (1981). Kluver-Bucy syndrome in Pick disease: Clinical and pathologic correlations. *Neurology*, *31*(11), 1415–1422. doi:10.1212/WNL.31.11.1415 PMID:7198189

Cummings, J. L., Morstorf, T., & Zhong, K. (2014). Alzheimer's disease drug-development pipeline: Few candidates, frequent failures. *Alzheimer's Research & Therapy*, *6*(4), 37. doi:10.1186/alzrt269 PMID:25024750

Curmi, P. A., Gavet, O., Charbaut, E., Ozon, S., Lachkar-Colmerauer, S., Manceau, V., ... Sobel, A. (1999). Stathmin and its phosphoprotein family: General properties, biochemical and functional interaction with tubulin. *Cell Structure and Function*, 24(5), 345–357. doi:10.1247/csf.24.345 PMID:15216892

Currais, A., Fischer, W., Maher, P., & Schubert, D. (2017). Intraneuronal protein aggregation as a trigger for inflammation and neurodegeneration in the aging brain. *The FASEB Journal*, *31*(1), 5–10. doi:10.1096/fj.201601184 PMID:28049155

Cushman, M., Johnson, B. S., King, O. D., Gitler, A. D., & Shorter, J. (2010). Prion-like disorders: Blurring the divide between transmissibility and infectivity. *Journal of Cell Science*, *123*(8), 1191–1201. doi:10.1242/jcs.051672 PMID:20356930

D'Souza, I., Poorkaj, P., Hong, M., Nochlin, D., Lee, V. M.-Y., Bird, T. D., & Schellenberg, G. D. (1999). Missense and silent tau gene mutations cause frontotemporal dementia with parkinsonism-chromosome 17 type, by affecting multiple alternative RNA splicing regulatory elements. *Proceedings of the National Academy of Sciences of the United States of America*, *96*(10), 5598–5603. doi:10.1073/pnas.96.10.5598 PMID:10318930

Da Cruz, S., & Cleveland, D. W. (2011). Understanding the role of TDP-43 and FUS/TLS in ALS and beyond. *Current Opinion in Neurobiology*, *21*(6), 904–919. doi:10.1016/j.conb.2011.05.029 PMID:21813273

Dai, M., & Masahiro, K. (2013). The Molecular Mechanisms of zinc neurotoxicity and the pathogenesis of vascular type senile dementia. *International Journal of Molecular Sciences*, *14*(11), 22067–22081. PMID:24213606

Daldin, M., Fodale, V., Cariulo, C., Azzollini, L., Verani, M., Martufi, P., ... Macdonald, D. (2017). Polyglutamine expansion affects huntingtin conformation in multiple Huntington's disease models. *Scientific Reports*, 7(1), 5070. doi:10.103841598-017-05336-7 PMID:28698602

Damiano, M., Galvan, L., Déglon, N., & Brouillet, E. (2010). Mitochondria in Huntington's disease. *Biochimica et Biophysica Acta*, *1802*(1), 52–61. doi:10.1016/j.bbadis.2009.07.012 PMID:19682570

Damiano, M., Starkov, A. A., Petri, S., Kipiani, K., Kiaei, M., Mattiazzi, M., ... Manfredi, G. (2006). Neural mitochondrial Ca2+ capacity impairment precedes the onset of motor symptoms in G93A Cu/Zn-superoxide dismutase mutant mice. *Journal of Neurochemistry*, *96*(5), 1349–1361. doi:10.1111/j.1471-4159.2006.03619.x PMID:16478527

Daneman, R., & Prat, A. (2015). The blood-brain barrier. *Cold Spring Harbor Perspectives in Biology*, 7(1), a020412. doi:10.1101/cshperspect.a020412 PMID:25561720

Danta, C. C., & Piplani, P. (2014). The discovery and development of new potential antioxidant agents for the treatment of neurodegenerative diseases. *Expert Opinion on Drug Discovery*, *9*(10), 1205–1222. doi:10.1517/17460441.2014.94 2218 PMID:25056182

Darios, F., Corti, O., Lücking, C. B., Hampe, C., Muriel, M. P., Abbas, N., & Brice, A. (2003). Parkin prevents mitochondrial swelling and cytochrome c release in mitochondria-dependent cell death. *Human Molecular Genetics*, *12*(5), 517–526. doi:10.1093/hmg/ddg044 PMID:12588799

Darvesh, S., & Freedman, M. (1996). Subcortical dementia: A neurobehavioral approach. *Brain and Cognition*, *31*(2), 230–249. doi:10.1006/brcg.1996.0043 PMID:8812000

Dauer, W., & Przedborski, S. (2003). Parkinson's Disease, Mechanisms and Models. *Neuron*, 39(6), 889–909. doi:10.1016/S0896-6273(03)00568-3 PMID:12971891

Davenport, R. J., Swingler, R. J., Chancellor, A. M., & Warlow, C. P. (1996). Avoiding false positive diagnoses of motor neuron disease: Lessons from the Scottish Motor Neuron Disease Register. *Journal of Neurology, Neurosurgery, and Psychiatry*, 60(2), 147–151. doi:10.1136/jnnp.60.2.147 PMID:8708642

Davey, G. P., & Clark, J. B. (1996). Threshold effects and control of oxidative phosphorylation in nonsynaptic rat brain mitochondria. *Journal of Neurochemistry*, *66*(4), 1617–1624. doi:10.1046/j.1471-4159.1996.66041617.x PMID:8627318

Davidson, P. W., Myers, G. J., & Weiss, B. (2004). Mercury exposure and child development outcomes. *Pediatrics*, *113*, 1023–1029. PMID:15060195

Davies, L., Wolska, B., Hilbich, C., Multhaup, G., Martins, R., Simms, G., ... Masters, C. L. (1988). A4 amyloid protein deposition and the diagnosis of Alzheimer's disease: Prevalence in aged brains determined by immunocytochemistry compared with conventional neuropathologic techniques. *Neurology*, *38*(11), 1688–1693. doi:10.1212/WNL.38.11.1688 PMID:3054625

Daviglus, M. L., Bell, C. C., Berrettini, W., Bowen, P. E., Connolly, E. S. Jr, & Cox, N. J. (2010). national institutes of health state-of-the-science conference statement: Preventing Alzheimer disease and cognitive decline. *Annals of Internal Medicine*, *153*(3), 176–181. doi:10.7326/0003-4819-153-3-201008030-00260 PMID:20547888

Davis, G. C., Williams, A. C., Markey, S. P., Ebert, M. H., Caine, E. D., Reichert, C. M., & Kopin, I. J. (1979). Chronic parkinsonism secondary to intravenous injection of meperidine analogues. *Psychiatry Research*, *1*(3), 249–254. doi:10.1016/0165-1781(79)90006-4 PMID:298352

Davis, R. E., & Williams, M. (2012). Mitochondrial function and dysfunction: An update. *The Journal of Pharmacology* and *Experimental Therapeutics*, *342*(3), 598–607. doi:10.1124/jpet.112.192104 PMID:22700430

Dawson, J. R. (1933). Cellular Inclusions in Cerebral Lesions of Lethargic Encephalitis. *The American Journal of Pathology*, *9*(1), 7–16.3.

Dawson, T. M., & Dawson, V. L. (2010). The Role of Parkin in Familial and Sporadic Parkinson's Disease. *Movement Disorders : Official Journal of the Movement Disorder Society*, 25(1), S32-S39.

Dawson, T. M., & Dawson, V. L. (2010). The Role of Parkin in Familial and Sporadic Parkinson's Disease. *Movement Disorders: Official Journal of the Movement Disorder Society*, 25(1), S32-S39.

Dawson, T. M., & Dawson, V. L. (2010). The role of parkin in familial and sporadic Parkinson's disease. *Journal of Movement Disorders*, 25(S1), S32–S39. doi:10.1002/mds.22798 PMID:20187240

De Baets, G., Reumers, J., Blanco, J. D., Dopazo, J., Schymkowitz, J., & Rousseau, F. (2011). An evolutionary trade-off between protein turnover rate and protein aggregation favors a higher aggregation propensity in fast degrading proteins. *PLoS Computational Biology*, 7(6), e1002090. doi:10.1371/journal.pcbi.1002090 PMID:21731483

de Carvalho, M., Dengler, R., Eisen, A., England, J. D., Kaji, R., Kimura, J., ... Swash, M. (2008). Electrodiagnostic criteria for diagnosis of ALS. *Clinical Neurophysiology*, *119*(3), 497–503. doi:10.1016/j.clinph.2007.09.143 PMID:18164242

De Castro, I. P., Costa, A., Lam, D., Tufi, R., Fedele, V., Moisoi, N., ... Martins, L. M. (2012). Genetic analysis of mitochondrial protein misfolding in Drosophila melanogaster. *Cell Death and Differentiation*, *19*(8), 1308–1316. doi:10.1038/ cdd.2012.5 PMID:22301916

de Jong, S. W., Huisman, M. H., Sutedja, N. A., van der Kooi, A. J., de Visser, M., Schelhaas, H. J., ... van den Berg, L. H. (2012). Smoking, alcohol consumption, and the risk of amyotrophic lateral sclerosis: A population-based study. *American Journal of Epidemiology*, *176*(3), 233–239. doi:10.1093/aje/kws015 PMID:22791740

de Kloet, E. R., Oitzl, M. S., & Joëls, M. (1999). Stress and cognition: Are corticosteroids good or bad guys? *Trends in Neurosciences*, 22(10), 422–426. doi:10.1016/S0166-2236(99)01438-1 PMID:10481183

de la Fuente-Fernández, R. (2012). Frontostriatal cognitive staging in Parkinson's disease. *Parkinson's Disease*, 2012, 561046. doi:10.1155/2012/561046 PMID:22191070

de la Monte, S. M., & Tong, M. (2014). Brain metabolic dysfunction at the core of Alzheimer's disease. *Biochemical Pharmacology*, 88(4), 548–559. doi:10.1016/j.bcp.2013.12.012 PMID:24380887

De Lau, L. M., & Breteler, M. M. (2006). Epidemiology of Parkinson's disease. *Lancet Neurology*, 5(6), 525–535. doi:10.1016/S1474-4422(06)70471-9 PMID:16713924

de Lau, L. M., Schipper, C. M., Hofman, A., Koudstaal, P. J., & Breteler, M. M. (2005). Prognosis of Parkinson disease: risk of dementia and mortality: the Rotterdam Study. *Archives of Neurology*, 62(8), 1265–1269. doi:10.1001/archneur.62.8.1265 PMID:16087767

De Leeuw, F. E., de Groot, J. C., Achten, E., Oudkerk, M., Ramos, L. M., Heijboer, R., ... Breteler, M. M. (2001). Prevalence of cerebral white matter lesions in elderly people: A population based magnetic resonance imaging study. The Rotterdam Scan Study. *Journal of Neurology, Neurosurgery, and Psychiatry*, *70*(1), 9–14. doi:10.1136/jnnp.70.1.9 PMID:11118240

de Moura, M. B., dos Santos, L. S., & Van Houten, B. (2010). Mitochondrial dysfunction in neurodegenerative diseases and cancer. *Environmental and Molecular Mutagenesis*, *51*, 391–405. PMID:20544881

de Oliveira Manoel, A. L., Goffi, A., Zampieri, F. G., Turkel-Parrella, D., Duggal, A., Marotta, T. R., ... Abrahamson, S. (2016). The critical care management of spontaneous intracranial hemorrhage: A contemporary review. *Critical Care (London, England)*, 20(1), 272. doi:10.118613054-016-1432-0 PMID:27640182

de Oliveira, D. M., Ferreira Lima, R. M., & El-Bachá, R. S. (2012). Brain rust: Recent discoveries on the role of oxidative stress in neurodegenerative diseases. *Nutritional Neuroscience*, *15*(3), 94–102. doi:10.1179/1476830511Y.0000000029 PMID:22583954

De Stefani, D., Raffaello, A., Teardo, E., Szabo, I., & Rizzuto, R. (2011). A forty kilodalton protein of the inner membrane is the mitochondrial calcium uniporter. *Nature*, 476(7360), 336–340. doi:10.1038/nature10230 PMID:21685888

De Vos, K. J., Mórotz, G. M., Stoica, R., Tudor, E. L., Lau, K. F., Ackerley, S., & Miller, C. C. (2011). VAPB interacts with the mitochondrial protein PTPIP51 to regulate calcium homeostasis. *Human Molecular Genetics*, *21*(6), 1299–1311. doi:10.1093/hmg/ddr559 PMID:22131369

Deane, R., Bell, R. D., Sagare, A., & Zlokovic, B. V. (2009). Clearance of amyloid-beta peptide across the blood-brain barrier: Implication for therapies in Alzheimer's disease. *CNS & Neurological Disorders - Drug Targets*, 8(1), 16–30. doi:10.2174/187152709787601867 PMID:19275634

Deas, E., Plun-Favreau, H., & Wood, N. W. (2009). PINK1 function in health and disease. *EMBO Molecular Medicine*, *1*(3), 152–165. doi:10.1002/emmm.200900024 PMID:20049715

Deas, E., Wood, N. W., & Plun-Favreau, H. (2011). Mitophagy and Parkinson's disease: The PINK1–parkin link. *Biochimica et Biophysica Acta (BBA)- Molecular Cell Research*, *1813*(4), 623–633.

Debette, S., Beiser, A., DeCarli, C., Au, R., Himali, J. J., Kelly-Hayes, M., ... Seshadri, S. (2010). Association of MRI markers of vascular brain injury with incident stroke, mild cognitive impairment, dementia, and mortality: The Framingham Offspring Study. *Stroke*, *41*(4), 600–606. doi:10.1161/STROKEAHA.109.570044 PMID:20167919

Debette, S., & Markus, H. S. (2010). The clinical importance of white matter hyperintensities on brain magnetic resonance imaging: Systematic review and meta-analysis. *BMJ (Clinical Research Ed.)*, *341*(jul26 1), c3666. doi:10.1136/bmj.c3666 PMID:20660506

Deborah, R. M., & Cathy, W. L. (2012). Ion channels and zinc: Mechanisms of neurotoxicity and neurodegeneration. *Journal of Toxicology*, 2012, 785647. PMID:22645609

Defazio, G., Berardelli, A., Fabbrini, G., Martino, D., Fincati, E., Fiaschi, A., ... Tinazzi, M. (2008). Pain as a nonmotor symptom of Parkinson disease: Evidence from a case-control study. *Archives of Neurology*, 65(9), 1191–1194. doi:10.1001/archneurol.2008.2 PMID:18779422

Dehay, B., Bové, J., Rodríguez-Muela, N., Perier, C., Recasens, A., Boya, P., & Vila, M. (2010). Pathogenic lysosomal depletion in Parkinson's disease. *The Journal of Neuroscience*, *30*(37), 12535–12544. doi:10.1523/JNEURO-SCI.1920-10.2010 PMID:20844148

Dehay, B., Decressac, M., Bourdenx, M., Guadagnino, I., Fernagut, P.-O., Tamburrino, A., ... Bezard, E. (2016). Targeting α-synuclein: Therapeutic options. *Movement Disorders*, *31*(6), 882–888. doi:10.1002/mds.26568 PMID:26926119 Delacourte, A., Flament, S., Dibe, E. M., Hublau, P., Sablonnière, B., Hémon, B., ... Défossez, A. (1990). Pathological proteins Tau 64 and 69 are specifically expressed in the somatodendritic domain of the degenerating cortical neurons during Alzheimer's disease. Demonstration with a panel of antibodies against Tau proteins. *Acta Neuropathologica*, *80*(2), 111–7. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/2117840

Delatycki, M. B., Williamson, R., & Forrest, S. M. (2000). Friedreich ataxia: An overview. *Journal of Medical Genetics*, 37(1), 1–8. doi:10.1136/jmg.37.1.1 PMID:10633128

DeLong, M. R. (1990, July). Primate models of movement disorders of basal ganglia origin. *Trends in Neurosciences*, 13(7), 281–285. doi:10.1016/0166-2236(90)90110-V PMID:1695404

DeMaagd, G., & Philip, A. (2015). Parkinson's Disease and Its Management: Part 1: Disease Entity, Risk Factors, Pathophysiology, Clinical Presentation, and Diagnosis. *P&T*, 40(8), 504–532. PMID:26236139

Den Brok, M. G., van Dalen, J. W., van Gool, W. A., Moll van Charante, E. P., de Bie, R. M., & Richard, E. (2015). Apathy in Parkinson's disease: A systematic review and meta-analysis. *Movement Disorders*, *30*(6), 759–769. doi:10.1002/ mds.26208 PMID:25787145

Deng, H., Dodson, M. W., Huang, H., & Guo, M. (2008). The Parkinson's disease genes pink1 and parkin promote mitochondrial fission and/or inhibit fusion in Drosophila. *Proceedings of the National Academy of Sciences of the United States of America*, *105*(38), 14503–14508. doi:10.1073/pnas.0803998105 PMID:18799731

Desport, J. C., Mabrouk, T., Bouillet, P., Perna, A., Preux, P. M., & Couratier, P. (2005). Complications and survival following radiologically and endoscopically-guided gastrostomy in patients with amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, 6(2), 88–93. doi:10.1080/14660820410021258a PMID:16036431

Desport, J. C., Preux, P. M., Truong, C. T., Courat, L., Vallat, J. M., & Couratier, P. (2000). Nutritional assessment and survival in ALS patients. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, *1*, 91–96. doi:10.1080/14660820050515386 PMID:11467055

Dettmer, U., Newman, A. J., Soldner, F., Luth, E. S., Kim, N. C., von Saucken, V. E., ... Selkoe, D. (2015). Parkinsoncausing α -synuclein missense mutations shift native tetramers to monomers as a mechanism for disease initiation. *Nature Communications*, 6(1), 7314. doi:10.1038/ncomms8314 PMID:26076669

Devi, L., Prabhu, B. M., Galati, D. F., Avadhani, N. G., & Anandatheerthavarada, H. K. (2006). Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheimer's disease brain is associated with mitochondrial dysfunction. *The Journal of Neuroscience*, *26*(35), 9057–9068. doi:10.1523/JNEUROSCI.1469-06.2006 PMID:16943564

Devi, L., Raghavendran, V., Prabhu, B. M., Avadhani, N. G., & Anandatheerthavarada, H. K. (2008). Mitochondrial import and accumulation of α-synuclein impair complex I in human dopaminergic neuronal cultures and Parkinson disease brain. *The Journal of Biological Chemistry*, 283(14), 9089–9100. doi:10.1074/jbc.M710012200 PMID:18245082

Devinsky, O., Vezzani, A., Najjar, S., De Lanerolle, N. C., & Rogawski, M. A. (2013). Glia and epilepsy: Excitability and inflammation. *Trends in Neurosciences*, *36*(3), 174–184. doi:10.1016/j.tins.2012.11.008 PMID:23298414

Dewey, R. B., & Jankovic, J. (1989). Hemiballism-hemichorea: Clinical and pharmacologic findings in 21 patients. *Archives of Neurology*, *46*(8), 862–867. doi:10.1001/archneur.1989.00520440044020 PMID:2757526

Dexter, D. T., Carayon, A., Javoy-Agid, F., Agid, Y., Wells, F. R., Daniel, S. E., ... Marsden, C. D. (1991). Alterations in the levels of iron, ferritin and other trace metals in parkinson's disease and other neurodegenerative diseases affecting the basal ganglia. *Brain*, *114*(4), 1953–1975. doi:10.1093/brain/114.4.1953 PMID:1832073

Dexter, D., Carter, C., Wells, F., Javoy-Agid, F., Agid, Y., Lees, A., ... Marsden, C. D. (1989). Basal lipid peroxidation in substantia nigra is increased in Parkinson's disease. *Journal of Neurochemistry*, *52*(2), 381–389. doi:10.1111/j.1471-4159.1989. tb09133.x PMID:2911023

Di Donato, I., Bianchi, S., De Stefano, N., Dichgans, M., Dotti, M. T., Duering, M., ... Federico, A. (2017). Cerebral Autosomal Dominant Arteriopathy with Subcortical Infarcts and Leukoencephalopathy (CADASIL) as a model of small vessel disease: Update on clinical, diagnostic, and management aspects. *BMC Medicine*, *15*(1), 41. doi:10.118612916-017-0778-8 PMID:28231783

Di Maio, R., Barrett, P. J., Hoffman, E. K., Barrett, C. W., Zharikov, A., Borah, A., ... Greenamyre, J. T. (2016). α -Synuclein binds to TOM20 and inhibits mitochondrial protein import in Parkinson's disease. *Science Translational Medicine*, 8(342), 342–378. doi:10.1126citranslmed.aaf3634 PMID:27280685

Di Monte, D., Jewell, S. A., Ekström, G., Sandy, M. S., & Smith, M. T. (1986). 1-Methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MPTP) and 1-methyl-4-phenylpyridine (MPP+) cause rapid ATP depletion in isolated hepatocytes. *Biochemical and Biophysical Research Communications*, *137*(1), 310–315. doi:10.1016/0006-291X(86)91211-8 PMID:3487319

Dias, V., Junn, E., & Mouradian, M. M. (2013). The role of oxidative stress in Parkinson's disease. *Journal of Parkinson's Disease*, *3*(4), 461–491. PMID:24252804

Dickerson, B. C., Goncharova, I., Sullivan, M., Forchetti, C., Wilson, R., Bennett, D., ... deToledo-Morrell, L. (2001). MRI-derived entorhinal and hippocampal atrophy in incipient and very mild Alzheimer's disease. *Neurobiology of Aging*, 22(5), 747–754. doi:10.1016/S0197-4580(01)00271-8 PMID:11705634

Dick, F. D. (2006). Parkinson's disease and pesticide exposures. *British Medical Bulletin*, 79(1), 219–231. doi:10.1093/ bmb/ldl018 PMID:17242039

Dickson, D. W. (1998). Pick's disease: A modern approach. *Brain Pathology (Zurich, Switzerland)*, 8(2), 339–354. doi:10.1111/j.1750-3639.1998.tb00158.x PMID:9546291

Dickson, D. W. (2010). Neuropathology of non-Alzheimer degenerative disorders. *International Journal of Clinical and Experimental Pathology*, *3*(1), 1. PMID:19918325

Dickson, D. W., Bergeron, C., Chin, S. S., Duyckaerts, C., Horoupian, D., Ikeda, K., ... Litvan, I. (2002). Office of Rare Diseases neuropathologic criteria for corticobasal degeneration. *Journal of Neuropathology and Experimental Neurology*, *61*(11), 935–946. doi:10.1093/jnen/61.11.935 PMID:12430710

Dickson, D. W., Davies, P., Mayeux, R., Crystal, H., Horoupian, D. S., Thompson, A., & Goldman, J. E. (1986). Diffuse Lewy body disease. Neuropathological and biochemical studies of six patients. *Acta Neuropathologica*, 75(1), 8–15. doi:10.1007/BF00686786 PMID:3434218

Dickson, D. W., Feany, M. B., Yen, S. H., Mattiace, L. A., & Davies, P. (1996). Cytoskeletal pathology in non-Alzheimer degenerative dementia: new lesions in diffuse Lewy body disease, Pick's disease, and corticobasal degeneration. In *New Trends in the Diagnosis and Therapy of Non-Alzheimer's Dementia* (pp. 31–46). Springer Vienna. doi:10.1007/978-3-7091-6892-9_2

Dickson, D. W., Fujishiro, H., DelleDonne, A., Menke, J., Ahmed, Z., Klos, K. J., ... Ahlskog, J. E. (2008). Evidence that incidental Lewy body disease is pre-symptomatic Parkinson's disease. *Acta Neuropathologica*, *115*(4), 437–444. doi:10.100700401-008-0345-7 PMID:18264713

Dickson, D. W., Lin, W., Liu, W. K., & Yen, S. H. (1999). Multiple system atrophy: A sporadic synucleinopathy. *Brain Pathology (Zurich, Switzerland)*, *9*(4), 721–732. doi:10.1111/j.1750-3639.1999.tb00553.x PMID:10517510

Dickson, D. W., Liu, W., Hardy, J., Farrer, M., Mehta, N., Uitti, R., ... Yen, S. H. (1999). Widespread alterations of alpha-synuclein in multiple system atrophy. *American Journal of Pathology*, *155*(4), 1241–1251. doi:10.1016/S0002-9440(10)65226-1 PMID:10514406

Diehl, J., Grimmer, T., Drzezga, A., Riemenschneider, M., Förstl, H., & Kurz, A. (2004). Cerebral metabolic patterns at early stages of frontotemporal dementia and semantic dementia. A PET study. *Neurobiology of Aging*, 25(8), 1051–1056. doi:10.1016/j.neurobiologing.2003.10.007 PMID:15212830

Dienel, G. A. (2013). Astrocytic energetics during excitatory neurotransmission: What are contributions of glutamate oxidation and glycolysis? *Neurochemistry International*, 63(4), 244–258. doi:10.1016/j.neuint.2013.06.015 PMID:23838211

Diener, H. C., Sacco, R. L., Yusuf, S., Cotton, D., Ounpuu, S., Lawton, W. A., ... Yoon, B. W. (2008). Prevention Regimen for Effectively Avoiding Second Strokes (PRoFESS) study group. Effects of aspirin plus extended-release dipyridamole versus clopidogrel and telmisartan on disability and cognitive function after recurrent stroke in patients with ischaemic stroke in the Prevention Regimen for Effectively Avoiding Second Strokes (PRoFESS) trial: A double-blind, active and placebo-controlled study. *Lancet Neurology*, 7(10), 875–884. doi:10.1016/S1474-4422(08)70198-4 PMID:18757238

Dirani, M., Nasreddine, W., Abdulla, F., & Beydoun, A. (2014). Seizure control and improvement of neurological dysfunction in Lafora disease with perampanel. *Epilepsy & Behavior Case Reports*, 2, 164–166. doi:10.1016/j.ebcr.2014.09.003 PMID:25667898

Distad, B. J., Meekins, G. D., Liou, L. L., Weiss, M. D., Carter, G. T., & Miller, R. G. (2008). Drug Therapy in Amyotrophic Lateral Sclerosis. *Physical Medicine and Rehabilitation Clinics of North America*, *19*(3), 633–651. doi:10.1016/j. pmr.2008.04.005 PMID:18625421

Ditsworth, D., Maldonado, M., McAlonis-Downes, M., Sun, S., Seelman, A., Drenner, K., ... Da Cruz, S. (2017). Mutant TDP-43 within motor neurons drives disease onset but not progression in amyotrophic lateral sclerosis. *Acta Neuropathologica*, *133*(6), 907–922. doi:10.100700401-017-1698-6 PMID:28357566

Dixit, S., Bernardo, A., Walker, J. M., Kennard, J. A., Kim, G. Y., Kessler, E. S., & Harrison, F. E. (2015). Vitamin C deficiency in the brain impairs cognition, increases amyloid accumulation and deposition, and oxidative stress in APP/ PSEN1 and normally aging mice. *ACS Chemical Neuroscience*, *6*(4), 570–581. doi:10.1021/cn500308h PMID:25642732

Doens, D., & Fernández, P. L. (2014). Microglia receptors and their implications in the response to amyloid β for Alzheimer's disease pathogenesis. *Journal of Neuroinflammation*, 11(1), 48. doi:10.1186/1742-2094-11-48 PMID:24625061

Dohi, K., Ohtaki, H., Nakamachi, T., Yofu, S., Satoh, K., Miyamoto, K., ... Aruga, T. (2010). Gp91 phox (NOX2) in classically activated microglia exacerbates traumatic brain injury. *Journal of Neuroinflammation*, 7(1), 41. doi:10.1186/1742-2094-7-41 PMID:20659322

Donaghy, P. C., & McKeith, I. G. (2014). The clinical characteristics of dementia with Lewy bodies and a consideration of prodromal diagnosis. *Alzheimer's Research & Therapy*, *6*(4), 46. doi:10.1186/alzrt274 PMID:25484925

Dong, S., Duan, Y., Hu, Y., & Zhao, Z. (2012). Advances in the pathogenesis of Alzheimer's disease: A re-evaluation of amyloid cascade hypothesis. *Translational Neurodegeneration*, *1*(1), 18. doi:10.1186/2047-9158-1-18 PMID:23210692

Dormann, D., & Haass, C. (2013). Fused in sarcoma (FUS): An oncogene goes awry in neurodegeneration. *Molecular* and Cellular Neurosciences, 56, 475–286. doi:10.1016/j.mcn.2013.03.006 PMID:23557964

Dormann, D., Rodde, R., Edbauer, D., Bentmann, E., Fischer, I., Hruscha, A., ... Haass, C. (2010). ALS-associated fused in sarcoma (FUS) mutations disrupt Transportin-mediated nuclear import. *The EMBO Journal*, 29(16), 2841–2857. doi:10.1038/emboj.2010.143 PMID:20606625

Doss-Pepe, E. W., Chen, L., & Madura, K. (2005). *α-synuclein* and *Parkin* contribute to the assembly of ubiquitin lysine 63-linked multiubiquitin chains. *The Journal of Biological Chemistry*, 280(17), 16619–16624. doi:10.1074/jbc. M413591200 PMID:15718234

Doty, R. L., Deems, D. A., & Stellar, S. (1988). Olfactory dysfunction in parkinsonism: A general deficit unrelated to neurologic signs, disease stage, or disease duration. *Neurology*, *38*(8), 1237–1244. doi:10.1212/WNL.38.8.1237 PMID:3399075

Dozono, K., Ishii, N., Nishihara, Y., & Horie, A. (1991). An autopsy study of the incidence of lacunes in relation to age, hypertension, and arteriosclerosis. *Stroke*, 22(8), 993–996. doi:10.1161/01.STR.22.8.993 PMID:1866767

Drachman, D. B., Frank, K., Dykes-Hoberg, M., Teismann, P., Almer, G., Przedborski, S., & Rothstein, J. D. (2002). Cyclooxygenase 2 inhibition protects motor neurons and prolongs survival in a transgenic mouse model of ALS. *Annals of Neurology*, *52*(6), 771–778. doi:10.1002/ana.10374 PMID:12447931

Drago, I., Pizzo, P., & Pozzan, T. (2011). After half a century mitochondrial calcium in- and efflux machineries reveal themselves. *European Molecular Biology Organization Journal*, *30*(20), 4119–4125. doi:10.1038/emboj.2011.337 PMID:21934651

Dröge, W. (2002). Free radicals in the physiological control of cell function. *Physiological Reviews*, 82(1), 47–95. doi:10.1152/physrev.00018.2001 PMID:11773609

Drose, S., Brandt, U., & Wittig, I. (2014). Mitochondrial respiratory chain complexes as sources and targets of thiol-based redox-regulation. *Biochimica et Biophysica Acta*, *1844*(8), 1344–1354. doi:10.1016/j.bbapap.2014.02.006 PMID:24561273

Dubois, B., Burn, D., Goetz, C., Aarsland, D., Brown, R. G., Broe, G. A., ... Korczyn, A. (2007). Diagnostic procedures for Parkinson's disease dementia: Recommendations from the movement disorder society task force. *Movement Disorders*, 22(16), 2314–2324. doi:10.1002/mds.21844 PMID:18098298

Duda, J. E., Giasson, B. I., Mabon, M. E., Lee, V. M. Y., & Trojanowski, J. Q. (2002). Novel antibodies to synuclein show abundant striatal pathology in Lewy body diseases. *Annals of Neurology*, *52*(2), 205–210. doi:10.1002/ana.10279 PMID:12210791

Duering, M., Righart, R., Wollenweber, F. A., Zietemann, V., Gesierich, B., & Dichgans, M. (2015). Acute infarcts cause focal thinning in remote cortex via degeneration of connecting fiber tracts. *Neurology*, *84*(4), 1685–1692. doi:10.1212/ WNL.000000000001502 PMID:25809303

Duering, M., Zieren, N., Hervé, D., Jouvent, E., Reyes, S., Peters, N., ... Dichgans, M. (2011). Strategic role of frontal white matter tracts in vascular cognitive impairment: A voxel-based lesion-symptom mapping study in CADASIL. *Brain*, *134*(8), 2366–2375. doi:10.1093/brain/awr169 PMID:21764819

Du, J. J., & Chen, S. D. (2017). Current Nondopaminergic Therapeutic Options for Motor Symptoms of Parkinson's Disease. *Chinese Medical Journal*, *130*(15), 1856–1866. doi:10.4103/0366-6999.211555 PMID:28748860

Dujardin, K., Defebvre, L., Krystkowiak, P., Degreef, J. F., & Destee, A. (2003). Executive function differences in multiple system atrophy and Parkinson's disease. *Parkinsonism & Related Disorders*, 9(4), 205–211. doi:10.1016/S1353-8020(02)00050-0 PMID:12618055

Du, L., Zhang, X., Han, Y. Y., Burke, N. A., Kochanek, P. M., Watkins, S. C., ... Clark, R. S. (2003). Intra-mitochondrial poly (ADP-ribosylation) contributes to NAD+ depletion and cell death induced by oxidative stress. *The Journal of Biological Chemistry*, 278(20), 18426–18433. doi:10.1074/jbc.M301295200 PMID:12626504

Duyckaerts, C., Delatour, B., & Potier, M. C. (2009). Classification and basic pathology of Alzheimer disease. *Acta Neuropathologica*, *118*(1), 5–36. doi:10.100700401-009-0532-1 PMID:19381658

Dwyer, B. E., Zacharski, L. R., Balestra, D. J., Lerner, A. J., Perry, G., Zhu, X., & Smith, M. A. (2009). Getting the iron out: Phlebotomy for Alzheimer's disease? *Medical Hypotheses*, 72(5), 504–509. doi:10.1016/j.mehy.2008.12.029 PMID:19195795

Dyken, P. R. (1985). Subacute sclerosing panencephalitis. Current status. Neurologic Clinics, 3(1), 179–196. PMID: 2581121

Dyken, P., & Krawiecki, N. (1983). Neurodegenerative diseases of infancy and childhood. *Annals of Neurology*, *13*(4), 351–364. doi:10.1002/ana.410130402 PMID:6301358

Ebrahimi-Fakhari, D., Wahlster, L., & McLean, P. J. (2012). Protein degradation pathways in Parkinson's disease: Curse or blessing. *Acta Neuropathologica*, *124*(2), 153–172. doi:10.100700401-012-1004-6 PMID:22744791

Echeverria, V., & Cuello, A. C. (2002). Intracellular A-beta amyloid, a sign for worse things to come? *Molecular Neurobiology*, *26*(2–3), 299–316. doi:10.1385/MN:26:2-3:299 PMID:12428762

Eder, K., Kish, S. J., Kirchgessner, M., & Ross, B. M. (1998). Brain phospholipids and fatty acids in Friedreich's ataxia and spinocerebellar atrophy type-1. *Movement Disorders*, *13*(5), 813–819. doi:10.1002/mds.870130510 PMID:9756151

Editors of Encyclopædia Britannica. (2016). Alzheimer Disease. In *Encyclopaedia Britannica* (pp. 1–14). Encyclopædia Britannica, Inc. Retrieved from https://www.britannica.com/science/Alzheimer-disease

Edwards, R. H. (1993). Neural degeneration and the transport of neurotransmitters. *Annals of Neurology*, *34*(5), 638–645. doi:10.1002/ana.410340504 PMID:7902065

Ehlers, S., & Gillberg, C. (1993). The epidemiology of Asperger syndrome: A total population study. *Journal of Child Psychology and Psychiatry, and Allied Disciplines, 34*(8), 1327–1350. doi:10.1111/j.1469-7610.1993.tb02094.x PMID:8294522

Eichner, T., & Radford, S. E. (2011). A diversity of assembly mechanisms of a generic amyloid fold. *Molecular Cell*, 43(1), 8–18. doi:10.1016/j.molcel.2011.05.012 PMID:21726806

Eidelberg, D., Sotrel, A., Joachim, C., Selkoe, D., Forman, A., Perl, D. P., & Pendlebury, W. W. (1987). Adult onset Hallervorden-Spatz disease with neurofibrillary pathology. A discrete clinico pathological entity. *Brain*, *110*(Pt 4), 993–1013. doi:10.1093/brain/110.4.993 PMID:2888513

Eikermann-Haerter, K., Yuzawa, I., Dilekoz, E., Joutel, A., Moskowitz, M. A., & Ayata, C. (2011). Cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy syndrome mutations increase susceptibility to spreading depression. *Annals of Neurology*, *69*(2), 413–418. doi:10.1002/ana.22281 PMID:21387384

Eisai. (2017). Major R and D Pipeline. Retrieved from http://www.eisai.com/pdf/eir/erepo/epipeline.pdf

Ekstrand, M. I., Terzioglu, M., Galter, D., Zhu, S., Hofstetter, C., Lindqvist, E., & Hoffer, B. (2007). Progressive Parkinsonism in mice with respiratory-chain-deficient dopamine neurons. *Proceedings of the National Academy of Sciences of the United States of America*, *104*(4), 1325–1330. doi:10.1073/pnas.0605208103 PMID:17227870

Elias, M. F., Elias, P. K., Sullivan, L. M., Wolf, P. A., & D'Agostino, R. B. (2005). Obesity, diabetes and cognitive deficit: The Framingham Heart Study. *Neurobiology of Aging*, *26*(1), 11–16. doi:10.1016/j.neurobiolaging.2005.08.019 PMID:16223549

Elias, W. J., Huss, D., Voss, T., Loomba, J., Khaled, M., Zadicario, E., ... Wintermark, M. (2013). A pilot study of focused ultrasound thalamotomy for essential tremor. *The New England Journal of Medicine*, *369*(7), 640–648. doi:10.1056/ NEJMoa1300962 PMID:23944301

Elkins, J. S., Longstreth, W. T. Jr, Manolio, T. A., Newman, A. B., Bhadelia, R. A., & Johnston, S. C. (2006). Education and the cognitive decline associated with MRI-defined brain infarct. *Neurology*, *67*(3), 435–440. doi:10.1212/01. wnl.0000228246.89109.98 PMID:16894104

Ellis, D. Z., Rabe, J., & Sweadner, K. J. (2003). Global loss of Na,K-ATPase and its nitric oxide-mediated regulation in a transgenic mouse model of amyotrophic lateral sclerosis. *The Journal of Neuroscience*, 23, 43–51. PMID:12514200

Ellis, R., Olichney, J., Thal, L., Mirra, S., Morris, J., Beekly, D., & Heyman, A. (1996). Cerebral amyloid angiopathy in the brains of patients with Alzheimer's disease The CERAD experience, part XV. *Neurology*, *46*(6), 1592–1596. doi:10.1212/WNL.46.6.1592 PMID:8649554

Ellis, T. M., Foote, K. D., Fernandez, H. H., Sudhyadhom, A., Rodriguez, R. L., Zeilman, P., ... Okun, M. S. (2008). Reoperation for suboptimal outcomes after deep brain stimulation surgery. *Neurosurgery*, *63*(4), 754–760. doi:10.1227/01. NEU.0000325492.58799.35 PMID:18981887

Elmallah, M. I., Borgmeyer, U., Betzel, C., & Redecke, L. (2013). Impact of methionine oxidation as an initial event on the pathway of human prion protein conversion. *Prion*, 7(5), 404–411. doi:10.4161/pri.26745 PMID:24121542

Elmore, S. (2007). Apoptosis: A Review of Programmed Cell Death. *Toxicologic Pathology*, 35(4), 495–516. doi:10.1080/01926230701320337 PMID:17562483

Elsabbagh, M., Divan, G., Koh, Y. J., Kim, Y. S., Kauchali, S., Marcín, C., ... Fombonne, E. (2012). Global prevalence of autism and other pervasive developmental disorders. *Autism Research*, *5*(3), 160–179. doi:10.1002/aur.239 PMID:22495912

Elson, J. L., Herrnstadt, C., Preston, G., Thal, L., Morris, C. M., Edwardson, J., ... Howell, N. (2006). Does the mitochondrial genome play a role in the etiology of Alzheimer's disease? *Human Genetics*, *119*(3), 241–254. doi:10.100700439-005-0123-8 PMID:16408223

Emerit, J. M., Edeas, F. B., & Bricaire, F. (2004). Neurodegenerative diseases and oxidative stress. *Biomedicine and Pharmacotherapy*, 58(1), 39–46. doi:10.1016/j.biopha.2003.11.004 PMID:14739060

Emery, N. J., & Clayton, N. S. (2001). Effects of experience and social context on prospective caching strategies by scrub jays. *Nature*, 414(6862), 443–446. doi:10.1038/35106560 PMID:11719804

Emmanouilidou, E., Stefanis, L., & Vekrellis, K. (2010). Cell-produced α-synuclein oligomers are targeted to, and impair, the 26S proteasome. *Neurobiology of Aging*, *31*(6), 953–968. doi:10.1016/j.neurobiolaging.2008.07.008 PMID:18715677

Emre, M. (2003). Dementia associated with Parkinson's disease. *Lancet Neurology*, 2(4), 229–237. doi:10.1016/S1474-4422(03)00351-X PMID:12849211

Emre, M., Aarsland, D., Brown, R., Burn, D. J., Duyckaerts, C., Mizuno, Y., ... Dubois, B. (2007). Clinical diagnostic criteria for dementia associated with Parkinson's disease. *Movement Disorders*, 22(12), 1689–1707. doi:10.1002/ mds.21507 PMID:17542011

Engelender, S. (2008). Ubiquitination of α -synuclein and autophagy in Parkinson's disease. *Autophagy*, 4(3), 372–374. doi:10.4161/auto.5604 PMID:18216494

Engel, J. Jr, Wiebe, S., French, J., Sperling, M., Williamson, P., Spencer, D., ... Enos, B. (2003). Practice parameter: Temporal lobe and localized neocortical resections for epilepsy. *Neurology*, *60*(4), 538–547. doi:10.1212/01. WNL.0000055086.35806.2D PMID:12601090

English, A. R., & Voeltz, G. K. (2013). Endoplasmic reticulum structure and interconnections with other organelles. *Cold Spring Harbor Perspectives in Biology*, *5*(4), a013227. doi:10.1101/cshperspect.a013227 PMID:23545422

Enstrom, A. M., Lit, L., Onore, C. E., Gregg, J. P., Hansen, R. L., Pessah, I. N., ... Zimmerman, A. W. (2005). Immunity, neuroglia and neuroinflammation in autism. *International Review of Psychiatry (Abingdon, England)*, *17*(6), 485–495. doi:10.1080/02646830500381930 PMID:16401547

Erkinjuntti, T., Inzitari, D., Pantoni, L., Wallin, A., Scheltens, P., Rockwood, K., ... Desmond, D. W. (2000). Research criteria for subcortical vascular dementia in clinical trials. *Journal of Neural Transmission (Vienna, Austria)*, *59*, 23–30. PMID:10961414

Ernst, R. L., Hay, J. W., Fenn, C., Tinklenberg, J., & Yesavage, J. A. (1997). Cognitive function and the costs of Alzheimer disease. An exploratory study. *Archives of Neurology*, *54*(6), 687–693. doi:10.1001/archneur.1997.00550180013006 PMID:9193203

Esler, W. P., & Wolfe, M. S. (2001). A portrait of Alzheimer secretases--new features and familiar faces. *Science*, 293(5534), 1449–1454. doi:10.1126cience.1064638 PMID:11520976

Evans, T. A., Siedlak, S. L., Lu, L., Fu, X., Wang, Z., McGinnis, W. R., ... Zhu, X. (2008). The autistic phenotype exhibits a remarkably localized modification of brain protein by products of free radical-induced lipid oxidation. *American Journal of Biochemistry and Biotechnology*, 4(2), 61–72. doi:10.3844/ajbbsp.2008.61.72

Exner, N., Treske, B., Paquet, D., Holmström, K., Schiesling, C., Gispert, S., & Krüger, R. (2007). Loss-of-function of human PINK1 results in mitochondrial pathology and can be rescued by Parkin. *The Journal of Neuroscience*, *27*(45), 12413–12418. doi:10.1523/JNEUROSCI.0719-07.2007 PMID:17989306

Fahn, S. (2003). Description of Parkinson's disease as a clinical syndrome. *Annals of the New York Academy of Sciences*, *991*(1), 1–14. doi:10.1111/j.1749-6632.2003.tb07458.x PMID:12846969

Fakhoury, M. (2015). Autistic spectrum disorders: A review of clinical features, theories and diagnosis. *International Journal of Developmental Neuroscience*, 43, 70–77. doi:10.1016/j.ijdevneu.2015.04.003 PMID:25862937

Fallis, B. A., & Hardiman, O. (2009). Aggregation of neurodegenerative disease in ALS kindreds. *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases, 10*(2), 95–98. doi:10.1080/17482960802209664 PMID:18608094

Fang, F., Bellocco, R., Hernan, M. A., & Ye, W. (2006). Smoking, snuff dipping and the risk of amyotrophic lateral sclerosis—a prospective cohort study. *Neuroepidemiology*, 27(4), 217–221. doi:10.1159/000096956 PMID:17106211

Fang, F., Hallmarker, U., James, S., Ingre, C., Michaelsson, K., Ahlbom, A., & Feychting, M. (2016). Amyotrophic lateral sclerosis among cross-country skiers in Sweden. *European Journal of Epidemiology*, *31*(3), 247–253. doi:10.100710654-015-0077-7 PMID:26220522

Fania, C., Arosio, B., Capitanio, D., Torretta, E., Gussago, C., Ferri, E., & Gelfi, C. (2017). Protein signature in cerebrospinal fluid and serum of Alzheimer's disease patients : The case of apolipoprotein A-1 proteoforms. *PLoS One*, *12*(6), 1–19. doi:10.1371/journal.pone.0179280 PMID:28628634

Fan, J., Hu, Z., Zeng, L., Lu, W., Tang, X., Zhang, J., & Li, T. (2008). Golgi apparatus and neurodegenerative diseases. *International Journal of Developmental Neuroscience*, 26(6), 523–534. doi:10.1016/j.ijdevneu.2008.05.006 PMID:18599251

Farg, M. A., Soo, K. Y., Warraich, S. T., Sundaramoorthy, V., Blair, I. P., & Atkin, J. D. (2012). Ataxin-2 interacts with FUS and intermediate-length polyglutamine expansions enhance FUS-related pathology in amyotrophic lateral sclerosis. *Human Molecular Genetics*, 22(4), 717–728. doi:10.1093/hmg/dds479 PMID:23172909

Farmer, C., Thurm, A., & Grant, P. (2013). Pharmacotherapy for the core symptoms in autistic disorder: Current status of the research. *Drugs*, 73(4), 303–314. doi:10.100740265-013-0021-7 PMID:23504356

Farrer, M. J. (2006). Genetics of Parkinson disease: Paradigm shifts and future prospects. *Nature Reviews. Genetics*, 7(4), 306–318. doi:10.1038/nrg1831 PMID:16543934

Fayet, G., Jansson, M., Sternberg, D., Moslemi, A. R., Blondy, P., Lombès, A., ... Oldfors, A. (2002). Ageing muscle: Clonal expansions of mitochondrial DNA point mutations and deletions cause focal impairment of mitochondrial function. *Neuromuscular Disorders*, *12*(5), 484–493. doi:10.1016/S0960-8966(01)00332-7 PMID:12031622

Fearnley, J. M., & Lees, A. J. (1991). Ageing and Parkinson's disease: Substantia nigra regional selectivity. *Brain*, *114*(5), 2283–2301. doi:10.1093/brain/114.5.2283 PMID:1933245

Febbraro, F., Giorgi, M., Caldarola, S., Loreni, F., & Romero-ramos, M. (2012). α-Synuclein expression is modulated at the translational level by iron. *Neuroreport*, 23(9), 576–580. doi:10.1097/WNR.0b013e328354a1f0 PMID:22581044

Feng, B., Ruiz, M. A., & Chakrabarti, S. (2013). Oxidative-stress-induced epigenetic changes in chronic diabetic complications- Review. *Canadian Journal of Physiology and Pharmacology*, *91*(3), 213–220. doi:10.1139/cjpp-2012-0251 PMID:23537434

Fenn, A. M., Henry, C. J., Huang, Y., Dugan, A., & Godbout, J. P. (2012). Lipopolysaccharide-induced interleukin (IL)-4 receptor-alpha expression and corresponding sensitivity to the M2 promoting effects of IL-4 are impaired in microglia of aged mice. *Brain, Behavior, and Immunity*, *26*(5), 766–777. doi:10.1016/j.bbi.2011.10.003 PMID:22024136

Fenoy, A. J., & Schiess, M. C. (2017, July). Deep Brain Stimulation of the Dentato-Rubro-Thalamic Tract: Outcomes of Direct Targeting for Tremor. *Neuromodulation*, 20(5), 429–436. doi:10.1111/ner.12585 PMID:28256785

Ferguson, T. A., & Elman, L. B. (2007). Clinical presentation and diagnosis of amyotrophic lateral sclerosis. *NeuroRehabilitation*, 22, 409–416. PMID:18198425

Ferman, T. J., Boeve, B. F., Smith, G. E., Silber, M. H., Lucas, J. A., Graff-Radford, N. R., ... Ivnik, R. J. (2002). Dementia with Lewy bodies may present as dementia and REM sleep behavior disorder without parkinsonism or hallucinations. *Journal of the International Neuropsychological Society*, 8(7), 907–914. doi:10.1017/S1355617702870047 PMID:12405541

Fernández-Checa, J. C., Fernández, A., Morales, A., Mari, M., Garcia-Ruiz, C., & Colell, A. (2010). Oxidative stress and altered mitochondrial function in neurodegenerative diseases: Lessons from mouse models. *CNS & Neurological Disorders - Drug Targets*, 9(4), 439–454. doi:10.2174/187152710791556113 PMID:20522012

Ferreira, M. E., de Vasconcelos, A. S., da Costa Vilhena, T., da Silva, T. L., da Silva Barbosa, A., Gomes, A. R., ... Percário, S. (2015). Oxidative Stress in Alzheimer's Disease: Should We Keep Trying Antioxidant Therapies? *Cellular and Molecular Neurobiology*, *35*(5), 595–614. doi:10.100710571-015-0157-y PMID:25616523

Ferro, J. M., Canhão, P., Stam, J., Bousser, M. G., Barinagarrementeria, F., Massaro, A., ... Kasner, S. E. (2009). Delay in the diagnosis of cerebral vein and dural sinus thrombosis: Influence on outcome. *Stroke*, *40*(9), 3133–3138. doi:10.1161/ STROKEAHA.109.553891 PMID:19608994

Findling, R. L. (2005). Pharmacologic treatment of behavioral symptoms in autism and pervasive developmental disorders. *The Journal of Clinical Psychiatry*, *66*(supplement 10), S26–S31. PMID:16401147

Finfer, S. R., & Cohen, J. (2001). Severe traumatic brain injury. *Resuscitation*, 48(1), 77–90. doi:10.1016/S0300-9572(00)00321-X PMID:11162885

Fink, A. L. (2006). The aggregation and fibrillation of alpha-synuclein. *Accounts of Chemical Research*, *39*(9), 628–634. doi:10.1021/ar050073t PMID:16981679

Finkbeiner, S. (2011). Huntington's Disease. *Cold Spring Harbor Perspectives in Biology*, *3*(6), a007476. doi:10.1101/ cshperspect.a007476 PMID:21441583

Finkel, T., & Holbrook, N. J. (2000). Oxidants, oxidative stress and the biology of ageing. *Nature*, 408(6809), 239–247. doi:10.1038/35041687 PMID:11089981

Fischer, U., Liu, Q., & Dreyfuss, G. (1997). The SMN–SIP1 complex has an essential role in spliceosomal snRNP biogenesis. *Cell*, 90(6), 1023–1029. doi:10.1016/S0092-8674(00)80368-2 PMID:9323130

Fisher, R. S., Acevedo, C., Arzimanoglou, A., Bogacz, A., Cross, J. H., Elger, C. E., ... Wiebe, S. (2014). A practical clinical definition of epilepsy. *Epilepsia*, 55(4), 475–482. doi:10.1111/epi.12550 PMID:24730690

Fisher, R. S., Cross, J. H., French, J. A., Higurashi, N., Hirsch, E., Jansen, F. E., ... Zuberi, S. M. (2017). Operational classification of seizure types by the International League Against Epilepsy: Position Paper of the ILAE Commission for Classification and Terminology. *Epilepsia*, 58(4), 522–530. doi:10.1111/epi.13670 PMID:28276060

Fjell, A. M., & Walhovd, K. B. (2010). Structural brain changes in aging: Courses, causes and cognitive consequences. *Reviews in the Neurosciences*, *21*(3), 187–221. doi:10.1515/REVNEURO.2010.21.3.187 PMID:20879692

Flood, F., Murphy, S., Cowburn, R. F., Lannfelt, L., Walker, B., & Johnston, J. A. (2005). Proteasome-mediated effects on amyloid precursor protein processing at the γ-secretase site. *The Biochemical Journal*, *385*(2), 545–550. doi:10.1042/BJ20041145 PMID:15473868

Fodero, L. R., Mok, S. S., Losic, D., Martin, L. L., Aguilar, M. I., Barrow, C. J., & Small, D. H. (2004). α 7-Nicotinic acetylcholine receptors mediate an A β 1– 42-induced increase in the level of acetylcholinesterase in primary cortical neurones. *Journal of Neurochemistry*, 88(5), 1186–1193. doi:10.1046/j.1471-4159.2003.02296.x PMID:15009674

Foix, C., & Nicolesco, J. (1925). Les noyaux gris centraux et la region mesencephalo-sous-optique: suivi d'un appendice sur l'anatomie pathologique de la maladie de Parkinson. Masson.

Follett, K. A., Weaver, F. M., Stern, M., Hur, K., Harris, C. L., Luo, P., ... Reda, D. J. (2010). Pallidal versus subthalamic deep-brain stimulation for Parkinson's disease. *The New England Journal of Medicine*, *362*(22), 2077–2091. doi:10.1056/ NEJMoa0907083 PMID:20519680

Folstein, S. E., Santangelo, S. L., Gilman, S. E., Piven, J., Landa, R. R., Lainhart, J., ... Wzorek, M. (1999). Predictors of cognitive test patterns in autism families. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, 40(7), 1117–1128. doi:10.1111/1469-7610.00528 PMID:10576540

Fombonne, E. (1999). The epidemiology of autism: A review. *Psychological Medicine*, 29(4), 769–786. doi:10.1017/S0033291799008508 PMID:10473304

Fombonne, E. (2003). Epidemiological surveys of autism and other pervasive developmental disorders: An update. *Journal of Autism and Developmental Disorders*, *33*(4), 365–382. doi:10.1023/A:1025054610557 PMID:12959416

Fombonne, E. (2011). Incidence and prevalence of pervasive developmental disorders. In E. Hollander, A. Kolevzon, & J. T. Coyle (Eds.), *Textbook of autism spectrum disorders* (pp. 117–136). Washington, DC: American Psychiatric Publishing, Inc. doi:10.1093/med/9780195371826.003.0007

Fonseca-Ornelas, L., Eisbach, S. E., Paulat, M., Giller, K., Fernández, C. O., Outeiro, T. F., ... Zweckstetter, M. (2014). Small molecule-mediated stabilization of vesicle-associated helical α-synuclein inhibits pathogenic misfolding and aggregation. *Nature Communications*, *5*, 5857. doi:10.1038/ncomms6857 PMID:25524885

Fontbonne, A., Berr, C., Ducimetière, P., & Alpérovitch, A. (2001). Changes in cognitive abilities over a 4-year period are unfavorably affected in elderly diabetic subjects. *Diabetes Care*, *24*(2), 366–370. doi:10.2337/diacare.24.2.366 PMID:11213894

Food and Drug Administration. (2002). Revised preventive measures to reduce the possible risk of transmission of Creutzfeldt-Jakob (CJD) disease and variant Creutzfeldt-Jakob disease (vCJD) by blood and blood products. FDA.

Forman, M. S., Trojanowski, J. Q., & Lee, V. M.-Y. (2004). Neurodegenerative diseases: A decade of discoveries paves the way for therapeutic breakthroughs. *Nature Medicine*, *10*(10), 1055–1063. doi:10.1038/nm1113 PMID:15459709

Fornai, F., Longone, P., Cafaro, L., Kastsiuchenka, O., Ferrucci, M., & Manca, M. L., ... Paparelli, A. (2008). Lithium delays progression of amyotrophic lateral sclerosis. *Proceedings of the National Academy of Sciences USA*, *105*, 2052–2057. 10.1073/pnas.0708022105

Fornai, F., Schlüter, O. M., Lenzi, P., Gesi, M., Ruffoli, R., Ferrucci, M., ... Battaglia, G. (2005). Parkinson-like syndrome induced by continuous MPTP infusion: Convergent roles of the ubiquitin-proteasome system and α -synuclein. *Proceedings of the National Academy of Sciences of the United States of America*, 102(9), 3413–3418. doi:10.1073/ pnas.0409713102 PMID:15716361

Forno, L. S. (1996). Neuropathology of Parkinson's disease. *Journal of Neuropathology and Experimental Neurology*, 55(30), 259–272.

Forno, L. S. (1996). Neuropathology of Parkinson's disease. *Journal of Neuropathology and Experimental Neurology*, 55(3), 259–272. doi:10.1097/00005072-199603000-00001 PMID:8786384

Foster, T. C. (2007). Calcium homeostasis and modulation of synaptic plasticity in the aged brain. *Aging Cell*, *6*(3), 319–325. doi:10.1111/j.1474-9726.2007.00283.x PMID:17517041

Fourcade, S., López-Erauskin, J., Galino, J., Duval, C., Naudi, A., Jove, M., ... Pujol, A. (2008). Early oxidative damage underlying neurodegeneration in X-adrenoleukodystrophy. *Human Molecular Genetics*, *17*(12), 1762–1773. doi:10.1093/hmg/ddn085 PMID:18344354

Fox, M. W., Ahlskog, J. E., & Kelly, P. J. (1991). Stereotactic ventrolateralis thalamotomy for medically refractory tremor in post-levodopa era Parkinson's disease patients. *Journal of Neurosurgery*, 75(5), 723–730. doi:10.3171/jns.1991.75.5.0723 PMID:1919694

Francis, P. T. (2005). The interplay of neurotransmitters in Alzheimer's disease. *CNS Spectrums*, *10*(11Suppl 18), 6–9. doi:10.1017/S1092852900014164 PMID:16273023

Fratiglioni, L., Launer, L., Andersen, K., Breteler, M., Copeland, J., Dartigues, J., ... Hofman, A. (2000). Incidence of dementia and major subtypes in Europe: A collaborative study of population-based cohorts. Neurologic Diseases in the Elderly Research Group. *Neurology*, *54*(11Suppl 5), S10–S15. PMID:10854355

Frederickson, C. J. (1989). Neurobiology of zinc and zinc-containing neurons. *International Review of Neurobiology*, *31*, 145–238. doi:10.1016/S0074-7742(08)60279-2 PMID:2689380

Frederickson, C. J., & Bush, A. I.Dilina do. (2001). Synaptically released zinc: Physiological functions and pathological effects. *Biometals*, *14*(3/4), 353–366. doi:10.1023/A:1012934207456 PMID:11831465

Frederickson, C. J., Giblin, L. J., Balaji, R. V., Masalha, R., Frederickson, C. J., Zeng, Y., ... Kreze, A. (2006). Synaptic release of zinc from brain slices: Factors governing release, imaging, and accurate calculation of concentration. *Journal of Neuroscience Methods*, *154*(1-2), 19–29. doi:10.1016/j.jneumeth.2005.11.014 PMID:16460810

Frederickson, C. J., Suh, S. W., Silva, D., Frederickson, C. J., & Thompson, R. B. (2000). Importance of zinc in the central nervous system: The zinc- containing neuron. *The Journal of Nutrition*, *130*(5), 1471S–1483S. doi:10.1093/jn/130.5.1471S PMID:10801962

Freed, C. R., Greene, P. E., Breeze, R. E., Tsai, W. Y., DuMouchel, W., Kao, R., ... Fahn, S. (2001). Transplantation of embryonic dopamine neurons for severe Parkinson's disease. *The New England Journal of Medicine*, *344*(10), 710–719. doi:10.1056/NEJM200103083441002 PMID:11236774

Freibaum, B. D., Lu, Y., Lopez-Gonzalez, R., Kim, N. C., Almeida, S., Lee, K. H., ... Wong, P. C. (2015). GGGGGCC repeat expansion in C9orf72 compromises nucleocytoplasmic transport. *Nature*, *525*(7567), 129–133. doi:10.1038/ nature14974 PMID:26308899

Freixes, M., Rodríguez, A., Dalfó, E., & Ferrer, I. (2006). Oxidation, glycoxidation, lipoxidation, nitration, and responses to oxidative stress in the cerebral cortex in Creutzfeldt–Jakob disease. *Neurobiology of Aging*, 27(12), 1807–1815. doi:10.1016/j.neurobiolaging.2005.10.006 PMID:16310893

Fridman, O. (1999). Hyperhomocysteinemia: atherothrombosis and neurotoxicity. *Acta physiologica, pharmacologica et therapeutica latinoamericana: órgano de la Asociación Latinoamericana de Ciencias Fisiológicas y [de] la Asociación Latinoamericana de Farmacología, 49*(1), 21-30.

Friedman-Levi, Y., Meiner, Z., Canello, T., Frid, K., Kovacs, G. G., Budka, H., & Gabizon, R. (2011). Fatal Prion Disease in a Mouse Model of Genetic E200K Creutzfeldt-Jakob Disease. *PLoS Pathogens*, 7(11), e1002350. doi:10.1371/journal.ppat.1002350 PMID:22072968

Frigerio, R., Fujishiro, H., Ahn, T.-B., Josephs, K. A., Maraganore, D. M., DelleDonne, A., ... Ahlskog, E. J. (2011). Incidental Lewy Body Disease: Do some cases represent a preclinical stage of Dementia with Lewy Bodies? *Neurobiology of Aging*, *32*(5), 857–863. doi:10.1016/j.neurobiolaging.2009.05.019 PMID:19560232

Frisoni, G. B., Fox, N. C., Jack, C. R., Scheltens, P., & Thompson, P. M. (2010). The clinical use of structural MRI in Alzheimer disease. *Nature Reviews. Neurology*, *6*(2), 67–77. doi:10.1038/nrneurol.2009.215 PMID:20139996

Frucht, S. J. (2013). The definition of dystonia: Current concepts and controversies. *Movement Disorders*, 28(7), 884–888. doi:10.1002/mds.25529 PMID:23893444

Fuchs, R. A., Eaddy, J. L., Su, Z. I., & Bell, G. H. (2007). Interactions of the basolateral amygdala with the dorsal hippocampus and dorsomedial prefrontal cortex regulate drug context-induced reinstatement of cocaine-seeking in rats. *The European Journal of Neuroscience*, 26(2), 487–498. doi:10.1111/j.1460-9568.2007.05674.x PMID:17650119

Fuentes, R., Petersson, P., & Nicolelis, M. A. (2010). Restoration of locomotive function in Parkinson's disease by spinal cord stimulation: Mechanistic approach. *The European Journal of Neuroscience*, *32*(7), 1100–1108. doi:10.1111/j.1460-9568.2010.07417.x PMID:21039949

Fujita, Y., Mizuno, Y., Takatama, M., & Okamoto, K. (2008). Anterior horn cells with abnormal TDP-43 immuno reactivities show fragmentation of the Golgi apparatus in ALS. *Journal of the Neurological Sciences*, 269(1), 30–34. doi:10.1016/j.jns.2007.12.016 PMID:18206910

Fujita, Y., Okamoto, K., Sakurai, A., Amari, M., Nakazato, Y., & Gonatas, N. K. (1999). Fragmentation of the Golgi apparatus of Betz cells in patients with amyotrophic lateral sclerosis. *Journal of the Neurological Sciences*, *163*(1), 81–85. doi:10.1016/S0022-510X(99)00014-3 PMID:10223416

Fujita, Y., Okamoto, K., Sakurai, A., Gonatas, N. K., & Hirano, A. (2000). Fragmentation of the Golgi apparatus of the anterior horn cells in patients with familial amyotrophic lateral sclerosis with SOD1 mutations and posterior column involvement. *Journal of the Neurological Sciences*, *174*(2), 137–140. doi:10.1016/S0022-510X(00)00265-3 PMID:10727699

Fujiwara, H., Hasegawa, M., Dohmae, N., Kawashima, A., Masliah, E., Goldberg, M. S., ... Iwatsubo, T. (2002). [alpha]-Synuclein is phosphorylated in synucleinopathy lesions. *Nature Cell Biology*, *4*(2), 160–164. doi:10.1038/ncb748 PMID:11813001

Fukaya, C., Sumi, K., Otaka, T., Obuchi, T., Kano, T., Kobayashi, K., ... Katayama, Y. (2010). Nexframe frameless stereotaxy with multitract microrecording: Accuracy evaluated by frame-based stereotactic X-ray. *Stereotactic and Functional Neurosurgery*, *88*(3), 163–168. doi:10.1159/000313868 PMID:20431327

Fukui, H., & Moraes, C. T. (2008). The mitochondrial impairment, oxidative stress and neurodegeneration connection: Reality or just an attractive hypothesis? *Trends in Neurosciences*, *31*(5), 251–256. doi:10.1016/j.tins.2008.02.008 PMID:18403030

Funayama, M., Ohe, K., Amo, T., Furuya, N., Yamaguchi, J., Saiki, S., ... Hattori, N. (2015). CHCHD2 mutations in autosomal dominant late-onset Parkinson's disease: A genome-wide linkage and sequencing study. *Lancet Neurology*, *14*(3), 274–282. doi:10.1016/S1474-4422(14)70266-2 PMID:25662902

Gabbita, S. P., Lovell, M. A., & Markesbery, W. R. (1998). Increased nuclear DNA oxidation in the brain in Alzheimer's disease. *Journal of Neurochemistry*, *71*(5), 2034–2040. doi:10.1046/j.1471-4159.1998.71052034.x PMID:9798928

Gadoth, N. (2011). Subacute Sclerosing Pan-Encephalitis (SSPE) – Past and Present. Pathogenesis of Encephalitis, 135-149.

Gai, W. P., Halliday, G. M., Blumbergs, P. C., Geffen, L. B., & Blessing, W. W. (1991). Substance P-containing neurons in the mesopontine tegmentum are severely affected in Parkinson's disease. *Brain*, *114*(5), 2253–2267. doi:10.1093/brain/114.5.2253 PMID:1718530

Galvin, J. E., & Balasubramaniam, M. (2013). Lewy Body Dementia: The Under-Recognized but Common FOE. *Cerebrum: The Dana Forum on Brain Science*, 2013, 13. PMID:24772233

Galvin, J. E., Duda, J. E., Kaufer, D. I., Lippa, C. F., Taylor, A., & Zarit, S. H. (2010). Lewy Body Dementia: Caregiver burden and unmet needs. *Alzheimer Disease and Associated Disorders*, 24(2), 177–181. doi:10.1097/WAD.0b013e3181c72b5d PMID:20505434

Galvin, J. E., Duda, J. E., Kaufer, D. I., Lippa, C. F., Taylor, A., & Zarit, S. H. (2010). Lewy body dementia: The caregiver experience of clinical care. *Parkinsonism & Related Disorders*, *16*(6), 388–392. doi:10.1016/j.parkreldis.2010.03.007 PMID:20434939

Gambetti, P., Mellman, W. J., & Gonatas, N. K. (1969). Familial spongy degeneration of the central nervous system(vanBogaert-Bertrant disease). *Acta Neuropathologica*, *12*(2), 103–115. doi:10.1007/BF00692500 PMID:5789730

Games, D., Valera, E., Spencer, B., Rockenstein, E., Mante, M., Adame, A., ... Masliah, E. (2014). Reducing C-terminaltruncated alpha-synuclein by immunotherapy attenuates neurodegeneration and propagation in Parkinson's disease-like models. *The Journal of Neuroscience*, *34*(28), 9441–9454. doi:10.1523/JNEUROSCI.5314-13.2014 PMID:25009275

Gandhi, P. N., Chen, S. G., & Wilson Delfosse, A. L. (2009). Leucine rich repeat kinase 2 (LRRK2): A key player in the pathogenesis of Parkinson's disease. *Journal of Neuroscience Research*, 87(6), 1283–1295. doi:10.1002/jnr.21949 PMID:19025767

Ganguly, P., Alam, S.F. (2015) Role of homocysteine in the development of cardiovascular disease. Nutrition Journal, 10.

Ganguly, G., Chakrabarti, S., Chatterjee, U., & Saso, L. (2017). Proteinopathy, oxidative stress and mitochondrial dysfunction: Cross talk in Alzheimer's disease and Parkinson's disease. *Drug Design, Development and Therapy*, *11*, 797–810. doi:10.2147/DDDT.S130514 PMID:28352155

Gant, J. C., Sama, M. M., Landfield, P. W., & Thibault, O. (2006). Early and simultaneous emergence of multiple hippocampal biomarkers of aging is mediated by Ca2+-induced Ca2+ release. *The Journal of Neuroscience*, *26*(13), 3482–3490. doi:10.1523/JNEUROSCI.4171-05.2006 PMID:16571755

Gao, H. L., Zheng, W., Xin, N., Chi, Z.-H., Wang, Z.-Y., Chen, J., & Wang, Z.-Y. (2009). Zinc deficiency reduces neurogenesis accompanied by neuronal apoptosis through caspase-dependent and- independent signaling pathways. *Neurotoxicity Research*, *16*(4), 416–425. doi:10.100712640-009-9072-7 PMID:19548052

García-Escudero, V., Martín-Maestro, P., Perry, G., & Avila, J. (2013). Deconstructing mitochondrial dysfunction in Alzheimer disease. *Oxidative Medicine and Cellular Longevity*. PMID:23840916

Garcia, G., Nanni, S., Figueira, I., Ivanov, I., McDougall, G. J., Stewart, D., ... Santos, C. N. (2017). Bioaccessible (poly) phenol metabolites from raspberry protect neural cells from oxidative stress and attenuate microglia activation. *Food Chemistry*, *215*, 274–283. doi:10.1016/j.foodchem.2016.07.128 PMID:27542476

Garcia-Perez, C., Roy, S. S., Naghdi, S., Lin, X., Davies, E., & Hajnóczky, G. (2012). Bid-induced mitochondrial membrane permeabilization waves propagated by local reactive oxygen species (ROS) signaling. *Proceedings of the National Academy of Sciences of the United States of America*, 109(12), 4497–4502. doi:10.1073/pnas.1118244109 PMID:22393005

Garcia-Ruiz, P. J., Chaudhuri, K. R., & Martinez-Martin, P. (2014). Non-motor symptoms of Parkinson's disease A review... from the past. *Journal of the Neurological Sciences*, *338*(1), 30–33. doi:10.1016/j.jns.2014.01.002 PMID:24433931

Gardet, A., Benita, Y., Li, C., Sands, B. E., Ballester, I., Stevens, C., ... Podolsky, D. K. (2010). LRRK2 is involved in the IFN-γ response and host response to pathogens. *Journal of Immunology (Baltimore, Md.: 1950)*, *185*(9), 5577–5585. doi:10.4049/jimmunol.1000548 PMID:20921534

Gardini, S., Concari, L., Pagliara, S., Ghetti, C., Venneri, A., & Caffarra, P. (2011). Visuo-spatial imagery impairment in posterior cortical atrophy: A cognitive and SPECT study. *Behavioural Neurology*, *24*(2), 123–132. doi:10.1155/2011/547451 PMID:21606573

Garg, R. (2002). Subacute sclerosing panencephalitis. *Postgraduate Medical Journal*, 78(916), 63–70. doi:10.1136/ pmj.78.916.63 PMID:11807185

Garske, T., & Ghani, A. C. (2010). Uncertainty in the tail of the variant Creutzfeldt-Jakob disease epidemic in the UK. *PLoS One*, *5*(12), e15626. doi:10.1371/journal.pone.0015626 PMID:21203419

Gauczynski, S., Nikles, D., El-Gogo, S., Papy-Garcia, D., Rey, C., Alban, S., & Weiss, S. (2006). The 37-kDa/67-kDa laminin receptor acts as a receptor for infectious prions and is inhibited by polysulfated glycanes. *The Journal of Infectious Diseases*, *194*(5), 702–709. doi:10.1086/505914 PMID:16897671

Gelb, D. J., Oliver, E., & Gilman, S. (1999). Diagnostic criteria for Parkinson disease. *Archives of Neurology*, 56(1), 33–39. doi:10.1001/archneur.56.1.33 PMID:9923759

Gentry, M. S., Worby, C. A., & Dixon, J. E. (2005). Insights into Lafora disease: Malin is an E3 ubiquitin ligase that ubiquitinates and promotes the degradation of laforin. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(24), 8501–8506. doi:10.1073/pnas.0503285102 PMID:15930137

Geraghty, M. E., Bates-Wall, J., Ratliff-Schaub, K., & Lane, A. E. (2010). Nutritional interventions and therapies in autism: a spectrum of what we know: part 2. *Infant, Child & Adolescent Nutrition*, 2(2), 120–133. doi:10.1177/1941406410366848

Gerdes, M., Solot, C., Wand, P. P., Moss, E., LaRossa, D., & Randall, P. (1999). Cognitive and behavior profile of pre-school children with chromosome 22q11.2 deletion. *American Journal of Medical Genetics*, 85(2), 127–133. doi: PMID:10406665

Gerfen, C. R., & Surmeier, D. J. (2011). Modulation of striatal projection systems by dopamine. *Annual Review of Neuroscience*, *34*(1), 441–466. doi:10.1146/annurev-neuro-061010-113641 PMID:21469956

Gerlach, M., Ben-Shachar, D., Riederer, P., & Youdim, M. B. (1994). Altered brain metabolism of iron as a cause of neurodegenerative diseases? *Journal of Neurochemistry*, *63*(3), 793–807. doi:10.1046/j.1471-4159.1994.63030793.x PMID:7519659

Germiniani, F. M., Miranda, A. P., Ferenczy, P., Munho, R. P., & Teive, H. A. (2012, July). Tourette's syndrome: From demonic possession and psychoanalysis to the discovery of gene. *Arquivos de Neuro-Psiquiatria*, 70(7), 547–549. doi:10.1590/S0004-282X2012000700014 PMID:22836463

Gerschman, R., Gilbert, D. L., Nye, S. W., Dwyer, P., & Fenn, W. O. (1954). Oxygen poisoning and x-irradiation: A mechanism in common. *Science*, *119*(3097), 623–626. doi:10.1126cience.119.3097.623 PMID:13156638

Geschwind, M. D., Kuo, A. L., Wong, K. S., Haman, A., Devereux, G., Raudabaugh, B. J., & Thai, J. N. (2013). Quinacrine treatment trial for sporadic Creutzfeldt-Jakob disease. *Neurology*, *81*(23), 2015–2023. doi:10.1212/WNL.0b013e3182a9f3b4 PMID:24122181

Ghadge, G. D., Lee, J. P., Bindokas, V. P., Jordan, J., Ma, L., Miller, R. J., & Roos, R. P. (1997). Mutant Superoxide Dismutase-1-Linked Familial Amyotrophic Lateral Sclerosis: Molecular Mechanisms of Neuronal Death and Protection. *The Journal of Neuroscience*, *17*(22), 8756–8766. PMID:9348345

Ghavami, S., Shojaei, S., Yeganeh, B., Ande, S. R., Jangamreddy, J. R., Mehrpour, M., ... Kashani, H. H. (2014). Autophagy and apoptosis dysfunction in neurodegenerative disorders. *Progress in Neurobiology*, *112*, 24–49. doi:10.1016/j. pneurobio.2013.10.004 PMID:24211851

Ghaziuddin, M., Ghaziuddin, N., & Greden, J. (2002). Depression in persons with autism: Implications for research and clinical care. *Journal of Autism and Developmental Disorders*, *32*(4), 299–306. doi:10.1023/A:1016330802348 PMID:12199134

Ghezzi, D., & Zeviani, M. (2012). Assembly factors of human mitochondrial respiratory chain complexes: physiology and pathophysiology. In *Mitochondrial Oxidative Phosphorylation* (pp. 65–106). Springer New York. doi:10.1007/978-1-4614-3573-0_4

Giacomello, M., Drago, I., Pizzo, P., & Pozzan, T. (2007). Mitochondrial Ca2+ as a key regulator of cell life and death. *Cell Death and Differentiation*, *14*(7), 1267–1274. doi:10.1038j.cdd.4402147 PMID:17431419

Gianaros, P. J., Jennings, J. R., Sheu, L. K., Greer, P. J., Kuller, L. H., & Matthews, K. A. (2007). Prospective reports of chronic life stress predict decreased grey matter volume in the hippocampus. *NeuroImage*, *35*(2), 795–803. doi:10.1016/j. neuroimage.2006.10.045 PMID:17275340

Giasson, B. I., Duda, J. E., Murray, I. V., Chen, Q., Souza, J. M., Hurtig, H. I., ... Lee, V. M. Y. (2000). Oxidative damage linked to neurodegeneration by selective α-synuclein nitration in synucleinopathy lesions. *Science*, *290*(5493), 985–989. doi:10.1126cience.290.5493.985 PMID:11062131

Gijselinck, I., Van Langenhove, T., van der Zee, J., Sleegers, K., Philtjens, S., Kleinberger, G., ... Van Broeckhoven, C. (2012). A C9orf72 promoter repeat expansion in a Flanders–Belgian cohort disorders of the frontotemporal lobar degeneration- amyotrophic lateral sclerosis spectrum: A gene identification study. *Lancet Neurology*, *11*(1), 54–65. doi:10.1016/S1474-4422(11)70261-7 PMID:22154785

Giladi, N., Boroojerdi, B., Korczyn, A. D., Burn, D. J., Clarke, C. E., & Schapira, A. H. (2007). Rotigotine transdermal patch in early Parkinson's disease: A randomized, double-blind, controlled study versus placebo and ropinirole. *Movement Disorders*, 22(16), 2398–2404. doi:10.1002/mds.21741 PMID:17935234

Gillberg, C., Ehlers, S., Schaumann, H., Jakobsson, G., Dahlgren, S. O., Lindblom, R., ... Blidner, E. (1990). Autism under age 4 years: A clinical study of 28 cases referred for autistic symptoms in infancy. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, *31*(6), 921–934. doi:10.1111/j.1469-7610.1990.tb00834.x PMID:2246342

Gillham, J. E., Carter, A. S., Volkmar, F. R., & Sparrow, S. S. (2000). Toward a developmental operational definition of autism. *Journal of Autism and Developmental Disorders*, *30*(4), 269–278. doi:10.1023/A:1005571115268 PMID:11039854

Gilman, S., Wenning, G. K., Low, P. A., Brooks, D. J., Mathias, C. J., Trojanowski, J. Q., ... Vidailhet, M. (2008). Second consensus statement on the diagnosis of multiple system atrophy. *Neurology*, *71*(9), 670–676. doi:10.1212/01. wnl.0000324625.00404.15 PMID:18725592

Giorgi, C., De Stefani, D., Bononi, A., Rizzuto, R., & Pinton, P. (2009). Structural and functional link between the mitochondrial network and the endoplasmic reticulum. *The International Journal of Biochemistry & Cell Biology*, *41*(10), 1817–1827. doi:10.1016/j.biocel.2009.04.010 PMID:19389485

Giri, M., Zhang, M., & Lü, Y. (2016). Genes associated with Alzheimer's disease: An overview and current status. *Clinical Interventions in Aging*, *11*, 665. doi:10.2147/CIA.S105769 PMID:27274215

Gitcho, M. A., Baloh, R. H., Chakraverty, S., Mayo, K., Norton, J. B., Levitch, D., ... Cairns, N. J. (2008). TDP-43 A315T mutation in familial motor neuron disease. *Annals of Neurology*, 63(4), 535–538. doi:10.1002/ana.21344 PMID:18288693

Glauser, L., Sonnay, S., Stafa, K., & Moore, D. J. (2011). Parkin promotes the ubiquitination and degradation of the mitochondrial fusion factor mitofusin 1. *Journal of Neurochemistry*, *118*(4), 636–645. doi:10.1111/j.1471-4159.2011.07318.x PMID:21615408

Glenner, G. G., & Wong, C. W. (1984a). Alzheimer's disease: Initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Biochemical and Biophysical Research Communications*, *120*(3), 885–890. doi:10.1016/S0006-291X(84)80190-4 PMID:6375662

Glenner, G. G., & Wong, C. W. (1984b). Alzheimer's disease and Down's syndrome: Sharing of a unique cerebrovascular amyloid fibril protein. *Biochemical and Biophysical Research Communications*, *122*(3), 1131–1135. doi:10.1016/0006-291X(84)91209-9 PMID:6236805

Glinka, Y., Gassen, M., & Youdim, M. B. H. (1997). Mechanism of 6-hydroxydopamine neurotoxicity. In *Advances in Research on Neurodegeneration* (pp. 55–66). Vienna: Springer. doi:10.1007/978-3-7091-6842-4_7

Glorioso, C., Oh, S., Douillard, G. G., & Sibille, E. (2011). Brain molecular aging, promotion of neurological disease and modulation by Sirtuin5 longevity gene polymorphism. *Neurobiology of Disease*, *41*(2), 279–290. doi:10.1016/j. nbd.2010.09.016 PMID:20887790

Godefroy, O., Leclercq, C., & Roussel, M. (2013). Vascular cognitive impairment in the stroke unit and after the acute stage. In O. Godefroy (Ed.), *Behavioral and cognitive neurology of stroke* (2nd ed.; pp. 22–31). Cambridge University press. doi:10.1017/CBO9781139058988.004

Goedert, M. (2004). Tau protein and neurodegeneration. *Seminars in Cell & Developmental Biology*, *15*(1), 45–49. doi:10.1016/j.semcdb.2003.12.015 PMID:15036206

Goedert, M., Crowther, R. A., & Spillantini, M. G. (1998). Tau mutations cause frontotemporal dementias. *Neuron*, 21(5), 955–958. doi:10.1016/S0896-6273(00)80615-7 PMID:9856453

Goedert, M., Jakes, R., & Crowther, R. A. (1999). Effects of frontotemporal dementia FTDP-17 mutations on heparininduced assembly of tau filaments. *FEBS Letters*, *450*(3), 306–311. doi:10.1016/S0014-5793(99)00508-6 PMID:10359094

Goedert, M., Klug, A., & Crowther, R. A. (2006). Tau protein, the paired helical filament and Alzheimer disease. *Journal of Alzheimer's Disease*, *9*(s3), 195–207. doi:10.3233/JAD-2006-9S323 PMID:16914859

Goetz, C. G., Emre, M., & Dubois, B. (2008). Parkinson's disease dementia: Definitions, guidelines, and research perspectives in diagnosis. *Annals of Neurology*, 64(S2), S81–S92. doi:10.1002/ana.21455 PMID:19127578

Goetz, C. G., Tilley, B. C., Shaftman, S. R., Stebbins, G. T., Fahn, S., Martinez-Martin, P., ... LaPelle, N. (2008). Movement Disorder Society-Sponsored Revision of the Unified Parkinson's Disease Rating Scale (MDS-UPDRS): Scale presentation and clinimetric testing results. *Movement Disorders*, 23(15), 2129–2170. doi:10.1002/mds.22340 PMID:19025984

Goker-Alpan, O., Giasson, B. I., Eblan, M. J., Nguyen, J., Hurtig, H. I., Lee, V. M., ... Sidransky, E. (2006). Glucocerebrosidase mutations are an important risk factor for Lewy body disorders. *Neurology*, 67(5), 908–910. doi:10.1212/01. wnl.0000230215.41296.18 PMID:16790605

Goldenberg, M. M. (2008). Medical Management of Parkinson's Disease. P&T, 33(10), 590-606. PMID:19750042

Goldman, J. G., Goetz, C. G., Brandabur, M., Sanfilippo, M., & Stebbins, G. T. (2008). Effects of dopaminergic medications on psychosis and motor function in dementia with Lewy bodies. *Movement Disorders*, 23(15), 2248–2250. doi:10.1002/mds.22322 PMID:18823039

Goldman, J. G., & Weintraub, D. (2015). Advances in the treatment of cognitive impairment in Parkinson's disease. *Movement Disorders*, *30*(11), 1471–1489. doi:10.1002/mds.26352 PMID:26297863

Goldman, J. G., Williams-Gray, C., Barker, R. A., Duda, J. E., & Galvin, J. E. (2014). The spectrum of cognitive impairment in Lewy body diseases. *Movement Disorders*, 29(5), 608–621. doi:10.1002/mds.25866 PMID:24757110

Goldstein, H. (2002). Communication intervention for children with autism: A review of treatment efficacy. *Journal of Autism and Developmental Disorders*, *32*(5), 373–396. doi:10.1023/A:1020589821992 PMID:12463516

Golikov, N. (1985). Adaptation in Farm Animals. Sofia, Bulgaria: Agropromizdat.

Gomberg, M. (1900). An instance of trivalent carbon: Triphenylmethyl. *Journal of the American Chemical Society*, 22(11), 757–771. doi:10.1021/ja02049a006

Gomez-Ospina, N. (2006 May 30). Arylsulfatase A Deficiency. In M. P. Adam, H. H. Ardinger, & R. A. Pagon (Eds.), *GeneReviews*® (pp. 1993–2017). Seattle, WA: University of Washington, Seattle. Available from https://www.ncbi.nlm. nih.gov/books/NBK1130/

Gómez-Tortosa, E., Newell, K., Irizarry, M. C., Sanders, J. L., & Hyman, B. T. (2000). α-Synuclein immunoreactivity in dementia with Lewy bodies: Morphological staging and comparison with ubiquitin immunostaining. *Acta Neuropathologica*, *99*(4), 352–357. doi:10.1007004010051135 PMID:10787032

Gomori, A. J., Partnow, M. J., Horoupian, D. S., & Hirano, A. (1973). The ataxic form of Creutzfeldt-Jakob disease. *Archives of Neurology*, 29(5), 318–323. doi:10.1001/archneur.1973.00490290058006 PMID:4582819

Gomperts, S. N. (2016). Lewy Body Dementias: Dementia With Lewy Bodies and Parkinson Disease Dementia. *Continuum: Lifelong Learning in Neurology*, 22(2), 435-463.

Gomperts, S. N. (2016). Lewy Body Dementias: Dementia With Lewy Bodies and Parkinson Disease Dementia. *Continuum (Minneapolis, Minn.)*, 22(2), 435–463. doi:10.1212/CON.000000000000309 PMID:27042903

Gomperts, S. N., Locascio, J. J., Marquie, M., Santarlasci, A. L., Rentz, D. M., Maye, J., ... Growdon, J. H. (2012). Brain amyloid and cognition in Lewy body diseases. *Movement Disorders*, 27(8), 965–973. doi:10.1002/mds.25048 PMID:22693110

Gonatas, J. O., Mezitis, S. G., Stieber, A., Fleischer, B., & Gonatas, N. K. (1989). MG-160. A novel sialoglycoprotein of the medial cisternae of the Golgi apparatus. *The Journal of Biological Chemistry*, 264(1), 646–653. PMID:2909545

Gonatas, N. K., Stieber, A., Mourelatos, Z., Chen, Y., Gonatas, J. O., Appel, S. H., ... Hauw, J. J. (1992). Fragmentation of the Golgi apparatus of motor neurons in amyotrophic lateral sclerosis. *American Journal of Pathology*, *140*(3), 731. PMID:1546747

Goñi, J., Esteban, F. J., de Mendizábal, N. V., Sepulcre, J., Ardanza-Trevijano, S., Agirrezabal, I., & Villoslada, P. (2008). A computational analysis of protein-protein interaction networks in neurodegenerative diseases. *BMC Systems Biology*, 2(1), 52. doi:10.1186/1752-0509-2-52 PMID:18570646

Gonzalez-Cuyar, L. F., Sonnen, J. A., Montine, K. S., Keene, C. D., & Montine, T. J. (2011). Role of Cerebrospinal Fluid and Plasma Biomarkers in the Diagnosis of Neurodegenerative Disorders and Mild Cognitive Impairment. *Current Neurology and Neuroscience Reports*, *11*(5), 455–463. doi:10.100711910-011-0212-0 PMID:21725901

Gorelick, P. B., Scuteri, A., Black, S. E., Decarli, C., Greenberg, S. M., Iadecola, C., ... Seshadri, S. (2011). Vascular contributions to cognitive impairment and dementia: A statement for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke*, *42*(9), 2672–2713. doi:10.1161/STR.0b013e3182299496 PMID:21778438

Gorlin, B. J., McAlpine, C.P, Garwick, A, Wieling, E. (2016). Severe Childhood Autism: The Family Lived Experience *J Pediatr Nurs.*, *31*(6), 580-597. doi: . 2016.09.00210.1016/j.pedn

Gorman, A. M. (2008). Neuronal cell death in neurodegenerative diseases: Recurring themes around protein handling. *Journal of Cellular and Molecular Medicine*, *12*(6a), 2263–2280. doi:10.1111/j.1582-4934.2008.00402.x PMID:18624755

Gosal, D., Ross, O. A., & Toft, M. (2006). Parkinson's disease: The genetics of a heterogeneous disorder. *European Journal of Neurology*, *13*(6), 616–627. doi:10.1111/j.1468-1331.2006.01336.x PMID:16796586

Gotthardt, K., Weyand, M., Kortholt, A., Van Haastert, P. J., & Wittinghofer, A. (2008). Structure of the Roc–COR domain tandem of C. tepidum, a prokaryotic homologue of the human LRRK2 Parkinson kinase. *The EMBO Journal*, 27(16), 2239–2249. doi:10.1038/emboj.2008.150 PMID:18650931

Gotz, J., Chen, F., van Dorpe, J., & Nitsch, R. M. (2001). Formation of Neurofibrillary Tangles in P301L Tau Transgenic Mice Induced by Abeta 42 Fibrils. *Science*, 293(5534), 1491–1495. doi:10.1126cience.1062097 PMID:11520988

Gouarné, C., Tracz, J., Paoli, M. G., Deluca, V., Seimandi, M., Tardif, G., ... Pruss, R. M. (2015). Protective role of olesoxime against wild-type α-synuclein-induced toxicity in human neuronally differentiated SHSY-5Y cells. *British Journal of Pharmacology*, *172*(1), 235–245. doi:10.1111/bph.12939 PMID:25220617

Gout, J.-F., Kahn, D., Duret, L., & Paramecium Post-Genomics, C. (2010). The Relationship among Gene Expression, the Evolution of Gene Dosage, and the Rate of Protein Evolution. *PLOS Genetics*, *6*(5), e1000944. doi:10.1371/journal. pgen.1000944 PMID:20485561

Gouveia, L. O., & De Carvalho, M. (2007). Young-onset sporadic amyotrophic lateral sclerosis: A distinct nosological entity? *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases*, 1–5. PMID:17852021

Gouw, A. A., Seewann, A., van der Flier, W. M., Barkhof, F., Rozemuller, A. M., Scheltens, P., & Geurts, J. J. (2011). Heterogeneity of small vessel disease: A systematic review of MRI and histopathology correlations. *Journal of Neurology, Neurosurgery, and Psychiatry*, 82(2), 126–135. doi:10.1136/jnnp.2009.204685 PMID:20935330

Gowers, W. R. (1898). A manual of diseases of the nervous system (Vol. 2). P. Blakiston, Son & Company.

Grabowski, G. A. (2008). Phenotype, diagnosis, and treatment of Gaucher's disease. *Lancet*, 372(9645), 1263–1271. doi:10.1016/S0140-6736(08)61522-6 PMID:19094956

Graham, J. G., & Oppenheimer, D. R. (1969). Orthostatic hypotension and nicotine sensitivity in a case of multiple system atrophy. *Journal of Neurology, Neurosurgery, and Psychiatry*, 32(1), 28–34. doi:10.1136/jnnp.32.1.28 PMID:5774131

Granata, T. (2012). Metabolic and degenerative disorders. *Handbook of Clinical Neurology*, *108*, 485–511. doi:10.1016/B978-0-444-52899-5.00045-9 PMID:22939050

Granot, E., & Kohen, R. (2004). Oxidative stress in childhood – in health and disease states. *Clinical Nutrition (Edinburgh, Lothian)*, 23(1), 3–11. doi:10.1016/S0261-5614(03)00097-9 PMID:14757387

Grasso, G., Sfacteria, A., Meli, F., Passalacqua, M., Fodale, V., Buemi, M., ... Tomasello, F. (2007). The role of erythropoietin in neuroprotection: Therapeutic perspectives. *Drug News & Perspectives*, 20(5), 315–320. doi:10.1358/ dnp.2007.20.5.1120219 PMID:17878959

Graziano, A. C. E., Pannuzzo, G., Avola, R., & Cardile, V. (2016, November 1). Chaperones as potential therapeutics for Krabbe disease. *Journal of Neuroscience Research*, *94*(11), 1220–1230. doi:10.1002/jnr.23755 PMID:27638605

Greenamyre, J. T., & Hastings, T. G. (2004). Parkinson's--divergent causes, convergent mechanisms. *Science*, *304*(5674), 1120–1122. doi:10.1126cience.1098966 PMID:15155938

Greenberg, S. M., & Vonsattel, J. P. (1997). Diagnosis of cerebral amyloid angiopathy. Sensitivity and specificity of cortical biopsy. *Stroke*, *28*(7), 1418–1422. doi:10.1161/01.STR.28.7.1418 PMID:9227694

Green, D. R., & Kroemer, G. (2004). The pathophysiology of mitochondrial cell death. *Science*, *305*(5684), 626–629. doi:10.1126cience.1099320 PMID:15286356

Greenfield, J. G., & Bosanquet, F. D. (1953). The brain-stem lesions in Parkinsonism. *Journal of Neurology, Neurosurgery, and Psychiatry*, *16*(4), 213–226. doi:10.1136/jnnp.16.4.213 PMID:13109537

Greenwood, P. M., & Parasuraman, R. (2010). Neuronal and cognitive plasticity: A neurocognitive framework for ameliorating cognitive aging. *Frontiers in Aging Neuroscience*, 2. PMID:21151819

Greggio, E., Jain, S., Kingsbury, A., Bandopadhyay, R., Lewis, P., Kaganovich, A., & Ahmad, R. (2006). Kinase activity is required for the toxic effects of mutant LRRK2/dardarin. *Neurobiology of Disease*, *23*(2), 329–341. doi:10.1016/j. nbd.2006.04.001 PMID:16750377

Greggio, E., Taymans, J. M., Zhen, E. Y., Ryder, J., Vancraenenbroeck, R., Beilina, A., ... Cookson, M. R. (2009). The Parkinson's disease kinase LRRK2 autophosphorylates its GTPase domain at multiple sites. *Biochemical and Biophysical Research Communications*, *389*(3), 449–454. doi:10.1016/j.bbrc.2009.08.163 PMID:19733152

Greggio, E., Zambrano, I., Kaganovich, A., Beilina, A., Taymans, J. M., Daniëls, V., ... Thomas, K. J. (2008). The Parkinson disease-associated leucine-rich repeat kinase 2 (LRRK2) is a dimer that undergoes intramolecular autophosphorylation. *The Journal of Biological Chemistry*, 283(24), 16906–16914. doi:10.1074/jbc.M708718200 PMID:18397888

Greicius, M. D., Krasnow, B., Reiss, A. L., & Menon, V. (2003). Functional connectivity in the resting brain: A network analysis of the default mode hypothesis. *Proceedings of the National Academy of Sciences of the United States of America*, 100(1), 253–258. doi:10.1073/pnas.0135058100 PMID:12506194

Grierson, A. J., & Miller, C. (2006). Axonal transport and amyotrophic lateral sclerosis. In Amyotrophic Lateral Sclerosis (2nd ed.). Informa Healthcare Journal.

Griess, B., Tom, E., Domann, F., & Teoh-Fitzgerald, M. (2017). Extracellular superoxide dismutase and its role in cancer. *Free Radical Biology & Medicine*, *112*, 464–479. doi:10.1016/j.freeradbiomed.2017.08.013 PMID:28842347

Griffith, J. S. (1967). Self-replication and scrapie. Nature, 215(5105), 1043–1044. doi:10.1038/2151043a0 PMID:4964084

Grilli, M., Goffi, F., Memo, M., & Spano, P. F. (1996). Interleukin-1beta and glutamate activate the NF kappa B/ Rel binding site from the regulatory region of the amyloid precursor protein gene in primary neuronal cultures. *The Journal of Biological Chemistry*, 271(25), 15002–15007. doi:10.1074/jbc.271.25.15002 PMID:8663145

Groeneveld, G. J., Veldink, J. H., van der Tweel, I., Kalmijn, S., Beijer, C., de Visser, M., ... van den Berg, L. H. (2003). A randomized sequential trial of creatine in amyotrophic lateral sclerosis. *Annals of Neurology*, *53*(4), 437–445. doi:10.1002/ ana.10554 PMID:12666111

Groeschel, S., Kehrer, C., Engel, C., & Dali, Í. (2011). Metachromatic leukodystrophy: Natural course of cerebral MRI changes in relation to clinical course. *Journal of Inherited Metabolic Disease*, *34*(5), 1095–1102. doi:10.100710545-011-9361-1 PMID:21698385

Gros-Louis, F., Gaspar, C., & Rouleau, G. A. (2006). Genetics of familial and sporadic amyotrophic lateral sclerosis. *Biochimica et Biophysica Acta (BBA)- Molecular Basis of Disease*, *1762*(11), 956–972. doi:10.1016/j.bbadis.2006.01.004

Grossman, M., Gross, R. G., Moore, P., Dreyfuss, M., McMillan, C. T., Cook, P. A., ... Siderowf, A. (2012). Difficulty processing temporary syntactic ambiguities in Lewy body spectrum disorder. *Brain and Language*, *120*(1), 52–60. doi:10.1016/j.bandl.2011.08.007 PMID:21962945

Guentchev, M., Groschup, M. H., Kordek, R., Liberski, P. P., & Budka, H. (1998). Severe, early and selective loss of a subpopulation of GABAergic inhibitory neurons in experimental transmissible spongiform encephalopathies. *Brain Pathology (Zurich, Switzerland)*, 8(4), 615–623. doi:10.1111/j.1750-3639.1998.tb00188.x PMID:9804371

Guentchev, M., Voigtländer, T., Haberler, C., Groschup, M. H., & Budka, H. (2000). Evidence for oxidative stress in experimental prion disease. *Neurobiology of Disease*, *7*(4), 270–273. doi:10.1006/nbdi.2000.0290 PMID:10964599

Guillozet, A. L., Weintraub, S., Mash, D. C., & Mesulam, M. M. (2003). Neurofibrillary Tangles, Amyloid, and Memory in Aging and Mild Cognitive Impairment. *Archives of Neurology*, *60*(5), 729. doi:10.1001/archneur.60.5.729 PMID:12756137

Gu, M., Gash, M., Mann, V., Javoy-Agid, F., Cooper, J., & Schapira, A. (1996). Mitochondrial defect in Huntington's disease caudate nucleus. *Annals of Neurology*, *39*(3), 385–389. doi:10.1002/ana.410390317 PMID:8602759

Gunter, T. E., & Sheu, S. S. (2009). Characteristics and possible functions of mitochondr ial Ca²⁺ transport mechanisms. *Biochimica et Biophysica Acta*, *1787*(11), 1291–1308. doi:10.1016/j.bbabio.2008.12.011 PMID:19161975

Guo, C., Sun, L., Chen, X., & Zhang, D. (2013). Oxidative stress, mitochondrial damage and neurodegenerative diseases. *Neural Regeneration Research*, 8(21), 2003. PMID:25206509

Guo, J. L., & Lee, V. M. Y. (2014). Cell-to-cell transmission of pathogenic proteins in neurodegenerative diseases. *Nature Medicine*, *20*(2), 130–138. doi:10.1038/nm.3457 PMID:24504409

Guo, W., Chen, Y., Zhou, X., Kar, A., Ray, P., Chen, X., ... Zhu, L. (2011). An ALS-associated mutation affecting TDP-43 enhances protein aggregation, fibril formation and neurotoxicity. *Nature Structural & Molecular Biology*, *18*(7), 822–830. doi:10.1038/nsmb.2053 PMID:21666678

Gurney, M. E., Cutting, F. B., Zhai, P., Doble, A., Taylor, C. P., Andrus, P. K., & Hall, E. D. (1996). Benefit of vitamin E, riluzole, and gabapentin in a transgenic model of familial amyotrophic lateral sclerosis. *Annals of Neurology*, *39*(2), 147–157. doi:10.1002/ana.410390203 PMID:8967745

Gurney, M. E., Pu, H., Chiu, A. Y., Dal Canto, M. C., Polchow, C. Y., & Alexander, D. D., ... Deng, H. X. (1994). Motor neuron degeneration in mice that express a human Cu,Zn superoxide dismutase mutation. *Science*, 264.

Gutman, J. (2002). Glutathione- Your Bodies Most Powerful Protector (3rd ed.). Montreal: Communications Kudoca Inc.

Gwon, J. G., Kwon, T. W., Cho, Y. P., Kang, D. W., Han, Y., & Noh, M. (2017). Analysis of Risk Factors for Cerebral Microinfarcts after Carotid Endarterectomy and the Relevance of Delayed Cerebral Infarction. *Journal of Clinical Neurology (Seoul, Korea)*, *13*(1), 32–37. doi:10.3988/jcn.2017.13.1.32 PMID:27730766

Haase, H., & Rink, L. (2009). Functional significance of zinc- related signaling pathways in immune cells. *Annual Review* of Nutrition, 29(1), 133–152. doi:10.1146/annurev-nutr-080508-141119 PMID:19400701

Habibyar, A. F., Sharma, N., & Khurana, N. (2016). PASS assisted prediction and pharmacological evaluation of hesperidin against scopolamine induced amnesia in mice. *European Journal of Pharmacology*, 789, 385–394. doi:10.1016/j. ejphar.2016.07.013 PMID:27397428

Hachinski, V. C., Lassen, N. A., & Marshall, J. (1974). Multi-infarct dementia. A cause of mental deterioration in the elderly. *Lancet*, 2(7874), 207–210. doi:10.1016/S0140-6736(74)91496-2 PMID:4135618

Hacke, W., Kaste, M., Bluhmki, E., Brozman, M., Dávalos, A., Guidetti, D., ... Toni, D. (2008). Thrombolysis with alteplase 3 to 4.5 hours after acute ischemic stroke. *The New England Journal of Medicine*, *359*(13), 1317–1329. doi:10.1056/ NEJMoa0804656 PMID:18815396

Haeusler, A. R., Donnelly, C. J., Periz, G., Simko, E. A., Shaw, P. G., Kim, M. S., ... Sattler, R. (2014). C9orf72 nucleotide repeat structures initiate molecular cascades of disease. *Nature*, *507*(7491), 195–200. doi:10.1038/nature13124 PMID:24598541

Hajo, H., & Lothar, R. (2009). The immune system and the impact of zinc during aging. *Immunity & Ageing*, 6(1), 9. doi:10.1186/1742-4933-6-9 PMID:19523191

Halasi, M., Váraljai, R., Benevolenskaya, E., & Gartel, A. L. (2016). A Novel Function of Molecular Chaperone HSP70 suppression of oncogenic foxm1 after proteotoxic stress. *The Journal of Biological Chemistry*, 291(1), 142–148. doi:10.1074/jbc.M115.678227 PMID:26559972

Hall, G. (2011). What is the Link Between Protein Aggregation and Interneuronal Lesion Propagation in Neurodegenerative Disease? In *Neurodegenerative Diseases-Processes, Prevention, Protection and Monitoring*. InTech.

Halliday, G., Hely, M., Reid, W., & Morris, J. (2008). The progression of pathology in longitudinally followed patients with Parkinson's disease. *Acta Neuropathologica*, *115*(4), 409–415. doi:10.100700401-008-0344-8 PMID:18231798

Halliwell, B. (1992). Reactive Oxygen Species and the Central Nervous System. *Journal of Neurochemistry*, 59(5), 1609–1623. doi:10.1111/j.1471-4159.1992.tb10990.x PMID:1402908

Halliwell, B. (2007). Biochemistry of oxidative stress. *Biochemistry Society Transanctions*, 35(5), 1147–1150. doi:10.1042/BST0351147 PMID:17956298

Halliwell, B., & Gutteridge, J. M. (1990). Role of free radicals and catalytic metal ions in human disease: An overview. *Methods in Enzymology*, *186*, 1–85. doi:10.1016/0076-6879(90)86093-B PMID:2172697

Hamano, K., Hayashi, M., Shioda, K., Fukatsu, R., & Mizutani, S. (2008). Mechanisms of neurodegeneration in mucopolysaccharidoses II and IIIB: Analysis of human brain tissue. *Acta Neuropathologica*, *115*(5), 547–559. doi:10.100700401-007-0325-3 PMID:18060551 Hambidge, M. (2000). Human zinc deficiency. *The Journal of Nutrition*, *130*(5), 1344S–1349S. doi:10.1093/jn/130.5.1344S PMID:10801941

Hamblet, N. S., Ragland, B., Ali, M., Conyers, B., & Castora, F. J. (2006). Mutations in mitochondrial-encoded cytochrome c oxidase subunits I, II, and III genes detected in Alzheimer's disease using single-strand conformation polymorphism. *Electrophoresis*, *27*(2), 398–408. doi:10.1002/elps.200500420 PMID:16358358

Hammerman, M. (1987). Insulin-like growth factors and aging. *Endocrinology and Metabolism Clinics of North America*, *16*(4), 995–1011. PMID:3322823

Handley, O. J., Naji, J. J., Dunnett, S. B., & Rosser, A. E. (2006). Pharmaceutical, cellular and genetic therapies for Huntington's disease. *Clinical Science*, *110*(1), 73–88. doi:10.1042/CS20050148 PMID:16336206

Haneef, S. S., & Doss, C. G. (2016). Personalized Pharmacoperones for Lysosomal Storage Disorder. *Advances in Protein Chemistry and Structural Biology Personalized Medicine*, *102*, 225–265. doi:10.1016/bs.apcsb.2015.10.001 PMID:26827607

Hanson, E., Kalish, L. A., Bunce, E., Curtis, C., McDaniel, S., Ware, J., & Petry, J. (2007). Use of complementary and alternative medicine among children diagnosed with autism spectrum disorder. *Journal of Autism and Developmental Disorders*, *37*(4), 628–636. doi:10.100710803-006-0192-0 PMID:16977497

Hardiman, O., van den Berg, L. H., & Kiernan, M. C. (2011). Clinical diagnosis and management of amyotrophic lateral sclerosis. *Nature Reviews. Neurology*, 7(11), 639–649. doi:10.1038/nrneurol.2011.153 PMID:21989247

Harding, A. (1984). The hereditary ataxias and related disorders. Edinburgh, UK: Churchill Livingstone.

Harding, A. J., Broe, G. A., & Halliday, G. M. (2002). Visual hallucinations in Lewy body disease relate to Lewy bodies in the temporal lobe. *Brain*, *125*(2), 391–403. doi:10.1093/brain/awf033 PMID:11844739

Hardingham, G. E., & Bading, H. (2010). Synaptic Versus Extrasynaptic NMDA Receptor Signalling: Implications for Neurodegenerative Disorders. *Nature Reviews. Neuroscience*, *11*(10), 682–696. doi:10.1038/nrn2911 PMID:20842175

Hardy, J. (2006). Alzheimer's disease: the amyloid cascade hypothesis: an update and reappraisal. *Journal of Alzheimer's Disease : JAD, 9*(3 Suppl), 151–3. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/16914853

Hardy, J., & Selkoe, D. J. (2002). The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science*, 297(5580), 353-356.

Hardy, J. A., & Higgins, G. A. (1992). Alzheimer's disease: The amyloid cascade hypothesis. *Science*, 256(5054), 184–185. doi:10.1126cience.1566067 PMID:1566067

Hardy, J., & Selkoe, D. J. (2002). The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science*, 297(5580), 353–356. doi:10.1126cience.1072994 PMID:12130773

Hariz, M. I., Krack, P., Alesch, F., Augustinsson, L. E., Bosch, A., Ekberg, R., ... Benabid, A.-L. (2008, June). Multicentre European study of thalamic stimulation for parkinsonian tremor: A 6 year follow-up. *Journal of Neurology, Neurosurgery, and Psychiatry*, *79*(6), 694–699. doi:10.1136/jnnp.2007.118653 PMID:17898034

Harman, D. (1956). Aging: A theory based on free radical and radiation chemistry. *Journal of Gerontology*, *11*(3), 298–300. doi:10.1093/geronj/11.3.298 PMID:13332224

Harman, D. (1972). The biologic clock: The mitochondria? *Journal of the American Geriatrics Society*, 20(4), 145–147. doi:10.1111/j.1532-5415.1972.tb00787.x PMID:5016631

Harman, D. (2006). Free radical theory of aging: An update. *Annals of the New York Academy of Sciences*, *1067*(1), 10–21. doi:10.1196/annals.1354.003 PMID:16803965

Harrington, J. W., Rosen, L., Garnecho, A., & Patrick, P. A. (2006). Parental perceptions and use of complementary and alternative medicine practices for children with autistic spectrum disorders in private practice. *Journal of Developmental and Behavioral Pediatrics*, 27(2Supplement2), S156–S161. doi:10.1097/00004703-200604002-00014 PMID:16685182

Harris, D. (2017). Study unravels mystery of how nerve cells are damaged in neurodegenerative diseases. *Medical and Life Sciences*, 1–2.

Harris, R. A. (2014). Spatial, temporal, and functional aspects of macrophages during "the good, the bad, and the ugly" phases of inflammation. *Frontiers in Immunology*, *5*, 612. doi:10.3389/fimmu.2014.00612 PMID:25520719

Hartl, F. U., Bracher, A., & Hayer-Hartl, M. (2011). Molecular chaperones in protein folding and proteostasis. *Nature*, 475(7356), 324–332. doi:10.1038/nature10317 PMID:21776078

Harwood, C. A., McDermott, C. J., & Shaw, P. J. (2009). Physical activity as an exogenous risk factor in motor neuron disease (MND): A review of the evidence. *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases*, *10*(4), 191–204. doi:10.1080/17482960802549739 PMID:19263258

Hasan, R., Rink, L., & Haase, H. (2012). Zinc signals in neutrophil granulocytes are required for the formation of neutrophil extra- cellular traps. *Innate Immunity*, *19*(3), 253–264. doi:10.1177/1753425912458815 PMID:23008348

Hasegawa, M., Smith, M. J., & Goedert, M. (1998). Tau proteins with FTDP-17 mutations have a reduced ability to promote microtubule assembly. *FEBS Letters*, 437(3), 207–210. doi:10.1016/S0014-5793(98)01217-4 PMID:9824291

Hassan, A., Whitwell, J. L., & Josephs, K. A. (2011). The corticobasal syndrome–Alzheimer's disease conundrum. *Expert Review of Neurotherapeutics*, *11*(11), 1569–1578. doi:10.1586/ern.11.153 PMID:22014136

Hassan, I. (2011). Consultation-liaison psychiatry and prevention of severe neuroleptic sensitivity reactions in dementia with Lewy bodies. *Australasian Psychiatry*, *19*(6), 536–537. doi:10.3109/10398562.2011.580750 PMID:22077306

Hassan, I., Chibber, S., Khan, A. A., & Naseem, I. (2013). Cisplatin-induced neurotoxicity *in vivo* can be alleviated by riboflavin under photoillumination. *Cancer Biotherapy & Radiopharmaceuticals*, 28(2), 160–168. doi:10.1089/ cbr.2012.1312 PMID:23215961

Hattori, N., & Mizuno, Y. (2004). Pathogenetic mechanisms of parkin in Parkinson's disease. *Lancet*, *364*(9435), 722–724. doi:10.1016/S0140-6736(04)16901-8 PMID:15325839

Hauser, R. A., Schapira, A. H., Rascol, O., Barone, P., Mizuno, Y., Salin, L., ... Poewe, W. (2010). Randomized, doubleblind, multicenter evaluation of pramipexole extended release once daily in early Parkinson's disease. *Movement Disorders*, 25(15), 2542–2549. doi:10.1002/mds.23317 PMID:20669317

Hayes, M. W., Fung, V. S., Kimber, T. E., & O'Sullivan, J. D. (2010). Current concepts in the management of Parkinson disease. *The Medical Journal of Australia*, *192*(3), 144–149. PMID:20121682

Hayley, S., Crocker, S. J., Smith, P. D., Shree, T., Jackson-Lewis, V., Przedborski, S., ... Park, D. S. (2004). Regulation of dopaminergic loss by Fas in a 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine model of Parkinson's disease. *The Journal of Neuroscience*, *24*(8), 2045–2053. doi:10.1523/JNEUROSCI.4564-03.2004 PMID:14985447

Head, M. W., & Ironside, J. W. (2012). Review: Creutzfeldt-Jakob disease: prion protein type, disease phenotype and agent strain. *Neuropathology and Applied Neurobiology*, *38*(4), 296–310. doi:10.1111/j.1365-2990.2012.01265.x PMID:22394291

Hebert, L. E., Beckett, L. A., Scherr, P. A., & Evans, D. A. (2001). Annual incidence of Alzheimer disease in the United States projected to the years 2000 through 2050. *Alzheimer Disease and Associated Disorders*, *15*(4), 169–173. doi:10.1097/00002093-200110000-00002 PMID:11723367

Hebert, L. E., Scherr, P. A., Bienias, J. L., Bennett, D. A., & Evans, D. A. (2003). Alzheimer disease in the US population: Prevalence estimates using the 2000 census. *Archives of Neurology*, *60*(8), 1119–1122. doi:10.1001/archneur.60.8.1119 PMID:12925369

Hedden, T., & Gabrieli, J. D. (2004). Insights into the ageing mind: A view from cognitive neuroscience. *Nature Reviews*. *Neuroscience*, *5*(2), 87–96. doi:10.1038/nrn1323 PMID:14735112

Hedrich, K., Eskelson, C., Wilmot, B., Marder, K., Harris, J., Garrels, J., ... Lang, A. E. (2004). Distribution, type, and origin of Parkin mutations: Review and case studies. *Movement Disorders*, *19*(10), 1146–1157. doi:10.1002/mds.20234 PMID:15390068

Heffernan, C., Jenkinson, C., Holmes, T., Macleod, H., Kinnear, W., Oliver, D., ... Ampong, M. A. (2006). Management of respiration in MND/ ALS patients: An evidence based review. *Amyotrophic Lateral Sclerosis; Official Publication of the World Federation of Neurology Research Group on Motor Neuron Diseases*, 7(1), 5–15. doi:10.1080/14660820510043235 PMID:16546753

Heinemann, U., Krasnianski, A., Meissner, B., Kallenberg, K., Kretzschmar, H. A., Schulz-Schaeffer, W., & Zerr, I. (2008). Brain biopsy in patients with suspected Creutzfeldt-Jakob disease. *Journal of Neurosurgery*, *109*(4), 735–741. doi:10.3171/JNS/2008/109/10/0735 PMID:18826363

Heiss, W. D., Rosenberg, G. A., Thiel, A., Berlot, R., & de Reuck, J. (2016). Neuroimaging in vascular cognitive impairment: A state-of-the-art review. *BMC Medicine*, *14*(1), 174. doi:10.118612916-016-0725-0 PMID:27806705

Hely, M. A., Reid, W. G., Adena, M. A., Halliday, G. M., & Morris, J. G. (2008). The Sydney multicenter study of Parkinson's disease: The inevitability of dementia at 20 years. *Movement Disorders*, 23(6), 837–844. doi:10.1002/mds.21956 PMID:18307261

Henderson, L., Gregory, J., Irving, K., & Swan, G. (2003). *The national diet & nutrition survey: adults aged 19 to 64 years*. London: Stationery Office.

Hening, W. A., Allen, R. P., Earley, C. J., Picchietti, D. L., & Silber, M. H. (2004). An update on the dopaminergic treatment of restless legs syndrome and periodic limb movement disorder. *Sleep*, 27(3), 560–583. doi:10.1093leep/27.3.560 PMID:15164915

Hensley, K., Mhatre, M., Mou, S., Pye, Q. N., Stewart, C., West, M., & Williamson, K. S. (2006). On the relation of oxidative stress to neuroinfl ammation: Lessons learned from the G93A-SOD1 mouse model of amyotrophic lateral sclerosis. *Antioxidants & Redox Signalling*, 8(11-12), 2075–2087. doi:10.1089/ars.2006.8.2075 PMID:17034351

Hernández, A. S., García, T. A., Torres, R. E., Pacheco, C. R., & Vázquez, R. J. J. (1999). Prions: Definition and diseases. *Anales de Medicina Interna (Madrid, Spain)*, *16*, 647–653. PMID:10686720

Hernandez, D. G., Reed, X., & Singleton, A. B. (2016). Genetics in Parkinson disease: Mendelian vs. non-Mendelian inheritance. *Journal of Neurochemistry*, *139*(Suppl 1), 59–74. doi:10.1111/jnc.13593 PMID:27090875

Hernandez, D., Ruiz, C. P., Crawley, A., Malkani, R., Werner, J., Gwinn-Hardy, K., ... Singleton, A. (2005). The dardarin G2019S mutation is a common cause of Parkinson's disease but not other neurodegenerative diseases. *Neuroscience Letters*, *389*(3), 137–139. doi:10.1016/j.neulet.2005.07.044 PMID:16102903

Herrera, A. S., Del, C. A., Esparza, M., Ashraf, M. G., Zamyatnin, A. A., & Aliev, G. (2015). Beyond mitochondria, what would be the energy source of the cell? *Central Nervous System Agents in Medicinal Chemistry*, *15*(1), 32–41. do i:10.2174/1871524915666150203093656 PMID:25645910

Herzog, J., Fietzek, U., Hamel, W., Morsnowski, A., Steigerwald, F., Schrader, B., ... Volkmann, J. (2004). Most effective stimulation site in subthalamic deep brain stimulation for Parkinson's disease. *Movement Disorders*, *19*(9), 1050–1054. doi:10.1002/mds.20056 PMID:15372594

Hibberd, C., Yau, J. L., & Seckl, J. R. (2000). Glucocorticoids and the ageing hippocampus. *Journal of Anatomy*, *197*(4), 553–562. doi:10.1046/j.1469-7580.2000.19740553.x PMID:11197528

Higgins, C. M., Jung, C., & Xu, Z. (2003). ALS-associated mutant SOD1 G93A causes mitochondrial vacuolation by expansion of the intermembrane space and by involvement of SOD1 aggregation and peroxisomes. *BMC Neuroscience*, *4*(1), 16. doi:10.1186/1471-2202-4-16 PMID:12864925

Hill, A. F., Joiner, S., Wadsworth, J. D., Sidle, K. C., Bell, J. E., Budka, H., & Collinge, J. (2003). Molecular classification of sporadic Creutzfeldt–Jakob disease. *Brain*, *126*(6), 1333–1346. doi:10.1093/brain/awg125 PMID:12764055

Hilz, M. J., Stemper, B., Heckmann, J. G., & Neundörfer, B. (2000). Mechanisms of cerebral autoregulation, assessment and interpretation by means of transcranial doppler sonography. *Fortschritte der Neurologie-Psychiatrie*, 68(9), 398–412. doi:10.1055-2000-11798 PMID:11037638

Hinault, M. P., Cuendet, A. F. H., Mattoo, R. U., Mensi, M., Dietler, G., Lashuel, H. A., & Goloubinoff, P. (2010). Stable α -synuclein oligomers strongly inhibit chaperone activity of the Hsp70 system by weak interactions with J-domain co-chaperones. *The Journal of Biological Chemistry*, 285(49), 38173–38182. doi:10.1074/jbc.M110.127753 PMID:20847048

Hirai, K., Aliev, G., Nunomura, A., Fujioka, H., Russell, R. L., Atwood, C. S., ... Shimohama, S. (2001). Mitochondrial abnormalities in Alzheimer's disease. *The Journal of Neuroscience*, *21*(9), 3017–3023. PMID:11312286

Hirano, T., Murakami, M., Fukada, T., Nishida, K., Yamasaki, S., & Suzuki, T. (2008). Roles of zinc and zinc signaling in immunity: Zinc as an intracellular signaling molecule. *Advances in Immunology*, *97*, 149–176. doi:10.1016/S0065-2776(08)00003-5 PMID:18501770

Hirsch, E. C., Graybiel, A. M., Duyckaerts, C., & Javoy-Agid, F. (1987). Neuronal loss in the pedunculopontine tegmental nucleus in Parkinson disease and in progressive supranuclear palsy. *Proceedings of the National Academy of Sciences of the United States of America*, 84(16), 5976–5980. doi:10.1073/pnas.84.16.5976 PMID:3475716

Hockenbery, D. M., Oltvai, Z. N., Yin, X.-M., Milliman, C. L., & Korsmeyer, S. J. (1993). Bcl-2 functions in an antioxidant pathway to prevent apoptosis. *Cell*, 75(2), 241–251. doi:10.1016/0092-8674(93)80066-N PMID:7503812

Hoehn, M. M., & Yahr, M. D. (1967). Parkinsonism onset, progression, and mortality. *Neurology*, *17*(5), 427–427. doi:10.1212/WNL.17.5.427 PMID:6067254

Hogarth, P., Gregory, A., Kruer, M. C., Sanford, L., Wagoner, W., Natowicz, M. R., ... Hayflick, S. J. (2013). New NBIA subtype: Genetic, clinical, pathologic, and radiographic features of MPAN. *Neurology*, *80*(3), 268–275. doi:10.1212/ WNL.0b013e31827e07be PMID:23269600

Hogl, B., Saletu, M., Brandauer, E., Glatzl, S., Frauscher, B., Seppi, K., ... Poewe, W. (2002). Modafinil for the treatment of daytime sleepiness in Parkinson's disease: A double-blind, randomized, crossover, placebo-controlled polygraphic trial. *Sleep*, 25(8), 905–909. doi:10.1093leep/25.8.62 PMID:12489899

Holdorff, B. (2006). Fritz heinrich lewy (1885-1950). *Journal of Neurology*, 253(5), 677–678. doi:10.100700415-006-0130-2 PMID:16767545

Hollmann, M., & Heinemann, S. (1994). Cloned glutamate receptors. *Annual Review of Neuroscience*, *17*(1), 31–108. doi:10.1146/annurev.ne.17.030194.000335 PMID:8210177

Holman, R. C., Belay, E. D., Christensen, K. Y., Maddox, R. A., Minino, A. M., Folkema, A. M., & Schonberger, L. B. (2010). Human Prion Diseases in the United States. *PLoS One*, 5(1), 1–8. doi:10.1371/journal.pone.0008521 PMID:20049325

Holmes, G., Sirven, J., & Fisher, R. S. (2014). *Temporal Lobe Epilepsy*. Epilepsy Foundation and Epilepsy Therapy Project. Available at: http://www.epilepsy.com/learn/types-epilepsy-syndromes/temporal-lobe-epilepsy

Honer, W. G. (2003). Pathology of presynaptic proteins in Alzheimer's disease: More than simple loss of terminals. *Neurobiology of Aging*, *24*(8), 1047–1062. doi:10.1016/j.neurobiolaging.2003.04.005 PMID:14643376

Hong, L., Huang, H.-C., & Jiang, Z.-F. (2014). Relationship between amyloid-beta and the ubiquitin–proteasome system in Alzheimer's disease. *Neurological Research*, *36*(3), 276–282. doi:10.1179/1743132813Y.0000000288 PMID:24512022

Hong, M., Zhukareva, V., Vogelsberg-Ragaglia, V., Wszolek, Z., Reed, L., Miller, B. I., & ... (1998). Mutationspecific functional impairments in distinct tau isoforms of hereditary FTDP-17. *Science*, 282(5395), 1914–1917. doi:10.1126cience.282.5395.1914 PMID:9836646

Horimoto, Y., Matsumoto, M., Nakazawa, H., Yuasa, H., Morishita, M., Akatsu, H., ... Kosaka, K. (2003). Cognitive conditions of pathologically confirmed dementia with Lewy bodies and Parkinson's disease with dementia. *Journal of the Neurological Sciences*, *216*(1), 105–108. doi:10.1016/S0022-510X(03)00220-X PMID:14607310

Horner, R., Carr, E., Strain, P., Todd, A., & Reed, H. (2002). Problem behavior interventions for young children with autism: A research synthesis. *Journal of Autism and Developmental Disorders*, *32*(5), 423–446. doi:10.1023/A:1020593922901 PMID:12463518

Hornykiewicz, O., & Kish, S. J. (1987). Biochemical pathophysiology of Parkinson's disease. *Advances in Neurology*, 45, 19–34. PMID:2881444

Hotz, C., Peerson, J. M., & Brown, K. H. (2003). Suggested lower cutoffs of serum zinc concentrations for assessing zinc status: Reanalysis of the second national health and nutrition examination survey data (1976–1980). *The American Journal of Clinical Nutrition*, 78(4), 756–764. doi:10.1093/ajcn/78.4.756 PMID:14522734

Houlden, H., Baker, M., Morris, H. R., MacDonald, N., Pickering-Brown, S., Adamson, J., & ... (2001). Corticobasal degeneration and progressive supranuclear palsy share a common tau haplotype. *Neurology*, *56*(12), 1702–1706. doi:10.1212/WNL.56.12.1702 PMID:11425937

Howland, D. S., Liu, J., She, Y., Goad, B., Maragakis, N. J., Kim, B., ... Rothstein, J. D. (2002). Focal loss of the glutamate transporter EAAT2 in a transgenic rat model of SOD1 mutant-mediated amyotrophic lateral sclerosis (ALS). *Proceedings of the National Academy of Sciences*, *99*, 1604–1609. 10.1073/pnas.032539299

Hroudová, J., Singh, N., & Fišar, Z. (2014). Mitochondrial dysfunctions in neurodegenerative diseases: Relevance to Alzheimer's disease. *BioMed Research International*, 2014, 1–9. doi:10.1155/2014/175062 PMID:24900954

Huang, L., & Tepaamorndech, S. (2013). The SLC30 family of zinc transporters are view of current understanding of their biological and pathophysiological roles. *Molecular Aspects of Medicine*, *34*(2-3), 548–560. doi:10.1016/j. mam.2012.05.008 PMID:23506888

Huang, W. J., Zhang, X., & Chen, W. W. (2016). Role of oxidative stress in Alzheimer's disease. *Biomedical Reports*, 4(5), 519–522. doi:10.3892/br.2016.630 PMID:27123241

Huang, X., Moir, R. D., Tanzi, R. E., Bush, A. I., & Rogers, J. T. (2004). Redox-active metals, oxidative stress, and Alzheimer's disease pathology. *Annals of the New York Academy of Sciences*, *1012*(1), 153–163. doi:10.1196/annals.1306.012 PMID:15105262

Huffman, L. C., Sutcliffe, T. L., Tanner, I. S. D., & Feldman, H. M. (2011). Management of symptoms in children with autism spectrum disorders: A comprehensive review of pharmacologic and complementary-alternative medicine treatments. *Journal of Developmental and Behavioral Pediatrics*, *32*(1), 56–68. doi:10.1097/DBP.0b013e3182040acf PMID:21160435

Hughes, J. T., Brownell, B., & Hewer, R. L. (1968). The peripheral sensory pathway in friedreich's ataxia. An examination by light and electron microscopy of the posterior nerve roots, posterior root ganglia, and peripheral sensory nerves in cases of friedreich's ataxia. *Brain*, *91*(4), 803–818. doi:10.1093/brain/91.4.803 PMID:4178703

Hu, H., & Li, M. (2016). Mitochondria-targeted antioxidant mitotempo protects mitochondrial function against amyloid beta toxicity in primary cultured mouse neurons. *Biochemical and Biophysical Research Communications*, 478(1), 174–180. doi:10.1016/j.bbrc.2016.07.071 PMID:27444386

Huisman, M. H., Seelen, M., de Jong, S. W., Dorresteijn, K. R., van Doormaal, P. T., van der Kooi, A. J., ... Veldink, J. H. (2013). Lifetime physical activity and the risk of amyotrophic lateral sclerosis. *Journal of Neurology, Neurosurgery, and Psychiatry*, *84*(9), 976–981. doi:10.1136/jnnp-2012-304724 PMID:23418211

Huisman, T. (2010). Diffusion-weighted and diffusion tensor imaging of the brain, made easy. *Cancer Imaging; the Official Publication of the International Cancer Imaging Society*, *10*(1A), S163–S171. doi:10.1102/1470-7330.2010.9023 PMID:20880787

Hu, Q., & Wang, G. (2016). Mitochondrial dysfunction in Parkinson's disease. *Translational Neurodegeneration*, 5(1), 14. doi:10.118640035-016-0060-6 PMID:27453777

Hurd, M. D., Martorell, P., Delavande, A., Mullen, K. J., & Langa, K. M. (2013). Monetary Costs of Dementia in the United States. *The New England Journal of Medicine*, *368*(14), 1326–1334. doi:10.1056/NEJMsa1204629PMID:23550670

Hurtado, D. E., Molina-Porcel, L., Iba, M., Aboagye, A. K., Paul, S. M., Trojanowski, J. Q., & Lee, V. M.-Y. (2010). Aβ Accelerates the Spatiotemporal Progression of Tau Pathology and Augments Tau Amyloidosis in an Alzheimer Mouse Model. *American Journal of Pathology*, *177*(4), 1977–1988. doi:10.2353/ajpath.2010.100346 PMID:20802182

Husebo, B. S., Ballard, C., Sandvik, R., Nilsen, O. B., & Aarsland, D. (2011). Efficacy of treating pain to reduce behavioural disturbances in residents of nursing homes with dementia: Cluster randomised clinical trial. *BMJ (Clinical Research Ed.)*, 343(jul15 1), d4065. doi:10.1136/bmj.d4065 PMID:21765198

Hussain, I., Powell, D., Howlett, D. R., Tew, D. G., Meek, T. D., Chapman, C., ... Christie, G. (1999). Identification of a Novel Aspartic Protease (Asp 2) as β-Secretase. *Molecular and Cellular Neurosciences*, *14*(6), 419–427. doi:10.1006/mcne.1999.0811 PMID:10656250

Huss, D. S., Dallapiazza, R. F., Shah, B. B., Harrison, M. B., Diamond, J., & Elias, W. J. (2015, December). Functional assessment and quality of life in essential tremor with bilateral or unilateral DBS and focused ultrasound thalamotomy. *Movement Disorders*, *30*(14), 1937–1943. doi:10.1002/mds.26455 PMID:26769606

Hutton, M., Lendon, C. L., Rizzu, P., Baker, M., Froelich, S., Houlden, H., ... Heutink, P. (1998). Association of missense and 5'-splice-site mutations in tau with the inherited dementia FTDP-17. *Nature*, *393*(6686), 702–705. doi:10.1038/31508 PMID:9641683

Huuskonen, J., Suuronen, T., Miettinen, R., van Groen, T., & Salminen, A. (2005). A refined in vitro model to study inflammatory responses in organotypic membrane culture of postnatal rat hippocampal slices. *Journal of Neuroinflammation*, *2*(1), 25. doi:10.1186/1742-2094-2-25 PMID:16285888

Hynd, M. R., Scott, H. L., & Dodd, P. R. (2004). Glutamate-mediated excitotoxicity and neurodegeneration in Alzheimer's disease. *Neurochemistry International*, 45(5), 583–595. doi:10.1016/j.neuint.2004.03.007 PMID:15234100

Hyun-Ju, S., Young-Eun, C., Taewan, K., Hong-In, S., & In-Sook, K. (2010). Zinc may increase bone formation through stimulating cell proliferation, alkaline phosphatase activity and collagen synthesis in osteoblastic MC3T3-E1 cells. *Nutrition Research and Practice*, *4*(5), 356–361. doi:10.4162/nrp.2010.4.5.356 PMID:21103080

Iacono, R. P., Shima, F., Lonser, R. R., Kuniyoshi, S., Maeda, G., & Yamada, S. (1995, June). The results, indications, and physiology of posteroventral pallidotomy for patients with Parkinson's disease. *Neurosurgery*, *36*(6), 1118–1125. doi:10.1227/00006123-199506000-00008 PMID:7643990

Iadecola, C. (2013). The pathobiology of vascular dementia. *Neuron*, 80(4), 844–866. doi:10.1016/j.neuron.2013.10.008 PMID:24267647

Ideker, T., & Sharan, R. (2008). Protein networks in disease. *Genome Research*, *18*(4), 644–652. doi:10.1101/gr.071852.107 PMID:18381899

Imam, S. Z., Karahalil, B., Hogue, B. A., Souza-Pinto, N. C., & Bohr, V. A. (2006). Mitochondrial and nuclear DNArepair capacity of various brain regions in mouse is altered in an age-dependent manner. *Neurobiology of Aging*, 27(8), 1129–1136. doi:10.1016/j.neurobiolaging.2005.06.002 PMID:16005114

Imlach, W. L., Beck, E. S., Choi, B. J., Lotti, F., Pellizzoni, L., & McCabe, B. D. (2012). SMN is required for sensorymotor circuit function in Drosophila. *Cell*, 151(2), 427–439. doi:10.1016/j.cell.2012.09.011 PMID:23063130

Indo, H. P., Davidson, M., Yen, H. C., Suenaga, S., Tomita, K., Nishii, T., ... Majima, H. J. (2007). Evidence of ROS generation by mitochondria in cells with impaired electron transport chain and mitochondrial DNA damage. *Mitochondrion*, *7*(1), 106–118. doi:10.1016/j.mito.2006.11.026 PMID:17307400

Ingram, E. M., & Spillantini, M. G. (2002). Tau gene mutations: Dissecting the pathogenesis of FTDP-17. *Trends in Molecular Medicine*, 8(12), 555–562. doi:10.1016/S1471-4914(02)02440-1 PMID:12470988

Inoue, K., O'Bryant, Z., & Xiong, Z. G. (2015). Zinc-permeable ion channels: Effects on intracellular zinc dynamics and potential physiological/pathophysiological significance. *Current Medicinal Chemistry*, 22(10), 1248–1257. doi:10 .2174/0929867322666150209153750 PMID:25666796

Institutes, G. (2015). Loss of cellular energy leads to neuronal dysfunction in neurodegenerative disease model: Scientists developed new tests to accurately measure brain's energy supply, which is essential for understanding how impairments in the system cause neurodegeneration. Retrieved from www.sciencedaily.com/releases/2015/09/150914215617.htm

Iqbal, K., Liu, F., Gong, C. X., & Grundke-Iqbal, I. (2010). Tau in Alzheimer disease and related tauopathies. *Current Alzheimer Research*, 7(8), 656–664. doi:10.2174/156720510793611592 PMID:20678074

Irvine, G. B., El-Agnaf, O. M., Shankar, G. M., & Walsh, D. M. (2008). Protein Aggregation in the Brain: The Molecular Basis for Alzheimer's and Parkinson's Diseases. *Molecular Medicine (Cambridge, Mass.)*, 14(7–8), 451–464. PMID:18368143

Irwin, D. J., White, M. T., Toledo, J. B., Xie, S. X., Robinson, J. L., Van Deerlin, V., ... Hurtig, H. I. (2012). Neuropathologic substrates of Parkinson disease dementia. *Annals of Neurology*, 72(4), 587–598. doi:10.1002/ana.23659 PMID:23037886

Ito, D., Seki, M., Tsunoda, Y., Uchiyama, H., & Suzuki, N. (2011). Nuclear transport impairment of amyotrophic lateral sclerosis-linked mutations in FUS/TLS. *Annals of Neurology*, *69*(1), 152–162. doi:10.1002/ana.22246 PMID:21280085

Ito, G., Okai, T., Fujino, G. O., Takeda, K., Ichijo, H., Katada, T., & Iwatsubo, T. (2007). GTP binding is essential to the protein kinase activity of LRRK2, a causative gene product for familial Parkinson's disease. *Biochemistry*, *46*(5), 1380–1388. doi:10.1021/bi061960m PMID:17260967

Ittner, L. M., Ke, Y. D., Delerue, F., Bi, M., Gladbach, A., van Eersel, J., ... Götz, J. (2010). Dendritic Function of Tau Mediates Amyloid-β Toxicity in Alzheimer's Disease Mouse Models. *Cell*, *142*(3), 387–397. doi:10.1016/j.cell.2010.06.036 PMID:20655099

Iuvone, T., Esposito, G., Esposito, R., Santamaria, R., Di Rosa, M., & Izzo, A. A. (2004). Neuroprotective effect of cannabidiol, a non-psychoactive component from Cannabis sativa, on β-amyloid-induced toxicity in PC12 cells. *Journal of Neurochemistry*, *89*(1), 134–141. doi:10.1111/j.1471-4159.2003.02327.x PMID:15030397

Ivan, C. S., Seshadri, S., Beiser, A., Au, R., Kase, C. S., Kelly-Hayes, M., & Wolf, P. A. (2004). Dementia after stroke: The Framingham Study. *Stroke*, *35*(6), 1264–1268. doi:10.1161/01.STR.0000127810.92616.78 PMID:15118167

Iwai, A., Masliah, E., Yoshimoto, M., Ge, N., Flanagan, L., de Silva, H. A., ... Saitoh, T. (1995). The precursor protein of non-A beta component of Alzheimer's disease amyloid is a presynaptic protein of the central nervous system. *Neuron*, *14*(2), 467–475. doi:10.1016/0896-6273(95)90302-X PMID:7857654

Iwasaki, Y., Mori, K., & Ito, M. (2012). Investigation of the clinical course and treatment of prion disease patients in the akinetic mutism state in Japan. *Clinical Neurology*, *52*(5), 314-319.

Izumihara, A., Ishihara, T., Iwamoto, N., Yamashita, K., & Ito, H. (1999). Postoperative outcome of 37 patients with lobar intracerebral hemorrhage related to cerebral amyloid angiopathy. *Stroke*, *30*(1), 29–33. doi:10.1161/01.STR.30.1.29 PMID:9880384

Jaarsma, D., Rognoni, F., van Duijn, W., Verspaget, H. W., Haasdijk, E. D., & Holstege, J. C. (2001). CuZn superoxide dismutase (SOD1) accumulates in vacuolated mitochondria in transgenic mice expressing amyotrophic lateral sclerosislinked SOD1 mutations. *Acta Neuropathologica*, *102*(4), 293–305. PMID:11603803

Jabbari, B. (2016). History of Botulinum Toxin Treatment in Movement Disorders. *Tremor and Other Hyperkinetic Movements (New York, N.Y.)*, 6, 394. PMID:27917308

Jabbour, J. T., Garcia, J. H., Lemmi, H., Ragland, J., Duenas, D. A., & Sever, J. L. (1969). Subacute sclerosing pan encephalitis. A multidisciplinary study of eight cases. *Journal of the American Medical Association*, 207(12), 2248–2254. doi:10.1001/jama.1969.03150250078007 PMID:5818397

Jack, C. R. Jr, Wiste, H. J., Weigand, S. D., Rocca, W. A., Knopman, D. S., Mielke, M. M., ... Petersen, R. C. (2014). Age-specific population frequencies of cerebral beta-amyloidosis and neurodegeneration among people with normal cognitive function aged 50-89 years: A cross-sectional study. *Lancet Neurology*, *13*(10), 997–1005. doi:10.1016/S1474-4422(14)70194-2 PMID:25201514

Jackson, G. S., & Collinge, J. (2001). The molecular pathology of CJD: Old and new variants. *Molecular Pathology*, *54*(6), 393. PMID:11724914

Jackson-Lewis, V., Jakowec, M., Burke, R. E., & Przedborski, S. (1995). Time course and morphology of dopaminergic neuronal death caused by the neurotoxin 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine. *Neurodegeneration*, *4*(3), 257–269. doi:10.1016/1055-8330(95)90015-2 PMID:8581558

Jackson, M. J., Vasilaki, A., & McArdle, A. (2016). Cellular mechanisms underlying oxidative stress in human exercise. *Free Radical Biology & Medicine*, *98*(13-17). PMID:26912036

Jackson, W. S., & Krost, C. (2014). Peculiarities of Prion Diseases. *PLoS Pathogens*, *10*(11), e1004451. doi:10.1371/journal.ppat.1004451 PMID:25411777

Jakhar, R., Paul, S., Bhardwaj, M., & Kang, S. C. (2017). Astemizole–Histamine induces Beclin-1-independent autophagy by targeting p53-dependent crosstalk between autophagy and apoptosis. *Cancer Letters*, *372*(1), 89–100. doi:10.1016/j. canlet.2015.12.024 PMID:26739061

James, N. G., Digman, M. A., Gratton, E., Barylko, B., Ding, X., Albanesi, J. P., ... Jameson, D. M. (2012). Number and brightness analysis of LRRK2 oligomerization in live cells. *Biophysical Journal*, *102*(11), L41–L43. doi:10.1016/j. bpj.2012.04.046 PMID:22713584

Jana, N. R. (2012). Protein homeostasis and aging: Role of ubiquitin protein ligases. *Neurochemistry International*, 60(5), 443–447. doi:10.1016/j.neuint.2012.02.009 PMID:22353631

Jankovic, J., McDermott, M., Carter, J., Gauthier, S., Goetz, C., Golbe, L., ... Stern, M. (1990). Variable expression of Parkinson's disease A base-line analysis of the DAT ATOP cohort. *Neurology*, 40(10), 1529–1529. doi:10.1212/ WNL.40.10.1529 PMID:2215943

Jansen, A. C., & Andermann, E. (2007, Dec. 28). Progressive Myoclonus Epilepsy, Lafora Type. In *Gene Reviews* (pp. 1993-2017). Seattle, WA: University of Washington. Available from: https://www.ncbi.nlm.nih.gov/books/NBK1389/

Jansen, A. C. (2010). Lafora disease. In K. Kompoliti & V. L. Metman (Eds.), *Encyclopedia of movement disorders* (pp. 113–116). Oxford, UK: Academic Press. doi:10.1016/B978-0-12-374105-9.00338-5

Janus, C., Pearson, J., McLaurin, J., Mathews, P. M., Jiang, Y., Schmidt, S. D., ... Westaway, D. (2000). A beta peptide immunization reduces behavioural impairment and plaques in a model of Alzheimer's disease. *Nature*, 408(6815), 979–982. doi:10.1038/35050110 PMID:11140685

Jastroch, M., Divakaruni, A. S., Mookerjee, S., Treberg, J. R., & Brand, M. D. (2010). Mitochondrial proton and electron leaks. *Essays in Biochemistry*, 47, 53–67. doi:10.1042/bse0470053 PMID:20533900

Javier, F., & Rojas-garc, R. (2016). ALS: A bucket of genes, environment, metabolism and unknown ingredients. *Progress in Neurobiology*. doi:10.1016/j.pneurobio.2016.05.004 PMID:27236050

Javitch, J. A., D'Amato, R. J., Strittmatter, S. M., & Snyder, S. H. (1985). Parkinsonism-inducing neurotoxin, N-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine: Uptake of the metabolite N-methyl-4-phenylpyridine by dopamine neurons explains selective toxicity. *Proceedings of the National Academy of Sciences of the United States of America*, 82(7), 2173–2177. doi:10.1073/pnas.82.7.2173 PMID:3872460

Jellinger, K. (1988). The pedunculopontine nucleus in Parkinson's disease, progressive supranuclear palsy and Alzheimer's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 51(4), 540–543. doi:10.1136/jnnp.51.4.540 PMID:3379428

Jellinger, K. (2002). Alzheimer disease and cerebrovascular pathology: An update. *Journal of Neural Transmission* (*Vienna, Austria*), 109(5), 813–836. doi:10.1007007020200068 PMID:12111471

Jellinger, K. A. (2009). A critical evaluation of current staging of alpha-synucleinpathology in Lewy body disorders. *Biochimica et Biophysica Acta*, *1792*(7), 730–734. doi:10.1016/j.bbadis.2008.07.006 PMID:18718530

Jellinger, K. A. (2009). Formation and development of Lewy pathology: A critical update. *Journal of Neurology*, 256(3), 270–279. doi:10.100700415-009-5243-y PMID:19711116

Jellinger, K. A. (2010). Basic mechanisms of neurodegeneration: A critical update. *Journal of Cellular and Molecular Medicine*, *14*(3), 457–487. PMID:20070435

Jellinger, K. A. (2012). Interaction between pathogenic proteins in neurodegenerative disorders. *Journal of Cellular and Molecular Medicine*, *16*(6), 1166–1183. doi:10.1111/j.1582-4934.2011.01507.x PMID:22176890

Jellinger, K. A. (2013). Pathology and pathogenesis of vascular cognitive impairment - a critical update. *Frontiers in Aging Neuroscience*, *10*(5), 17. PMID:23596414

Jellinger, K. A., & Attems, J. (2015). Challenges of multimorbidity of the aging brain: A critical update. *Journal of Neural Transmission (Vienna, Austria)*, *122*(4), 505–521. doi:10.100700702-014-1288-x PMID:25091618

Jellinger, K. A., & Mitter-Ferstl, E. (2003). The impact of cerebrovascular lesions in Alzheimer disease. *Journal of Neurology*, 250(9), 1050–1055. doi:10.100700415-003-0142-0 PMID:14504965

Jellinger, K. A., Seppi, K., Wenning, G. K., & Poewe, W. (2002). Impact of coexistent Alzheimer pathology on the natural history of Parkinson's disease. *Journal of Neural Transmission (Vienna, Austria)*, *109*(3), 329–339. doi:10.1007007020200027 PMID:11956955

Jenkins, B. G., Koroshetz, W. J., Beal, M. F., & Rosen, B. R. (1993). Evidence for impairment of energy metabolism in vivo in Huntington's disease using localized 1H NMR spectroscopy. *Neurology*, *43*(12), 2689–2695. doi:10.1212/WNL.43.12.2689 PMID:8255479

Jeppesen, D. K., Bohr, V. A., & Stevnsner, T. (2011). DNA repair deficiency in neurodegeneration. *Progress in Neurobiology*, 94(2), 166–200. doi:10.1016/j.pneurobio.2011.04.013 PMID:21550379

Jéssica, B. B., Juliana, S. S., & Ana, R. S. O. (2017). Zinc and oxidative stress: Current Mechanisms. *Antioxidant*, 6(4), 24. doi:10.3390/antiox6020024

Jia, J., Ma, L., Wu, M., Zhang, L., Zhang, X., Zhai, Q., ... Xiong, L. (2014). Anandamide protects HT22 cells exposed to hydrogen peroxide by inhibiting CB1 receptor-mediated type 2 NADPH oxidase. *Oxidative Medicine and Cellular Longevity*. PMID:25136404

Jiang, J., Zhu, Q., Gendron, T. F., Saberi, S., McAlonis-Downes, M., Seelman, A., ... Schulte, D. (2016). Gain of toxicity from ALS/FTD-linked repeat expansions in C9ORF72 is alleviated by antisense oligonucleotides targeting GGGGCC-containing RNAs. *Neuron*, *90*(3), 535–550. doi:10.1016/j.neuron.2016.04.006 PMID:27112497

Jimenez-Del-Rio, M., & Velez-Pardo, C. (2012). The bad, the good, and the ugly about oxidative stress. *Oxidative Medicine and Cellular Longevity*, 2012, 1–13. doi:10.1155/2012/163913 PMID:22619696

Jindal, K., & Bansal, A. (2016). APOEɛ2 is Associated with Milder Clinical and Pathological Alzheimer's Disease. *Annals of Neurosciences*, 23(2), 112–112. doi:10.1159/000443572 PMID:27647961

Jin, H., Kanthasamy, A., Ghosh, A., Anantharam, V., Kalyanaraman, B., & Kanthasamy, A. G. (2014). Mitochondriatargeted antioxidants for treatment of Parkinson's disease: Preclinical and clinical outcomes. *Molecular Basis of Disease*, *1842*(8), 1282–1294. doi:10.1016/j.bbadis.2013.09.007 PMID:24060637

Jin, L. W., Masliah, E., Iimoto, D., Deteresa, R., Mallory, M., Sundsmo, M., ... Saitoh, T. (1996). Neurofibrillary tangleassociated alteration of stathmin in Alzheimer's disease. *Neurobiology of Aging*, *17*(3), 331–341. doi:10.1016/0197-4580(96)00021-8 PMID:8725893

Johnson, B. S., Snead, D., Lee, J. J., McCaffery, J. M., Shorter, J., & Gitler, A. D. (2009). TDP-43 is intrinsically aggregation-prone, and amyotrophic lateral sclerosis-linked mutations accelerate aggregation and increase toxicity. *The Journal of Biological Chemistry*, 284(30), 20329–20339. doi:10.1074/jbc.M109.010264 PMID:19465477

Johnson, K. A., Conn, P. J., & Niswender, C. M. (2009). Glutamate receptors as therapeutic targets for Parkinson's disease. *CNS & Neurological Disorders - Drug Targets*, 8(6), 475–491. doi:10.2174/187152709789824606 PMID:19702565

Johnson, K. A., Fox, N. C., Sperling, R. A., & Klunk, W. E. (2012). Brain Imaging in Alzheimer Disease. *Cold Spring Harbor Perspectives in Medicine*, 2(4), a006213. doi:10.1101/cshperspect.a006213 PMID:22474610

Johnson, W. M., Wilson-Delfosse, A. L., & Mieyal, J. J. (2012). Dysregulation of glutathione homeostasis in neurodegenerative diseases. *Nutrients*, 4(10), 1399–1440. doi:10.3390/nu4101399 PMID:23201762

Johnston, J. A., Dalton, M. J., Gurney, M. E., & Kopito, R. R. (2000). Formation of high molecular weight complexes of mutant Cu, Zn-superoxide dismutase in a mouse model for familial amyotrophic lateral sclerosis. *Proceedings of the National Academy of Sciences of the United States of America*, 97(23), 12571–12576. doi:10.1073/pnas.220417997 PMID:11050163

Johri, A., & Beal, M. F. (2012). Mitochondrial dysfunction in neurodegenerative diseases. *The Journal of Pharmacology* and *Experimental Therapeutics*, *342*(3), 619–630. doi:10.1124/jpet.112.192138 PMID:22700435

Joosten, E. (2001). Homocysteine, vascular dementia and Alzheimer's disease. *Clinical Chemistry and Laboratory Medicine*, *39*(8), 717–720. doi:10.1515/CCLM.2001.119 PMID:11592440

Josephs, K. A., Matsumoto, J. Y., & Lindor, N. M. (2004). Heterozygous Niemann- Pick disease type C presenting with tremor. *Neurology*, *63*(11), 2189–2190. doi:10.1212/01.WNL.0000145710.25588.2F PMID:15596783

Joshi, G., & Bekier, M. E., & II, Y. W. (2015). Golgi fragmentation in Alzheimer's disease. *Frontiers in Neuroscience*, 24(9), 340. PMID:26441511

Joshi, Y. B., & Praticò, D. (2014). Lipid peroxidation in psychiatric illness: Overview of clinical evidence. *Oxidative Medicine and Cellular Longevity*. PMID:24868318

Jost, W. H. (1997). Gastrointestinal motility problems in patients with Parkinson's disease. *Drugs & Aging*, 10(4), 249–258. doi:10.2165/00002512-199710040-00002 PMID:9108986

Joutel, A., Corpechot, C., Ducros, A., Vahedi, K., Chabriat, H., Mouton, P., ... Tournier-Lasserve, E. (1996). Notch3 mutations in CADASIL, a hereditary adult-onset condition causing stroke and dementia. *Nature*, *383*(6602), 707–710. doi:10.1038/383707a0 PMID:8878478

Joutel, A., Monet-Leprêtre, M., Gosele, C., Baron-Menguy, C., Hammes, A., Schmidt, S., ... Hubner, N. (2010). Cerebrovascular dysfunction and microcirculation rarefaction precede white matter lesions in a mouse genetic model of cerebral ischemic small vessel disease. *The Journal of Clinical Investigation*, *120*(2), 433–445. doi:10.1172/JCI39733 PMID:20071773

Joutel, A., Vahedi, K., Corpechot, C., Troesch, A., Chabriat, H., Vayssière, C., ... Tournier-Lasserve, E. (1997). Strong clustering and stereotyped nature of Notch3 mutations in CADASIL patients. *Lancet*, *350*(9090), 1511–1515. doi:10.1016/S0140-6736(97)08083-5 PMID:9388399

Jovicic, A., Mertens, J., Boeynaems, S., Bogaert, E., Chai, N., Yamada, S. B., ... Gitler, A. D. (2015). Modifie.rs of C9orf72 dipeptide repeat toxicity connect nucleocytoplasmic transport defects to FTD/ALS Nature. Neuroscience, 18, 1226–1229. PMID:26308983

Jucker, M., & Walker, L. C. (2011). Pathogenic protein seeding in Alzheimer disease and other neurodegenerative disorders. *Annals of Neurology*, 70(4), 532–540. doi:10.1002/ana.22615 PMID:22028219

Junn, E., Jang, W. H., Zhao, X., Jeong, B. S., & Mouradian, M. M. (2009). Mitochondrial localization of DJ-1 leads to enhanced neuroprotection. *Journal of Neuroscience Research*, 87(1), 123–129. doi:10.1002/jnr.21831 PMID:18711745

Junqué, C., Ramírez-Ruiz, B., Tolosa, E., Summerfield, C., Martí, M. J., Pastor, P., ... Mercader, J. M. (2005). Amygdalar and hippocampal MRI volumetric reductions in Parkinson's disease with dementia. *Movement Disorders*, 20(5), 540–544. doi:10.1002/mds.20371 PMID:15645532

Kadowaki, H., Nishitoh, H., Urano, F., Sadamitsu, C., Matsuzawa, A., Takeda, K., ... Ichijo, H. (2005). Amyloid β induces neuronal cell death through ROS-mediated ASK1 activation. *Cell Death and Differentiation*, *12*(1), 19–24. doi:10.1038j. cdd.4401528 PMID:15592360

Kagi, G., Bhatia, K. P., & Tolosa, E. (2010). The role of DAT-SPECT in movement disorders. *Journal of Neurology, Neurosurgery, and Psychiatry*, *81*(1), 5–12. doi:10.1136/jnnp.2008.157370 PMID:20019219

Kahle, P. J. (2008). alpha-Synucleinopathy models and human neuropathology: Similarities and differences. *Acta Neuropathologica*, *115*(1), 87–95. doi:10.100700401-007-0302-x PMID:17932682

Kahmann, L., Uciechowski, P., Warmuth, S., Plümäkers, B., Gressner, A. M., Malavolta, M., ... Rink, L. (2008). Zinc supplementation in the elderly reduces spontaneous inflammatory cytokine release and restores T-cell functions. *Rejuvenation Research*, *11*(1), 227–237. doi:10.1089/rej.2007.0613 PMID:18279033

Kaiser, D. M., Acharya, M., Leighton, P. L., Wang, H., Daude, N., Wohlgemuth, S., ... Allison, W. T. (2012). Amyloid beta precursor protein and prion protein have a conserved interaction affecting cell adhesion and CNS development. *PLoS One*, *7*(12), e51305. doi:10.1371/journal.pone.0051305 PMID:23236467

Kalinderi, K., Bostantjopoulou, S., & Fidani, L. (2016). The genetic background of Parkinson's disease: Current progress and future prospects. *Acta Neurologica Scandinavica*, *134*(5), 314–326. doi:10.1111/ane.12563 PMID:26869347

Kalra, S., Bergeron, C., & Lang, A. E. (1996). Lewy body disease and dementia: A review. *Archives of Internal Medicine*, *156*(5), 487–493. doi:10.1001/archinte.1996.00440050031004 PMID:8604954

Kannan, K., & Jain, S. K. (2000). Oxidative stress and apoptosis. *Pathophysiology*, 7(3), 153–163. doi:10.1016/S0928-4680(00)00053-5 PMID:10996508

Kann, M. G. (2007). Protein interactions and disease: Computational approaches to uncover the etiology of diseases. *Briefings in Bioinformatics*, 8(5), 333–346. doi:10.1093/bib/bbm031 PMID:17638813

Kantor, B., McCown, T., Leone, P., & Gray, S. J. (2014). Clinical Applications Involving CNS Gene Transfer. *Advances in Genetics*, 87, 71–124. doi:10.1016/B978-0-12-800149-3.00002-0 PMID:25311921

Karachi, C., Grabli, D., Bernard, F. A., Tandé, D., Wattiez, N., Belaid, H., ... Hartmann, A. (2010). Cholinergic mesencephalic neurons are involved in gait and postural disorders in Parkinson disease. *The Journal of Clinical Investigation*, *120*(8), 2745–2754. doi:10.1172/JCI42642 PMID:20628197

Karapetyan, Y. E., Sferrazza, G. F., Zhou, M., Ottenberg, G., Spicer, T., Chase, P., & Lasmezas, C. I. (2013). Unique drug screening approach for prion diseases identifies tacrolimus and astemizole as antiprion agents. *Proceedings of the National Academy of Sciences of the United States of America*, *110*(17), 7044–7049. doi:10.1073/pnas.1303510110 PMID:23576755

Karbowski, M., & Youle, R. J. (2003). Dynamics of mitochondrial morphology in healthy cells and during apoptosis. *Cell Death and Differentiation*, *10*(8), 870–880. doi:10.1038j.cdd.4401260 PMID:12867994

Karecla, P. I., & Kreis, T. E. (1992). Interaction of membranes of the Golgi complex with microtubules *in vitro*. *European Journal of Cell Biology*, *57*(2), 139–146. PMID:1387362

Karimzadeh, P., & Jafari, N., Nejad biglari, H., Rahimian, E., Ahmadabadi, F., Nemati, H., ... Mollamohammadi, M. (2014). The Clinical Features and Diagnosis of Canavan's Disease: A Case Series of Iranian Patients. *Iranian Journal of Child Neurology*, 8(4), 66–71. PMID:25657773

Karve, T. M., & Cheema, A. K. (2011). Small changes huge impact: The role of protein posttranslational modifications in cellular homeostasis and disease. *Journal of Amino Acids*, 2011, 207691. doi:10.4061/2011/207691 PMID:22312457

Kasarskis, E. J., Scarlata, D., Hill, R., Fuller, C., Stambler, N., & Cedarbaum, J. M. (1999). A retrospective study of percutaneous endoscopic gastrostomy in ALS patients during the BDNF and CNTF trials. *Journal of the Neurological Sciences*, *169*(1-2), 118–125. doi:10.1016/S0022-510X(99)00230-0 PMID:10540019

Katsuse, O., Iseki, E., Marui, W., & Kosaka, K. (2003). Developmental stages of cortical Lewy bodies and their relation to axonal transport blockage in brains of patients with dementia with Lewy bodies. *Journal of the Neurological Sciences*, 211(1), 29–35. doi:10.1016/S0022-510X(03)00037-6 PMID:12767494

Katzenschlager, R., & Lees, A. J. (2002). Treatment of Parkinson's disease: Levodopa as the first choice. *Journal of Neurology*, 249. PMID:12375059

Kaul, R., Gao, G. P., Matalon, R., Aloya, M., Su, Q., Jin, M., ... Clarke, J. T. (1996). Identification and expression of eight novel mutations among non-Jewish patients with Canavan disease. *American Journal of Human Genetics*, 59(1), 95–102. PMID:8659549

Kaur, S. J., McKeown, S. R., & Rashid, S. (2016). Mutant SOD1 mediated pathogenesis of amyotrophic lateral sclerosis. *Gene*, 577(2), 109–118. doi:10.1016/j.gene.2015.11.049 PMID:26657039

Kaur, U., Banerjee, P., Bir, A., Sinha, M., Biswas, A., & Chakrabarti, S. (2015). Reactive oxygen species, redox signaling and neuroinflammation in Alzheimer's disease: The NF-κB connection. *Current Topics in Medicinal Chemistry*, *15*(5), 446–457. doi:10.2174/1568026615666150114160543 PMID:25620241

Keage, H.A., Carare, R.O., Friedland, R.P., Ince, P.G., Love, S., Nicoll, J.A., ... Brayne, C. (2009). Population studies of sporadic cerebral amyloid angiopathy and dementia: a systematic review. *BMC Neurology*, *13*.

Keeney, P. M., Xie, J., Capaldi, R. A., & Bennett, J. P. (2006). Parkinson's disease brain mitochondrial complex I has oxidatively damaged subunits and is functionally impaired and misassembled. *The Journal of Neuroscience*, *26*(19), 5256–5264. doi:10.1523/JNEUROSCI.0984-06.2006 PMID:16687518

Kehagia, A. A., Barker, R. A., & Robbins, T. W. (2013). Cognitive impairment in Parkinson's disease: The dual syndrome hypothesis. *Neurodegenerative Diseases*, *11*(2), 79–92. doi:10.1159/000341998 PMID:23038420

Keller, J. N., Schmitt, F. A., Scheff, S. W., Ding, Q., Chen, Q., Butterfield, D. A., & Markesbery, W. R. (2005). Evidence of increased oxidative damage in subjects with mild cognitive impairment. *Neurology*, *64*(7), 1152–1156. doi:10.1212/01. WNL.0000156156.13641.BA PMID:15824339

Kelly, E. J., Quaife, C. J., Froelick, G. J., & Palmiter, R. D. (1996). Metallothione in I and II protect against zinc deficiency and zinc toxicity in mice. *The Journal of Nutrition*, *126*(7), 1782–1790. PMID:8683339

Kelman, C., Ramakrishnan, V., Davies, A., & Holloway, K. (2010). Analysis of stereotactic accuracy of the cosmanrobert-wells frame and nexframe frameless systems in deep brain stimulation surgery. *Stereotactic and Functional Neurosurgery*, 88(5), 288–295. doi:10.1159/000316761 PMID:20588080

Kenny, E. R., Burton, E. J., & O'Brien, J. T. (2008). A volumetric magnetic resonance imaging study of entorhinal cortex volume in dementia with Lewy bodies. *Dementia and Geriatric Cognitive Disorders*, 26(3), 218–225. doi:10.1159/000153432 PMID:18781072

Kepe, V., Bordelon, Y., & Boxer, A. (2013). PET imaging of neuropathology in tauopathies: Progressive supranuclear palsy. *Journal of Alzheimer's Disease*, *36*(1), 145–153. PMID:23579330

Kerbeshian, J., Burd, L., & Avery, K. (2011). Pharmacotherapy of autism: A review and clinical approach. *Journal of Developmental and Physical Disabilities*, *13*(3), 199–228. doi:10.1023/A:1016686802786

Kerman, A., Liu, H. N., Croul, S., Bilbao, J., Rogaeva, E., Zinman, L., ... Chakrabartty, A. (2010). Amyotrophic lateral sclerosis is a non-amyloid disease in which extensive misfolding of SOD1 is unique to the familial form. *Acta Neuropathologica*, *119*(3), 335–344. doi:10.100700401-010-0646-5 PMID:20111867

Kern, J. K., Geier, D. A., Sykes, C. K., & Geier, M. R. (2013). Evidences of neurodegeneration in autisum spectrum disorder. *Translational Neurodegeneration*, 2(1), 17. doi:10.1186/2047-9158-2-17 PMID:23925007

Kern, J. K., & Jones, A. M. (2006). Evidence of toxicity, oxidative stress, and neuronal insult in autism. *Journal of Toxicology and Environmental Health. Part B, Critical Reviews*, 9(6), 485–499. doi:10.1080/10937400600882079 PMID:17090484

Khandelwal, P. J., Herman, A. M., Hoe, H. S., Rebeck, G. W., & Moussa, C. E. H. (2011). Parkin mediates beclindependent autophagic clearance of defective mitochondria and ubiquitinated Aβ in AD models. *Human Molecular Genetics*, 20(11), 2091–2102. doi:10.1093/hmg/ddr091 PMID:21378096

Khan, S. M., Cassarino, D. S., Abramova, N. N., Keeney, P. M., Borland, M. K., Trimmer, P. A., ... Westaway, D. (2005). Hypersensitivity of DJ-1-deficient mice to 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyrindine (MPTP) and oxidative stress. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(14), 5215–5220. doi:10.1073/ pnas.0501282102 PMID:15784737

Khan, S., Gill, S. S., Mooney, L., White, P., Whone, A., Brooks, D. J., & Pavese, N. (2010). Combined pedunculopontinesubthalamic stimulation in Parkinson disease. *Neurology*, 78(14), 1090–1095. doi:10.1212/WNL.0b013e31824e8e96 PMID:22402859

Khan, T. A., Hassan, I., Ahmad, A., Perveeen, A., Aman, S., Quddusi, S., ... Aliev, G. (2016). Recent updates on the dynamic association between oxidative stress and neurodegenerative disorders. *CNS & Neurological Disorders - Drug Targets*, *15*, 310–320. doi:10.2174/1871527315666160202124518 PMID:26831262

Kieburtz, K. (2011). Twice-daily, low-dose pramipexole in early Parkinson's disease: A randomized, placebo-controlled trial. *Movement Disorders*, 26(1), 37–44. doi:10.1002/mds.23396 PMID:20925067

Kiernan, M. C., Vucic, S., Cheah, B. C., Turner, M. R., Eisen, A., Hardiman, O., ... Zoing, M. C. (2011). Amyotrophic lateral sclerosis. *Lancet*, *377*(9769), 942–955. doi:10.1016/S0140-6736(10)61156-7 PMID:21296405

Killiany, R., Hyman, B., Gomez-Isla, T., Moss, M., Kikinis, R., Jolesz, F., ... Albert, M. (2002). MRI measures of entorhinal cortex vs hippocampus in preclinical AD. *Neurology*, 58(8), 1188–1196. doi:10.1212/WNL.58.8.1188 PMID:11971085

Kim, B., Yang, M. S., Choi, D., Kim, J. H., Kim, H. S., Seol, W., ... Joe, E. H. (2012). Impaired inflammatory responses in murine Lrrk2-knockdown brain microglia. *PLoS One*, 7(4), e34693. doi:10.1371/journal.pone.0034693 PMID:22496842

Kim, C., Ho, D. H., Suk, J. E., You, S., Michael, S., Kang, J., ... Lee, S. J. (2013). Neuron-released oligomeric α-synuclein is an endogenous agonist of TLR2 for paracrine activation of microglia. *Nature Communications*, *4*, 1562. doi:10.1038/ ncomms2534 PMID:23463005

Kim, I., Rodriguez-Enriquez, S., & Lemasters, J. J. (2007). Selective degradation of mitochondria by mitophagy. *Archives of Biochemistry and Biophysics*, 462(2), 245–253. doi:10.1016/j.abb.2007.03.034 PMID:17475204

Kim, J., Wigram, T. C., & Gold, C. (2008). The effects of improvisational music therapy on joint attention behaviors in autistic children: A randomized controlled study. *Journal of Autism and Developmental Disorders*, *38*(9), 1758–1766. doi:10.100710803-008-0566-6 PMID:18592368

Kim, N.-H., Park, S.-J., Jin, J.-K., Kwon, M.-S., Choi, E.-K., Carp, R. I., & Kim, Y.-S. (2000). Increased ferric iron content and iron-induced oxidative stress in the brains of scrapie-infected mice. *Brain Research*, 884(1), 98–103. doi:10.1016/S0006-8993(00)02907-3 PMID:11082491

Kim, T. D., Choi, E., Rhim, H., Paik, S. R., & Yang, C. H. (2004). α-synuclein has structural and functional similarities to small heat shock proteins. *Biochemical and Biophysical Research Communications*, *324*(4), 1352–1359. doi:10.1016/j. bbrc.2004.09.208 PMID:15504363

Kim, Y. E., Hipp, M. S., Bracher, A., Hayer-Hartl, M., & Ulrich Hartl, F. (2013). Molecular chaperone functions in protein folding and proteostasis. *Annual Review of Biochemistry*, 82(1), 323–355. doi:10.1146/annurev-biochem-060208-092442 PMID:23746257

Kim, Y. S., Leventhal, B. L., Koh, Y. J., Fombonne, E., Laska, E., Lim, E. C., ... Grinker, R. R. (2011). Prevalence of autism spectrum disorders in a total population sample. *The American Journal of Psychiatry*, *168*(9), 904–912. doi:10.1176/appi.ajp.2011.10101532 PMID:21558103

King, M. E., Kan, H.-M., Baas, P. W., Erisir, A., Glabe, C. G., & Bloom, G. S. (2006). Tau-dependent microtubule disassembly initiated by prefibrillar β-amyloid. *The Journal of Cell Biology*, *175*(4), 541–546. doi:10.1083/jcb.200605187 PMID:17101697

Kinumi, T., Kimata, J., Taira, T., Ariga, H., & Niki, E. (2004). Cysteine-106 of DJ-1 is the most sensitive cysteine residue to hydrogen peroxide-mediated oxidation in vivo in human umbilical vein endothelial cells. *Biochemical and Biophysical Research Communications*, *317*(3), 722–728. doi:10.1016/j.bbrc.2004.03.110 PMID:15081400

Kiss, Z. H., Wilkinson, M., Krcek, J., Suchowersky, O., Hu, B., Murphy, W. F., ... Tasker, R. R. (2003, October). Is the target for thalamic deep brain stimulation the same as for thalamotomy? *Movement Disorders*, *18*(10), 1169–1175. doi:10.1002/mds.10524 PMID:14534922

Kitada, T., Asakawa, S., Hattori, N., Matsumine, H., Yamamura, Y., Minoshima, S., ... Shimizu, N. (1998). Mutations in the *Parkin* gene cause autosomal recessive juvenile Parkinsonism. *Nature*, *392*(6676), 605–608. doi:10.1038/33416 PMID:9560156

Klein, C., & Westenberger, A. (2012). Genetics of Parkinson's Disease. *Cold Spring Harbor Perspectives in Medicine*, 2(1), a008888. doi:10.1101/cshperspect.a008888 PMID:22315721

Klein, J. C., Eggers, C., Kalbe, E., Weisenbach, S., Hohmann, C., Vollmar, S., ... Hilker, R. (2010). Neurotransmitter changes in dementia with Lewy bodies and Parkinson disease dementia in vivo. *Neurology*, 74(11), 885–892. doi:10.1212/ WNL.0b013e3181d55f61 PMID:20181924

Klivenyi, P., Ferrante, R. J., Matthews, R. T., Bogdanov, M. B., Klein, A. M., Andreassen, O. A., ... Beal, M. F. (1999). Neuroprotective effects of creatine in a transgenic animal model of amyotrophic lateral sclerosis. *Nature Medicine*, *5*(3), 347–350. doi:10.1038/6568 PMID:10086395

Klivenyi, P., Siwek, D., Gardian, G., Yang, L., Starkov, A., Cleren, C., ... Beal, M. F. (2006). Mice lacking alpha-synuclein are resistant to mitochondrial toxins. *Neurobiology of Disease*, 21(3), 541–548. doi:10.1016/j.nbd.2005.08.018 PMID:16298531

Knaap, M. S., Ramesh, V., Schiffmann, R., Blaser, S., Kyllerman, M., Gholkar, A., ... Salomons, G. S. (2006). Alexander disease: Ventricular garlands and abnormalities of the medulla and spinal cord. *Neurology*, *66*(4), 494–498. doi:10.1212/01. wnl.0000198770.80743.37 PMID:16505300

Knudsen, K. A., Rosand, J., Karluk, D., & Greenberg, S. M. (2001). Clinical diagnosis of cerebral amyloid angiopathy: Validation of the Boston criteria. *Neurology*, *56*(4), 537–539. doi:10.1212/WNL.56.4.537 PMID:11222803

Knutson, B., Adams, C. M., Fong, G. W., & Hommer, D. (2001). Anticipation of increasing monetary reward selectively recruits nucleus accumbens. *The Journal of Neuroscience*, *21*, RC159. PMID:11459880

Koellhoffer, E., McCullough, L., & Ritzel, R. (2017). Old maids: Aging and its impact on microglia function. *International Journal of Molecular Sciences*, *18*(4), 769. doi:10.3390/ijms18040769 PMID:28379162

Koh, J. Y. (2005). Endogenous zinc in neurological diseases. *Journal of Clinical Neurology (Seoul, Korea)*, 1(2), 121–133. doi:10.3988/jcn.2005.1.2.121 PMID:20396459

Koh, J. Y., Suh, S. W., Gwag, B. J., He, Y. Y., Hsu, C. Y., & Choi, D. W. (1996). The role of zinc in selective neuronal death after transient global cerebral ischemia. *Science*, 272(5264), 1013–1016. doi:10.1126cience.272.5264.1013 PMID:8638123

Kohlschütter, A. (2013). Lysosomal leukodystrophies. Handbook of Clinical Neurology Pediatric Neurology Part III, 1611-1618.

Kolesnikova, E. É. (2013). Mitochondrial dysfunction and molecular bases of neurodegenerative diseases. *Neurophysiology*, *45*(1), 89–102. doi:10.100711062-013-9341-1

Kolevzon, A., Gross, R., & Reichenberg, A. (2007). Prenatal and perinatal risk factors for autism: A review and integration of findings. *Archives of Pediatrics & Adolescent Medicine*, *161*(4), 326–333. doi:10.1001/archpedi.161.4.326 PMID:17404128

Kong, J., & Xu, Z. (1998). Massive mitochondrial degeneration in motor neurons triggers the onset of amyotrophic lateral sclerosis in mice expressing a mutant SOD1. *The Journal of Neuroscience*, *18*(9), 3241–3250. PMID:9547233

Koppers, M., Blokhuis, A. M., Westeneng, H. J., Terpstra, M. L., Zundel, C. A., Vieira de Sa, R., ... Veldink, J. H. (2015). C9orf72 ablation in mice does not cause motor neuron degeneration or motor deficits. *Annals of Neurology*, 78(3), 426–438. doi:10.1002/ana.24453 PMID:26044557

Kordinas, V., Ioannidis, A., & Chatzipanagiotou, S. (2016). The Telomere/Telomerase System in Chronic Inflammatory Diseases. Cause or Effect? *Genes*, 7(9), 60. doi:10.3390/genes7090060 PMID:27598205

Kordower, J. H., Chu, Y., Hauser, R. A., Freeman, T. B., & Olanow, C. W. (2008). Lewy body-like pathology in long-term embryonic nigral transplants in Parkinson's disease. *Nature Medicine*, *14*(5), 504–506. doi:10.1038/nm1747 PMID:18391962

Korsmeyer, S. J., Wei, M. C., Saito, M. T., Weiler, S., Oh, K. J., & Schlesinger, P. H. (2000). Pro-apoptotic cascade activates BID, which oligomerizes BAK or BAX into pores that result in the release of cytochrome c. *Cell Death and Differentiation*, 7(12), 1166–1173. doi:10.1038j.cdd.4400783 PMID:11175253

Kosaka, K. (1990). Diffuse Lewy body disease in Japan. *Journal of Neurology*, 237(3), 197–204. doi:10.1007/BF00314594 PMID:2196340

Kosaka, K. (2014). Lewy body disease and dementia with Lewy bodies. *Proceedings of the Japan Academy. Series B, Physical and Biological Sciences*, *90*(8), 301–306. doi:10.2183/pjab.90.301 PMID:25311140

Kosaka, K., Iseki, E., Odawara, T., & Yamamoto, T. (1996). Cerebral type of Lewy body disease. *Neuropathology*, *16*(1), 32–35. doi:10.1111/j.1440-1789.1996.tb00152.x

Kosaka, K., Yoshimura, M., Ikeda, K., & Budka, H. (1983). Diffuse type of Lewy body disease: Progressive dementia with abundant cortical Lewy bodies and senile changes of varying degree--a new disease? *Clinical Neuropathology*, *3*(5), 185–192. PMID:6094067

Kovacs, G. G. (2016). Molecular pathological classification of neurodegenerative diseases: Turning towards precision medicine. *International Journal of Molecular Sciences*, *17*(2), 189. doi:10.3390/ijms17020189 PMID:26848654

Kovacs, G. G., Alafuzoff, I., Al-Sarraj, S., Arzberger, T., Bogdanovic, N., Capellari, S., ... Budka, H. (2008). Mixed brain pathologies in dementia: The BrainNet Europe consortium experience. *Dementia and Geriatric Cognitive Disorders*, 26(4), 343–350. doi:10.1159/000161560 PMID:18849605

Kovacs, G. G., Botond, G., & Budka, H. (2010). Protein coding of neurodegenerative dementias: The neuropathological basis of biomarker diagnostics. *Acta Neuropathologica*, *119*(4), 389–408. doi:10.100700401-010-0658-1 PMID:20198481

Kovacs, G. G., & Budka, H. (2008). Prion diseases: From protein to cell pathology. *American Journal of Pathology*, 172(3), 555–565. doi:10.2353/ajpath.2008.070442 PMID:18245809

Kovacs, G. G., & Budka, H. (2010). Current concepts of neuropathological diagnostics in practice: Neurodegenerative diseases. *Clinical Neuropathology*, *29*(5), 271–288. doi:10.5414/NPP29271 PMID:20860890

Kovacs, G. G., Murrell, J. R., Horvath, S., Haraszti, L., Majtenyi, K., Molnar, M. J., ... Spina, S. (2009). TARDBP variation associated with frontotemporal dementia, supranuclear gaze palsy, and chorea. *Movement Disorders*, 24(12), 1843–1847. doi:10.1002/mds.22697 PMID:19609911

Kozma, C. (1998). On cognitive variability in velocardiofacial syndrome: Profound mental retardation and autism. *American Journal of Medical Genetics*, *81*(3), 269–270. doi: PMID:9603617

Krauthammer, M., Kaufmann, C. A., Gilliam, T. C., & Rzhetsky, A. (2004). Molecular triangulation: Bridging linkage and molecular-network information for identifying candidate genes in Alzheimer's disease. *Proceedings of the National Academy of Sciences of the United States of America*, 101(42), 15148–15153. doi:10.1073/pnas.0404315101 PMID:15471992

Kraytsberg, Y., Kudryavtseva, E., McKee, A. C., Geula, C., Kowall, N. W., & Khrapko, K. (2006). Mitochondrial DNA deletions are abundant and cause functional impairment in aged human substantia nigra neurons. *Nature Genetics*, *38*(5), 518–520. doi:10.1038/ng1778 PMID:16604072

Kreitzer, A. C., & Malenka, R. C. (2008). Striatal plasticity and basal ganglia circuit function. *Neuron*, *60*(4), 543–554. doi:10.1016/j.neuron.2008.11.005 PMID:19038213

Kress, Y., Gaskin, F., Brosnan, C. F., & Levine, S. (1981). Effects of zinc on the cytoskeletal proteins in the central nervous system of the rat. *Brain Research*, 220(1), 139–149. doi:10.1016/0006-8993(81)90217-1 PMID:6974032

Krishnamurthi, R. V., Feigin, V. L., Forouzanfar, M. H., Mensah, G. A., Connor, M., Bennett, D. A., ... Murray, C. (2013). Global and regional burden of first-ever ischaemic and haemorrhagic stroke during 1990-2010: Findings from the Global Burden of Disease Study 2010. *The Lancet. Global Health*, *1*(5), e259–e281. doi:10.1016/S2214-109X(13)70089-5 PMID:25104492

Krishnaswamy, S., Verdile, G., Groth, D., Kanyenda, L., & Martins, R. N. (2009). The structure and function of Alzheimer's gamma secretase enzyme complex. *Critical Reviews in Clinical Laboratory Sciences*, *46*(5–6), 282–301. doi:10.3109/10408360903335821 PMID:19958215

Krüger, R., Kuhn, W., Müller, T., Woitalla, D., Graeber, M., Kösel, S., ... Riess, O. (1998). AlaSOPro mutation in the gene encoding α-synuclein in Parkinson's disease. *Nature Genetics*, *18*(2), 106–108. doi:10.1038/ng0298-106 PMID:9462735

Kuchino, Y., Mori, F., Kasai, H., Inoue, H., Iwai, S., Miura, K., ... Nishimura, S. (1987). Misreading of DNA templates containing 8-hydroxydeoxyguanosine at the modified base and at adjacent residues. *Nature*, *327*(6117), 77–79. doi:10.1038/327077a0 PMID:3574469

Kudo, M., Brem, M. S., & Canfield, W. M. (2006). Mucolipidosis II (I-Cell Disease) and Mucolipidosis IIIA (Classical Pseudo-Hurler Polydystrophy) Are Caused by Mutations in the GlcNAc-Phosphotransferase α/β –Subunits Precursor Gene. *American Journal of Human Genetics*, 78(3), 451–463. doi:10.1086/500849 PMID:16465621

Kuhn, P.-H., Wang, H., Dislich, B., Colombo, A., Zeitschel, U., Ellwart, J. W., ... Lichtenthaler, S. F. (2010). ADAM10 is the physiologically relevant, constitutive α -secretase of the amyloid precursor protein in primary neurons. *The EMBO Journal*, 29(17), 3020–3032. doi:10.1038/emboj.2010.167 PMID:20676056

Kumar, A., & Bansal, A. (2017). Integrated bioinformatics analysis of differentially expressed genes (DEGS) of Alzheimer's disease (AD) datasets from gene expression omnibus (GEO). *Alzheimer's & Dementia: The Journal of the Alzheimer's Association*, *13*(7), 953. doi:10.1016/j.jalz.2017.06.1270

Kumar, A., & Singh, A. (2015). A review on mitochondrial restorative mechanism of antioxidants in Alzheimer's disease and other neurological conditions. *Frontiers in Pharmacology*, 6. PMID:26441662

Kumar, A., & Singh, T. R. (2017). A New Decision Tree to Solve the Puzzle of Alzheimer's Disease Pathogenesis Through Standard Diagnosis Scoring System. *Interdisciplinary Sciences, Computational Life Sciences, 9*(1), 107–115. doi:10.100712539-016-0144-0 PMID:26792126

Kumar, H., Lim, H.-W., More, S. V., Kim, B.-W., Koppula, S., Kim, I. S., & Choi, D.-K. (2012). The Role of Free Radicals in the Aging Brain and Parkinson's Disease: Convergence and Parallelism. *International Journal of Molecular Sciences*, *13*(8), 10478–10504. doi:10.3390/ijms130810478 PMID:22949875

Kumar, K. R., Djarmati-Westenberger, A., & Grunewald, A. (2011). Genetics of Parkinson's disease. *Seminars in Neurology*, *31*(5), 433–440. doi:10.1055-0031-1299782 PMID:22266881

Kumar, R., Lozano, A. M., Montgomery, E., & Lang, A. E. (1998). Pallidotomy and deep brain stimulation of the pallidum and subthalamic nucleus in advanced Parkinson's disease. *Movement Disorders*, *13*(Suppl 1), 73–82. PMID:9613722

Kumar, V., Bansal, A., & Chauhan, R. S. (2017). Modular Design of Picroside-II Biosynthesis Deciphered through NGS Transcriptomes and Metabolic Intermediates Analysis in Naturally Variant Chemotypes of a Medicinal Herb, Picrorhiza kurroa. *Frontiers in Plant Science*, 8. doi:10.3389/fpls.2017.00564 PMID:28443130

Kunikowska, G., & Jenner, P. (2001). 6-Hydroxydopamine-lesioning of the nigrostriatal pathway in rats alters basal ganglia mRNA for copper, zinc-and manganese-superoxide dismutase, but not glutathione peroxidase. *Brain Research*, *922*(1), 51–64. doi:10.1016/S0006-8993(01)03149-3 PMID:11730701

Kuo, H.-K., & Lipsitz, L. A. (2004). Cerebral white matter changes and geriatric syndromes: Is there a link? *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, *59*(8), M818–M826. doi:10.1093/gerona/59.8.M818 PMID:15345732

Kupsch, A., Kuehn, A., Klaffke, S., Meissner, W., Harnack, D., Winter, C., ... Trottenberg, T. (2003). Deep brain stimulation in dystonia. *Journal of Neurology*, 250(S1Suppl 1), I47–I52. doi:10.100700415-003-1110-2 PMID:12761637

Kurland, L. T. (1988). Amyotrophic lateral sclerosis and Parkinson's disease complex on Guam linked to an environmental neurotoxin. *Trends in Neurosciences*, *11*(2), 51–54. doi:10.1016/0166-2236(88)90163-4 PMID:2465598

Kuroda, Y., Mitsui, T., Kunishige, M., Shono, M., Akaike, M., Azuma, H., & Matsumoto, T. (2006). Parkin enhances mitochondrial biogenesis in proliferating cells. *Human Molecular Genetics*, *15*(6), 883–895. doi:10.1093/hmg/ddl006 PMID:16449237

Kurth, M. C., Adler, C. H., Hilaire, M. S., Singer, C., Waters, C., LeWitt, P., ... Yoo, K. (1997). Tolcapone Improves Motor Function and Reduces Levodopa Requirement in Patients with Parkinson's Disease Experiencing Motor Fluctuations A Multicenter, Double-Blind, Randomized, Placebo-Controlled Trial. *Neurology*, *48*(1), 81–87. doi:10.1212/ WNL.48.1.81 PMID:9008498

Kurtz, A. L., & Kaufer, D. I. (2011). Dementia in Parkinson's disease. *Current Treatment Options in Neurology*, *13*(3), 242–254. doi:10.100711940-011-0121-1 PMID:21461668

Kuther, G., & Struppler, A. (1987). Therapeutic trial with N-acetylcysteine in amyotrophic lateral sclerosis. *Advances in Experimental Medicine and Biology*, 209, 281–284. PMID:3577918

Kuusisto, E., Parkkinen, L., & Alafuzoff, I. (2003). Morphogenesis of Lewy bodies: Dissimilar incorporation of α-synuclein, ubiquitin, and p62. *Journal of Neuropathology and Experimental Neurology*, 62(12), 1241–1253. doi:10.1093/jnen/62.12.1241 PMID:14692700

Kuwajima, M., Dehoff, M. H., Furuichi, T., Worley, P. F., Hall, R. A., & Smith, Y. (2007). Localization and expression of group I metabotropic glutamate receptors in the mouse striatum, globus pallidus, and subthalamic nucleus: Regulatory effects of MPTP treatment and constitutive Homer deletion. *The Journal of Neuroscience*, *27*(23), 6249–6260. doi:10.1523/JNEUROSCI.3819-06.2007 PMID:17553998

Kuwana, T., Mackey, M. R., Perkins, G., Ellisman, M. H., Latterich, M., Schneiter, R., ... Newmeyer, D. D. (2002). Bid, Bax, and lipids cooperate to form supramolecular openings in the outer mitochondrial membrane. *Cell*, *111*(3), 331–342. doi:10.1016/S0092-8674(02)01036-X PMID:12419244

Kwan, P., Arzimanoglou, A., Berg, A. T., Brodie, M. J., Allen Hauser, W., Mathern, G., ... French, J. (2010). Definition of drug resistant epilepsy: Consensus proposal by the ad hoc Task Force of the ILAE Commission on Therapeutic Strategies. *Epilepsia*, *51*(6), 1069–1077. doi:10.1111/j.1528-1167.2009.02397.x PMID:19889013

Kwiatkowski, T. J., Bosco, D. A., Leclerc, A. L., Tamrazian, E., Vanderburg, C. R., Russ, C., & Valdmanis, P. (2009). Mutations in the FUS/TLS gene on chromosome 16 cause familial amyotrophic lateral sclerosis. *Science*, *323*(5918), 1205–1208. doi:10.1126cience.1166066 PMID:19251627

Kwon, H. M., Lynn, M. J., Turan, T. N., Derdeyn, C. P., Fiorella, D., Lane, B. F., ... Chimowitz, M. I. (2016). Frequency, Risk Factors, and Outcome of Coexistent Small Vessel Disease and Intracranial Arterial Stenosis: Results From the Stenting and Aggressive Medical Management for Preventing Recurrent Stroke in Intracranial Stenosis (SAMMPRIS) Trial. *JAMA Neurology*, *73*(1), 36–42. doi:10.1001/jamaneurol.2015.3145 PMID:26618534

Kwon, I., Xiang, S., Kato, M., Wu, L., Theodoropoulos, P., Wang, T., ... McKnight, S. L. (2014). Poly-dipeptides encoded by the C9orf72 repeats bind nucleoli, impede RNA biogenesis, and kill cells. *Science*, *345*(6201), 1139–1145. doi:10.1126cience.1254917 PMID:25081482

Kwon, M. J. (2016). Neurodegenerative disorders of childhood. In R. Kliegman, B. Stanton, J. W. St. Geme, N. F. Schor, & R. E. Behrman (Eds.), *Nelson text book of Pediatrics* (20th ed.; pp. 2910–2918). Philadelphia, PA: Elsevier.

Lacomblez, L., Bensimon, G., Leigh, P. N., Guillet, P., Powe, L., Durrleman, S., ... Meininger, V. (1996). A confirmatory dose ranging study of riluzole in ALS. *Neurology*, 47(4), S242–S250. doi:10.1212/WNL.47.6_Suppl_4.242S PMID:8959996

Lagier-Tourenne, C., Baughn, M., Rigo, F., Sun, S., Liu, P., Li, H. R., ... Katz, M. (2013). Targeted degradation of sense and antisense C9orf72 RNA foci as therapy for ALS and frontotemporal degeneration. *Proceedings of the National Academy of Sciences*, *110*, E4530–E4539.

Lagier-Tourenne, C., Polymenidou, M., & Cleveland, D. W. (2010). TDP-43 and FUS/TLS: Emerging roles in RNA processing and neurodegeneration. *Human Molecular Genetics*, 19(R1), R46–R64. doi:10.1093/hmg/ddq137 PMID:20400460

Lagier-Tourenne, C., Polymenidou, M., Hutt, K. R., Vu, A. Q., Baughn, M., Huelga, S. C., ... Yeo, G. W. (2012). Divergent roles of ALS-linked proteins FUS/TLS and TDP-43 intersect in processing long pre-mRNAs. *Nature Neuroscience*, *15*(11), 1488–1497. doi:10.1038/nn.3230 PMID:23023293

Lagouge, M., & Larsson, N. G. (2013). The role of mitochondrial DNA mutations and free radicals in disease and ageing. *Journal of Internal Medicine*, 273(6), 529–543. doi:10.1111/joim.12055 PMID:23432181

Laitinen, L. V., Bergenheim, A. T., & Hariz, M. I. (1992, January). Leksell's posteroventral pallidotomy in the treatment of Parkinson's disease. *Journal of Neurosurgery*, 76(1), 53–61. doi:10.3171/jns.1992.76.1.0053 PMID:1727169

Lam, P. Y., Yin, F., Hamilton, R. T., Boveris, A., & Cadenas, E. (2009). Elevated neuronal nitric oxide synthase expression during ageing and mitochondrial energy production. *Free Radical Research*, *43*(5), 431–439. doi:10.1080/10715760902849813 PMID:19347761

Lang, A. E., Duff, J., Saint-Cyr, J. A., Trepanier, L., Gross, R. E., Lombardi, W., ... Lozano, A. M. (1999, September). Posteroventral medial pallidotomy in Parkinson's disease. *Journal of Neurology*, *246*(S2Suppl 2), II28–II41. doi:10.1007/BF03161079 PMID:10526000

Lang, A. E., Lozano, A. M., Montgomery, E., Duff, J., Tasker, R., & Hutchinson, W. (1997). Posteroventral medial pallidotomy in advanced Parkinson's disease. *The New England Journal of Medicine*, *337*(15), 1036–1042. doi:10.1056/ NEJM199710093371503 PMID:9321531

Lang, C. J., Heckmann, J. G., & Neundörfer, B. (1998). Creutzfeldt-Jakob disease via dural and corneal transplants. *Journal of the Neurological Sciences*, *160*(2), 128–139. doi:10.1016/S0022-510X(98)00226-3 PMID:9849795

Langston, J. W., Ballard, P., Tetrud, J. W., & Irwin, I. (1983). Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. *Science*, *219*(4587), 979–980. doi:10.1126cience.6823561 PMID:6823561

Lansbury, P. T., & Lashuel, H. A. (2006). A century-old debate on protein aggregation and neurodegeneration enters the clinic. *Nature*, 443(7113), 774–779. doi:10.1038/nature05290 PMID:17051203

Larson, P. S. (2014). Deep brain stimulation for movement disorders. *Neurotherapeutics; the Journal of the American Society for Experimental NeuroTherapeutics*, *11*(3), 465–474. doi:10.100713311-014-0274-1 PMID:24833244

Larsson, N. G. (2010). Somatic mitochondrial DNA mutations in mammalian aging. *Annual Review of Biochemistry*, 79(1), 683–706. doi:10.1146/annurev-biochem-060408-093701 PMID:20350166

Larsson, N. G., Wang, J., Wilhelmsson, H., Oldfors, A., Rustin, P., Lewandoski, M., & Clayton, D. A. (1998). Mitochondrial transcription factor A is necessary for mtDNA maintenance and embryogenesis in mice. *Nature Genetics*, *18*(3), 231–236. doi:10.1038/ng0398-231 PMID:9500544

Lau, A., & Tymianski, M. (2010). Glutamate receptors, neurotoxicity and neurodegeneration. *Pflügers Archiv*, 460(2), 525–542. doi:10.100700424-010-0809-1 PMID:20229265

Lau, F. C., Shukitt-Hale, B., & Joseph, J. A. (2007). Nutritional intervention in brain aging: Reducing the effects of inflammation and oxidative stress. *Sub-Cellular Biochemistry*, 42, 299–318. doi:10.1007/1-4020-5688-5_14 PMID:17612057

Launer, L. J., Hughes, T. M., & White, L. R. (2011). Microinfarcts, brain atrophy, and cognitive function: The Honolulu Asia Aging Study Autopsy Study. *Annals of Neurology*, 70(5), 774–780. doi:10.1002/ana.22520 PMID:22162060

Laura, W., Heather, M. C., & David, A. H. (2007). The cellular prion protein (PrPC): Its physiological function and role in disease. *Biochimica et Biophysica Acta*, 1772(6), 629–644. doi:10.1016/j.bbadis.2007.02.011 PMID:17451912

Lawrence, J., Parmenter, T., & McDonald, T. (2013). Failure to manage constipation in Parkinson's disease. A review of medical services: A patients perspective. *Movement Disorders*, 28(1), 187.

Lazarov, O., Robinson, J., Tang, Y.-P., Hairston, I. S., Korade-Mirnics, Z., Lee, V. M.-Y., ... Sisodia, S. S. (2005). Environmental enrichment reduces Aβ levels and amyloid deposition in transgenic mice. *Cell*, *120*(5), 701–713. doi:10.1016/j. cell.2005.01.015 PMID:15766532

Lederer, C. W., Torrisi, A., Pantelidou, M., Santama, N., & Cavallaro, S. (2007). Pathways and genes differentially expressed in the motor cortex of patients with sporadic amyotrophic lateral sclerosis. *BMC Genomics*, 8(1), 26. doi:10.1186/1471-2164-8-26 PMID:17244347

Lee, C., & Yu, M. H. (2005). Protein folding and diseases. *Journal of Biochemistry and Molecular Biology*, 38(3), 275–280. PMID:15943901

Lee, D. W., & Andersen, J. K. (2006). Role of HIF-1 in iron regulation: Potential therapeutic strategy for neurodegenerative disorders. *Current Molecular Medicine*, 6(8), 883–893. doi:10.2174/156652406779010849 PMID:17168739

Lee, D.-H., Gold, R., & Linker, R. A. (2012). Mechanisms of oxidative damage in multiple sclerosis and neurodegenerative diseases: Therapeutic modulation via fumaric acid esters. *International Journal of Molecular Sciences*, *13*(9), 11783–11803. doi:10.3390/ijms130911783 PMID:23109883

Lee, G., Newman, S. T., Gard, D. L., Band, H., & Panchamoorthy, G. (1998). Tau interacts with src-family non-receptor tyrosine kinases. *Journal of Cell Science*, *111*(Pt 21), 3167–3177. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/9763511 PMID:9763511

Lee, H. J., Suk, J. E., Lee, K. W., Park, S. H., Blumbergs, P. C., Gai, W. P., & Lee, S. J. (2011). Transmission of synucleinopathies in the enteric nervous system of A53T alpha-synuclein transgenic mice. *Experimental Neurobiology*, 20(4), 181–188. doi:10.5607/en.2011.20.4.181 PMID:22355263

Lee, H., Zhu, X., Castellani, R. J., Nunomura, A., Perry, G., & Smith, M. A. (2007). Amyloid-beta in Alzheimer disease: The null versus the alternate hypotheses. *The Journal of Pharmacology and Experimental Therapeutics*, *321*(3), 823–829. doi:10.1124/jpet.106.114009 PMID:17229880

Lee, J. M., Grabb, M. C., Zipfel, G. J., & Choi, D. W. (2000). Brain tissue responses to ischemia. *The Journal of Clinical Investigation*, *106*(6), 723–731. doi:10.1172/JCI11003 PMID:10995780

Lee, J. S., & Lee, S. J. (2016). Mechanism of Anti-α-Synuclein Immunotherapy. *Journal of Movement Disorders*, 9(1), 14–19. doi:10.14802/jmd.15059 PMID:26828212

Lee, J., Kim, C. H., Kim, D. G., & Ahn, Y. S. (2009). Zinc inhibits amyloid beta production from Alzheimer's amyloid precursor protein in SH-SY5Y Cells. *The Korean Journal of Physiology & Pharmacology; Official Journal of the Korean Physiological Society and the Korean Society of Pharmacology, 13*(3), 195–200. doi:10.4196/kjpp.2009.13.3.195 PMID:19885037

Lee, J., Kosaras, B., Del Signore, S. J., Cormier, K., McKee, A., Ratan, R. R., ... Ryu, H. (2011). Modulation of lipid peroxidation and mitochondrial function improves neuropathology in Huntington's disease mice. *Acta Neuropathologica*, *121*(4), 487–498. doi:10.100700401-010-0788-5 PMID:21161248

Lee, M., Kwon, Y. T., Li, M., Peng, J., Friedlander, R. M., & Tsai, L.-H. (2000). Neurotoxicity induces cleavage of p35 to p25 by calpain. *Nature*, *405*(6784), 360–364. doi:10.1038/35012636 PMID:10830966

Lee, S. E., Rabinovici, G. D., Mayo, M. C., Wilson, S. M., Seeley, W. W., DeArmond, S. J., ... Miller, B. L. (2011). Clinicopathological correlations in corticobasal degeneration. *Annals of Neurology*, *70*(2), 327–340. doi:10.1002/ana.22424 PMID:21823158

Lee, S. J., Kim, S. J., Kim, I. K., Ko, J., Jeong, C. S., Kim, G. H., & Cha, S. S. (2003). Crystal structures of human DJ-1 and Escherichia coli Hsp31, which share an evolutionarily conserved domain. *The Journal of Biological Chemistry*, 278(45), 44552–44559. doi:10.1074/jbc.M304517200 PMID:12939276

Lees, A. J., Hardy, J., & Revesz, T. (2009). Parkinson's disease. *Lancet*, 373(9680), 2055–2066. doi:10.1016/S0140-6736(09)60492-X PMID:19524782

Lee, V. M., Giasson, B. I., & Trojanowski, J. Q. (2004). More than just two peas in a pod: Common amyloidogenic properties of tau and α -synuclein in neurodegenerative diseases. *Trends in Neurosciences*, 27(3), 129–134. doi:10.1016/j. tins.2004.01.007 PMID:15036877

Lee, V. M., Goedert, M., & Trojanowski, J. Q. (2001). Neurodegenerative tauopathies. *Annual Review of Neuroscience*, 24(1), 1121–1159. doi:10.1146/annurev.neuro.24.1.1121 PMID:11520930

Lee, V. M.-Y., Kenyon, T. K., & Trojanowski, J. Q. (2005). Transgenic animal models of tauopathies. *Biochimica et Biophysica Acta*, *1739*(2–3), 251–259. doi:10.1016/j.bbadis.2004.06.014 PMID:15615643

Leger, G. C., & Johnson, N. (2007). A review on primary progressive aphasia. *Neuropsychiatric Disease and Treatment*, *3*(6), 745–752. PMID:19300609

Lehericy, S., Baulac, M., Chiras, J., Pierot, L., Martin, N., Pillon, B., ... Marsault, C. (1994). Amygdalohippocampal MR volume measurements in the early stages of Alzheimer disease. *AJNR. American Journal of Neuroradiology*, *15*(5), 929–937. PMID:8059663

Lehmann, M., Crutch, S. J., Ridgway, G. R., Ridha, B. H., Barnes, J., Warrington, E. K., ... Fox, N. C. (2011). Cortical thickness and voxel-based morphometry in posterior cortical atrophy and typical Alzheimer's disease. *Neurobiology of Aging*, *32*(8), 1466–1476. doi:10.1016/j.neurobiolaging.2009.08.017 PMID:19781814

Leigh, P. N., Abrahams, S., Al-Chalabi, A., Ampong, M. A., Goldstein, L. H., Johnson, J., ... Rio, A. (2003). The management of motor neurone disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 74(4), 32–47. PMID:14645465

Leigh, P. N., & Ray-Chaudhuri, K. (1994). Motor neuron disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, *57*(8), 886–896. doi:10.1136/jnnp.57.8.886 PMID:8057109

Lei, P., Ayton, S., Bush, A. I., & Adlard, P. A. (2011). GSK-3 in Neurodegenerative Diseases. *International Journal of Alzheimer's Disease*, 2011, 189246. doi:10.4061/2011/189246 PMID:21629738

Lemasters, J. J. (2005). Selective mitochondrial autophagy, or mitophagy, as a targeted defense against oxidative stress, mitochondrial dysfunction, and aging. *Rejuvenation Research*, 8(1), 3–5. doi:10.1089/rej.2005.8.3 PMID:15798367

Lenaz, G. (1998). Role of mitochondria in oxidative stress and ageing. *Biochimica et Biophysica Acta*, *1366*(1-2), 53–67. doi:10.1016/S0005-2728(98)00120-0 PMID:9714734

Lenglet, C., Abosch, A., Yacoub, E., De Martino, F., Sapiro, G., & Harel, N. (2012). Comprehensive in vivo mapping of the human basal ganglia and thalamic connectome in individuals using 7T MRI. *PLoS One*, 7(1), e29153. doi:10.1371/journal.pone.0029153 PMID:22235267

Leone, P., Shera, D., McPhee, S. W. J., Francis, J. S., Kolodny, E. H., Bilaniuk, L. T., ... Janson, C. G. (2012). Long-Term Follow-Up After Gene Therapy for Canavan Disease. *Science Translational Medicine*, *4*(165).

Leone, P., Janson, C. G., McPhee, S. J., & During, M. J. (1999). Global CNS gene transfer for a childhood neurogenetic enzyme deficiency: Canavan disease. *Current Opinion in Molecular Therapeutics*, 1(4), 487–492. PMID:11713764

Leong, S. L., Cappai, R., Barnham, K. J., & Pham, C. L. (2009). Modulation of alpha-synuclein aggregation by dopamine: A review. *Neurochemical Research*, *34*(10), 1838–1846. doi:10.100711064-009-9986-8 PMID:19444607

Leroy, J. G., Cathey, S., & Friez, M. J. (2008 Aug 26). Mucolipidosis II. In M. P. Adam, H. H. Ardinger, & R. A. Pagon (Eds.), GeneReviews (pp. 1993–2017). Academic Press.

Leroy, E., Boyer, R., Auburger, G., Leube, B., Ulm, G., Mezey, E., ... Polymeropoulos, M. H. (1998). The ubiquitin pathway in Parkinson's disease. *Nature*, *395*(6701), 451–452. doi:10.1038/26652 PMID:9774100

Lesage, S., & Brice, A. (2009). Parkinson's disease: From monogenic forms to genetic susceptibility factors. *Human Molecular Genetics*, *18*(1), 48–59. doi:10.1093/hmg/ddp012 PMID:19297401

Lesage, S., Drouet, V., Majounie, E., Deramecourt, V., Jacoupy, M., Nicolas, A., ... Brice, A. (2016). Loss of VPS13C function in autosomal-recessive parkinsonism causes mitochondrial dysfunction and increases PINK1/Parkin-dependent mitophagy. *American Journal of Human Genetics*, 98(3), 500–513. doi:10.1016/j.ajhg.2016.01.014 PMID:26942284

Leszek, J., Barreto, G. E., Gasiorowski, K., Koutsouraki, E., Avila-Rodrigues, M., & Aliev, G. (2016). Inflammatory mechanisms and oxidative stress as key factors responsible for progression of neurodegeneration: Role of brain innate immune system. *CNS & Neurological Disorders - Drug Targets*, *15*(3), 329–336. doi:10.2174/1871527315666160202 125914 PMID:26831258

Leuner, K., Schütt, T., Kurz, C., Eckert, S. H., Schiller, C., Occhipinti, A., ... Palmiter, R. D. (2012). Mitochondrionderived reactive oxygen species lead to enhanced amyloid beta formation. *Antioxidants & Redox Signalling*, *16*(12), 1421–1433. doi:10.1089/ars.2011.4173 PMID:22229260

Leveille, F., El Gaamouch, F., Gouix, E., Lecocq, M., Lobner, D., Nicole, O., & Buisson, A. (2008). Neuronal viability is controlled by a functional relation between synaptic and extrasynaptic NMDA receptors. *Federation of American Society for Experimental Biology Journal*, 22(12), 4258–4271. doi:10.1096/fj.08-107268 PMID:18711223

Levenson, C. W., & Morris, D. (2011). Zinc and neurogenesis: Making new neurons from development to adulthood. *Advances in Nutrition*, 2(2), 96–100. doi:10.3945/an.110.000174 PMID:22332038

Levine, M. S., Cepeda, C., Hickey, M. A., Fleming, S. M., & Chesselet, M.-F. (2004). Genetic mouse models of Huntington's and Parkinson's diseases: Illuminating but imperfect. *Trends in Neurosciences*, 27(11), 691–697. doi:10.1016/j. tins.2004.08.008 PMID:15474170

Levin, J., Kurz, A., Arzberger, T., Giese, A., & Höglinger, G. U. (2016). The Differential Diagnosis and Treatment of Atypical Parkinsonism. *Deutsches Ärzteblatt International*, *113*(5), 61–69. PMID:26900156

Lev, N., Ickowicz, D., Melamed, E., & Offen, D. (2008). Oxidative insults induce DJ-1 upregulation and redistribution: Implications for neuroprotection. *Neurotoxicology*, 29(3), 397–405. doi:10.1016/j.neuro.2008.01.007 PMID:18377993

Lev, N., Roncevic, D., Ickowicz, D., Melamed, E., & Offen, D. (2006). Role of DJ-1 in Parkinson's disease. *Journal of Molecular Neuroscience*, 29(3), 215–225. doi:10.1385/JMN:29:3:215 PMID:17085780

Levy, G., Schupf, N., Tang, M. X., Cote, L. J., Louis, E. D., Mejia, H., ... Marder, K. (2002). Combined effect of age and severity on the risk of dementia in Parkinson's disease. *Annals of Neurology*, *51*(6), 722–729. doi:10.1002/ana.10219 PMID:12112078

Levy, S. E., & Hyman, S. L. (2003). Use of complementary and alternative treatments: For children with autistic spectrum disorders is increasing. *Pediatric Annals*, *32*(10), 685–691. doi:10.3928/0090-4481-20031001-10 PMID:14606219

Levy, S. E., & Hyman, S. L. (2008). Levy., Hyman SL. Complementary and alternative medicine treatments for children with autism spectrum disorders. *Child and Adolescent Psychiatric Clinics of North America*, *17*(4), 803–820. doi:10.1016/j. chc.2008.06.004 PMID:18775371

Levy, S. F., LeBoeuf, A. C., Massie, M. R., Jordan, M. A., Wilson, L., & Feinstein, S. C. (2005). Three- and Four-repeat Tau Regulate the Dynamic Instability of Two Distinct Microtubule Subpopulations in Qualitatively Different Manners Implications for Neurodegeneration. *The Journal of Biological Chemistry*, *280*(14), 13520–13528. doi:10.1074/jbc. M413490200 PMID:15671021

Lewis, J., Dickson, D. W., Lin, W. L., Chisholm, L., Corral, A., Jones, G., ... McGowan, E. (2001). Enhanced Neurofibrillary Degeneration in Transgenic Mice Expressing Mutant Tau and APP. *Science*, 293(5534), 1487–1491. doi:10.1126cience.1058189 PMID:11520987

Lewis, J., Melrose, H., Bumcrot, D., Hope, A., Zehr, C., Lincoln, S., ... Farrer, M. J. (2008). In vivo silencing of alphasynuclein using naked siRNA. *Molecular Neurodegeneration*, *3*(1), 19. doi:10.1186/1750-1326-3-19 PMID:18976489

LeWitt, P. A., Rezai, A. R., Leehey, M. A., Ojemann, S. G., Flaherty, A. W., Eskandar, E. N., ... Feigin, A. (2011, April). AAV2-GAD gene therapy for advanced Parkinson's disease: A double-blind, sham-surgery controlled, randomised trial. *Lancet Neurology*, *10*(4), 309–319. doi:10.1016/S1474-4422(11)70039-4 PMID:21419704

Lichtenthaler, S. F. (2012). Alpha-secretase cleavage of the amyloid precursor protein: Proteolysis regulated by signaling pathways and protein trafficking. *Current Alzheimer Research*, 9(2), 165–177. doi:10.2174/156720512799361655 PMID:21605033

Lierberman, A. (2006). Are dementia and depression in Parkinson's disease related? *Journal of the Neurological Sciences*, 248(4), 138–142. doi:10.1016/j.jns.2006.05.022 PMID:16814323

Li, F., Calingasan, N. Y., Yu, F., Mauck, W. M., Toidze, M., Almeida, C. G., ... Lin, M. T. (2004). Increased plaque burden in brains of APP mutant MnSOD heterozygous knockout mice. *Journal of Neurochemistry*, 89(5), 1308–1312. doi:10.1111/j.1471-4159.2004.02455.x PMID:15147524

Li, J. Y., Englund, E., Holton, J. L., Soulet, D., Hagell, P., Lees, A. J., ... Widner, H. (2008). Lewy bodies in grafted neurons in subjects with Parkinson's disease suggest host-to-graft disease propagation. *Nature Medicine*, *14*(5), 501–503. doi:10.1038/nm1746 PMID:18391963

Li, J.-Q., Tan, L., & Yu, J.-T. (2014). The role of the LRRK2 gene in Parkinsonism. *Molecular Neurodegeneration*, 9(1), 47. doi:10.1186/1750-1326-9-47 PMID:25391693

Lilly. (2017). Clinical Development Pipeline. Retrieved March 7, 2017, from https://www.lilly.com/pipeline/index.html

Lim, J., & Yue, Z. (2015). Neuronal aggregates: Formation, clearance and spreading. *Developmental Cell*, *32*(4), 491–501. doi:10.1016/j.devcel.2015.02.002 PMID:25710535

Limviphuvadh, V., Tanaka, S., Goto, S., Ueda, K., & Kanehisa, M. (2007). The commonality of protein interaction networks determined in neurodegenerative disorders (NDDs). *Bioinformatics (Oxford, England)*, 23(16), 2129–2138. doi:10.1093/bioinformatics/btm307 PMID:17553855

Lin, C. H., Wu, R. M., Chang, H. Y., Chiang, Y. T., & Lin, H. H. (2013). Preceding pain symptoms and Parkinson's disease: A nationwide population-based cohort study. *European Journal of Neurology*, *20*(10), 1398–1404. doi:10.1111/ ene.12197 PMID:23679105

Lindquist, S., Krobitsch, S., Li, L., & Sondheimer, N. (2001). Investigating protein conformation–based inheritance and disease in yeast. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, *356*(1406), 169–176. doi:10.1098/rstb.2000.0762 PMID:11260797

Lindsay, J., Laurin, D., & Verreault, R. (2002). Risk factors for Alzheimer's disease: A prospective analysis from the Canadian Study of Health and Aging. *American Journal of Epidemiology*, *156*(5), 445–453. doi:10.1093/aje/kwf074 PMID:12196314

Lindström, V., Fagerqvist, T., Nordström, E., Eriksson, F., Lord, A., Tucker, S., ... Ingelsson, M. (2014). Immunotherapy targeting α-synuclein protofibrils reduced pathology in (Thy-1)-h[A30P] α-synuclein mice. *Neurobiology of Disease*, *69*, 134–143. doi:10.1016/j.nbd.2014.05.009 PMID:24851801

Ling, H. (2016). Clinical Approach to Progressive Supranuclear Palsy. *Journal of Movement Disorders*, 9(1), 3–13. doi:10.14802/jmd.15060 PMID:26828211

Ling, S. C., Polymenidou, M., & Cleveland, D. W. (2013). Converging mechanisms in ALS and FTD: Disrupted RNA and protein homeostasis. *Neuron*, 79(3), 416–438. doi:10.1016/j.neuron.2013.07.033 PMID:23931993

Linhares, M. N., & Tasker, R. R. (2000, February). Microelectrode-guided thalamotomy for Parkinson's disease. *Neurosurgery*, 46(2), 390–395. doi:10.1097/00006123-200002000-00024 PMID:10690728

Lin, M. T., & Beal, M. F. (2006). Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature*, 443(7113), 787–795. doi:10.1038/nature05292 PMID:17051205

Lin, M. T., Simon, D. K., Ahn, C. H., Kim, L. M., & Beal, M. F. (2002). High aggregate burden of somatic mtDNA point mutations in aging and Alzheimer's disease brain. *Human Molecular Genetics*, *11*(2), 133–145. doi:10.1093/hmg/11.2.133 PMID:11809722

Linn, J., Halpin, A., Demaerel, P., Ruhland, J., Giese, A. D., Dichgans, M., ... Greenberg, S. M. (2010). Prevalence of superficial siderosis in patients with cerebral amyloid angiopathy. *Neurology*, 74(17), 1346–1350. doi:10.1212/WNL.0b013e3181dad605 PMID:20421578

Lin, Y., Bloodgood, B. L., Hauser, J. L., Lapan, A. D., Koon, A. C., Kim, T. K., ... Greenberg, M. E. (2008). Activity dependent regulation of inhibitory synapse development by Npas4. *Nature*, 455(7217), 1198–1204. doi:10.1038/ nature07319 PMID:18815592

Liou, Y.-C., Sun, A., Ryo, A., Zhou, X. Z., Yu, Z.-X., Huang, H.-K., ... Lu, K. P. (2003). Role of the prolyl isomerase Pin1 in protecting against age-dependent neurodegeneration. *Nature*, 424(6948), 556–561. doi:10.1038/nature01832 PMID:12891359

Lippa, C. F., Duda, J. E., Grossman, M., Hurtig, H. I., Aarsland, D., Boeve, B. F., ... Wszolek, Z. K. (2007). DLB and PDD boundary issues: Diagnosis, treatment, molecular pathology, and biomarkers. *Neurology*, *68*(11), 812–819. doi:10.1212/01.wnl.0000256715.13907.d3 PMID:17353469

Lipsman, N., Schwartz, M. L., Huang, Y., Lee, L., Sankar, T., Chapman, M., ... Lozano, A. M. (2013, May). MR-guided focused ultrasound thalamotomy for essential tremor: A proof-of-concept study. *Lancet Neurology*, *12*(5), 462–468. doi:10.1016/S1474-4422(13)70048-6 PMID:23523144

Li, R., Messing, A., Goldman, J. E., & Brenner, M. (2002). GFAP mutations in Alexander disease. *International Journal of Developmental Neuroscience*, 20(3–5), 259–268. doi:10.1016/S0736-5748(02)00019-9 PMID:12175861

Little, S., Pogosyan, A., Neal, S., Zavala, B., Zrinzo, L., Hariz, M., ... Brown, P. (2013, September). Adaptive deep brain stimulation in advanced Parkinson disease. *Annals of Neurology*, 74(3), 449–457. doi:10.1002/ana.23951 PMID:23852650

516

Liu, X. B., Wang, J. A., Yu, S. P., Keogh, C. L. & Wei, L. (2008). Therapeutic strategy of erythropoietin in neurological disorders. *Central Nervous System and Neurological Disorders - Drug Targets*, *7*, 227-234.

Liu, B., Chen, Y., & Clair, D. K. S. (2008). ROS and p53: A versatile partnership. *Free Radical Biology & Medicine*, 44(8), 1529–1535. doi:10.1016/j.freeradbiomed.2008.01.011 PMID:18275858

Liu, J., Lillo, C., Jonsson, P. A., Velde, C. V., Ward, C. M., Miller, T. M., ... Andersen, P. M. (2004). Toxicity of familial ALS-linked SOD1 mutants from selective recruitment to spinal mitochondria. *Neuron*, 43(1), 5–17. doi:10.1016/j. neuron.2004.06.016 PMID:15233913

Liu, Y., Fallon, L., Lashuel, H. A., Liu, Z., & Lansbury, P. T. Jr. (2002). The UCHL1 gene encodes two opposing enzymatic activities that affect alpha-synuclein degradation and Parkinson's disease susceptibility. *Cell*, *111*(2), 209–218. doi:10.1016/S0092-8674(02)01012-7 PMID:12408865

Li, X. J., & Li, S. (2011). Proteasomal dysfunction in aging and Huntington disease. *Neurobiology of Disease*, 43(1), 4–8. doi:10.1016/j.nbd.2010.11.018 PMID:21145396

Li, X., Chauhan, A., Sheikh, A. M., Patil, S., Chauhan, V., Li, X. M., ... Malik, M. (2009). Elevated immune response in the brain of autistic patients. *Journal of Neuroimmunology*, 207(1-2), 111–116. doi:10.1016/j.jneuroim.2008.12.002 PMID:19157572

Li, Y., Dunn, L., Greggio, E., Krumm, B., Jackson, G. S., Cookson, M. R., ... Deng, J. (2009). The R1441C mutation alters the folding properties of the ROC domain of LRRK2. *Biochimica et Biophysica Acta*, *1792*(12), 1194–1197. doi:10.1016/j.bbadis.2009.09.010 PMID:19781641

Li, Y., Tomiyama, H., Sato, K., Hatano, Y., Yoshino, H., Atsumi, M., & Toda, T. (2005). Clinicogenetic study of PINK1 mutations in autosomal recessive early-onset Parkinsonism. *Neurology*, *64*(11), 1955–1957. doi:10.1212/01. WNL.0000164009.36740.4E PMID:15955953

Lobo, A., Launer, L., Fratiglioni, L., Andersen, K., Di Carlo, A., Breteler, M., ... Martinez-Lage, J. (2000). Prevalence of dementia and major subtypes in Europe: A collaborative study of population-based cohorts. *Neurology*, *54*(5), S4. PMID:10854354

Lockrow, J. P., Fortress, A. M., & Granholm, A. C. E. (2012). Age-related neurodegeneration and memory loss in Down syndrome. *Current Gerontology and Geriatrics Research*. PMID:22545043

Loerch, P. M., Lu, T., Dakin, K. A., Vann, J. M., Isaacs, A., Geula, C., ... Li, C. (2008). Evolution of the aging brain transcriptome and synaptic regulation. *PLoS One*, *3*(10), e3329. doi:10.1371/journal.pone.0003329 PMID:18830410

Loes, D. J., Peters, C., & Krivit, W. (1999, February). Globoid Cell Leukodystrophy: Distinguishing Early- Onset from Late-Onset Disease Using a Brain MR Imaging Scoring Method. *AJNR. American Journal of Neuroradiology*, 316–323. PMID:10094363

Logothetis, N. K., Pauls, J., Augath, M., Trinath, T., & Oeltermann, A. (2001). Neurophysiological investigation of the basis of the fMRI signal. *Nature*, *412*(6843), 150–157. doi:10.1038/35084005 PMID:11449264

Logroscino, G., Traynor, B. J., Hardiman, O., Chio, A., Couratier, P., Mitchell, J. D., ... Beghi, E. (2008). Descriptive epidemiology of amyotrophic lateral sclerosis: New evidence and unsolved issues. *Journal of Neurology, Neurosurgery, and Psychiatry*, *79*(1), 6–11. doi:10.1136/jnnp.2006.104828 PMID:18079297

Lolekha, P., Rasheed, A., & Yotsarawat, C. (2015). *Creutzfeldt-Jakob Disease in a Tertiary Care Hospital in Thailand:* A Case Series and Review of the Literature. Academic Press.

London, E. A., & Etzel, R. A. (2000). The environment as an etiologic factor in autism: A new direction for research. *Environmental Health Perspectives*, *108*(s3Suppl 3), 401–404. doi:10.1289/ehp.00108s3401 PMID:10852835

Lopes, M. A., Richardson, M. P., Abela, E., Rummel, C., Schindler, K., Goodfellow, M., & Terry, J. R. (2017). An optimal strategy for epilepsy surgery: Disruption of the rich-club? *PLoS Computational Biology*, *13*(8), e1005637. doi:10.1371/journal.pcbi.1005637 PMID:28817568

Lo, R. Y., Tanner, C. M., Albers, K. B., Leimpeter, A. D., Fross, R. D., Bernstein, A. L., ... Van Den Eeden, S. K. (2009). Clinical features in early Parkinson disease and survival. *Archives of Neurology*, *66*(11), 1353–1358. doi:10.1001/archneurol.2009.221 PMID:19901166

Lord, C. (1995). Follow up of two year olds referred for possible autism. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, *36*(8), 1365–1382. doi:10.1111/j.1469-7610.1995.tb01669.x PMID:8988272

Lord, C., & McGee, J. P. (2001). Educating Children With Autism. Washington, DC, USA: National Academy Press.

Lord, C., Risi, S., Lambrecht, L. E. H., Leventhal, B. L., & DiLavore, P. C. (2000). The Autism Diagnostic Observation Schedule-Generic: A standard measure of social and communication deficits associated with the spectrum of autism. *Journal of Autism and Developmental Disorders*, *30*(3), 205–223. doi:10.1023/A:1005592401947 PMID:11055457

Lord, C., Rutter, M., & Le Couteur, A. (1994). Autism Diagnostic Interview-Revised. A revised version of a diagnostic interview for caregivers of individuals with possible pervasive developmental disorders. *Journal of Autism and Developmental Disorders*, 24(5), 659–685. doi:10.1007/BF02172145 PMID:7814313

Lord, C., Volkmar, F., & Lombroso, P. J. (2002). Genetics of childhood disorders: XLII. Autism, Part 1: Diagnosis and assessment in autistic spectrum disorders (Development and Neurobiology). *Journal of the American Academy of Child and Adolescent Psychiatry*, *41*(9), 1134–1136. doi:10.1097/00004583-200209000-00015 PMID:12218436

Losada-Barreiro, S., & Bravo-Díaz, C. (2017). Free radicals and polyphenols: The redox chemistry of neurodegenerative diseases. *European Journal of Medicinal Chemistry*, *133*, 379–402. doi:10.1016/j.ejmech.2017.03.061 PMID:28415050

Lotfipour, A. K., Wharton, S., Schwarz, S. T., Gontu, V., Schafer, A., Peters, A. M., ... Bajaj, N. P. S. (2012, January). High resolution magnetic susceptibility mapping of the substantia nigra in Parkinson's disease. *Journal of Magnetic Resonance Imaging*, *35*(1), 48–55. doi:10.1002/jmri.22752 PMID:21987471

Lotharius, J., & Brundin, P. (2002). Pathogenesis of parkinson's disease: Dopamine, vesicles and [alpha]-synuclein. *Nature Reviews. Neuroscience*, *3*(12), 932–942. doi:10.1038/nrn983 PMID:12461550

Louis, E. D. (2014). Essential Tremor: From Bedside to Bench and Back to Bedside. *Current Opinion in Neurology*, 27(4), 461–467. doi:10.1097/WCO.0000000000115 PMID:24950011

Louise, C. S. (2000). Alzheimer's amyloid fibrils: Structure and assembly. *Biochimica et Biophysica Acta*, 1502(1), 16–30. doi:10.1016/S0925-4439(00)00029-6 PMID:10899428

Lovell, M. A., Smith, J. L., & Markesbery, W. R. (2006). Elevated zinc transporter-6 in mild cognitive impairment, Alzheimer disease, and pick disease. *Journal of Neuropathology and Experimental Neurology*, *65*(5), 489–498. doi:10.1097/01. jnen.0000229237.98124.91 PMID:16772872

Lu, B. (2009). Mitochondrial dynamics and neurodegeneration. *Current Neurology and Neuroscience Reports*, 9(3), 212–219. doi:10.100711910-009-0032-7 PMID:19348710

Lucetti, C., Logi, C., Del Dotto, P., Berti, C., Ceravolo, R., Baldacci, F., ... Bonuccelli, U. (2010). Levodopa response in dementia with lewy bodies: A 1-year follow-up study. *Parkinsonism & Related Disorders*, *16*(8), 522–526. doi:10.1016/j. parkreldis.2010.06.004 PMID:20615745

518

Lugowska, A., Berger, J., Tylki-Szymańska, A., Löschl, B., Molzer, B., Zobel, M., & Czartoryska, B. (2005). Molecular and phenotypic characteristics of metachromatic leukodystrophy patients from Poland. *Clinical Genetics*, *68*(1), 48–54. doi:10.1111/j.1399-0004.2005.00451.x PMID:15952986

Lukiw, W. J. (2012). Amyloid beta (Aβ) peptide modulators and other current treatment strategies for Alzheimer's disease (AD). *Expert opinion on emerging drugs*. Retrieved from http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3399957/

Luna, J., Logroscino, G., Couratier, P., & Marin, B. (2017). Current issues in ALS epidemiology: Variation of ALS occurrence between populations and physical activity as a risk factor. *Revue Neurologique*, *173*(5), 244–253. doi:10.1016/j. neurol.2017.03.035 PMID:28477849

Luoma, P., Melberg, A., Rinne, J. O., Kaukonen, J. A., Nupponen, N. N., Chalmers, R. M., ... Majamaa, K. (2004). Parkinsonism, premature menopause, and mitochondrial DNA polymerase γ mutations: Clinical and molecular genetic study. *Lancet*, *364*(9437), 875–882. doi:10.1016/S0140-6736(04)16983-3 PMID:15351195

Lushchak, V. I. (2014). Free radicals, reactive oxygen species, oxidative stress and its classification. *Chemico-Biological Interactions*, 224, 164–175. doi:10.1016/j.cbi.2014.10.016 PMID:25452175

Lustbader, J. W., Cirilli, M., Lin, C., Xu, H. W., Takuma, K., Wang, N., & Trinchese, F. (2004). ABAD directly links AB to mitochondrial toxicity in Alzheimer's disease. *Science*, *304*(5669), 448–452. doi:10.1126cience.1091230 PMID:15087549

Luth, E. S., Stavrovskaya, I. G., Bartels, T., Kristal, B. S., & Selkoe, D. J. (2014). Soluble, prefibrillar α-synuclein oligomers promote complex I-dependent, Ca2+-induced mitochondrial dysfunction. *The Journal of Biological Chemistry*, 289(31), 21490–21507. doi:10.1074/jbc.M113.545749 PMID:24942732

Maas, A. I., Stocchetti, N., & Bullock, R. (2008). Moderate and severe traumatic brain injury in adults. *Lancet Neurology*, 7(8), 728–741. doi:10.1016/S1474-4422(08)70164-9 PMID:18635021

Macario, A. J., Cappello, F., Zummo, G., & Conway de Macario, E. (2010). Chaperonopathies of senescence and the scrambling of interactions between the chaperoning and the immune systems. *Annals of the New York Academy of Sciences*, *1197*(1), 85–93. doi:10.1111/j.1749-6632.2010.05187.x PMID:20536837

Macedo, B., & Cordeiro, Y. (2017). Unraveling Prion Protein Interactions with Aptamers and Other PrP-Binding Nucleic Acids. *International Journal of Molecular Sciences*, *18*(5). Retrieved from http://www.ncbi.nlm.nih.gov/pmc/articles/ PMC5454936/ PMID:28513534

Machado, A., Rezai, A. R., Kopell, B. H., Gross, R. E., Sharan, A. D., & Benabid, A. L. (2006). Deep brain stimulation for Parkinson's disease: Surgical technique and perioperative management. *Movement Disorders*, 21(S14Suppl 14), S247–S258. doi:10.1002/mds.20959 PMID:16810722

MacKenzie-Graham, A., Tinsley, M. R., Shah, K. P., Aguilar, C., Strickland, L. V., Boline, J., ... Jacobs, R. E. (2006). Cerebellar cortical atrophy in experimental autoimmune encephalomyelitis. *NeuroImage*, *32*(3), 1016–1023. doi:10.1016/j. neuroimage.2006.05.006 PMID:16806982

Magrassi, L., Leto, K., & Rossi, F. (2013). Lifespan of neurons is uncoupled from organismal lifespan. *Proceedings of the National Academy of Sciences of the United States of America*, *110*(11), 4374–4379. doi:10.1073/pnas.1217505110 PMID:23440189

Magrinelli, F., Picelli, A., Tocco, P., Federico, A., Roncari, L., Smania, N., ... Tamburin, S. (2016). Pathophysiology of motor dysfunction in Parkinson's disease as the rationale for drug treatment and rehabilitation. *Parkinson's Disease*. PMID:27366343

Maheshwari, A., Fischer, M., Gambetti, P., Parker, A., Ram, A., Soto, C., & Mead, S. (2015). Recent US case of variant Creutzfeldt-Jakob disease—global implications. *Emerging Infectious Diseases*, 21(5), 750–759. doi:10.3201/eid2105.142017 PMID:25897712

Mahley, R. W., Weisgraber, K. H., & Huang, Y. (2006). Apolipoprotein E4: A causative factor and therapeutic target in neuropathology, including Alzheimer's disease. *Proceedings of the National Academy of Sciences of the United States of America*, *103*(15), 5644–5651. doi:10.1073/pnas.0600549103 PMID:16567625

Mahmood, A., Berry, J., Wenger, D. A., Escolar, M., Sobeih, M., Raymond, G., & Eichler, F. S. (2009). Metachromatic Leukodystrophy: A Case of Triplets with the Late Infantile Variant and a Systematic Review of the Literature. *Journal of Child Neurology*, *25*(5), 572–580. doi:10.1177/0883073809341669 PMID:20038527

Ma, J., & Lindquist, S. (2002). Conversion of PrP to a self-perpetuating PrPSc-like conformation in the cytosol. *Science*, 298(5599), 1785–1788. doi:10.1126cience.1073619 PMID:12386336

Ma, J., Wollmann, R., & Lindquist, S. (2002). Neurotoxicity and neurodegeneration when PrP accumulates in the cytosol. *Science*, *298*(5599), 1781–1785. doi:10.1126cience.1073725 PMID:12386337

Majersik, J. J. (2017). Inherites and Uncommon Causes of Stroke. Continuum, 23(1), 211-237. PMID:28157751

Majounie, E., Renton, A. E., Mok, K., Dopper, E. G., Waite, A., Rollinson, S., ... Traynor, B. J. (2012). Frequency of the C9orf72 hexanucleotide repeat expansion in patients with amyotrophic lateral sclerosis and frontotemporal dementia: A crosssectional study. *Lancet Neurology*, *11*(4), 323–330. doi:10.1016/S1474-4422(12)70043-1 PMID:22406228

Mak, C. M., Sheng, B., Lee, H. H., Lau, K. K., Chan, W. T., Lam, C. W., & Chan, Y. W. (2011). Young-onset parkinsonism in a Hong Kong Chinese man with adult-onset Hallervorden-Spatz syndrome. *The International Journal of Neuroscience*, *121*(4), 224–227. doi:10.3109/00207454.2010.542843 PMID:21198414

Malhotra, R., & Avidan, A. Y. (2012). Neurodegenerative disease and REM behavior disorder. *Current Treatment Options in Neurology*, *14*(5), 474–492. doi:10.100711940-012-0194-5 PMID:22879077

Malkus, K. A., Tsika, E., & Ischiropoulos, H. (2009). Oxidative modifications, mitochondrial dysfunction, and impaired protein degradation in Parkinson's disease: How neurons are lost in the Bermuda triangle. *Molecular Neurodegeneration*, *4*(1), 24. doi:10.1186/1750-1326-4-24 PMID:19500376

Ma, M. W., Wang, J., Zhang, Q., Wang, R., Dhandapani, K. M., Vadlamudi, R. K., & Brann, D. W. (2017). NADPH oxidase in brain injury and neurodegenerative disorders. *Molecular Neurodegeneration*, *12*(1), 7. doi:10.118613024-017-0150-7 PMID:28095923

Mancuso, M., Calsolaro, V., Orsucci, D., Carlesi, C., Choub, A., Piazza, S., & Siciliano, G. (2009). Mitochondria, Cognitive Impairment, and Alzheimer's Disease. *International Journal of Alzheimer's Disease*, *951548*. doi:10.4061/2009/951548 PMID:20798880

Mancuso, M., Orsucci, D., Calsolaro, V., Choub, A., & Siciliano, G. (2009). Coenzyme Q10 and Neurological Diseases. *Pharmaceuticals*, 2(3), 134–149. doi:10.3390/ph203134 PMID:27713230

Mancuso, M., Orsucci, D., Siciliano, G., & Murri, L. (2008). Mitochondria, mitochondrial DNA and Alzheimer's disease. What comes first? *Current Alzheimer Research*, *5*(5), 457–468. doi:10.2174/156720508785908946 PMID:18855587

Manczak, M., Calkins, M. J., & Reddy, P. H. (2011). Impaired mitochondrial dynamics and abnormal interaction of amyloid beta with mitochondrial protein Drp1 in neurons from patients with Alzheimer's disease: Implications for neuronal damage. *Human Molecular Genetics*, 20(13), 2495–2509. doi:10.1093/hmg/ddr139 PMID:21459773

Manczak, M., Jung, Y., Park, B. S., Partovi, D., & Reddy, P. H. (2005). Time course of mitochondrial gene expressions in mice brains: Implications for mitochondrial dysfunction, oxidative damage, and cytochrome c in aging. *Journal of Neurochemistry*, *92*(3), 494–504. doi:10.1111/j.1471-4159.2004.02884.x PMID:15659220

Manczak, M., Mao, P., Calkins, M. J., Cornea, A., Reddy, A. P., Murphy, M. P., ... Reddy, P. H. (2010). Mitochondriatargeted antioxidants protect against amyloid-β toxicity in Alzheimer's disease neurons. *Journal of Alzheimer's Disease*, 20(2), S609–S631. doi:10.3233/JAD-2010-100564 PMID:20463406

Manczak, M., Park, B. S., Jung, Y., & Reddy, P. H. (2004). Differential expression of oxidative phosphorylation genes in patients with Alzheimer's disease. *Neuromolecular Medicine*, 5(2), 147–162. doi:10.1385/NMM:5:2:147 PMID:15075441

Mandler, M., Valera, E., Rockenstein, E., Weninger, H., Patrick, C., Adame, A., ... Masliah, E. (2014). Next-generation active immunization approach for synucleinopathies: Implications for Parkinson's disease clinical trials. *Acta Neuropathologica*, *127*(6), 861–879. doi:10.100700401-014-1256-4 PMID:24525765

Mandzia, J. L., Smith, E. E., Horton, M., Hanly, P., Barber, P. A., Godzwon, C., ... Coutts, S. B. (2016). Imaging and baseline predictors of cognitive performance in minor ischemic stroke and patients with transient ischemic attack at 90 days. *Stroke*, *47*(3), 726–731. PMID:26846862

Mann, D. M., Jones, D., South, P. W., Snowden, J. S., & Neary, D. (1992). Deposition of amyloid beta protein in non-Alzheimer dementias: Evidence for a neuronal origin of parenchymal deposits of beta protein in neurodegenerative disease. *Acta Neuropathologica*, 83(4), 415–419. doi:10.1007/BF00713534 PMID:1575018

Manning, C. (2004). Beyond memory: Neuropsychologic features in differential diagnosis of dementia. *Clinics in Geriatric Medicine*, 20(1), 45–58. doi:10.1016/j.cger.2003.10.002 PMID:15062486

Manoharan, S., Guillemin, G. J., Abiramasundari, R. S., Essa, M. M., Akbar, M., & Akbar, M. D. (2016). The Role of Reactive Oxygen Species in the Pathogenesis of Alzheimer's Disease, Parkinson's Disease, and Huntington's Disease: A Mini Review. *Oxidative Medicine and Cellular Longevity*, 2016, 8590578. doi:10.1155/2016/8590578 PMID:28116038

Mao, P., & Reddy, P. H. (2010). Is multiple sclerosis a mitochondrial disease? *Biochimica et Biophysica Acta*, *1802*(1), 66–79. doi:10.1016/j.bbadis.2009.07.002 PMID:19607913

Maraganore, D. M., Wilkes, K., Lesnick, T. G., Strain, K. J., De Andrade, M., Rocca, W. A., ... Farrer, M. J. (2004). A limited role for DJ1 in Parkinson disease susceptibility. *Neurology*, *63*(3), 550–553. doi:10.1212/01.WNL.0000133402.78621. AD PMID:15304593

Maraldi, T. (2013). Natural compounds as modulators of NADPH oxidases. *Oxidative Medicine and Cellular Longevity*. PMID:24381714

Marder, K., Logroscino, G., Alfaro, B., Mejia, H., Halim, A., Louis, E., ... Mayeux, R. (1998). Environmental risk factors for Parkinson's disease in an urban multiethnic community. *Neurology*, *50*(1), 279–281. doi:10.1212/WNL.50.1.279 PMID:9443493

Marín, I., van Egmond, W. N., & van Haastert, P. J. (2008). The Roco protein family: A functional perspective. *The FASEB Journal*, 22(9), 3103–3110. doi:10.1096/fj.08-111310 PMID:18523161

Marino, M. J., Valenti, O., & Conn, P. J. (2003). Glutamate receptors and Parkinson's disease. *Drugs & Aging*, 20(5), 377–397. doi:10.2165/00002512-200320050-00006 PMID:12696997

Marklund, S. L., Westman, N. G., Lundgren, E., & Ross, G. (1982). Copper- and zinc-containing superoxide dismutase, manganese-containing superoxide dismutase, catalase, and glutathione peroxidase in nrmal and neoplastic human cell lines and normal human tissues. *Cancer Research*, *42*, 1955–1961. PMID:7066906

Mark, P. M. (2004). Metal-catalyzed disruption of membrane protein and lipid signaling in the pathogenesis of neurodegenerative disorders. *Annals of the New York Academy of Sciences*, *1012*(1), 37–50. doi:10.1196/annals.1306.004 PMID:15105254

Mark, R. J., Ashford, J. W., Goodman, Y., & Mattson, M. P. (1995). Anticonvulsants attenuate amyloid β-peptide neurotoxicity, Ca 2+ deregulation, and cytoskeletal pathology. *Neurobiology of Aging*, *16*(2), 187–198. doi:10.1016/0197-4580(94)00150-2 PMID:7777136

Marks, W. J. Jr, Bartus, R. T., Siffert, J., Davis, C. S., Lozano, A., Boulis, N., ... Olanow, C. W. (2010, December). Gene delivery of AAV2-neurturin for Parkinson's disease: A double-blind, randomised, controlled trial. *Lancet Neurology*, *9*(12), 1164–1172. doi:10.1016/S1474-4422(10)70254-4 PMID:20970382

Maroteaux, L., Campanelli, J. T., & Scheller, R. H. (1988). Synuclein: A neuron-specific protein localized to the nucleus and presynaptic nerve terminal. *The Journal of Neuroscience*, *8*, 2804–2815. PMID:3411354

Marsden, C. D. (1994). Parkinson's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 57(6), 672–681. doi:10.1136/jnnp.57.6.672 PMID:7755681

Marshall, R. S., Festa, J. R., Cheung, Y. K., Chen, R., Pavol, M. A., Derdeyn, C. P., ... Lazar, R. M. (2012). Cerebral hemodynamics and cognitive impairment: Baseline data from the RECON trial. *Neurology*, *78*(4), 250–255. doi:10.1212/ WNL.0b013e31824365d3 PMID:22238418

Marstrand, J., Garde, E., Rostrup, E., Ring, P., Rosenbaum, S., Mortensen, E. L., & Larsson, H. (2002). Cerebral perfusion and cerebrovascular reactivity are reduced in white matter hyperintensities. *Stroke*, *33*(4), 972–976. doi:10.1161/01. STR.0000012808.81667.4B PMID:11935046

Martinez-Vicente, M., Talloczy, Z., Kaushik, S., Massey, A. C., Mazzulli, J., Mosharov, E. V., ... Dauer, W. (2008). Dopamine-modified α-synuclein blocks chaperone-mediated autophagy. *The Journal of Clinical Investigation*, *118*(2), 777. PMID:18172548

Martin, L. J. (2010). Mitochondrial and cell death mechanisms in neurodegenerative diseases. *Pharmaceuticals*, *3*(4), 839–915. doi:10.3390/ph3040839 PMID:21258649

Martin, L. J., Pan, Y., Price, A. C., Sterling, W., Copeland, N. G., Jenkins, N. A., ... Lee, M. K. (2006). Parkinson's disease α -synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death. *The Journal of Neuroscience*, 26(1), 41–50. doi:10.1523/JNEUROSCI.4308-05.2006 PMID:16399671

Martin, L., Latypova, X., & Terro, F. (2011). Post-translational modifications of tau protein: Implications for Alzheimer's disease. *Neurochemistry International*, 58(4), 458–471. doi:10.1016/j.neuint.2010.12.023 PMID:21215781

Martin, L., Nikolaus-von, S., & Harald, W. (2012). Zinc diet and Alzheimer's disease: A systematic review. *Nutritional Neuroscience*, *15*(5), 1–12. PMID:22305647

Martin, S. J., Grimwood, P. D., & Morris, R. G. M. (2000). Synaptic plasticity and memory: An evaluation of the hypothesis. *Annual Review of Neuroscience*, 23(1), 649–711. doi:10.1146/annurev.neuro.23.1.649 PMID:10845078

Marx, F. P., Holzmann, C., Strauss, K. M., Li, L., Eberhardt, O., Gerhardt, E., ... Engelender, S. (2003). Identification and functional characterization of a novel R621C mutation in the synphilin-1 gene in Parkinson's disease. *Human Molecular Genetics*, *12*(11), 1223–1231. doi:10.1093/hmg/ddg134 PMID:12761037

Ma, S. L., Pastorino, L., Zhou, X. Z., & Lu, K. P. (2012). Prolyl Isomerase Pin1 Promotes Amyloid Precursor Protein (APP) Turnover by Inhibiting Glycogen Synthase Kinase-3β (GSK3β) Activity novel mechanism for pin1 to protect against Alzheimer disease. *The Journal of Biological Chemistry*, 287(10), 6969–6973. doi:10.1074/jbc.C111.298596 PMID:22184106

Ma, S. Y., Röytt, M., Collan, Y., & Rinne, J. O. (1999b). Unbiased morphometrical measurements show loss of pigmented nigral neurones with ageing. *Neuropathology and Applied Neurobiology*, 25(5), 394–399. doi:10.1046/j.1365-2990.1999.00202.x PMID:10564529

Mashhoon, Y., Wells, A. M., & Kantak, K. M. (2010). Interaction of the rostral basolateral amygdala and prelimbic prefrontal cortex in regulating reinstatement of cocaine-seeking behavior. *Pharmacology, Biochemistry, and Behavior*, *96*(3), 347–353. doi:10.1016/j.pbb.2010.06.005 PMID:20600250

Masliah, E., Rockenstein, E., Adame, A., Alford, M., Crews, L., Hashimoto, M., ... Schenk, D. (2005). Effects of alpha-synuclein immunization in a mouse model of Parkinson's disease. *Neuron*, 46(6), 857–868. doi:10.1016/j.neuron.2005.05.010 PMID:15953415

Massano, J., & Bhatia, K. P. (2012). Clinical approach to Parkinson's disease: Features, diagnosis, and principles of management. *Cold Spring Harbor Perspectives in Medicine*, 2(6), a008870. doi:10.1101/cshperspect.a008870 PMID:22675666

Massano, J., Costa, F., & Nadais, G. (2008). Teaching neuroImage: MRI in multiple system atrophy: "Hot cross bun" sign and hyperintense rim bordering the putamina. *Neurology*, 71(15), e38. doi:10.1212/01.wnl.0000327520.99034.28 PMID:18838658

Masters, C. L., Simms, G., Weinman, N. A., Multhaup, G., McDonald, B. L., & Beyreuther, K. (1985). Amyloid plaque core protein in Alzheimer disease and Down syndrome. *Proceedings of the National Academy of Sciences of the United States of America*, 82(12), 4245–4249. doi:10.1073/pnas.82.12.4245 PMID:3159021

Mastrianni, J. A. (2010). The genetics of prion diseases. *Genetics in Medicine*, *12*(4), 187–195. doi:10.1097/ GIM.0b013e3181cd7374 PMID:20216075

Matalon, R., & Matalon, K. M. (2015). Canavan Disease. In Rosenberg's Molecular and Genetic Basis of Neurological and Psychiatric Disease (5th ed.; pp. 695-701). Boston: Academic.

Matalon, R., & Michals-Matalon, K. (1999 Sep 16). Canavan Disease. In M. P. Adam, H. H. Ardinger, R. A. Pagon, & ... (Eds.), *GeneReviews* (pp. 1993–2017). Seattle, WA: University of Washington. Available from https://www.ncbi. nlm.nih.gov/books/NBK1234/

Math, P. C., & Gordon, J. L. (1997). Zinc Metabolism in the brain: Relevance to human neurodegenerative disorders. *Neurobiology of Disease*, *4*(3-4), 137–169. doi:10.1006/nbdi.1997.0163 PubMed

Matthews, F. E., Brayne, C., Lowe, J., McKeith, I., Wharton, S. B., & Ince, P. (2009). Epidemiological pathology of dementia: Attributable-risks at death in the Medical Research Council Cognitive Function and Ageing Study. *PLoS Medicine*, *6*(11), e1000180. doi:10.1371/journal.pmed.1000180 PMID:19901977

Mattiazzi, M., D'Aurelio, M., Gajewski, C. D., Martushova, K., Kiaei, M., Beal, M. F., & Manfredi, G. (2002). Mutated human SOD1 causes dysfunction of oxidative phosphorylation in mitochondria of transgenic mice. *The Journal of Biological Chemistry*, 277(33), 29626–29633. doi:10.1074/jbc.M203065200 PMID:12050154

Mattila, M. L., Kielinen, M., Linna, S. L., Jussila, K., Ebeling, H., Bloigu, R., ... Moilanen, I. (2011). Autism spectrum disorders according to DSM-IV-TR and comparison with DSM-5 draft criteria: An epidemiological study. *Journal of the American Academy of Child and Adolescent Psychiatry*, *50*(6), 583–592. doi:10.1016/j.jaac.2011.04.001 PMID:21621142

Mattson, M. P. (2004). Pathways towards and away from Alzheimer's disease. *Nature*, 430(7000), 631–639. doi:10.1038/ nature02621 PMID:15295589

Mattson, M. P. (2008). Glutamate and neurotrophic factors in neuronal plasticity and disease. *Annals of the New York Academy of Sciences*, *1144*(1), 97–112. doi:10.1196/annals.1418.005 PMID:19076369

Matyja, E., Taraszewska, A., Nagańska, E., Grieb, P., & Rafałowska, J. (2008). CDP-choline protects motor neurons against apoptotic changes in a model of chronic glutamate excitotoxicity *in vitro. Folia Neuropathologica*, *46*, 139–148. PMID:18587708

Maurer, I., Zierz, S., & Möller, H. J. (2000). A selective defect of cytochrome c oxidase is present in brain of Alzheimer disease patients. *Neurobiology of Aging*, 21(3), 455–462. doi:10.1016/S0197-4580(00)00112-3 PMID:10858595

Ma, V. Y., Chan, L., & Carruthers, K. J. (2014). Incidence, prevalence, costs, and impact on disability of common conditions requiring rehabilitation in the United States: Stroke, spinal cord injury, traumatic brain injury, multiple sclerosis, osteoarthritis, rheumatoid arthritis, limb loss, and back pain. *Archives of Physical Medicine and Rehabilitation*, *95*(5), 986–995. doi:10.1016/j.apmr.2013.10.032 PMID:24462839

Mazzini, L., Corra, T., Zaccala, M., Mora, G., Del Piano, M., & Galante, M. (1995). Percutaneous endoscopic gastrostomy and enteral nutrition in amyotrophic lateral sclerosis. *Journal of Neurology*, 242(10), 695–698. doi:10.1007/ BF00866922 PMID:8568533

Mazzio, E. A., Reams, R. R., & Soliman, K. F. (2004). The role of oxidative stress, impaired glycolysis and mitochondrial respiratory redox failure in the cytotoxic effects of 6-hydroxydopamine in vitro. *Brain Research*, *1004*(1), 29–44. doi:10.1016/j.brainres.2003.12.034 PMID:15033417

Mazzulli, J. R., Xu, Y. H., Sun, Y., Knight, A. L., McLean, P. J., Caldwell, G. A., ... Krainc, D. (2011). Gaucher disease glucocerebrosidase and α -synuclein form a bidirectional pathogenic loop in synucleinopathies. *Cell*, *146*(1), 37–52. doi:10.1016/j.cell.2011.06.001 PMID:21700325

McCall, K. A., Huang, C., & Fierke, C. A. (2000, May 01). Function and mechanism of zinc metalloenzymes. *The Journal of Nutrition*, *130*(5), 1437S–1446S. doi:10.1093/jn/130.5.1437S PubMed

McClain, C. J., McClain, M., Barve, S., & Boosalis, M. G. (2002). Trace metals and the elderly. *Clinics in Geriatric Medicine*, *18*(4), 801–818. doi:10.1016/S0749-0690(02)00040-X PMID:12608504

McConnell, S. (2002). Interventions to facilitate social interaction for young children with autism: Review of available research and recommendations for educational intervention and future research. *Journal of Autism and Developmental Disorders*, *32*(5), 351–372. doi:10.1023/A:1020537805154 PMID:12463515

McCoy, M. K., & Cookson, M. R. (2012). Mitochondrial quality control and dynamics in parkinson's disease. *Antioxidants & Redox Signalling*, *16*(9), 869–882. doi:10.1089/ars.2011.4019 PMID:21568830

McCoy, M. K., Martinez, T. N., Ruhn, K. A., Szymkowski, D. E., Smith, C. G., Botterman, B. R., ... Tansey, M. G. (2006). Blocking soluble tumor necrosis factor signaling with dominant-negative tumor necrosis factor inhibitor attenuates loss of dopaminergic neurons in models of Parkinson's disease. *The Journal of Neuroscience*, *26*(37), 9365–9375. doi:10.1523/JNEUROSCI.1504-06.2006 PMID:16971520

McDonald-McGinn, D. M., Kirschner, R., Goldmuntz, E., Sullivan, K., Eicher, P., & Gerdes, M. (1999). The Philadelphia story: The 22q11.2 deletion: Report on 250 patients. *Genetic Counseling (Geneva, Switzerland)*, 10, 11–24. PMID:10191425

McDonald-McGinn, D. M., Tonnesen, M. K., Laufer-Cahana, A., Finucane, B., Driscoll, D. A., Emanuel, B. S., & Zackai, E. H. (2001). Phenotype of the 22q11.2 deletion in individuals identified through an affected relative: Cast a wide FISHing net! *Genetics in Medicine*, *3*(1), 23–29. doi:10.1097/00125817-200101000-00006 PMID:11339373

McEntee, W. J., & Crook, T. H. (1993). Glutamate: Its role in learning, memory, and the aging brain. *Psychopharmacology*, *111*(4), 391–401. doi:10.1007/BF02253527 PMID:7870979

McEwen, B. S. (2002). Sex, stress and the hippocampus: Allostasis, allostatic load and the aging process. *Neurobiology* of Aging, 23(5), 921–939. doi:10.1016/S0197-4580(02)00027-1 PMID:12392796

McGleenon, B. M., Dynan, K. B., & Passmore, A. P. (1999). Acetylcholinesterase inhibitors in Alzheimer's disease. *British Journal of Clinical Pharmacology*, *48*(4), 471–480. doi:10.1046/j.1365-2125.1999.00026.x PMID:10583015

McKee, A. C., & Daneshvar, D. H. (2015). The neuropathology of traumatic brain injury. *Handbook of Clinical Neurology*, *127*, 45–66. doi:10.1016/B978-0-444-52892-6.00004-0 PMID:25702209

McKeith, I. G., Ballard, C. G., Perry, R. H., Ince, P. G., O'brien, J. T., Neill, D., ... Swann, A. (2000). Prospective validation of consensus criteria for the diagnosis of dementia with Lewy bodies. *Neurology*, *54*(5), 1050–1058. doi:10.1212/ WNL.54.5.1050 PMID:10720273

McKeith, I. G., Dickson, D. W., Lowe, J., Emre, M., O'Brien, J. T., Feldman, H., ... Yamada, M. (2005). Diagnosis and management of dementia with Lewy bodies: Third report of the DLB Consortium. *Neurology*, 65(12), 1863–1872. doi:10.1212/01.wnl.0000187889.17253.b1 PMID:16237129

McKeith, I. G., Galasko, D., Kosaka, K., Perry, E. K., Dickson, D. W., Hansen, L. A., ... Lennox, G. (1996). Consensus guidelines for the clinical and pathologic diagnosis of dementia with Lewy bodies (DLB) Report of the consortium on DLB international workshop. *Neurology*, *47*(5), 1113–1124. doi:10.1212/WNL.47.5.1113 PMID:8909416

McKeith, I., Del Ser, T., Spano, P., Emre, M., Wesnes, K., Anand, R., ... Spiegel, R. (2000). Efficacy of rivastigmine in dementia with Lewy bodies: A randomised, double-blind, placebo-controlled international study. *Lancet*, *356*(9247), 2031–2036. doi:10.1016/S0140-6736(00)03399-7 PMID:11145488

McNaught, K. S. P., Belizaire, R., Isacson, O., Jenner, P., & Olanow, C. W. (2003). Altered proteasomal function in sporadic Parkinson's disease. *Experimental Neurology*, *179*(1), 38–46. doi:10.1006/exnr.2002.8050 PMID:12504866

McNaught, K. S. P., & Jenner, P. (2001). Proteasomal function is impaired in substantia nigra in Parkinson's disease. *Neuroscience Letters*, 297(3), 191–194. doi:10.1016/S0304-3940(00)01701-8 PMID:11137760

McPheeters, M. L., Warren, Z., Sathe, N., Bruzek, J. L., Krishnaswami, S., Jerome, R. N., & Veenstra-Vanderweele, J. (2011). A systematic review of medical treatments for children with autism spectrum disorders. *Pediatrics*, *127*(5), e1312–e1321. doi:10.1542/peds.2011-0427 PMID:21464191

Mead, S., Mahal, S. P., Beck, J., Campbell, T., Farrall, M., Fisher, E., & Collinge, J. (2001). Sporadic—but not variant— Creutzfeldt-Jakob disease is associated with polymorphisms upstream of PRNP exon 1. *American Journal of Human Genetics*, *69*(6), 1225–1235. doi:10.1086/324710 PMID:11704923

Mead, S., Poulter, M., Uphill, J., Beck, J., Webb, T., Campbell, T., & Collinge, J. (2013). Identifies Genetic Risk Factors For Plaques In Variant Creutzfeldt-Jakob Plaques In Sporadic Creutzfeldt-Jakob. *Neurology*, *2*(4), 127–1135.

Mecocci, P., Beal, M. F., Cecchetti, R., Polidori, M. C., Cherubini, A., Chionne, F., ... Senin, U. (1997). Mitochondrial membrane fluidity and oxidative damage to mitochondrial DNA in aged and AD human brain. *Molecular and Chemical Neuropathology*, *31*(1), 53–64. doi:10.1007/BF02815160 PMID:9271005

Mehdorn, H. M. (2016, June). Deep brain stimulation for dystonia: Review of the literature. *Journal of Neurosurgical Sciences*, *60*(2), 199–210. PMID:26977634

Meltzer, C. C., Smith, G., DeKosky, S. T., Pollock, B. G., Mathis, C. A., Moore, R. Y., ... Reynolds, C. F. (1998). Serotonin in aging, late-life depression, and Alzheimer's disease: The emerging role of functional imaging. *Neuropsychopharmacology*, *18*(6), 407–430. doi:10.1016/S0893-133X(97)00194-2 PMID:9571651

Mendez, M. F., & Shapira, J. S. (2011). Loss of emotional insight in behavioral variant frontotemporal dementia or "frontal anosodiaphoria". *Consciousness and Cognition*, 20(4), 1690–1696. doi:10.1016/j.concog.2011.09.005 PMID:21959203

Meraz-Ríos, M. A., Franco-Bocanegra, D., & Rios, D. T. & Campos-Peña, V. (2014). Early onset Alzheimer's disease and oxidative stress. *Oxidative Medicine and Cellular Longevity*. PMID:24669286

Mercado, G., Castillo, V., Vidal, R., & Hetz, C. (2015). ER proteostasis disturbances in Parkinson's disease : Novel insights. *Frontiers in Aging Neuroscience*, 7(March), 1–5. PMID:25870559

Messing, A., Brenner, M., Feany, M. B., Nedergaard, M., & Goldman, J. E. (2012). Alexander Disease. *The Journal of Neuroscience*, 32(15), 5017–5023. doi:10.1523/JNEUROSCI.5384-11.2012 PMID:22496548

Michelle, S. H., Laura, J. B., & Paul, Q. T. (2000). Endogenous mechanisms of neuroprotection: Role of zinc, copper, and carnosine. *Brain Research*, 852(1), 56–61. doi:10.1016/S0006-8993(99)02215-5 PMID:10661495

Middleton, L. E., Lam, B., Fahmi, H., Black, S. E., McIlroy, W. E., Stuss, D. T., ... Turner, G. R. (2014). Frequency of domain-specific cognitive impairment in sub-acute and chronic stroke. *NeuroRehabilitation*, *34*(2), 305–312. PMID:24401826

Mietelska-Porowska, A., Wasik, U., Goras, M., Filipek, A., & Niewiadomska, G. (2014). Tau Protein Modifications and Interactions: Their Role in Function and Dysfunction. *International Journal of Molecular Sciences*, *15*(3), 4671–4713. doi:10.3390/ijms15034671 PMID:24646911

Migliaccio, R., Agosta, F., Toba, M. N., Samri, D., Corlier, F., de Souza, L. C., ... Bartolomeo, P. (2012). Brain networks in posterior cortical atrophy: A single case tractography study and literature review. *Cortex*, 48(10), 1298–1309. doi:10.1016/j.cortex.2011.10.002 PMID:22099855

Miklya, I., Göltl, P., Hafenscher, F., & Pencz, N. (2014). The role of parkin in Parkinson's disease. *Neuropsychopharmacologia Hungarica: a Magyar Pszichofarmakologiai Egyesulet lapja= official journal of the Hungarian Association of Psychopharmacology, 16*(2), 67-76.

Milakovic, T., & Johnson, G. V. (2005). Mitochondrial respiration and ATP production are significantly impaired in striatal cells expressing mutant huntingtin. *The Journal of Biological Chemistry*, 280(35), 30773–30782. doi:10.1074/jbc.M504749200 PMID:15983033

Milhavet, O., & Lehmann, S. (2002). Oxidative stress and the prion protein in transmissible spongiform encephalopathies. *Brain Research. Brain Research Reviews*, *38*(3), 328–339. doi:10.1016/S0165-0173(01)00150-3 PMID:11890980

Miller, S. (2010). Aging and decision making. Academic Press.

Miller, G. W. (2007). Paraquat: The red herring of Parkinson's disease research. *Toxicological Sciences*, *100*(1), 1–2. doi:10.1093/toxsci/kfm223 PMID:17934192

Miller, R. G., Jackson, C. E., Kasarskis, E. J., England, J. D., Forshew, D., Johnson, W., ... Wooley, S. C. (2009). Practice Parameter update: The care of the patient with amyotrophic lateral sclerosis: Drug, nutritional, and respiratory therapies (an evidence-based review): Report of the Quality Standards Subcommittee of the American Academy of Neurology. *Neurology*, *73*(15), 1218–1226. doi:10.1212/WNL.0b013e3181bc0141 PMID:19822872

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Miller, R. G., Moore, D. H., Gelinas, D. F., Dronsky, V., Mendoza, M., Barohn, R. J., ... Olney, R. (2001). Phase III randomized trial of gabapentin in patients with amyotrophic lateral sclerosis. *Neurology*, *56*(7), 843–848. doi:10.1212/ WNL.56.7.843 PMID:11294919

Mills, R. D., Sim, C. H., Mok, S. S., Mulhern, T. D., Culvenor, J. G., & Cheng, H. C. (2008). Biochemical aspects of the neuroprotective mechanism of PTEN induced kinase 1 (PINK1). *Journal of Neurochemistry*, *105*(1), 18–33. doi:10.1111/j.1471-4159.2008.05249.x PMID:18221368

Minassian, B. A. (2001). Lafora's disease: Towards a clinical, pathologic, and molecular synthesis. *Pediatric Neurology*, 25(1), 21–29. doi:10.1016/S0887-8994(00)00276-9 PMID:11483392

Minton, A. P. (2005). Influence of macromolecular crowding upon the stability and state of association of proteins: Predictions and observations. *Journal of Pharmaceutical Sciences*, 94(8), 1668–1675. doi:10.1002/jps.20417 PMID:15986476

Misonou, H., Morishima-kawashima, M., & Ihara, Y. (2000). Oxidative stress induces intracellular accumulation of amyloid beta-protein (Abeta) in human neuroblastoma cells. *Biochemistry*, *39*(23), 6951–6959. doi:10.1021/bi000169p PMID:10841777

Mitsumoto, H., & Rabkin, J. G. (2007). Palliative care for patients with amyotrophic lateral sclerosis: Prepare for the worst and hope for the best. *Journal of the American Medical Association*, 298(2), 207–216. doi:10.1001/jama.298.2.207 PMID:17622602

Mizielinska, S., Gronke, S., Niccoli, T., Ridler, C. E., Clayton, E. L., Devoy, A., ... Pietrzyk, J. (2014). C9orf72 repeat expansions cause neurodegeneration in Drosophila through arginine-rich proteins. *Science*, *345*(6201), 1192–1194. doi:10.1126cience.1256800 PMID:25103406

Mizuno, Y., Hattori, N., Kitada, T., Matsumine, H., Mori, H., Shimura, H., ... Shimizu, N. (2000). Familial Parkinson's disease. Alpha-synuclein and parkin. *Advances in Neurology*, *86*, 13–21. PMID:11553970

Mocchegiani, E., Bertoni-Freddari, C., Marcellini, F., & Malavolta, M. (2005). Brain, aging and neu-rodegeneration:role of zinc ion availability. *Progress in Neurobiology*, 75(6), 367–390. doi:10.1016/j.pneurobio.2005.04.005 PMID:15927345

Mocchegiani, E., Burkle, A., & Fulop, T. (2008). Zinc and ageing (ZINCAGEProject). *Experimental Gerontology*, *43*(5), 361–362. doi:10.1016/j.exger.2008.03.009 PMID:18417310

Moehle, M. S., Webber, P. J., Tse, T., Sukar, N., Standaert, D. G., DeSilva, T. M., ... West, A. B. (2012). LRRK2 inhibition attenuates microglial inflammatory responses. *The Journal of Neuroscience*, *32*(5), 1602–1611. doi:10.1523/ JNEUROSCI.5601-11.2012 PMID:22302802

Moghal, S., Rajput, A. H., D'Arcy, C., & Rajput, R. (1994). Prevalence of movement disorders in elderly community residents. *Neuroepidemiology*, *13*(4), 175–178. doi:10.1159/000110376 PMID:8090259

Mohri, I., Taniike, M., Taniguchi, H., Kanekiyo, T., Aritake, K., Inui, T., ... Urade, Y. (2006). *Prostaglandin D 2 -Mediated Microglia / Astrocyte Interaction Enhances Astrogliosis and Demyelination in twitcher*. Academic Press.

Moisan, F., Spinosi, J., Dupupet, J. L., Delabre, L., Mazurie, J. L., Goldberg, M., ... Elbaz, A. (2011). The relation between type of farming and prevalence of Parkinson's disease among agricultural workers in five French districts. *Movement Disorders*, *26*(2), 271–279. doi:10.1002/mds.23370 PMID:21412834

Molliex, A., Temirov, J., Lee, J., Coughlin, M., Kanagaraj, A. P., Kim, H. J., ... Taylor, J. P. (2015). Phase separation by low complexity domains promotes stress granule assembly and drives pathological fibrillization. *Cell*, *163*(1), 123–133. doi:10.1016/j.cell.2015.09.015 PMID:26406374

Moll, L., El-Ami, T., & Cohen, E. (2014). Selective manipulation of aging: A novel strategy for the treatment of neurodegenerative disorders. *Swiss Medical Weekly*, *144*, w13917. PMID:24526357

Molloy, S., McKeith, I. G., O'brien, J. T., & Burn, D. J. (2005). The role of levodopa in the management of dementia with Lewy bodies. *Journal of Neurology, Neurosurgery, and Psychiatry*, 76(9), 1200–1203. doi:10.1136/jnnp.2004.052332 PMID:16107351

Monaghan, T. S., & Delanty, N. (2010). Lafora disease: Epidemiology, pathophysiology and management. *CNS Drugs*, 24(7), 549–561. doi:10.2165/11319250-00000000-00000 PMID:20527995

Monchi, O., Petrides, M., Mejia-Constain, B., & Strafella, A. P. (2007). Cortical activity in Parkinson's disease during executive processing depends on striatal involvement. *Brain*, *130*(1), 233–244. doi:10.1093/brain/awl326 PMID:17121746

Monte, De Ia., S. M., Lu, B. X., Sohn, Y. K., Etienne, D., Kraft, J., Ganju, N., & Wands, J. R. (2000). Aberrant expression of nitric oxide synthase III in Alzheimer's disease: relevance to cerebral vasculopathy and neurodegeneration. *Neurobiology of Aging*, *21*(2), 309-319.

Montgomery, E. B. Jr. (2013, April). Subthalamic versus globus pallidus deep brain stimulation. *Lancet Neurology*, *12*(4), 329. doi:10.1016/S1474-4422(13)70045-0 PMID:23518323

Montie, H. L., & Durcan, T. M. (2013). The cell and molecular biology of neurodegenerative diseases : An overview. *Frontiers in Neurology*, 4(November), 1–2. PMID:24348458

Montuschi, A., Iazzolino, B., Calvo, A., Moglia, C., Lopiano, L., Restagno, G., ... Chiò, A. (2015). Cognitive correlates in amyotrophic lateral sclerosis: A population-based study in Italy. *Journal of Neurology, Neurosurgery, and Psychiatry*, *86*(2), 168–173. doi:10.1136/jnnp-2013-307223 PMID:24769471

Moody, D. M., Thore, C. R., Anstrom, J. A., Challa, V. R., Langefeld, C. D., & Brown, W. R. (2004). Quantification of afferent vessels shows reduced brain vascular density in subjects with leukoaraiosis. *Radiology*, 233(3), 883–890. doi:10.1148/radiol.2333020981 PMID:15564412

Moorhouse, P., & Rockwood, K. (2013). Vascular cognitive impairment in the memory clinic. In O. Godefroy (Ed.), *Behavioral and cognitive neurology of stroke* (2nd ed.; pp. 9–21). Cambridge University Press. doi:10.1017/CBO9781139058988.003

Morán, M., Moreno-Lastres, D., Marín-Buera, L., Arenas, J., Martín, M. A., & Ugalde, C. (2012). Mitochondrial respiratory chain dysfunction: Implications in neurodegeneration. *Free Radical Biology & Medicine*, *53*(3), 595–609. doi:10.1016/j.freeradbiomed.2012.05.009 PMID:22595027

Moreno-Gonzalez, I., & Soto, C. (2011). Misfolded protein aggregates: Mechanisms, structures and potential for disease transmission. *Seminars in Cell & Developmental Biology*, 22(5), 482–487. doi:10.1016/j.semcdb.2011.04.002 PMID:21571086

Moreton, F. C., Razvi, S. S., Davidson, R., & Muir, K. W. (2014). Changing clinical patterns and increasing prevalence in CADASIL. *Acta Neurologica Scandinavica*, *130*(3), 197–203. doi:10.1111/ane.12266 PMID:24840674

Morgan, C., Gupta, M., El-Feky, W., Shamim, S., & Opatowsky, M. (2009). Creutzfeldt-Jakob disease: Case discussion and imaging review. *Proceedings - Baylor University. Medical Center*, 22(1), 69–71. doi:10.1080/08998280.2009.119 28476 PMID:19169404

Morgan, J. T., Chana, G., Pardo, C. A., Achim, C., Semendeferi, K., Buckwalter, J., ... Everall, I. P. (2010). Microglial activation and increased microglial density observed in the dorsolateral prefrontal cortex in autism. *Biological Psychiatry*, *68*(4), 368–376. doi:10.1016/j.biopsych.2010.05.024 PMID:20674603

528

Mori, E., Ikeda, M., & Kosaka, K. (2012). Donepezil for dementia with Lewy bodies: A randomized, placebo-controlled trial. *Annals of Neurology*, 72(1), 41–52. doi:10.1002/ana.23557 PMID:22829268

Moro, E., Gross, R. E., & Krauss, J. K. (2013, June). What's new in surgical treatment for dystonia? *Movement Disorders*, 28(7), 1013–1020. doi:10.1002/mds.25550 PMID:23893457

Moro, E., Hamani, C., Poon, Y. Y., Al-Khairallah, T., Dostrovsky, J. O., Hutchison, W. D., & Lozano, A. M. (2010, January 01). (2010-a). Unilateral pedunculopontine stimulation improves falls in Parkinson's disease. *Brain*, *133*(Pt 1), 215–224. doi:10.1093/brain/awp261 PMID:19846583

Moro, E., Lozano, A. M., Pollak, P., Agid, Y., Rehncrona, S., Volkmann, J., ... Lang, A. E. (2010, April 15). (2010-b). Long-term results of a multicenter study on subthalamic and pallidal stimulation in Parkinson's disease. *Movement Disorders*, 25(5), 578–586. doi:10.1002/mds.22735 PMID:20213817

Morris, C. M., Candy, J. M., Oakley, A. E., Bloxham, C. A., & Edwardson, J. A. (1992). Histochemical distribution of non-haem iron in the human brain. *Acta Anatomica*, *144*(3), 235–257. doi:10.1159/000147312 PMID:1529678

Morris, G. P., Clark, I. A., & Vissel, B. (2014). Inconsistencies and Controversies Surrounding the Amyloid Hypothesis of Alzheimer's Disease. *Acta Neuropathologica Communications*, 2. Retrieved from http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4207354/ PMID:25231068

Mosconi, L., de Leon, M., Murray, J., Lu, J., Javier, E., McHugh, P., & Swerdlow, R. H. (2011). Reduced mitochondria cytochrome oxidase activity in adult children of mothers with Alzheimer's disease. *Journal of Alzheimer's Disease*, 27(3), 483–490. PMID:21841246

Moulton, P. V., & Yang, W. (2012). Air pollution, oxidative stress, and Alzheimer's disease. *Journal of Environmental and Public Health*, 2012, 472751. doi:10.1155/2012/472751 PMID:22523504

Mounsey, R. B., & Teismann, P. (2011). Mitochondrial dysfunction in Parkinson's disease: Pathogenesis and neuroprotection. *Parkinson's Disease*. PMID:21234411

Mourelatos, Z., Adler, H., Hirano, A., Donnenfeld, H., Gonatas, J. O., & Gonatas, N. K. (1990). Fragmentation of the Golgi apparatus of motor neurons in amyotrophic lateral sclerosis revealed by organelle-specific antibodies. *Proceedings of the National Academy of Sciences of the United States of America*, 87(11), 4393–4395. doi:10.1073/pnas.87.11.4393 PMID:2349244

Muchowski, P. J., & Wacker, J. L. (2005a). Modulation of neurodegeneration by molecular chaperones. *Nature Reviews*. *Neuroscience*, *6*(1), 11–22. doi:10.1038/nrn1587 PMID:15611723

Mucke, L., Abraham, C. R., Ruppe, M. D., Rockenstein, E. M., Toggas, S. M., Mallory, M., ... Masliah, E. (1995). Protection against HIV-1 gp120-induced brain damage by neuronal expression of human amyloid precursor protein. *The Journal of Experimental Medicine*, *181*(4), 1551–6. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/7699335

Muhammad, S. A. Z., & Sandrine, T. (2012). Nutrition, adult hippocampal neurogenesis and mental health. *British Medical Bulletin*, 103(1), 89–114. doi:10.1093/bmb/lds021 PMID:22833570

Munhoz, R. P., Cerasa, A., & Okun, M. S. (2014). Surgical treatment of dyskinesia in Parkinson's disease. *Frontiers in Neurology*, *5*, 65. doi:10.3389/fneur.2014.00065 PMID:24808889

Muoio, V., Persson, P. B., & Sendeski, M. M. (2014). The neurovascular unit - concept review. *Acta Physiologica (Oxford, England)*, 210(4), 790–798. doi:10.1111/apha.12250 PMID:24629161

Murphy, D. D., Rueter, S. M., Trojanowski, J. Q., & Lee, V. M. Y. (2000). Synucleins are developmentally expressed, and α -synuclein regulates the size of the presynaptic vesicular pool in primary hippocampal neurons. *The Journal of Neuroscience*, 20(9), 3214–3220. PMID:10777786

Murphy, J., Henry, R., & Lomen-Hoerth, C. (2007). Establishing subtypes of the continuum of frontal lobe impairment in amyotrophic lateral sclerosis. *Archives of Neurology*, *64*(3), 330–334. doi:10.1001/archneur.64.3.330 PMID:17353375

Murphy, M. P. (2009). How mitochondria produce reactive oxygen species. *The Biochemical Journal*, 417(1), 1–13. doi:10.1042/BJ20081386 PMID:19061483

Murphy, M. P., & LeVine, H. (2010). Alzheimer's Disease and the β-Amyloid Peptide. *Journal of Alzheimer's disease*. *JAD*, *19*(1), 311. doi:10.3233/JAD-2010-1221 PMID:20061647

Murray, C. J. L., Vos, T., Lozano, R., Naghavi, M., Flaxman, A. D., Michaud, C., ... Lopez, A. D. (2012). Disability adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: A systematic analysis for the Global Burden of Disease Study 2010. *Lancet*, *380*(9859), 2197–2223. doi:10.1016/S0140-6736(12)61689-4 PMID:23245608

Murray, R. K., & Davis, J. C. (2003). Harper's Illustrated Biochemistry. New York: McGraw-Hill Medical.

Musatov, A., & Robinson, N. C. (2012). Susceptibility of mitochondrial electron-transport complexes to oxidative damage. Focus on cytochrome c oxidase. *Free Radical Research*, *46*(11), 1313–1326. doi:10.3109/10715762.2012.71727 3 PMID:22856385

Myers, S. M., Johnson, C. P., & Lipkin, P. H. (2007). Management of children with autism spectrum disorders. *Pediatrics*, *120*(5), 1162–1182. doi:10.1542/peds.2007-2362 PMID:17967921

Nagańska, E., Taraszewska, A., Matyja, E., Grieb, P., & Rafałowska, J. (2010). Neuroprotective effect of erythropoietin in amyotrophic lateral sclerosis (ALS) model in vitro. Ultrastructural study. *Folia Neuropathologica*, *48*, 35–44. PMID:20383809

Na, H. K., Park, J. H., Kim, J. H., Kim, H. J., Kim, S. T., Werring, D. J., ... Na, D. L. (2015). Cortical superficial siderosis: A marker of vascular amyloid in patients with cognitive impairment. *Neurology*, *84*(8), 849–855. doi:10.1212/ WNL.000000000001288 PMID:25632096

Nakamura, K. (2013). α-Synuclein and mitochondria: Partners in crime? *Neurotherapeutics; the Journal of the American Society for Experimental NeuroTherapeutics, 10*(3), 391–399. doi:10.100713311-013-0182-9 PMID:23512373

Nakamura, K., Christine, C. W., Starr, P. A., & Marks, W. J. Jr. (2007). Effects of unilateral subthalamic and pallidal deep brain stimulation on fine motor functions in Parkinson's disease. *Movement Disorders*, 22(5), 619–626. doi:10.1002/mds.21300 PMID:17230483

Nakano, I., & Hirano, A. (1984). Parkinson's disease: Neuron loss in the nucleus basalis without concomitant Alzheimer's disease. *Annals of Neurology*, *15*(5), 415–418. doi:10.1002/ana.410150503 PMID:6732189

Nakano, Y., Hirayama, K., & Terao, K. (1987). Hepatic ultrastructural changes and liver dysfunction in amyotrophic lateral sclerosis. *Archives of Neurology*, 44(1), 103–106. doi:10.1001/archneur.1987.00520130079022 PMID:3800708

Namavar, Y., Barth, P. G., & Baas, F. (2011). Classification, diagnosis and potential mechanisms in pontocerebellar hypoplasia. *Orphanet Journal of Rare Diseases*, *6*(1), 50. doi:10.1186/1750-1172-6-50 PMID:21749694

Näntö-Salonen, K., Muller, H. L., Hoffman, A. R., Vu, T. H., & Rosenfeld, R. G. (1993). Mechanisms of thyroid hormone action on the insulin-like growth factor system: All thyroid hormone effects are not growth hormone mediated. *Endocrinology*, *132*(2), 781–788. doi:10.1210/endo.132.2.7678799 PMID:7678799

Napolitano, A., Pezzella, A., & Prota, G. (1999). New reaction pathways of dopamine under oxidative stress conditions: Nonenzymatic iron-assisted conversion to norepinephrine and the neurotoxins 6-hydroxydopamine and 6,7-dihydroxytet-rahydroisoquinoline. *Chemical Research in Toxicology*, *12*(11), 1090–1097. doi:10.1021/tx990079p PMID:10563835

Narendra, D. P., Kane, L. A., Hauser, D. N., Fearnley, I. M., & Youle, R. J. (2010). p62/SQSTM1 is required for Parkininduced mitochondrial clustering but not mitophagy; VDAC1 is dispensable for both. *Autophagy*, *6*(8), 1090–1106. doi:10.4161/auto.6.8.13426 PMID:20890124

Narendra, D., Tanaka, A., Suen, D. F., & Youle, R. J. (2008). Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. *The Journal of Cell Biology*, *183*(5), 795–803. doi:10.1083/jcb.200809125 PMID:19029340

Navarro, A., & Boveris, A. (2007a). Brain mitochondrial dysfunction in aging: conditions that improve survival, neurological performance and mitochondrial function. *Frontiers in Bioscience: A Journal & Virtual Library*, *12*, 1154-1163.

Navarro, A., & Boveris, A. (2007b). The mitochondrial energy transduction system and the aging process. *American Journal of Physiology. Cell Physiology*, 292(2), C670–C686. doi:10.1152/ajpcell.00213.2006 PMID:17020935

Navarro, A., & Boveris, A. (2010). Brain Mitochondrial Dysfunction in Aging, Neurodegeneration, and Parkinson's Disease. *Frontiers in Aging Neuroscience*, *2*, 34. PMID:20890446

Navarro, A., Gomez, C., López-Cepero, J. M., & Boveris, A. (2004). Beneficial effects of moderate exercise on mice aging: Survival, behavior, oxidative stress, and mitochondrial electron transfer. *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology*, 286(3), R505–R511. doi:10.1152/ajpregu.00208.2003 PMID:14615275

Navlakha, S., & Kingsford, C. (2010). The power of protein interaction networks for associating genes with diseases. *Bioinformatics (Oxford, England)*, 26(8), 1057–1063. doi:10.1093/bioinformatics/btq076 PMID:20185403

Nearly 1 in 6 of world's population suffer from neurological disorders – UN report. (2007). Retrieved from http://www. un.org/apps/news/story.asp?cr=neurological&newsid=21689#.WhBcS2iCzIU

Neary, D., Snowden, J. S., Gustafson, L., Passant, U., Stuss, D., Black, S., ... Benson, D. F. (1998). Frontotemporal lobar degeneration: A consensus on clinical diagnostic criteria. *Neurology*, *51*(6), 1546–1554. doi:10.1212/WNL.51.6.1546 PMID:9855500

Neary, D., Snowden, J. S., & Mann, D. M. (1993). The clinical pathological correlates of lobar atrophy. *Dementia (Basel, Switzerland)*, 4(3-4), 154–159. PMID:8401784

Nègre-Pagès, L., Regragui, W., Bouhassira, D., Grandjean, H., & Rascol, O. (2008). Chronic pain in Parkinson's disease: The cross-sectional French DoPaMiP survey. *Movement Disorders*, 23(10), 1361–1369. doi:10.1002/mds.22142 PMID:18546344

Nelson, L. M., McGuire, V., Longstreth, W. T., & Matkin, C. (2000). Population-based case-control study of amyotrophic lateral sclerosis in western Washington State. I. Cigarette smoking and alcohol consumption. *American Journal of Epidemiology*, 151(2), 156–163. doi:10.1093/oxfordjournals.aje.a010183 PMID:10645818

Neudorfer, O., Giladi, N., Elstein, D., Abrahamov, A., Turezkite, T., Aghai, E., ... Zimran, A. (1996). Occurrence of Parkinson's syndrome in type I Gaucher disease. *QJM: an International Journal of Medicine*, *89*(9), 691–694. doi:10.1093/ qjmed/89.9.691 PMID:8917744

Neumann, M., Bentmann, E., Dormann, D., Jawaid, A., DeJesus-Hernandez, M., Ansorge, O., ... Mackenzie, I. R. A. (2011). FET proteins TAF15 and EWS are selective markers that distinguish FTLD with FUS pathology from amyo-trophic lateral sclerosis with FUS mutations. *Brain*, *134*(Pt 9), 2595–2609. doi:10.1093/brain/awr201 PMID:21856723

Neumann, M., Rademakers, R., Roeber, S., Baker, M., Kretzschmar, H., & Mackenzie, I. R. (2009). A new subtype of frontotemporal lobar degeneration with FUS pathology. *Brain*, *132*(11), 2922–2931. doi:10.1093/brain/awp214 PMID:19674978

Neumann, M., Sampathu, D. M., Kwong, L. K., Truax, A. C., Micsenyi, M. C., Chou, T. T., ... Lee, V. M.-Y. (2006). Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science*, *314*(5796), 130–133. doi:10.1126cience.1134108 PMID:17023659

NeuroGenetic Pharmaceuticals. (2017). *Products and Technology*. Retrieved March 7, 2017, from http://neurogenet-icpharmaceuticals.com/

Newmeyer, D. D., & Ferguson-Miller, S. (2003). Mitochondria: Releasing power for life and unleashing the machineries of death. *Cell*, *112*(4), 481–490. doi:10.1016/S0092-8674(03)00116-8 PMID:12600312

Newschaffer, C. J., Croen, L. A., Daniels, J., Giarelli, E., Grether, J. K., Levy, S. E., ... Windham, G. C. (2007). The epidemiology of autism spectrum disorders. *Annual Review of Public Health*, 28(1), 235–258. doi:10.1146/annurev. publhealth.28.021406.144007 PMID:17367287

Nguyen, B. M., Kim, D., Bricker, S., Bongard, F., Neville, A., Putnam, B., ... Plurad, D. (2014). Effect of marijuana use on outcomes in traumatic brain injury. *The American Surgeon*, *80*(10), 979–983. PMID:25264643

Nguyen, T.-P., & Ho, T.-B. (2012). Detecting disease genes based on semi-supervised learning and protein-protein interaction networks. *Artificial Intelligence in Medicine*, 54(1), 63–71. doi:10.1016/j.artmed.2011.09.003 PMID:22000346

Nguyen, T.-P., Liu, W., & Jordán, F. (2011). Inferring pleiotropy by network analysis: Linked diseases in the human PPI network. *BMC Systems Biology*, *5*(1), 179. doi:10.1186/1752-0509-5-179 PMID:22034985

Nicholls, D. G., & Budd, S. L. (2000). Mitochondria and neuronal survival. *Physiological Reviews*, 80(1), 315–360. doi:10.1152/physrev.2000.80.1.315 PMID:10617771

Niciu, M. J., Kelmendi, B., & Sanacora, G. (2012). Overview of glutamatergic neurotransmission in the nervous system. *Pharmacology, Biochemistry, and Behavior, 100*(4), 656–664. doi:10.1016/j.pbb.2011.08.008 PMID:21889952

Nicklas, W. J., Youngster, S. K., Kindt, M. V., & Heikkila, R. E. (1987). IV. MPTP, MPP+ and mitochondrial function. *Life Sciences*, 40(8), 721–729. doi:10.1016/0024-3205(87)90299-2 PMID:3100899

Niklasso, L., Rasmussen, P., Oskarsdottir, S., & Gillberg, C. (2001). Neuropsychiatric disorders in the 22q11 deletion syndrome. *Genetics in Medicine*, *3*(1), 79–84. doi:10.1097/00125817-200101000-00017 PMID:11339385

Niklasson, L., Rasmussen, P., Oskarsdottir, S., & Gillberg, C. (2002). Chromosome 22q11 deletion syndrome (CATCH 22): Neuropsychiatric and neuropsychological aspects. *Developmental Medicine and Child Neurology*, 44(01), 44–50. doi:10.1017/S0012162201001645 PMID:11811651

Nilsson, P., Iwata, N., Muramatsu, S., Tjernberg, L. O., Winblad, B., & Saido, T. C. (2010). Gene therapy in Alzheimer's disease – potential for disease modification. *Journal of Cellular and Molecular Medicine*, *14*(4), 741–757. doi:10.1111/j.1582-4934.2010.01038.x PMID:20158567

Niranjan, B. G., Bhat, N. K., & Avadhani, N. G. (1982). Preferential attack of mitochondrial DNA by aflatoxin B1 during hepatocarcinogenesis. *Science*, *215*(4528), 73–75. doi:10.1126cience.6797067 PMID:6797067

Nisbet, R. M., Polanco, J.-C., Ittner, L. M., & Götz, J. (2015). Tau aggregation and its interplay with amyloid-β. *Acta Neuropathologica*, *129*(2), 207–220. doi:10.100700401-014-1371-2 PMID:25492702

532

Nishikawa, K., Li, H., Kawamura, R., Osaka, H., Wang, Y. L., Hara, Y., ... Aoki, S. (2003). Alterations of structure and hydrolase activity of parkinsonism-associated human ubiquitin carboxyl-terminal hydrolase L1 variants. *Biochemical and Biophysical Research Communications*, 304(1), 176–183. doi:10.1016/S0006-291X(03)00555-2 PMID:12705903

Nishioka, K., Hayashi, S., Farrer, M. J., Singleton, A. B., Yoshino, H., Imai, H., ... Mizoguchi, K. (2006). Clinical heterogeneity of α -synuclein gene duplication in Parkinson's disease. *Annals of Neurology*, 59(2), 298–309. doi:10.1002/ ana.20753 PMID:16358335

Niu, J., Yu, M., Wang, C., & Xu, Z. (2012). Leucine-rich repeat kinase 2 disturbs mitochondrial dynamics via Dynamin-like protein. *Journal of Neurochemistry*, *122*(3), 650–658. doi:10.1111/j.1471-4159.2012.07809.x PMID:22639965

Nixon, R. A., & Yang, D.-S. (2012). Autophagy and neuronal cell death in neurological disorders. *Cold Spring Harbor Perspectives in Biology*, 4(10), a008839. doi:10.1101/cshperspect.a008839 PMID:22983160

Núñez-Santana, F. L., Oh, M. M., Antion, M. D., Lee, A., Hell, J. W., & Disterhoft, J. F. (2014). Surface L-type Ca2+ channel expression levels are increased in aged hippocampus. *Aging Cell*, *13*(1), 111–120. doi:10.1111/acel.12157 PMID:24033980

Nussbaum, J. M., Schilling, S., Cynis, H., Silva, A., Swanson, E., Wangsanut, T., ... Bloom, G. S. (2012). Prion-like behaviour and tau-dependent cytotoxicity of pyroglutamylated amyloid-β. *Nature*, *485*(7400), 651–655. doi:10.1038/ nature11060 PMID:22660329

Nussbaum, R. L., & Ellis, C. E. (2003). Alzheimer's disease and Parkinson's disease. *The New England Journal of Medicine*, 348(14), 1356–1364. doi:10.1056/NEJM2003ra020003 PMID:12672864

Nuvolone, M., Schmid, N., Miele, G., Sorce, S., Moos, R., Schori, C., & Aguzzi, A. (2017). Cystatin F is a biomarker of prion pathogenesis in mice. *PLoS One*, *1*(10), 1–25. PMID:28178353

O'Brien, J. T., & Thomas, A. (2015). Vascular dementia. *Lancet*, 386(10004), 1698–1706. doi:10.1016/S0140-6736(15)00463-8 PMID:26595643

O'Brien, R. J., & Wong, P. C. (2011). Amyloid precursor protein processing and Alzheimer's disease. *Annual Review of Neuroscience*, *34*(1), 185–204. doi:10.1146/annurev-neuro-061010-113613 PMID:21456963

O'Donnell, M. J., Chin, S. L., Rangarajan, S., Xavier, D., Liu, L., Zhang, H., ... Yusuf, S. (2016). Global and regional effects of potentially modifiable risk factors associated with acute stroke in 32 countries (INTERSTROKE), a case-control study. *Lancet*, *388*(10046), 761–775. doi:10.1016/S0140-6736(16)30506-2 PMID:27431356

Obwegeser, A. A., Uitti, R. J., Turk, M. F., Strongosky, A. J., & Wharen, R. E. (2000). Thalamic stimulation for the treatment of midline tremors in essential tremor patients. *Neurology*, *54*(12), 2342–2344. doi:10.1212/WNL.54.12.2342 PMID:10881269

Oczkowska, A., Kozubski, W., Lianeri, M., & Dorszewska, J. (2013). Mutations in PRKN and SNCA Genes Important for the Progress of Parkinson's Disease. *Current Genomics*, *14*(8), 502–517. doi:10.2174/13892029146661312102058 39 PMID:24532983

Oddo, S. (2008). The ubiquitin-proteasome system in Alzheimer's disease. *Journal of Cellular and Molecular Medicine*, *12*(2), 363–373. doi:10.1111/j.1582-4934.2008.00276.x PMID:18266959

Oddo, S., Caccamo, A., Shepherd, J. D., Murphy, M. P., Golde, T. E., Kayed, R., ... LaFerla, F. M. (2003). Triple-transgenic model of Alzheimer's disease with plaques and tangles: Intracellular Abeta and synaptic dysfunction. *Neuron*, *39*(3), 409–421. doi:10.1016/S0896-6273(03)00434-3 PMID:12895417

Odekerken, V. J., van Laar, T., Staal, M. J., Mosch, A., Hoffmann, C. F., Nijssen, P. C., ... de Bie, R. M. A. (2013, January). Subthalamic nucleus versus globus pallidus bilateral deep brain stimulation for advanced Parkinson's disease (NSTAPS study): A randomised controlled trial. *Lancet Neurology*, *12*(1), 37–44. doi:10.1016/S1474-4422(12)70264-8 PMID:23168021

Oestreicher, E., Sengstock, G. J., Riederer, P., Olanow, C. W., Dunn, A. J., & Arendash, G. W. (1994). Degeneration of nigrostriatal dopaminergic neurons increases iron within the substantia nigra: A histochemical and neurochemical study. *Brain Research*, *660*(1), 8–18. doi:10.1016/0006-8993(94)90833-8 PMID:7828004

Ofman, R., Dijkstra, I. M. E., Van Roermund, C. W. T., Burger, N., Turkenburg, M., Van Cruchten, A., ... Kemp, S. (2010). The role of ELOVL1 in very long-chain fatty acid homeostasis and X-linked adrenoleukodystrophy. *EMBO Molecular Medicine*, 2(3), 90–97. doi:10.1002/emmm.201000061 PMID:20166112

Ogawa, S., Tank, D. W., Menon, R., Ellermann, J. M., Kim, S. G., Merkle, H., & Ugurbil, K. (1992). Intrinsic signal changes accompanying sensory stimulation: Functional brain mapping with magnetic resonance imaging. *Proceedings of the National Academy of Sciences of the United States of America*, 89(13), 5951–5955. doi:10.1073/pnas.89.13.5951 PMID:1631079

Ogilvie, C. M., Moore, J., Daker, M., Palferman, S., & Docherty, Z. (2009). Chromosome 22q11 deletions are not found in autistic patients identified using strict diagnostic criteria. International Molecular Genetics Study of Autism Consortium. *American Journal of Medical Genetics*, *96*(1), 15–17. doi: PMID:10686546

Ohnishi, T., Matsuda, H., Hashimoto, T., Kunihiro, T., Nishikawa, M., Uema, T., & Sasaki, M. (2000). Abnormal regional cerebral blood flow in childhood autism. *Brain*, *123*(9), 1838–1844. doi:10.1093/brain/123.9.1838 PMID:10960047

Ohta, Y., Yashiro, K., Ohashi, K., & Imai, Y. (2012). Disruption of non-enzymatic antioxidant defense systems in the brain of rats with water-immersion restraint stress. *Journal of Clinical Biochemistry and Nutrition*, *51*(2), 136–142. doi:10.3164/jcbn.11-14 PMID:22962533

Ohyagi, Y., Yamada, T., Nishioka, K., Clarke, N. J., Tomlinson, A. J., Naylor, S., ... Younkin, S. G. (2000). Selective increase in cellular A β 42 is related to apoptosis but not necrosis. *Neuroreport*, *11*(1), 167–171. doi:10.1097/00001756-200001170-00033 PMID:10683851

Okazaki, H., Lipkin, L. E., & Aronson, S. M. (1961). Diffuse intracytoplasmic ganglionic inclusions (Lewy type) associated with progressive dementia and quadriparesis in flexion. *Journal of Neuropathology and Experimental Neurology*, 20(2), 237–244. doi:10.1097/00005072-196104000-00007 PMID:13730588

Okun, M. S., & Foote, K. D. (2010, December). Parkinson's disease DBS: What, when, who and why? The time has come to tailor DBS targets. *Expert Review of Neurotherapeutics*, *10*(12), 1847–1857. doi:10.1586/ern.10.156 PMID:21384698

Olanow, C. W., Goetz, C. G., Kordower, J. H., Stoessl, A. J., Sossi, V., Brin, M. F., ... Freeman, T. B. (2003, September). A double-blind controlled trial of bilateral fetal nigral transplantation in Parkinson's disease. *Annals of Neurology*, *54*(3), 403–414. doi:10.1002/ana.10720 PMID:12953276

Olanow, C. W., & Tatton, W. G. (1999). Etiology and pathogenesis of Parkinson's disease. *Annual Review of Neuroscience*, 22(1), 123–144. doi:10.1146/annurev.neuro.22.1.123 PMID:10202534

Oliveira, A. S. B., & Pereira, R. D. B. (2009). Amytrophic lateral Sclerosis (ALS): Three letters that change people's life. For ever. *Arquivos de Neuro-Psiquiatria*, 67(3a), 750–782. doi:10.1590/S0004-282X2009000400040 PMID:19722069

Oliveira, J. (2010). Nature and cause of mitochondrial dysfunction in Huntington's disease: Focusing on huntingtin and the striatum. *Journal of Neurochemistry*, *114*(1), 1–12. PMID:20403078

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Olkowski, Z. L. (1998). Mutant AP endonuclease in patients with amyotrophic lateral sclerosis. *Neuroreport*, 9(2), 239–242. doi:10.1097/00001756-199801260-00012 PMID:9507962

Ondo, W. (2016). Essential Tremor: What We Can Learn from Current Pharmacotherapy. *Tremor and Other Hyperkinetic Movements (New York, N.Y.)*, *6*, 356. PMID:26989572

Orchard, J. P., & Tolar, J. (2010, January). Transplant Outcomes in Leukodystrophies. *Seminars in Hematology*, 47(1), 70–78. doi:10.1053/j.seminhematol.2009.10.006 PMID:20109614

Orosz, F., Kovács, G. G., Lehotzky, A., Oláh, J., Vincze, O., & Ovádi, J. (2004). TPPP/p25: from unfolded protein to misfolding disease: prediction and experiments. *Biology of the Cell*, *96*(9), 701–711. doi:10.1016/j.biolcel.2004.08.002 PMID:15567525

Orr, A. L., Li, S., Wang, C. E., Li, H., Wang, J., Rong, J., & Li, X. J. (2008). N-terminal mutant huntingtin associates with mitochondria and impairs mitochondrial trafficking. *The Journal of Neuroscience*, 28(11), 2783–2792. doi:10.1523/JNEUROSCI.0106-08.2008 PMID:18337408

Ortolano, S., Vieitez, I., Agis-Balboa, R. C., & Spuch, C. (2014). Loss of GABAergic cortical neurons underlies the neuropathology of Lafora disease. *Molecular Brain*, 7(1), 7. doi:10.1186/1756-6606-7-7 PMID:24472629

Ota, M., Yasuno, F., Ito, H., Seki, C., Nozaki, S., Asada, T., & Suhara, T. (2006). Age-related decline of dopamine synthesis in the living human brain measured by positron emission tomography with L-[β-11 C] DOPA. *Life Sciences*, 79(8), 730–736. doi:10.1016/j.lfs.2006.02.017 PMID:16580023

Oti, M., Snel, B., Huynen, M. A., & Brunner, H. G. (2006). Predicting disease genes using protein-protein interactions. *Journal of Medical Genetics*, *43*(8), 691–698. doi:10.1136/jmg.2006.041376 PMID:16611749

O-Uchi, J., Pan, S., & Sheu, S.-S. (2012). Perspectives on: SGP symposium on mitochondrial physiology and medicine: molecular identities of mitochondrial Ca^{2+} influx mechanism: updated passwords for accessing mitochondrial Ca^{2+} -linked health and disease. *The Journal of General Physiology*, *139*(6), 435–443. doi:10.1085/jgp.201210795 PMID:22641638

Ou, S. H., Wu, F., Harrich, D., & ... (1995). Cloning and characterization of a novel cellular protein, TDP-43, that binds to human immunodeficiency virus type 1 TAR DNA sequence motifs. *Journal of Virology*, *69*(6), 3584–3596. PMID:7745706

Ovbiagele, B., Goldstein, L. B., Higashida, R. T., Howard, V. J., Johnston, S. C., Khavjou, O. A., ... Trogdon, J. G. (2013). Forecasting the future of stroke in the United States: A policy statement from the American Heart Association and American Stroke Association. *Stroke*, 44(8), 2361–2375. doi:10.1161/STR.0b013e31829734f2 PMID:23697546

Owsley, C., & McGwin, G. Jr. (2010). Vision and Driving. *Vision Research*, 50(23), 2348–2361. doi:10.1016/j. visres.2010.05.021 PMID:20580907

Oyewole, A. O., & Birch-Machin, M. A. (2015). Mitochondria-targeted antioxidants. *The FASEB Journal*, 29(12), 4766–4771. doi:10.1096/fj.15-275404 PMID:26253366

Pacher, P., Beckman, J. S., & Liaudet, L. (2007). Nitric oxide and peroxynitrite in health and disease. *Physiological Reviews*, 87(1), 315–424. doi:10.1152/physrev.00029.2006 PMID:17237348

Pagliarini, D. J., & Rutter, J. (2013). Hallmarks of a new era in mitochondrial biochemistry. *Genes & Development*, 27(24), 2615–2627. doi:10.1101/gad.229724.113 PMID:24352419

Paisan-Ruiz, C., Lang, A. E., Kawarai, T., Sato, C., Salehi-Rad, S., Fisman, G. K., & Rogaeva, E. (2005). LRRK2 gene in Parkinson disease Mutation analysis and case control association study. *Neurology*, *65*(5), 696–700. doi:10.1212/01. WNL.0000167552.79769.b3 PMID:16157901

Paloczi, J., Varga, Z. V., Hasko, G., & Pacher, P. (in press). Neuroprotection in Oxidative Stress-Related Neurodegenerative Diseases: Role of Endocannabinoid System Modulation. *Antioxidants & Redox Signalling*. doi:10.1089/ars.2017.7144

Palop, J. J., Chin, J., & Mucke, L. (2006). A network dysfunction perspective on neurodegenerative diseases. *Nature*, 443(7113), 768–773. doi:10.1038/nature05289 PMID:17051202

Pamplona, R., Naudí, A., Gavín, R., Pastrana, M. A., Sajnani, G., Ilieva, E. V., ... Requena, J. R. (2008). Increased oxidation, glycoxidation, and lipoxidation of brain proteins in prion disease. *Free Radical Biology & Medicine*, 45(8), 1159–1166. doi:10.1016/j.freeradbiomed.2008.07.009 PMID:18703134

Panda, S. K., Wefers, B., Ortiz, O., Floss, T., Schmid, B., Haass, C., ... Kuhn, R. (2013). Highly efficient targeted mutagenesis in mice using TALENs. *Genetics*, 195(3), 703–713. doi:10.1534/genetics.113.156570 PMID:23979585

Panday, A., Sahoo, M. K., Osorio, D., & Batra, S. (2015). NADPH oxidases: An overview from structure to innate immunity-associated pathologies. *Cellular & Molecular Immunology*, *12*(1), 5–23. doi:10.1038/cmi.2014.89 PMID:25263488

Panegyres, P. K., & Armari, E. (2013). Therapies for human prion diseases. *American Journal of Neurodegenerative Disease*, 2(3), 176. PMID:24093082

Panigrahi, P. P., & Singh, T. R. (2012). Computational analysis for functional and evolutionary aspects of BACE-1 and associated Alzheimer's related proteins. *International Journal of Computational Intelligence Studies*, *1*(4), 322–332. doi:10.1504/IJCISTUDIES.2012.050355

Panigrahi, P. P., & Singh, T. R. (2013). Computational studies on Alzheimer's disease associated pathways and regulatory patterns using microarray gene expression and network data: Revealed association with aging and other diseases. *Journal of Theoretical Biology*, *334*, 109–121. doi:10.1016/j.jtbi.2013.06.013 PMID:23811083

Panov, A. V., Gutekunst, C.-A., Leavitt, B. R., Hayden, M. R., Burke, J. R., Strittmatter, W. J., & Greenamyre, J. T. (2002). Early mitochondrial calcium defects in Huntington's disease are a direct effect of polyglutamines. *Nature Neuroscience*, *5*(8), 731–736. doi:10.1038/nn884 PMID:12089530

Pantoni, L. (2017). Have Stroke Neurologists Entered the Arena of Stroke-Related Cognitive Dysfunctions? Not Yet, but They Should! *Stroke*, *48*(6), 1441–1442. doi:10.1161/STROKEAHA.117.016869 PMID:28487330

Paoletti, P., Vergnano, A. M., Barbour, B., & Casado, M. (2009). Zinc at glutamatergic synapses. *Neuroscience*, *158*(1), 126–136. doi:10.1016/j.neuroscience.2008.01.061 PMID:18353558

Papadia, S., Soriano, F. X., Léveillé, F., Martel, M. A., Dakin, K. A., Hansen, H. H., ... Hardingham, G. E. (2008). Synaptic NMDA receptor activity boosts intrinsic antioxidant defenses. *Nature Neuroscience*, *11*(4), 476–487. doi:10.1038/ nn2071 PMID:18344994

Pappolla, M. A., Omar, R. A., Kim, K. S., & Robakis, N. K. (1992). Immunohistochemical evidence of oxidative [corrected] stress in Alzheimer's disease. *American Journal of Pathology*, *140*(3), 621–628. Retrieved from http://www.ncbi. nlm.nih.gov/pubmed/1372157 PMID:1372157

Paradies, G., Paradies, V., de Benedictis, V., Ruggiero, F. M., & Petrosillo, G. (2014). Functional role of cardiolipin in mitochondrial bioenergetics. *Biochimica et Biophysica Acta*, *1837*(4), 408–417. doi:10.1016/j.bbabio.2013.10.006 PMID:24183692

Parchi, P., Capellari, S., Chen, S. G., Petersen, R. B., Gambetti, P., Kopp, N., ... Kretzschmar, H. (1997). Typing prion isoforms. *Nature*, *386*(6622), 232–234. doi:10.1038/386232a0 PMID:9069279

Parchi, P., Castellani, R., Capellari, S., Ghetti, B., Young, K., Chen, S. G., ... Trojanowski, J. Q. (1996). Molecular basis of phenotypic variability in sporadc creudeldt-jakob disease. *Annals of Neurology*, *39*(6), 767–778. doi:10.1002/ ana.410390613 PMID:8651649

Pardo, B., Mena, M. A., Casarejos, M. J., Paino, C. L., & De Yébenes, J. G. (1995). Toxic effects of L-DOPA on mesencephalic cell cultures: Protection with antioxidants. *Brain Research*, 682(1), 133–143. doi:10.1016/0006-8993(95)00341-M PMID:7552304

Parihar, M. S., Parihar, A., Fujita, M., Hashimoto, M., & Ghafourifar, P. (2008). Mitochondrial association of alphasynuclein causes oxidative stress. *Cellular and Molecular Life Sciences*, 65(7), 1272–1284. doi:10.100700018-008-7589-1 PMID:18322646

Parihar, M. S., Parihar, A., Fujita, M., Hashimoto, M., & Ghafourifar, P. (2009). Alpha-synuclein overexpression and aggregation exacerbates impairment of mitochondrial functions by augmenting oxidative stress in human neuroblastoma cells. *The International Journal of Biochemistry & Cell Biology*, *41*(10), 2015–2024. doi:10.1016/j.biocel.2009.05.008 PMID:19460457

Parker, W. D., Boyson, S. J., & Parks, J. K. (1989). Abnormalities of the electron transport chain in idiopathic Parkinson's disease. *Annals of Neurology*, *26*(6), 719–723. doi:10.1002/ana.410260606 PMID:2557792

Parkinson, J. (1817). An Essay on the Shaking Palsy. London: Whittingham and Rowland.

Parkinson, J. (1817). An Essay on the Shaking Palsy. London: Sherwood, Neely, and Jones.

Park, J., Kim, S. Y., Cha, G. H., Lee, S. B., Kim, S., & Chung, J. (2005). Drosophila DJ-1 mutants show oxidative stresssensitive locomotive dysfunction. *Gene*, *361*, 133–139. doi:10.1016/j.gene.2005.06.040 PMID:16203113

Pase, M. P., Satizabal, C. L., & Seshadri, S. (2017). Role of Improved Vascular Health in the Declining Incidence of Dementia. *Stroke*, *48*(7), 2013–2020. doi:10.1161/STROKEAHA.117.013369 PMID:28596460

Pasinelli, P., & Brown, R. H. (2006). Molecular biology of amyotrophic lateral sclerosis: Insights from genetics. *Nature Reviews. Neuroscience*, 7(9), 710–723. doi:10.1038/nrn1971 PMID:16924260

Passero, S., Burgalassi, L., D'Andrea, P., & Battistini, N. (1995). Recurrence of bleeding in patients with primary intracerebral hemorrhage. *Stroke*, *26*(7), 1189–1192. doi:10.1161/01.STR.26.7.1189 PMID:7604411

Patel, N. K., Heywood, P., O'Sullivan, K., McCarter, R., Love, S., & Gill, S. S. (2003, May). Unilateral subthalamotomy in the treatment of Parkinson's disease. *Brain*, *126*(Pt 5), 1136–1145. doi:10.1093/brain/awg111 PMID:12690053

Paterna, J. C., Leng, A., Weber, E., Feldon, J., & Büeler, H. (2007). DJ-1 and Parkin modulate dopamine-dependent behavior and inhibit MPTP-induced nigral dopamine neuron loss in mice. *Molecular Therapy*, *15*(4), 698–704. doi:10.1038j. mt.6300067 PMID:17299411

Paulson, H. L. (2009). The spinocerebellar ataxias. *Journal of Neuro-Ophthalmology*, 29(3), 227–237. doi:10.1097/WNO0b013e3181b416de PMID:19726947

Paulson, H. L., Das, S. S., Crino, P. B., Perez, M. K., Patel, S. C., Gotsdiner, D., ... Pittman, R. N. (1997). Machado-Joseph disease gene product is a cytoplasmic protein widely expressed in brain. *Annals of Neurology*, *41*(4), 453–462. doi:10.1002/ana.410410408 PMID:9124802

Pavlica, S., & Gebhardt, R. (2000). Comparison of uptake and neuroprotective potential of seven zinc-salts. *Neurochemistry International*, *56*(1), 84–93. doi:10.1016/j.neuint.2009.09.005 PMID:19782114

Paxinou, E., Chen, Q., Weisse, M., Giasson, B. I., Norris, E. H., Rueter, S. M., ... Ischiropoulos, H. (2001). Induction of alpha-synuclein aggregation by intracellular nitrative insult. *Journal of Neurology*, *15*, 8053–8061. PMID:11588178

Payne, B. A. I., & Chinnery, P. F. (2015). Mitochondrial dysfunction in aging: Much progress but many unresolved questions. *Biochimica et Biophysica Acta*, *1847*(11), 1347–1353. doi:10.1016/j.bbabio.2015.05.022 PMID:26050973

Pearce, L. A., McClure, L. A., Anderson, D. C., Jacova, C., Sharma, M., Hart, R. G., & Benavente, O. R. (n.d.). SPS3 Investigators. Effects of long-term blood pressure lowering and dual antiplatelet treatment on cognitive function in patients with recent lacunar stroke: A secondary analysis from the SPS3 randomised trial. *Lancet Neurology*, *13*(7), 1177–1185. PMID:25453457

Pearson, K. J., Baur, J. A., Lewis, K. N., Peshkin, L., Price, N. L., Labinskyy, N., ... Jamieson, H. A. (2008). Resveratrol delays age-related deterioration and mimics transcriptional aspects of dietary restriction without extending life span. *Cell Metabolism*, 8(2), 157–168. doi:10.1016/j.cmet.2008.06.011 PMID:18599363

Pedersen, W. A., Fu, W., Keller, J. N., Markesbery, W. R., Appel, S., Smith, R. G., ... Mattson, M. P. (1998). Protein modification by the lipid peroxidation product 4-hydroxynonenal in the spinal cords of amyotrophic lateral sclerosis patients. *Annals of Neurology*, *44*(5), 819–824. doi:10.1002/ana.410440518 PMID:9818940

Peila, R., Rodriguez, B. L., & Launer, L. J. (2002). Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: The Honolulu-Asia Aging Study. *Diabetes*, *51*(4), 1256–1262. doi:10.2337/diabetes.51.4.1256 PMID:11916953

Pellegrino, M. W., & Haynes, C. M. (2015). Mitophagy and the mitochondrial unfolded protein response in neurodegeneration and bacterial infection. *BMC Biology*, *13*(1), 22. doi:10.118612915-015-0129-1 PMID:25857750

Pellicano, C., Benincasa, D., Pisani, V., Buttarelli, F. R., Giovannelli, M., & Pontieri, F. E. (2007). Prodromal non-motor symptoms of Parkinson's disease. *Neuropsychiatric Disease and Treatment*, *3*(1), 145–152. doi:10.2147/nedt.2007.3.1.145 PMID:19300544

Pendlebury, S. T. (2009). Stroke-related dementia: Rates, risk factors and implications for future research. *Maturitas*, 64(3), 165–171. doi:10.1016/j.maturitas.2009.09.010 PMID:19818568

Pendlebury, S. T., & Rothwell, P. M. (2009). Prevalence, incidence, and factors associated with pre-stroke and poststroke dementia: A systematic review and meta-analysis. *Lancet Neurology*, 8(11), 1006–1018. doi:10.1016/S1474-4422(09)70236-4 PMID:19782001

Penner, L. A., Blair, I. V., Albrecht, T. L., & Dovidio, J. F. (2014). Reducing racial health care disparities: A social psychological analysis. *Policy Insights from the Behavioral and Brain Sciences*, 1(1), 204–212. doi:10.1177/2372732214548430 PMID:25705721

Peres, R., De Guio, F., Chabriat, H., & Jouvent, E. (2016). Alterations of the cerebral cortex in sporadic small vessel disease: A systematic review of in vivo MRI data. *Journal of Cerebral Blood Flow and Metabolism*, *36*(4), 681–695. doi:10.1177/0271678X15625352 PMID:26787108

Perez-Lloret, S., & Rascol, O. (2010). Dopamine receptor agonists for the treatment of early or advanced Parkinson's disease. *CNS Drugs*, 24(11), 941–968. doi:10.2165/11537810-000000000-00000 PMID:20932066

Pérez-Suárez, J., Torres Díaz,, C.V., López Manzanares,, L., Navas García,, M., Pastor,, J., & Barrio Fernández,, P., & de Sola, R. (2017). Radiofrequency Lesions through Deep Brain Stimulation Electrodes in Movement Disorders: Case Report and Review of the Literature. *Stereotactic and Functional Neurosurgery*, *95*(3), 137–141. doi:10.1159/000454891 PMID:28433987

Perier, C., Tieu, K., Guégan, C., Caspersen, C., Jackson-Lewis, V., Carelli, V., ... Vila, M. (2005). Complex I deficiency primes Bax-dependent neuronal apoptosis through mitochondrial oxidative damage. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(52), 19126–19131. doi:10.1073/pnas.0508215102 PMID:16365298

Perl, D. P. (2010). Neuropathology of Alzheimer â€TM s Disease. *The Mount Sinai Journal of Medicine, New York*, 77(1), 32–42. doi:10.1002/msj.20157 PMID:20101720

Perry, D. K., Smyth, M. J., Stennicke, H. R., Salvesen, G. S., Duriez, P., Poirier, G. G., & Hannun, Y. A. (1997). Zinc is a potent inhibitor of the apoptotic protease, caspase-3. A novel target for zinc in the inhibition of apoptosis. *The Journal of Biological Chemistry*, 272(30), 18530–18533. doi:10.1074/jbc.272.30.18530 PMID:9228015

Perry, E. K., Curtis, M., Dick, D. J., Candy, J. M., Atack, J. R., Bloxham, C. A., ... Perry, R. H. (1985). Cholinergic correlates of cognitive impairment in Parkinson's disease: Comparisons with Alzheimer's disease. *Journal of Neurology, Neurosurgery, and Psychiatry*, 48(5), 413–421. doi:10.1136/jnnp.48.5.413 PMID:3998751

Perry, G., Friedman, R., Shaw, G., & Chau, V. (1987). Ubiquitin is detected in neurofibrillary tangles and senile plaque neurites of Alzheimer disease brains. *Proceedings of the National Academy of Sciences of the United States of America*, 84(9), 3033–3036. doi:10.1073/pnas.84.9.3033 PMID:3033674

Perry, G., Sayre, L. M., Atwood, C. S., Castellani, R. J., Cash, A. D., & Rottkamp, C. A. (2002). The role of iron and copper in the aetiology of neurodegenerative disorders. *Central Nervous System Drugs*, *16*(5), 339–352. PMID:11994023

Perry, V. H., & O'Connor, V. (2010). The role of microglia in synaptic stripping and synaptic degeneration: A revised perspective. *ASN Neuro*, *2*(5), e00047. doi:10.1042/AN20100024 PMID:20967131

Persidsky, Y., Ramirez, S. H., Haorah, J., & Kanmogne, G. D. (2006). Blood-brain barrier: Structural components and function under physiologic and pathologic conditions. *Journal of Neuroimmune Pharmacology*, *1*(3), 223–236. doi:10.100711481-006-9025-3 PMID:18040800

Perumal, A. S., Gopal, V. B., Tordzro, W. K., Cooper, T. B., & Cadet, J. L. (1992). Vitamin E attenuates the toxic effects of 6-hydroxydopamine on free radical scavenging systems in rat brain. *Brain Research Bulletin*, 29(5), 699–701. doi:10.1016/0361-9230(92)90142-K PMID:1422867

Peters, N., Opherk, C., Bergmann, T., Castro, M., Herzog, J., & Dichgans, M. (2005). Spectrum of mutations in biopsyproven CADASIL: Implications for diagnostic strategies. *Archives of Neurology*, *62*(7), 1091–1094. doi:10.1001/archneur.62.7.1091 PMID:16009764

Peters, R. (2006). Ageing and the brain. *Postgraduate Medical Journal*, 82(964), 84–88. doi:10.1136/pgmj.2005.036665 PMID:16461469

Petit, A., Kawarai, T., Paitel, E., Sanjo, N., Maj, M., Scheid, M., ... Wang, L. (2005). Wild-type PINK1 prevents basal and induced neuronal apoptosis, a protective effect abrogated by Parkinson disease-related mutations. *The Journal of Biological Chemistry*, 280(40), 34025–34032. doi:10.1074/jbc.M505143200 PMID:16079129

Petkov, C. I., Wu, C. C., Eberling, J. L., Mungas, D., Zrelak, P. A., Yonelinas, A. P., ... Jagust, W. J. (2004). Correlates of memory function in community-dwelling elderly: The importance of white matter hyperintensities. *Journal of the International Neuropsychological Society*, *10*(3), 371–381. doi:10.1017/S1355617704103056 PMID:15147595

Petrova, M., Mehrabian-Spasova, S., Aarsland, D., Raycheva, M., & Traykov, L. (2015). Clinical and Neuropsychological Differences between Mild Parkinson's Disease Dementia and Dementia with Lewy Bodies. *Dementia and Geriatric Cognitive Disorders. Extra*, 5(2), 212–220. doi:10.1159/000375363 PMID:26195977 Pettorruso, M., De Risio, L., Martinotti, G., Di Nicola, M., Ruggeri, F., & Conte, G. (2014). ... Janiri, L. (2014). Targeting the Glutamatergic System to Treat Pathological Gambling: Current Evidence and Future Perspectives. *BioMed Research International*. PMID:25013755

Pfeiffer, R. F. (2007). Multiple system atrophy. *Handbook of Clinical Neurology*, 84, 305–326. doi:10.1016/S0072-9752(07)84046-2 PMID:18808955

Philo, J. S., & Arakawa, T. (2009). Mechanisms of protein aggregation. *Current Pharmaceutical Biotechnology*, *10*(4), 348–351. doi:10.2174/138920109788488932 PMID:19519409

Piazza, O., Sirén, A. L., & Ehrenreich, H. (2004). Soccer, neurotrauma and amyotrophic lateral sclerosis: Is there a connection? *Current Medical Research and Opinion*, 20(4), 505–508. doi:10.1185/030079904125003296 PMID:15119987

Pickrell, A. M., & Youle, R. J. (2015). The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease. *Neuron*, 85(2), 257–273. doi:10.1016/j.neuron.2014.12.007 PMID:25611507

Picone, P., Nuzzo, D., Caruana, L., Scafidi, V., & Di Carlo, M. (2014). Mitochondrial dysfunction: Different routes to Alzheimer's disease therapy. *Oxidative Medicine and Cellular Longevity*. PMID:25221640

Pietro, M., Britne, S., & Juan, C. C. (2012). Motor control abnormalities in Parkinson's disease. *Cold Spring Harbor Perspectives in Medicine*, 2, a009282. PMID:22675667

Pigott, K., Rick, J., Xie, S. X., Hurtig, H., Chen-Plotkin, A., Duda, J. E., ... Weintraub, D. (2015). Longitudinal study of normal cognition in Parkinson disease. *Neurology*, 85(15), 1276–1282. doi:10.1212/WNL.0000000000002001 PMID:26362285

Piven, J., Palmer, P., Jacobi, D., Childress, D., & Arndt, S. (1997). Broader autism phenotype: Evidence from a family history study of multiple-incidence autism families. *The American Journal of Psychiatry*, *154*(2), 185–190. doi:10.1176/ ajp.154.2.185 PMID:9016266

Plotegher, N., Gratton, E., & Bubacco, L. (2014). Number and Brightness analysis of alpha-synuclein oligomerization and the associated mitochondrial morphology alterations in live cells. *Biochimica et Biophysica Acta (BBA)-General Subjects, 1840*(6).

Plum, L. M., Rink, L., & Haase, H. (2010). The essential toxin: Impact of zinc on human health. *International Journal of Environmental Research and Public Health*, 7(4), 1342–1365. doi:10.3390/ijerph7041342 PMID:20617034

Pocchiari, M., & Ladogana, A. (2015). Rethinking of doxycycline therapy in Creutzfeldt-Jakob disease. *JOURNAL OF Neurology. Neurosurgery and Psychiatry*, 0(0), 2014–2015.

Poewe, W. (2008). Non-motor symptoms in Parkinson's disease. *European Journal of Neurology*, 15(s1), 14–20. doi:10.1111/j.1468-1331.2008.02056.x PMID:18353132

Poewe, W., & Mahlknecht, P. (2009). The clinical progression of Parkinson's disease. *Parkinsonism & Related Disorders*, 15, S28–S32. doi:10.1016/S1353-8020(09)70831-4 PMID:20123553

Polidori, M. C., Meccoci, P., Browne, S. E., Senin, U., & Beal, M. F. (1999). Oxidative damage to mitochondrial DNA in Huntington's disease parietal cortex. *Neuroscience Letters*, 272(1), 53–56. doi:10.1016/S0304-3940(99)00578-9 PMID:10507541

Pollak, P., Benabid, A. L., Gross, C., Gao, D. M., Laurent, A., Benazzouz, A., & (1993). Effects of the stimulation of the subthalamic nucleus in Parkinson disease. *Revue Neurologique*, *149*(3), 175–176. PMID:8235208

Pollock, A., St George, B., Fenton, M., & Firkins, L. (2014). Top 10 research priorities relating to life after strokeconsensus from stroke survivors, caregivers, and health professionals. *International Journal of Stroke*, *9*(3), 313–320. doi:10.1111/j.1747-4949.2012.00942.x PMID:23227818

Polymenidou, M., & Cleveland, D. W. (2017). Biological spectrum of Amyotrophic lateral sclerosis. *Cold Spring Harbor Perspectives in Medicine*, 7(11), a024133. doi:10.1101/cshperspect.a024133 PMID:28062558

Polymenidou, M., Lagier-Tourenne, C., Hutt, K. R., Bennett, C. F., Cleveland, D. W., & Yeo, G. W. (2012). Misregulated RNA processing in amyotrophic lateral sclerosis. *Brain Research*, *1462*, 3–15. doi:10.1016/j.brainres.2012.02.059 PMID:22444279

Polymenidou, M., Lagier-Tourenne, C., Hutt, K. R., Huelga, S. C., Moran, J., Liang, T. Y., ... Cleveland, D. W. (2011). Long pre-mRNA depletion and RNA missplicing contribute to neuronal vulnerability from loss of TDP-43. *Nature Neuroscience*, *14*(4), 459–468. doi:10.1038/nn.2779 PMID:21358643

Polymeropoulos, M. H., Lavedan, C., Leroy, E., Ide, S. E., Dehejia, A., Dutra, A., ... Stenroos, E. S. (1997). Mutation in the α -synuclein gene identified in families with Parkinson's disease. *Science*, 276(5321), 2045–2047. doi:10.1126cience.276.5321.2045 PMID:9197268

Ponte, M. L. (2006). Insights into the Management of Emerging Infections : Regulating Variant Creutzfeldt-Jakob Disease Transfusion Risk in the UK and the US. Academic Press.

Pont-Sunyer, C., Hotter, A., Gaig, C., Seppi, K., Compta, Y., Katzenschlager, R., ... Tolosa, E. (2015). The Onset of Nonmotor Symptoms in Parkinson's disease (The ONSET PD Study). *Movement Disorders*, *30*(2), 229–237. doi:10.1002/mds.26077 PMID:25449044

Poole, A. C., Thomas, R. E., Andrews, L. A., McBride, H. M., Whitworth, A. J., & Pallanck, L. J. (2008). The PINK1/ Parkin pathway regulates mitochondrial morphology. *Proceedings of the National Academy of Sciences of the United States of America*, *105*(5), 1638–1643. doi:10.1073/pnas.0709336105 PMID:18230723

Popescu, A., Lippa, C. F., Lee, V. M. Y., & Trojanowski, J. Q. (2004). Lewy bodies in the amygdala: Increase of α -synuclein aggregates in neurodegenerative diseases with tau-based inclusions. *Archives of Neurology*, *61*(12), 1915–1919. doi:10.1001/archneur.61.12.1915 PMID:15596612

Popović, I. M., Serić, V., & Demarin, V. (2007). Mild cognitive impairment in symptomatic and asymptomatic cerebrovascular disease. *Journal of the Neurological Sciences*, 257(1-2), 185–193. doi:10.1016/j.jns.2007.01.029 PMID:17328916

Poser, S., Zerr, I., Schroeter, A., Otto, M., Giese, A., Steinhoff, B. J., & Kretzschmar, H. A. (2000). Clinical and differential diagnosis of Creutzfeldt-Jakob disease. *Archives of Virology. Supplementum*, 2(16), 153–159. PMID:11214918

Posner, M. I., & Rothbart, M. K. (2007). *Educating the human brain*. American Psychological Association. doi:10.1037/11519-000

Postina, R. (2008). A closer look at alpha-secretase. *Current Alzheimer Research*, 5(2), 179–186. doi:10.2174/156720508783954668 PMID:18393803

Postuma, R. B., Berg, D., Stern, M., Poewe, W., Olanow, C. W., Oertel, W., ... Deuschl, G. (2015). MDS clinical diagnostic criteria for Parkinson's disease. *Movement Disorders*, *30*(12), 1591–1601. doi:10.1002/mds.26424 PMID:26474316

Postuma, R. B., Gagnon, J. F., Pelletier, A., & Montplaisir, J. (2013). Prodromal autonomic symptoms and signs in Parkinson's disease and dementia with Lewy bodies. *Movement Disorders*, 28(5), 597–604. doi:10.1002/mds.25445 PMID:23554107

Prasad, A. S. (2009). Impact of the discovery of human zinc deficiency on health. *Journal of the American College of Nutrition*, 28(3), 257–265. doi:10.1080/07315724.2009.10719780 PMID:20150599

Praticò, D., Uryu, K., Leight, S., Trojanoswki, J. Q., & Lee, V. M.-Y. (2001). Increased lipid peroxidation precedes amyloid plaque formation in an animal model of Alzheimer amyloidosis. *The Journal of Neuroscience*, *21*(12), 4183–4187. PMID:11404403

Price, C. J., & Friston, K. J. (2002). Degeneracy and cognitive anatomy. *Trends in Cognitive Sciences*, *6*(10), 416–421. doi:10.1016/S1364-6613(02)01976-9 PMID:12413574

Pringsheim, T., Jette, N., Frolkis, A., & Steeves, T. D. L. (2014). The prevalence of Parkinson's disease: A systematic review and meta-analysis. *Movement Disorders*, 29(13), 1583–1590. doi:10.1002/mds.25945 PMID:24976103

Pruisner, S. (2001). Shattuck lecture-neurodegenerative diseases and prions. *The New England Journal of Medicine*, 344(20), 1516–1526. doi:10.1056/NEJM200105173442006 PMID:11357156

Prusiner, S. B. (1991). Molecular biology of prion diseases. *Science*, 252(5012), 1515–1522. doi:10.1126cience.1675487 PMID:1675487

Prusiner, S. B. (2013). Biology and genetics of prions causing neurodegeneration. *Annual Review of Genetics*, 47(1), 601–623. doi:10.1146/annurev-genet-110711-155524 PMID:24274755

Prust, M., Wang, J., Morizono, H., Messing, A., Brenner, M., Gordon, E., ... Vanderver, A. (2011). *GFAP* mutations, age at onset, and clinical subtypes in Alexander disease. *Neurology*, 77(13), 1287–1294. doi:10.1212/WNL.0b013e3182309f72 PMID:21917775

Przedborski, S., Jackson-Lewis, V., Naini, A. B., Jakowec, M., Petzinger, G., Miller, R., & Akram, M. (2001). The parkinsonian toxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP): A technical review of its utility and safety. *Journal of Neurochemistry*, *76*(5), 1265–1274. doi:10.1046/j.1471-4159.2001.00183.x PMID:11238711

Przedborski, S., Vila, M., & Jackson-Lewis, V. (2003). Neurodegeneration: What is it and where are we? *The Journal of Clinical Investigation*, *111*(1), 3–10. doi:10.1172/JCI200317522 PMID:12511579

Purves, D., Augustine, G. J., Fitzpatrick, D., Katz, L. C., LaMantia, A.-S., McNamara, J. O., & Williams, S. M. (2001). *Circuits within the basal ganglia system*. Academic Press.

Pyle, A., Foltynie, T., Tiangyou, W., Lambert, C., Keers, S. M., Allcock, L. M., ... Barker, R. (2005). Mitochondrial DNA haplogroup cluster UKJT reduces the risk of PD. *Annals of Neurology*, *57*(4), 564–567. doi:10.1002/ana.20417 PMID:15786469

Qin, B., Cartier, L., Dubois-dauphin, M., Li, B., Serrander, L., & Krause, K. H. (2006). A key role for the microglial NADPH oxidase in APP-dependent killing of neurons. *Neurobiology of Aging*, 27(11), 1577–1587. doi:10.1016/j.neurobiolaging.2005.09.036 PMID:16260066

Qin, W., Haroutunian, V., Katsel, P., Cardozo, C. P., Ho, L., Buxbaum, J. D., & Pasinetti, G. M. (2009). PGC-1α expression decreases in the Alzheimer disease brain as a function of dementia. *Archives of Neurology*, *66*(3), 352–361. doi:10.1001/archneurol.2008.588 PMID:19273754

Qiu, C., Kivipelto, M., & von Strauss, E. (2009). Epidemiology of Alzheimer's disease: Occurrence, determinants, and strategies toward intervention. *Dialogues in Clinical Neuroscience*, *11*(2), 111. PMID:19585947

Qui, J., Tan, Y. W., Hagenston, A. M., Martel, M. A., Kneisel, N., Skehel, P. A., ... Hardingham, G. E. (2013). Mitochondrial calcium uniporter Mcu controls excitotoxicity and is transcriptionally repressed by neuroprotective nuclear calcium signals. *Nature Communications*, *4*, 2034. PMID:23774321

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Quintanilla, R. A., Jin, Y. N., Bernhardi, R. V., Gail, V., & Johnson, W. (2013). Mitochondrial permeability transition pore induces mitochondria injury in Huntington disease. *Molecular Neurodegeneration*, 8(1), 45. doi:10.1186/1750-1326-8-45 PMID:24330821

Quirk, G. J., & Bee, J. S. (2006). Prefrontal involvement in the regulation of emotion: Convergence of rat and human studies. *Current Opinion in Neurobiology*, *16*(6), 723–727. doi:10.1016/j.conb.2006.07.004 PMID:17084617

Quirk, G. J., & Mueller, D. (2008). Neural mechanisms of extinction learning and retrieval. *Neuropsychopharmacology*, 33(1), 56–72. doi:10.1038j.npp.1301555 PMID:17882236

Qureshi, A. I., Mendelow, A. D., & Hanley, D. F. (2009). Intracerebral haemorrhage. *Lancet*, *373*(9675), 1632–1644. doi:10.1016/S0140-6736(09)60371-8 PMID:19427958

Rabin, S. J., Kim, J. M., Baughn, M., Libby, R. T., Kim, Y. J., Fan, Y., ... Ravits, J. (2010). Sporadic ALS has compartmentspecific aberrant exon splicing and altered cell-matrix adhesion biology. *Human Molecular Genetics*, *19*(2), 313–328. doi:10.1093/hmg/ddp498 PMID:19864493

Rademakers, R., Neumann, M., & Mackenzie, I. R. (2012). Advances in understanding the molecular basis of frontotemporal dementia. *Nature Reviews. Neurology*, 8(8), 423–434. doi:10.1038/nrneurol.2012.117 PMID:22732773

Rademakers, R., & van Blitterswijk, M. (2013). Motor neuron disease in 2012: Novel causal genes and disease modifiers. *Nature Reviews. Neurology*, *9*(2), 63–64. doi:10.1038/nrneurol.2012.276 PMID:23318296

Rahimi, J., & Kovacs, G. G. (2014). Prevalence of mixed pathologies in the aging brain. *Alzheimer's Research & Therapy*, 6(9), 82. doi:10.118613195-014-0082-1 PMID:25419243

Rahman, K. (2007). Studies on free radicals, antioxidants, and co-factors. *Clinical Interventions in Aging*, *2*, 219–236. PMID:18044138

Rajagopalan, S., & Andersen, J. K. (2001). Alpha synuclein aggregation: Is it the toxic gain of function responsible for neurodegeneration in Parkinson's disease? *Mechanisms of Ageing and Development*, *122*(14), 1499–1510. doi:10.1016/S0047-6374(01)00283-4 PMID:11511392

Rajamohamedsait, H. B., & Sigurdsson, E. M. (2012). Histological Staining of Amyloid and Pre-Amyloid Peptides and Proteins in Mouse Tissue. *Methods in Molecular Biology (Clifton, N.J.), 849*, 411–424. doi:10.1007/978-1-61779-551-0_28 PMID:22528106

Rajeswaran, J., & Bennett, C. N. (2013). The Neuropsychology of Stress. *Stress and Work: Perspectives on Understanding and Managing Stress*, 13.

Rajput, A. H., Rozdilsky, B., & Rajput, A. (1991). Accuracy of clinical diagnosis in Parkinsonism – a prospective study. *The Canadian Journal of Neurological Sciences*, *18*(3), 275–278. doi:10.1017/S0317167100031814 PMID:1913360

Rakhit, R., Cunningham, P., Furtos-Matei, A., Dahan, S., Qi, X. F., Crow, J. P., ... Chakrabartty, A. (2002). Oxidationinduced misfolding and aggregation of superoxide dismutase and its implications for amyotrophic lateral sclerosis. *The Journal of Biological Chemistry*, 277(49), 47551–47556. doi:10.1074/jbc.M207356200 PMID:12356748

Ramanathan, A., Nelson, A. R., Sagare, A. P., & Zlokovic, B. V. (2015). Impaired vascular-mediated clearance of brain amyloid beta in Alzheimer's disease: The role, regulation and restoration of LRP1. *Frontiers in Aging Neuroscience*, *7*, 136. doi:10.3389/fnagi.2015.00136 PMID:26236233

Ramirez-Bermudez, J. (2012). Alzheimer's disease: Critical notes on the history of a medical concept. *Archives of Medical Research*, 43(8), 595–599. doi:10.1016/j.arcmed.2012.11.008 PMID:23178566

Ramsay, R. R., Kowal, A. T., Johnson, M. K., Salach, J. I., & Singer, T. P. (1987). The inhibition site of MPP+, the neurotoxic bioactivation product of 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine is near the Q-binding site of NADH dehydrogenase. *Archives of Biochemistry and Biophysics*, 259(2), 645–649. doi:10.1016/0003-9861(87)90531-5 PMID:2827583

Rapoport, M., Dawson, H. N., Binder, L. I., Vitek, M. P., & Ferreira, A. (2002). Tau is essential to -amyloid-induced neurotoxicity. *Proceedings of the National Academy of Sciences of the United States of America*, 99(9), 6364–6369. doi:10.1073/pnas.092136199 PMID:11959919

Ravaglia, G., Forti, P., Maioli, F., Nesi, B., Pratelli, L., Savarino, L., ... Cavalli, G. (2000). Blood micronutrient and thyroid hormone concentrations in the oldest-old. *The Journal of Clinical Endocrinology and Metabolism*, 85(6), 2260–2265. doi:10.1210/jcem.85.6.6627 PMID:10852460

Raz, N., & Rodrigue, K. M. (2006). Differential aging of the brain: Patterns, cognitive correlates and modifiers. *Neuroscience and Biobehavioral Reviews*, *30*(6), 730–748. doi:10.1016/j.neubiorev.2006.07.001 PMID:16919333

Raz, N., Rodrigue, K. M., & Haacke, E. M. (2007). Brain Aging and Its Modifiers: Insights from in Vivo Neuromorphometry and Susceptibility Weighted Imaging. *Annals of the New York Academy of Sciences*, *1097*(1), 84–93. doi:10.1196/ annals.1379.018 PMID:17413014

Reaume, A. G., Elliott, J. L., Hoffman, E. K., Kowall, N. W., Ferrante, R. J., Siwek, D. F., ... Snider, W. D. (1996). Motor neurons in Cu/Zn superoxide dismutase-deficient mice develop normally but exhibit enhanced cell death after axonal injury. *Nature Genetics*, *13*(1), 43–47. doi:10.1038/ng0596-43 PMID:8673102

Rebrin, I., & Sohal, R. S. (2008). Pro-oxidant shift in glutathione redox state during aging. *Advanced Drug Delivery Reviews*, 60(13), 1545–1552. doi:10.1016/j.addr.2008.06.001 PMID:18652861

Recasens, A., & Dehay, B. (2014). Alpha-synuclein spreading in Parkinson's disease. *Frontiers in Neuroanatomy*, 8. PMID:25565982

Recchia, A., Debetto, P., Negro, A., Guidolin, D., Skaper, S. D., & Giusti, P. (2004). α-Synuclein and Parkinson's disease. *The FASEB Journal*, *18*(6), 617–626. doi:10.1096/fj.03-0338rev PMID:15054084

Reddy, P. H. (2007). Mitochondrial dysfunction in aging and Alzheimer's disease: Strategies to protect neurons. *Anti*oxidants & Redox Signalling, 9(10), 1647–1658. doi:10.1089/ars.2007.1754 PMID:17696767

Reddy, P. H. (2008). Mitochondrial medicine for aging and neurodegenerative diseases. *Neuromolecular Medicine*, *10*(4), 291–315. doi:10.100712017-008-8044-z PMID:18566920

Reddy, P. H., & Beal, M. F. (2005). Are mitochondria critical in the pathogenesis of Alzheimer's disease? *Brain Research. Brain Research Reviews*, *49*(3), 618–632. doi:10.1016/j.brainresrev.2005.03.004 PMID:16269322

Reddy, P. H., & Beal, M. F. (2008). Amyloid beta, mitochondrial dysfunction and synaptic damage: Implications for cognitive decline in aging and Alzheimer's disease. *Trends in Molecular Medicine*, *14*(2), 45–53. doi:10.1016/j.mol-med.2007.12.002 PMID:18218341

Reddy, P. H., McWeeney, S., Park, B. S., Manczak, M., Gutala, R. V., Partovi, D., ... Mori, M. (2004). Gene expression profiles of transcripts in amyloid precursor protein transgenic mice: Up-regulation of mitochondrial metabolism and apoptotic genes is an early cellular change in Alzheimer's disease. *Human Molecular Genetics*, *13*(12), 1225–1240. doi:10.1093/hmg/ddh140 PMID:15115763

Reddy, P. H., & Reddy, T. P. (2011). Mitochondria as a therapeutic target for aging and neurodegenerative diseases. *Current Alzheimer Research*, *8*, 393–409. doi:10.2174/156720511795745401 PMID:21470101

Redeker, V., Lachkar, S., Siavoshian, S., Charbaut, E., Rossier, J., Sobel, A., & Curmi, P. A. (2000). Probing the Native Structure of Stathmin and Its Interaction Domains with Tubulin Combined Use of Limited Proteolysis, Size Exclusion Chromatography, and Mass Spectrometry. *The Journal of Biological Chemistry*, 275(10), 6841–6849. doi:10.1074/ jbc.275.10.6841 PMID:10702243

Reid, E., Kloos, M., Ashley-Koch, A., Hughes, L., Bevan, S., Svenson, I. K., ... Pericak-Vance, M. A. (2002). A kinesin heavy chain (KIF5A) mutation in hereditary spastic paraplegia (SPG10). *American Journal of Human Genetics*, *71*(5), 1189–1194. doi:10.1086/344210 PMID:12355402

Reijmer, Y. D., Fotiadis, P., Martinez-Ramirez, S., Salat, D. H., Schultz, A., Shoamanesh, A., ... Greenberg, S. M. (2015). Structural network alterations and neurological dysfunction in cerebral amyloid angiopathy. *Brain*, *138*(1), 179–188. doi:10.1093/brain/awu316 PMID:25367025

Reijnders, J. S., Scholtissen, B., Weber, W. E., Aalten, P., Verhey, F. R., & Leentjens, A. F. (2010). Neuroanatomical correlates of apathy in Parkinson's disease: A magnetic resonance imaging study using voxel-based morphometry. *Movement Disorders*, 25(14), 2318–2325. doi:10.1002/mds.23268 PMID:20669264

Reiman, E. M., Caselli, R. J., Yun, L. S., Chen, K., Bandy, D., Minoshima, S., ... Osborne, D. (1996). Preclinical evidence of Alzheimer's disease in persons homozygous for the ɛ4 allele for apolipoprotein E. *The New England Journal of Medicine*, 334(12), 752–758. doi:10.1056/NEJM199603213341202 PMID:8592548

Remelli, M., Peana, M., Medici, S., Delogu, L. G., & Zoroddu, M. A. (2013). Interaction of divalent cations with peptide fragments from Parkinson's disease genes. *Dalton Transactions (Cambridge, England)*, 42(17), 5964–5974. doi:10.1039/C2DT32222F PMID:23202360

Remy, P., Doder, M., Lees, A., Turjanski, N., & Brooks, D. (2005). Depression in Parkinson's disease: Loss of dopamine and noradrenaline innervation in the limbic system. *Brain*, *128*(6), 1314–1322. doi:10.1093/brain/awh445 PMID:15716302

Rendón, W. O., Martínez-Alonso, E., Tomás, M., Martínez-Martínez, N., & Martínez-Menárguez, J. A. (2013). Golgi fragmentation is Rab and SNARE dependent in cellular models of Parkinson's disease. *Histochemistry and Cell Biology*, *139*(5), 671–684. doi:10.100700418-012-1059-4 PMID:23212845

Renton, A. E., Chio, A., & Traynor, B. J. (2014). State of play in amyotrophic lateral sclerosis genetics. *Nature Neuroscience*, *17*(1), 17–23. doi:10.1038/nn.3584 PMID:24369373

Renton, A. E., Majounie, E., Waite, A., Simón-Sánchez, J., Rollinson, S., Gibbs, J. R., ... Traynor, B. J. (2011). A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron*, 72(2), 257–268. doi:10.1016/j.neuron.2011.09.010 PMID:21944779

Repetto, M. G., Domínguez, R. O., Marschoff, E. R., & Serra, J. A. (2012). Free radicals, oxidative stress and oxidative damage in Parkinson's disease. In J. Dushanova (Ed.), *Mechanisms in Parkinson's Disease – Models and Treatments*. InTech Open.

Requena, J. R., Groth, D., Legname, G., Stadtman, E. R., Prusiner, S. B., & Levine, R. L. (2001). Copper-catalyzed oxidation of the recombinant SHa (29–231) prion protein. *Proceedings of the National Academy of Sciences of the United States of America*, 98(13), 7170–7175. doi:10.1073/pnas.121190898 PMID:11404462

Revuelta, G. J., & Lippa, C. F. (2009). Dementia with Lewy bodies and Parkinson's disease dementia may best be viewed as two distinct entities. *International Psychogeriatrics*, 21(2), 213. doi:10.1017/S1041610208008600 PMID:19173761

Rezai, A. R., Machado, A. G., Deogaonkar, M., Azmi, H., Kubu, C., & Boulis, N. M. (2008, February). Surgery for movement disorders. *Neurosurgery*, *62*(Suppl 2), 809–838. doi:10.1227/01.neu.0000316285.52865.53 PMID:18596424

Rezaie, P., Cairns, N. J., Chadwick, A., & Lantos, P. L. (1996). Lewy bodies are located preferentially in limbic areas in diffuse Lewy body disease. *Neuroscience Letters*, 212(2), 111–114. doi:10.1016/0304-3940(96)12775-0 PMID:8832651

Rezania, K., Yan, J., Dellefave, L., Deng, H. X., Siddique, N., & Pascuzzi, T. (2003). A rare Cu/Zn superoxide dismutase mutation causing familial amyotrophic lateral sclerosis with variable age of onset, incomplete penetrance and a sensory neuropathy. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, 4(3), 162–166. doi:10.1080/ aml.4.3.162.166 PMID:13129803

Rhodes-Kropf, J., Cheng, H., Castillo, E. H., & Fulton, A. T. (2011). Managing the patient with dementia in long-term care. *Clinics in Geriatric Medicine*, 27(2), 135–152. doi:10.1016/j.cger.2011.01.001 PMID:21641502

Riachi, N. J., LaManna, J. C., & Harik, S. I. (1989). Entry of 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine into the rat brain. *The Journal of Pharmacology and Experimental Therapeutics*, 249(3), 744–748. PMID:2786562

Richard, I. H., Papka, M., Rubio, A., & Kurlan, R. (2002). Parkinson's disease and dementia with Lewy bodies: One disease or two? *Movement Disorders*, *17*(6), 1161–1165. doi:10.1002/mds.10274 PMID:12465052

Richer, P., & Meige, H. (1895). Etude morphologique sur la maladie de Parkinson. *Nouvelle iconographie de la Salpêtriere, 8,* 361-371.

Richter, C., Park, J. W., & Ames, B. N. (1988). Normal oxidative damage to mitochondrial and nuclear DNA is extensive. *Proceedings of the National Academy of Sciences of the United States of America*, 85(17), 6465–6467. doi:10.1073/pnas.85.17.6465 PMID:3413108

Riederer, P., & Laux, G. (2011). MAO-inhibitors in Parkinson's Disease. *Experimental Neurobiology*, 20(1), 1–17. doi:10.5607/en.2011.20.1.1 PMID:22110357

Riesner, D. (2003). Biochemistry and structure of PrPC and PrPSc. *British Medical Bulletin*, 66(1), 21–33. doi:10.1093/bmb/66.1.21 PMID:14522846

Riggs, J. E. (1996). Amyotrophic lateral sclerosis, heterogeneous susceptibility, trauma, and epidemiology. *Archives of Neurology*, *53*(3), 225–227. doi:10.1001/archneur.1996.00550030031019 PMID:8651874

Riikonen, R. (2017). Insulin-Like Growth Factors in the Pathogenesis of Neurological Diseases in Children. *International Journal of Molecular Sciences*, *18*(10), 2056. doi:10.3390/ijms18102056 PMID:28954393

Rikans, L. E., & Hornbrook, K. R. (1997). Lipid peroxidation, antioxidant protection and aging. *Biochimica et Biophysica Acta*, *1362*(2-3), 116–127. doi:10.1016/S0925-4439(97)00067-7 PMID:9540842

Rinaldi, P., Polidori, M. C., Metastasio, A., Mariani, E., Mattioli, P., Cherubini, A., ... Mecocci, P. (2003). Plasma antioxidants are similarly depleted in mild cognitive impairment and in Alzheimer's disease. *Neurobiology of Aging*, 24(7), 915–919. doi:10.1016/S0197-4580(03)00031-9 PMID:12928050

Rineer, S., Finucane, B., & Simon, E. W. (1998). Autistic symptoms among children and young adults with isodicentric chromosome 15. *American Journal of Medical Genetics*, 74, 121–128. PMID:9754629

Rinne, U., Sonninen, V., & Siirtola, T. (1970). L-dopa treatment in Parkinson's disease. *European Neurology*, 4(6), 348–369. doi:10.1159/000113990 PMID:4932969

Risacher, S. L., & Saykin, A. J. (2013). Neuroimaging Biomarkers of Neurodegenerative Diseases and Dementia. *Seminars in Neurology*, *33*(4), 386–416. doi:10.1055-0033-1359312 PMID:24234359

Rizzardini, M., Chiesa, R., Angeretti, N., Lucca, E., Salmona, M., Forloni, G., & Cantoni, L. (1997). Prion Protein Fragment 106–126 Differentially Induces Heme Oxygenase-1 mRNA in Cultured Neurons and Astroglial Cells. *Journal of Neurochemistry*, 68(2), 715–720. doi:10.1046/j.1471-4159.1997.68020715.x PMID:9003061

Rizzuto, R., De Stefani, D., Raffaello, A., & Mammucari, C. (2012). Mitochondria as sensors and regulators of calcium signalling. *Nature Reviews. Molecular Cell Biology*, *13*(9), 566–578. doi:10.1038/nrm3412 PMID:22850819

Robins, D. L., Fein, D., Barton, M. L., & Green, J. A. (2001). The Modified Checklist for Autism in Toddlers: An initial study investigating the early detection of autism and pervasive developmental disorders. *Journal of Autism and Developmental Disorders*, *31*(2), 131–144. doi:10.1023/A:1010738829569 PMID:11450812

Robinson, E. B., Lichtenstein, P., Anckarsäter, H., Happé, F., & Ronald, A. (2013). Examining and interpreting the female protective effect against autistic behavior. *Proceedings of the National Academy of Sciences of the United States of America*, *110*(13), 5258–5262. doi:10.1073/pnas.1211070110 PMID:23431162

Rochester, L., Yarnall, A. J., Baker, M. R., David, R. V., Lord, S., Galna, B., & Burn, D. J. (2012). Cholinergic dysfunction contributes to gait disturbance in early Parkinson's disease. *Brain*, *135*(9), 2779–2788. doi:10.1093/brain/aws207 PMID:22961550

Rochon, P. A., Stukel, T. A., Sykora, K., Gill, S., Garfinkel, S., Anderson, G. M., ... Bronskill, S. E. (2005). Atypical antipsychotics and parkinsonism. *Archives of Internal Medicine*, *165*(16), 1882–1888. doi:10.1001/archinte.165.16.1882 PMID:16157833

Rock, R. B., Gekker, G., Hu, S., Sheng, W. S., Cheeran, M., Lokensgard, J. R., & Peterson, P. K. (2004). Role of microglia in central nervous system infections. *Clinical Microbiology Reviews*, *17*(4), 942–964. doi:10.1128/CMR.17.4.942-964.2004 PMID:15489356

Rodriguez, D. (2013). Leukodystrophies with astrocytic dysfunction. Handbook of Clinical Neurology Pediatric Neurology Part III, 1619-1628.

Rodriguez, D., Gauthier, F., Bertini, E., Bugiani, M., Brenner, M., N'guyen, S., ... Boespflug-Tanguy, O. (2001). Infantile Alexander Disease: Spectrum of GFAP Mutations and Genotype-Phenotype Correlation. *American Journal of Human Genetics*, *69*(5), 1134–1140. doi:10.1086/323799 PMID:11567214

Rogers, J. D., Brogan, D., & Mirra, S. S. (1985). The nucleus basalis of Meynert in neurological disease: A quantitative morphological study. *Annals of Neurology*, *17*(2), 163–170. doi:10.1002/ana.410170210 PMID:3883886

Roger, V. L., Go, A. S., Lloyd-Jones, D. M., Adams, R. J., Berry, J. D., Brown, T. M., ... Wylie-Rosett, J. (2011). Heart disease and stroke statistics—2011 update: A report from the american heart association. *Circulation*, 23(4), e18–e209. doi:10.1161/CIR.0b013e3182009701 PMID:21160056

Rojas, P., Rojas, C., Ebadi, M., Montes, S., Monroy-Noyola, A., & Serrano-García, N. (2004). EGb761 pretreatment reduces monoamine oxidase activity in mouse corpus striatum during 1-methyl-4-phenylpyridinium neurotoxicity. *Neurochemical Research*, *29*(7), 1417–1423. doi:10.1023/B:NERE.0000026406.64547.93 PMID:15202774

Rolinski, M., Fox, C., Maidment, I., & McShane, R. (2012). Cholinesterase inhibitors for dementia with Lewy bodies, Parkinson's disease dementia and cognitive impairment in Parkinson's disease. *The Cochrane Library*. PMID:22419314

Roman, G. C. (1987). Senile dementia of the Binswanger type. A vascular form of dementia in the elderly. *Journal of the American Medical Association*, 258(13), 1782–1788. doi:10.1001/jama.1987.03400130096040 PMID:3625988

Romano, A., Serviddio, G., Calcagnini, S., Villani, R., Giudetti, A. M., Cassano, T., & Gaetani, S. (2017). Linking lipid peroxidation and neuropsychiatric disorders: Focus on 4-hydroxy-2-nonenal. *Free Radical Biology & Medicine*, *111*, 281–293. doi:10.1016/j.freeradbiomed.2016.12.046 PMID:28063940

Romero, J. R., & Wolf, P. A. (2013). Epidemiology of Stroke: Legacy of the Framingham Heart Study. *Global Heart*, 8(1), 67–75. doi:10.1016/j.gheart.2012.12.007 PMID:23527318

Roos, R. A. (2010). Huntington's disease: A clinical review. Orphanet Journal of Rare Diseases, 5(1), 40. doi:10.1186/1750-1172-5-40 PMID:21171977

Roozendaal, B. (2002). Stress and memory: Opposing effects of glucocorticoids on memory consolidation and memory retrieval. *Neurobiology of Learning and Memory*, 78(3), 578–595. doi:10.1006/nlme.2002.4080 PMID:12559837

Rosenberg, G. A. (2014). Blood-Brain Barrier Permeability in Aging and Alzheimer's Disease. *The Journal of Prevention of Alzheimer's Disease*, *1*(3), 138–139. doi:10.14283/jpad.2014.25 PMID:26301207

Rosenberg, J. B., Kaminsky, S. M., Aubourg, P., Crystal, R. G., & Sondhi, D. (2016, November 1). Gene therapy for metachromatic leukodystrophy. *Journal of Neuroscience Research*, 94(11), 1169–1179. doi:10.1002/jnr.23792 PMID:27638601

Rosenbloom, M. H., Tartaglia, M. C., Forner, S. A., Wong, K. K., Kuo, A., Johnson, D. Y., & Geschwind, M. D. (2015). Metabolic disorders with clinical and radiologic features of sporadic Creutzfeldt-Jakob disease. *Neurology. Clinical Practice*, *5*(2), 108–115. doi:10.1212/CPJ.00000000000114 PMID:26137419

Rosen, D. R., Siddiquef, T., Patterson, D., Figlewicz, D. A., Sapp, P., Hentatif, A., ... Cayabyabi, A. (1993). Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic. *Nature*, *362*(6415), 4. doi:10.1038/362059a0 PMID:8446170

Rosen, K. M., Moussa, C. E. H., Lee, H. K., Kumar, P., Kitada, T., Qin, G., ... Querfurth, H. W. (2010). Parkin reverses intracellular β-amyloid accumulation and its negative effects on proteasome function. *Journal of Neuroscience Research*, 88(1), 167–178. doi:10.1002/jnr.22178 PMID:19610108

Rosenthal, E., Brennan, L., Xie, S., Hurtig, H., Milber, J., Weintraub, D., ... Siderowf, A. (2010). Association between cognition and function in patients with Parkinson disease with and without dementia. *Movement Disorders*, 25(9), 1170–1176. doi:10.1002/mds.23073 PMID:20310053

Rosin, B., Slovik, M., Mitelman, R., Rivlin-Etzion, M., Haber, S. N., Israel, Z., ... Bergman, H. (2011, October). Closed-loop deep brain stimulation is superior in ameliorating parkinsonism. *Neuron*, 72(2), 370–384. doi:10.1016/j. neuron.2011.08.023 PMID:22017994

Ross, G. W., & Petrovitch, H. (2001). Current evidence for neuroprotective effects of nicotine and caffeine against Parkinson's disease. *Drugs & Aging*, *18*(11), 797–806. doi:10.2165/00002512-200118110-00001 PMID:11772120

Rossignol, D. A. (2009). Novel and emerging treatments for autism spectrum disorders: A systematic review. *Annals of Clinical Psychiatry*, 21(4), 213–236. PMID:19917212

Rossi, S. C., Gorman, N., & Wetterhahn, K. E. (1988). Mitochondrial reduction of the carcinogen chromate: Formation of chromium (V). *Chemical Research in Toxicology*, *1*(2), 101–107. doi:10.1021/tx00002a003 PMID:2979716

Ross, M. A., Miller, R. G., Berchert, L., Parry, G., Barohn, R. J., Armon, C., ... McGuire, D. (1998). Toward earlier diagnosis of amyotrophic lateral sclerosis: Revised criteria. rhCNTF ALS Study Group. *Neurology*, *50*(3), 768–772. doi:10.1212/WNL.50.3.768 PMID:9521272

Rothfuss, O., Fischer, H., Hasegawa, T., Maisel, M., Leitner, P., Miesel, F., ... Patenge, N. (2009). Parkin protects mitochondrial genome integrity and supports mitochondrial DNA repair. *Human Molecular Genetics*, *18*(20), 3832–3850. doi:10.1093/hmg/ddp327 PMID:19617636

Rothstein, J. D. (2009). Current hypotheses for the underlying biology of amyotrophic lateral sclerosis. *Annals of Neurology*, 65(1). PMID:19191304

Roubertie, A., Semprino, M., Chaze, A. M., Rivier, F., Humbertclaude, V., Cheminal, R., ... Echenne, B. (2001). Neurological presentation of three patients with 22q11 deletion (CATCH 22q11.2 deletion syndrome). *Brain & Development*, 23(8), 810–814. doi:10.1016/S0387-7604(01)00258-3 PMID:11720799

Rowland, A. A., & Voeltz, G. K. (2012). Endoplasmic reticulum–mitochondria contacts: Function of the junction. *Nature Reviews. Molecular Cell Biology*, *13*(10), 607–625. doi:10.1038/nrm3440 PMID:22992592

Rowland, L. P., & Shneider, N. A. (2001). Amyotrophic lateral sclerosis. *The New England Journal of Medicine*, 344(22), 1688–1700. doi:10.1056/NEJM200105313442207 PMID:11386269

Rubenstein, R., Petersen, R. B., & Wisniewski, T. (2016). Diagnosis of Prion Diseases. In *Manual of Molecular and Clinical Laboratory Immunology* (8th ed.; pp. 682–702). American Society of Microbiology.

Rubin, J. E., McIntyre, C. C., Turner, R. S., & Wichmann, T. (2012). Basal ganglia activity patterns in parkinsonism and computational modeling of their downstream effects. *The European Journal of Neuroscience*, *36*(2), 2213–2228. doi:10.1111/j.1460-9568.2012.08108.x PMID:22805066

Rubinsztein, D. C., Mariño, G., & Kroemer, G. (2011). Autophagy and aging. *Cell*, *146*(5), 682–695. doi:10.1016/j. cell.2011.07.030 PMID:21884931

Rudd, P. M., Wormald, M. R., Wing, D. R., Prusiner, S. B., & Dwek, R. A. (2001). Prion glycoprotein: Structure, dynamics, and roles for the sugars. *Biochemistry*, 40(13), 3759–3766. doi:10.1021/bi002625f PMID:11300755

Rutter, M., Le Couteur, A., & Lord, C. (2003). *Autism Diagnostic Interview-Revised*. Los Angeles, CA: Western Psychological Services.

Ryan, C. L., Baranowski, D. C., Chitramuthu, B. P., Malik, S., Li, Z., Cao, M., ... Bateman, A. (2009). Progranulin is expressed within motor neurons and promotes neuronal cell survival. *BMC Neuroscience*, *10*(1), 130. doi:10.1186/1471-2202-10-130 PMID:19860916

Sadigh-Eteghad, S., Sabermarouf, B., Majdi, A., Talebi, M., Farhoudi, M., & Mahmoudi, J. (2015). Amyloid-beta: a crucial factor in Alzheimer's disease. *Medical Principles and Practice : International Journal of the Kuwait University*. *Health Science Centre*, 24(1), 1–10. doi:10.1159/000369101

Sæmundsen, E., Magnusson, P., Smari, J., & Sigurdardottir, S. (2003). Autism Diagnostic Interview-Revised and the Childhood Autism Rating Scale: Convergence and discrepancy in diagnosing autism. *Journal of Autism and Developmental Disorders*, *33*(3), 319–327. doi:10.1023/A:1024410702242 PMID:12908834

Safar, J. G., Geschwind, M. D., Deering, C., Didorenko, S., Sattavat, M., Sanchez, H., & Miller, B. L. (2005). Diagnosis of human prion disease. *Proceedings of the National Academy of Sciences of the United States of America*, *102*(9), 3501–3506. doi:10.1073/pnas.0409651102 PMID:15741275

Sah, P., Faber, E. S., Lopez De Armentia, M., & Power, J. (2003). The amygdaloid complex: Anatomy and physiology. *Physiological Reviews*, *83*(3), 803–834. doi:10.1152/physrev.00002.2003 PMID:12843409

Saibil, H. (2013). Chaperone machines for protein folding, unfolding and disaggregation. *Nature Reviews. Molecular Cell Biology*, *14*(10), 630–642. doi:10.1038/nrm3658 PMID:24026055

Sajdel-Sulkowska, E. M., Xu, M., & Koibuchi, N. (2009). Increase in cerebellar neurotrophin-3 and oxidative stress markers in autism. *Cerebellum (London, England)*, 8(3), 366–372. doi:10.100712311-009-0105-9 PMID:19357934

Sajdyk, T. J., & Shekhar, A. (2000). Sodium lactate elicits anxiety in rats after repeated GABA receptor blockade in the basolateral amygdala. *European Journal of Pharmacology*, *394*(2-3), 265–273. doi:10.1016/S0014-2999(00)00128-X PMID:10771292

Sakai, N. (2009, August). Pathogenesis of leukodystrophy for Krabbe disease: Molecular mechanism and clinical treatment. *Brain & Development*, *31*(7), 485–487. doi:10.1016/j.braindev.2009.03.001 PMID:19332366

Sakamoto, F., Shiraishi, S., Yoshida, M., Tomiguchi, S., Hirai, T., Namimoto, T., ... Yamashita, Y. (2014). Diagnosis of dementia with Lewy bodies: Diagnostic performance of combined ¹²³I-IMP brain perfusion SPECT and ¹²³I-MIBG myocardial scintigraphy. *Annals of Nuclear Medicine*, 28(3), 203–211. doi:10.100712149-013-0796-3 PMID:24363079

Sakurai, A., Okamoto, K., Fujita, Y., Nakazato, Y., Wakabayashi, K., Takahashi, H., & Gonatas, N. K. (2000). Fragmentation of the Golgi apparatus of the ballooned neurons in patients with corticobasal degeneration and Creutzfeldt-Jakob disease. *Acta Neuropathologica*, *100*(3), 270–274. doi:10.1007004010000182 PMID:10965796

Sakurai, A., Okamoto, K., Yaguchi, M., Fujita, Y., Mizuno, Y., Nakazato, Y., & Gonatas, N. K. (2002). Pathology of the inferior olivary nucleus in patients with multiple system atrophy. *Acta Neuropathologica*, *103*(6), 550–554. doi:10.100700401-001-0500-x PMID:12012086

Salat, D., Noyce, A. J., Schrag, A., & Tolosa, E. (2016). Challenges of modifying disease progression in prediagnostic Parkinson's disease. *Lancet Neurology*, 1–12. PMID:26993435

Salat, D., & Tolosa, E. (2013). Levodopa in the treatment of Parkinson's disease: Current status and new developments. *Journal of Parkinson's Disease*, *3*(3), 255–269. PMID:23948989

Samii, A., Nutt, J. G., & Ransom, B. R. (2004). Parkinson's disease. *Lancet (London, England)*, *363*(9423), 1783–1793. doi:10.1016/S0140-6736(04)16305-8 PMID:15172778

Samuel, W., Caligiuri, M., Galasko, D., Lacro, J., Marini, M., McClure, F. S., ... Jeste, D. V. (2000). Better cognitive and psychopathologic response to donepezil in patients prospectively diagnosed as dementia with Lewy bodies: A preliminary study. *International Journal of Geriatric Psychiatry*, *15*(9), 794–802. doi: PMID:10984725

Sanders, L. H., McCoy, J., Hu, X., Mastroberardino, P. G., Dickinson, B. C., Chang, C. J., ... Greenamyre, J. T. (2014). Mitochondrial DNA damage: Molecular marker of vulnerable nigral neurons in Parkinson's disease. *Neurobiology of Disease*, *70*, 214–223. doi:10.1016/j.nbd.2014.06.014 PMID:24981012

Sandi, C. (1998). The role and mechanisms of action of glucocorticoid involvement in memory storage. *Neural Plasticity*, *6*(3), 41–52. doi:10.1155/NP.1998.41 PMID:9920681

Sandi, C. (2004). Stress, cognitive impairment and cell adhesion molecules. *Nature Reviews. Neuroscience*, 5(12), 917. doi:10.1038/nrn1555 PMID:15550947

Sandvik, U., Koskinen, L. O., Lundquist, A., & Blomstedt, P. (2012, April). Thalamic and subthalamic deep brain stimulation for essential tremor: Where is the optimal target? *Neurosurgery*, *70*(4), 840–845. doi:10.1227/NEU.0b013e318236a809 PMID:22426044

Santamato, A., Ranieri, M., Cinone, N., Stuppiello, L. A., Valeno, G., & De Sanctis, J. L. (2015). ... Panza, F. (2015). Postural and Balance Disorders in Patients with Parkinson's Disease: A Prospective Open-Label Feasibility Study with Two Months of Action Observation Treatment. *Parkinson's Disease*. PMID:26798551

Sanz-Blasco, S., Valero, R. A., Rodríguez-Crespo, I., Villalobos, C., & Núñez, L. (2008). Mitochondrial Ca2+ overload underlies Aβ oligomers neurotoxicity providing an unexpected mechanism of neuroprotection by NSAIDs. *PLoS One*, *3*(7), e2718. doi:10.1371/journal.pone.0002718 PMID:18648507

Sá-Pereira, I., Brites, D., & Brito, M. A. (2012). Neurovascular unit: A focus on pericytes. *Molecular Neurobiology*, 45(2), 327–347. doi:10.100712035-012-8244-2 PMID:22371274

Sarnataro, D., Pepe, A., & Zurzolo, C. (in press). Cell Biology of Prion Protein. *Progress in Molecular Biology and Translational Science*. PMID:28838675

Sasaki, S., & Iwata, M. (1996). Impairment of fast axonal transport in the proximal axons of anterior horn neurons in amyotrophic lateral sclerosis. *Neurology*, 47(2), 535–540. doi:10.1212/WNL.47.2.535 PMID:8757033

Sas, K., Robotka, H., Toldi, J., & Vécsei, L. (2007). Mitochondria, metabolic disturbances, oxidative stress and the kynurenine system, with focus on neurodegenerative disorders. *Journal of the Neurological Sciences*, 257(1), 221–239. doi:10.1016/j.jns.2007.01.033 PMID:17462670

Sathiyamoorthy, S., Tan, X., & Tan, E. K. (2014). Lewy Body in Parkinson's Disease: Causes or Scars of Neurodegeneration? *Austin Journal of Clinical Neurology*, *1*(3), 1011.

Savica, R., Rocca, W. A., & Ahlskog, J. E. (2010). When does Parkinson disease start? *Archives of Neurology*, 67(7), 798–801. doi:10.1001/archneurol.2010.135 PMID:20625084

Sayre, L. M., Smith, M. A., & Perry, G. (2001). Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Current Medicinal Chemistry*, 8(7), 721–738. doi:10.2174/0929867013372922 PMID:11375746

Scahill, R. I., Schott, J. M., Stevens, J. M., Rossor, M. N., & Fox, N. C. (2002). Mapping the evolution of regional atrophy in Alzheimer's disease: Unbiased analysis of fluid-registered serial MRI. *Proceedings of the National Academy of Sciences of the United States of America*, 99(7), 4703–4707. doi:10.1073/pnas.052587399 PMID:11930016

Schapansky, J., Nardozzi, J. D., Felizia, F., & LaVoie, M. J. (2014). Membrane recruitment of endogenous LRRK2 precedes its potent regulation of autophagy. *Human Molecular Genetics*, 23(16), 4201–4214. doi:10.1093/hmg/ddu138 PMID:24682598

Schapira, A. H. V. (2008). Mitochondrial dysfunction in neurodegenerative diseases. *Neurochemical Research*, 33(12), 2502–2509. doi:10.100711064-008-9855-x PMID:18998208

Schapira, A. H. V. (2012). Targeting mitochondria for neuroprotection in Parkinson's disease. *Antioxidants & Redox Signalling*, *16*(9), 965–973. doi:10.1089/ars.2011.4419 PMID:22229791

Schapira, A. H. V., Cooper, J. M., Dexter, D., Clark, J. B., Jenner, P., & Marsden, C. D. (1990). Mitochondrial complex I deficiency in Parkinson's disease. *Journal of Neurochemistry*, *54*(3), 823–827. doi:10.1111/j.1471-4159.1990.tb02325.x PMID:2154550

Schapira, A. H. V., Olanow, C. W., Greenamyre, J. T., & Bezard, E. (2014). Slowing of neurodegeneration in Parkinson's disease and Huntington's disease: Future therapeutic perspectives. *Lancet*, *384*(9942), 545–555. doi:10.1016/S0140-6736(14)61010-2 PMID:24954676

Schechtman, M. A. (2007). Scientifically unsupported therapies in the treatment of young children with autism spectrum disorders. *Pediatric Annals*, *36*(8), 497–505. doi:10.3928/0090-4481-20070801-12 PMID:17849608

Scheibel, T., Bloom, J., & Lindquist, S. L. (2004). The elongation of yeast prion fibers involves separable steps of association and conversion. *Proceedings of the National Academy of Sciences of the United States of America*, *101*(8), 2287–2292. doi:10.1073/pnas.0308754101 PMID:14983002

Schenk, D. B., Koller, M., Ness, D. K., Griffith, S. G., Grundman, M., Zago, W., ... Kinney, G. G. (2017). First-in-human assessment of PRX002, an anti–α-synuclein monoclonal antibody, in healthy volunteers. *Movement Disorders*, *32*(2), 211–218. doi:10.1002/mds.26878 PMID:27886407

Scheperjans, F., Aho, V., Pereira, P. A., Koskinen, K., Paulin, L., Pekkonen, E., ... Auvinen, P. (2015, March). Gut microbiota are related to Parkinson's disease and clinical phenotype. *Movement Disorders*, *30*(3), 350–358. doi:10.1002/mds.26069 PMID:25476529

Scheuner, D., Eckman, C., Jensen, M., Song, X., Citron, M., Suzuki, N., ... Larson, E. (1996). Secreted amyloid β -protein similar to that in the senile plaques of Alzheimer's disease is increased *in vivo* by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nature Medicine*, 2(8), 864–870. doi:10.1038/nm0896-864 PMID:8705854

Schildknecht, S., Gerding, H. R., Karreman, C., Drescher, M., Lashuel, H. A., Outeiro, T. F., ... Leist, M. (2013). Oxidative and nitrative alpha-synuclein modifications and proteostatic stress: Implications for disease mechanisms and interventions in synucleinopathies. *Journal of Neurochemistry*, *125*(4), 491–511. doi:10.1111/jnc.12226 PMID:23452040

Schmahmann, J. D., Smith, E. E., Eichler, F. S., & Filley, C. M. (2008, October). Cerebral white matter: Neuroanatomy, clinical neurology, and neurobehavioral correlates. *Annals of the New York Academy of Sciences*, *1142*(1), 266–309. doi:10.1196/annals.1444.017 PMID:18990132

Schmidt, R., de Reus, M. A., Scholtens, S. H., van den Berg, L. H., & van den Heuvel, M. P. (2016). Simulating disease propagation across white matter connectome reveals anatomical substrate for neuropathology staging in amyotrophic lateral sclerosis. *NeuroImage*, *124*, 762–769. doi:10.1016/j.neuroimage.2015.04.005 PMID:25869856

Schmidt, R., Verstraete, E., de Reus, M. A., Veldink, J. H., van den Berg, L. H., & van den Heuvel, M. P. (2014). Correlation between structural and functional connectivity impairment in amyotrophic lateral sclerosis. *Human Brain Mapping*, *35*(9), 4386–4395. doi:10.1002/hbm.22481 PMID:24604691

Schmitt, F. A., Davis, D. G., Wekstein, D. R., Smith, C. D., Ashford, J. W., & Markesbery, W. R. (2000). Preclinal AD revisited: Neuropathology of cognitively normal older adults. *Neurology*, *55*(3), 370–376. doi:10.1212/WNL.55.3.370 PMID:10932270

Schneeberger, A., Tierney, L., & Mandler, M. (2016). Active immunization therapies for Parkinson's disease and multiple system atrophy. *Movement Disorders*, *31*(2), 214–224. doi:10.1002/mds.26377 PMID:26260853

Schneider, S. A., & Obeso, J. A. (2014). Clinical and pathological features of Parkinson's disease. In Behavioral Neurobiology of Huntington's Disease and Parkinson's Disease. Springer Berlin Heidelberg. doi:10.1007/7854_2014_317

Schneider, J. A., Arvanitakis, Z., Bang, W., & Bennett, D. A. (2007). Mixed brain pathologies account for most dementia cases in community-dwelling older persons. *Neurology*, *69*(24), 2197–2204. doi:10.1212/01.wnl.0000271090.28148.24 PMID:17568013

Schneider, J. A., Wilson, R. S., Bienias, J. L., Evans, D. A., & Bennett, D. A. (2004). Cerebral infarctions and the likelihood of dementia from Alzheimer disease pathology. *Neurology*, *62*(7), 148–155. doi:10.1212/01.WNL.0000118211.78503. F5 PMID:15079015

Scholten, N., Pfaff, H., Lehmann, H. C., Fink, G. R., & Karbach, U. (2013). Thrombolysis for acute stroke--a nationwide analysis of regional medical care. *Fortschritte der Neurologie-Psychiatrie*, *81*(10), 579–585. PMID:24081518

Schönberger, K., Ludwig, M. S., Wildner, M., & Weissbrich, B. (2013). Epidemiology of Subacute Sclerosing Panencephalitis (SSPE) in Germany from 2003 to 2009: A Risk Estimation. *PLoS One*, 8(7), e68909. doi:10.1371/journal. pone.0068909 PMID:23874807

Schreibman, L. (2000). Intensive behavioral/psychoeducational treatments for autism: Research needs and future directions. *Journal of Autism and Developmental Disorders*, *30*(5), 373–378. doi:10.1023/A:1005535120023 PMID:11098871

Schuler, F., & Casida, J. E. (2001). Functional coupling of PSST and ND1subunits in NADH:ubiquinone oxidoreductase established by photoaffinity labeling. *Biochimica et Biophysica Acta*, *1506*(1), 79–87. doi:10.1016/S0005-2728(01)00183-9 PMID:11418099

Schulze, T., Ahel, M., Ahlheim, J., Aït-Aïssa, S., Brion, F., Di Paolo, C., ... Hollert, H. (2017). Assessment of a novel device for onsite integrative large-volume solid phase extraction of water samples to enable a comprehensive chemical and effect-based analysis. *The Science of the Total Environment*, *581*, 350–358. doi:10.1016/j.scitotenv.2016.12.140 PMID:28062104

Schulz-Schaeffer, W. J. (2010). The synaptic pathology of α -synuclein aggregation in dementia with Lewy bodies, Parkinson's disease and Parkinson's disease dementia. *Acta Neuropathologica*, *120*(2), 131–143. doi:10.100700401-010-0711-0 PMID:20563819

Schumanon, C.M., & Amaral, D.G. (2006). Stereological analysis of amygdala neuron number in autism. *J Neurosci*, 26, 7674–7679.

Schüpbach, W. M., Rau, J., Knudsen, K., Volkmann, J., Krack, P., Timmermann, L., & ... (2013). Neurostimulation for Parkinson's disease with early motor complications. *The New England Journal of Medicine*, *368*(7), 610–622. doi:10.1056/NEJMoa1205158 PMID:23406026

Schuster-Böckler, B., & Bateman, A. (2008). Protein interactions in human genetic diseases. *Genome Biology*, 9(1), R9. doi:10.1186/gb-2008-9-1-r9 PMID:18199329

Scotcher, K. P., Irwin, I., DeLanney, L. E., Langston, J. W., & Monte, D. (1990). Effects of 1-Methyl-4-Phenyl-1, 2, 3, 6-Tetrahydropyridine and 1-Methyl-4-Phenylpyridinium Ion on ATP Levels of Mouse Brain Synaptosomes. *Journal of Neurochemistry*, *54*(4), 1295–1301. doi:10.1111/j.1471-4159.1990.tb01962.x PMID:2313288

Seabrook, G. R., Ray, W. J., Shearman, M., & Hutton, M. (2007). Beyond amyloid: The next generation of Alzheimer's disease therapeutics. *Molecular Interventions*, 7(5), 261–270. doi:10.1124/mi.7.5.8 PMID:17932415

Seidel, K., Mahlke, J., Siswanto, S., Krüger, R., Heinsen, H., Auburger, G., ... den Dunnen, W. (2015). The brainstem pathologies of Parkinson's disease and dementia with Lewy bodies. *Brain Pathology (Zurich, Switzerland)*, 25(2), 121–135. doi:10.1111/bpa.12168 PMID:24995389

Sejvar, J. J., Holman, R. C., Bresee, J. S., Kochanek, K. D., & Schonberger, L. B. (2005). Amyotrophic lateral sclerosis mortality in the United States, 1979–2001. *Neuroepidemiology*, 25(3), 144–152. doi:10.1159/000086679 PMID:15990445

Selikhova, M., Williams, D. R., Kempster, P. A., Holton, J. L., Revesz, T., & Lees, A. J. (2009). A clinico-pathological study of subtypes in Parkinson's disease. *Brain*, *132*(11), 2947–2957. doi:10.1093/brain/awp234 PMID:19759203

Selkoe, D. (1990). Amyloid β-protein deposition as a seminal pathogenic event in AD: An hypothesis. *Neurobiology of Aging*, *11*(3), 299. doi:10.1016/0197-4580(90)90746-M

Selkoe, D. J. (2001). Alzheimer's disease: Genes, proteins, and therapy. *Physiological Reviews*, 81(2), 741–766. doi:10.1152/physrev.2001.81.2.741 PMID:11274343

Selkoe, D. J. (2002). Alzheimer's disease is a synaptic failure. *Science*, 298(5594), 789–791. doi:10.1126cience.1074069 PMID:12399581

Selkoe, D. J., & Schenk, D. (2003). Alzheimer's disease: Molecular understanding predicts amyloid-based therapeutics. *Annual Review of Pharmacology and Toxicology*, *43*(1), 545–584. doi:10.1146/annurev.pharmtox.43.100901.140248 PMID:12415125

Selley, M., Close, D., & Stern, S. (2002). The effect of increased concentrations of homocysteine on the concentration of (E)-4-hydroxy-2-nonenal in the plasma and cerebrospinal fluid of patients with Alzheimer's disease. *Neurobiology of Aging*, *23*(3), 383–388. doi:10.1016/S0197-4580(01)00327-X PMID:11959400

Sen, S., Webber, P. J., & West, A. B. (2009). Dependence of leucine-rich repeat kinase 2 (LRRK2) kinase activity on dimerization. *The Journal of Biological Chemistry*, 284(52), 36346–36356. doi:10.1074/jbc.M109.025437 PMID:19826009

Sensi, S. L., Ton-That, D., Sullivan, P. G., Jonas, E. A., Gee, K. R., Kaczmarek, L. K., & Weiss, J. H. (2003). Modulation of mitochondrial function by endogenous Zn²⁺ pools. *Proceedings of the National Academy of Sciences of the United States of America*, *100*(10), 6157–6162. doi:10.1073/pnas.1031598100 PMID:12724524

Seppi, K., Weintraub, D., Coelho, M., Perez-Lloret, S., Fox, S. H., Katzenxchlager, R., ... Sampaio, C. (2011). The Movement Disorder Society Evidence-Based Medicine Review Update: Treatments for the non-Motor symptoms of Parkinson's disease. *Movement Disorders*, 26(3), 42–80. doi:10.1002/mds.23884 PMID:22021174

Serpell, L. C., & Smith, J. M. (2000). Direct visualisation of the β-sheet structure of synthetic Alzheimer's amyloid. *Journal of Molecular Biology*, 299(1), 225–231. doi:10.1006/jmbi.2000.3650 PMID:10860734

Serraz, B., Grand, T., & Paoletti, P. (2016). Altered zinc sensitivity of NMDA receptors harboring clinically-relevant mutations. *Neuropharmacology*, *109*, 196–204. doi:10.1016/j.neuropharm.2016.06.008 PMID:27288002

Settembre, C., Fraldi, A., Jahreiss, L., Spampanato, C., Venturi, C., Medina, D., ... Ballabio, A. (2008). A block of autophagy in lysosomal storage disorders. *Human Molecular Genetics*, *17*(1), 119–129. doi:10.1093/hmg/ddm289 PMID:17913701

Severin, S. E., & Boldyrev, A. A. (1991). Effect of carnosine, a specific component of striated muscle, on muscles and other tissues. *Biomedical Science*, *2*, 91–94. PMID:1912249

Seward, M. E., Swanson, E., Norambuena, A., Reimann, A., Cochran, J. N., Li, R., ... Bloom, G. S. (2013). Amyloidsignals through tau to drive ectopic neuronal cell cycle re-entry in Alzheimer's disease. *Journal of Cell Science*, *126*(5), 1278–1286. doi:10.1242/jcs.1125880 PMID:23345405

Shachar, T., Lo-Bianco, C., Recchia, A., Wiessner, C., Raas-Rosthschild, A., & Futerman, A. H. (2011). Lysosomal Storage Disorders and Parkinson's Disease: Gaucher Disease and Beyond. *Movement Disorders*, 29(9), 1593–1604. doi:10.1002/mds.23774 PMID:21618611

Shah, J., & Zeier, J. (2013). Long-distance communication and signal amplification in systemic acquired resistance. *Frontiers in Plant Science*, *4*, 30. doi:10.3389/fpls.2013.00030 PMID:23440336

Shannon, K. M. (1998). Ballism. In J. Jankovic & E. Tolosa (Eds.), *Parkinson's disease and movement disorders. 3* (pp. 365–375). Baltimore, MD: Williams and Wilkins.

Shao, J., & Diamond, M. I. (2007). Polyglutamine diseases: Emerging concepts in pathogenesis and therapy. *Human Molecular Genetics*, *16*(R2), R115–R123. doi:10.1093/hmg/ddm213 PMID:17911155

Sharma, D. R., Sunkaria, A., Wani, W. Y., Sharma, R. K., Kandimalla, R. J., Bal, A., & Gill, K. D. (2013). Aluminium induced oxidative stress results in decreased mitochondrial biogenesis via modulation of PGC-1α expression. *Toxicology and Applied Pharmacology*, 273(2), 365–380. doi:10.1016/j.taap.2013.09.012 PMID:24084166

Sharma, M., Tiwari, M., & Tiwari, R. K. (2015). Hyperhomocysteinemia: Impact on Neurodegenerative Diseases. *Basic & Clinical Pharmacology & Toxicology*, *117*(5), 287–296. doi:10.1111/bcpt.12424 PMID:26036286

554

Sharma, S., Moon, C. S., Khogali, A., Haidous, A., Chabenne, A., Ojo, C., ... Ebadi, M. (2013, September). Biomarkers in Parkinson's disease (recent update). *Neurochemistry International*, *63*(3), 201–229. doi:10.1016/j.neuint.2013.06.005 PMID:23791710

Shaw, A. S., Ampong, M. A., Rio, A., McClure, J., Leigh, P. N., & Sidhu, P. S. (2004). Entristar skin-level gastrostomy tube: Primary placement with radiologic guidance in patients with amyotrophic lateral sclerosis. *Radiology*, *233*(2), 392–399. doi:10.1148/radiol.2332031487 PMID:15459322

Shaw, P.J. (1999). Motor neurone disease. *British Medical Journal*, *318*(7191), 1118–1121. doi:10.1136/bmj.318.7191.1118 PMID:10213726

Shaw, P. J., Chinnery, R. M., Thagesen, H., Borthwick, G. M., & Ince, P. G. (1997). Immunocytochemical study of the distribution of the free radical scavenging enzymes Cu/Zn superoxide dismutase (SOD1); MN superoxide dismutase (MN SOD) and catalase in the normal human spinal cord and in motor neuron disease. *Journal of the Neurological Sciences*, *147*(2), 115–125. doi:10.1016/S0022-510X(96)05316-6 PMID:9106116

Shelkovnikova, T., Kulikova, A., Tsvetkov, F., Peters, O., Bachurin, S., Bukhman, V., & Ninkina, N. (2012). Proteinopathies--forms of neurodegenerative disorders with protein aggregation-based pathology. *Molekuliarnaia biologiia*, 46(3), 402–415. PMID:22888630

Shelkovnikova, T., Kulikova, A., Tsvetkov, P. O., Peters, O., Bachurin, S., Buchman, V. L., & Ninkina, N. (2012). Proteinopathies, neurodegenerative disorders with protein aggregation-based pathology. *Molecular Biology*, *46*(3), 362–374. doi:10.1134/S0026893312020161 PMID:22888630

Sheng, B., Wang, X., Su, B., Lee, H. G., Casadesus, G., Perry, G., & Zhu, X. (2012). Impaired mitochondrial biogenesis contributes to mitochondrial dysfunction in Alzheimer's disease. *Journal of Neurochemistry*, *120*(3), 419–429. doi:10.1111/j.1471-4159.2011.07581.x PMID:22077634

Shepherd, J. D., & Huganir, R. L. (2007). The cell biology of synaptic plasticity: AMPA receptor trafficking. *Annual Review of Cell and Developmental Biology*, 23(1), 613–643. doi:10.1146/annurev.cellbio.23.090506.123516 PMID:17506699

Shepherd, J., Blauw, G. J., Murphy, M. B., Bollen, E. L., Buckley, B. M., Cobbe, S. M., ... Jukema, J. W. (2002). Pravastatin in elderly individuals at risk of vascular disease (PROSPER): A randomised controlled trial. *Lancet*, *360*(9346), 1623–1630. doi:10.1016/S0140-6736(02)11600-X PMID:12457784

Sherer, T. B., Betarbet, R., Kim, J. H., & Greenamyre, J. T. (2003c). Selective microglial activation in the rat rotenone model of Parkinson's disease. *Neuroscience Letters*, *341*(2), 87–90. doi:10.1016/S0304-3940(03)00172-1PMID:12686372

Sherer, T. B., Betarbet, R., Testa, C. M., Seo, B. B., Richardson, J. R., Kim, J. H., ... Greenamyre, J. T. (2003a). Mechanism of toxicity in rotenone models of Parkinson's disease. *The Journal of Neuroscience*, 23(34), 10756–10764. PMID:14645467

Sherer, T. B., Kim, J. H., Betarbet, R., & Greenamyre, J. T. (2003b). Subcutaneous rotenone exposure causes highly selective dopaminergic degeneration and α -synuclein aggregation. *Experimental Neurology*, *179*(1), 9–16. doi:10.1006/exnr.2002.8072 PMID:12504863

Shibata, N. (2001). Transgenic mouse model for familial amyotrophic lateral sclerosis with superoxide dismutase 1 mutation. *Neuropathology*, 21(1), 82–92. doi:10.1046/j.1440-1789.2001.00361.x PMID:11304046

Shibata, N., Hirano, A., Yamamoto, T., Kato, Y., & Kobayashi, M. (2000). Superoxide dismutase-1 mutation-related neurotoxicity in familial amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, *1*(3), 143–161. doi:10.1080/14660820050515151 PMID:11464949

Shimada, H., Hirano, S., Shinotoh, H., Aotsuka, A., Sato, K., Tanaka, N., ... Hattori, T. (2009). Mapping of brain acetylcholinesterase alterations in Lewy body disease by PET. *Neurology*, *73*(4), 273–278. doi:10.1212/WNL.0b013e3181ab2b58 PMID:19474411

Shimizu, K., Ohtaki, K., Matsubara, K., Aoyama, K., Uezono, T., Saito, O., ... Shiono, H. (2001). Carrier-mediated processes in blood–brain barrier penetration and neural uptake of paraquat. *Brain Research*, *906*(1), 135–142. doi:10.1016/S0006-8993(01)02577-X PMID:11430870

Shin, S.-G., Kim, J. Y., Chung, H. Y., & Jeong, J. C. (2005). Zingerone as an antioxidant against peroxynitrite. *Journal of Agricultural and Food Chemistry*, 53(19), 7617–7622. doi:10.1021/jf051014x PMID:16159194

Shipton, O. A., Leitz, J. R., Dworzak, J., Acton, C. E. J., Tunbridge, E. M., Denk, F., ... Vargas-Caballero, M. (2011). Tau Protein Is Required for Amyloid -Induced Impairment of Hippocampal Long-Term Potentiation. *The Journal of Neuroscience*, *31*(5), 1688–1692. doi:10.1523/JNEUROSCI.2610-10.2011 PMID:21289177

Shors, T. J. (2006). Stressful Experience and Learning Across the Lifespan. *Annual Review of Psychology*, *57*(1), 55–85. doi:10.1146/annurev.psych.57.102904.190205 PMID:16318589

Shorter, J., & Lindquist, S. (2005). Prions as adaptive conduits of memory and inheritance. *Nature Reviews. Genetics*, *6*(6), 435–450. doi:10.1038/nrg1616 PMID:15931169

Shprintzen, R. J. (2006). Velo-cardio-facial syndrome: A distinctive behavioral phenotype. *Mental Retardation and Developmental Disabilities Research Reviews*, 6(2), 142–147. doi: PMID:10899808

Shu, S. K., Michelson, D. J., & Ashwal, S. (2017). Cognitive and Motor Regression. In Swaiman's Pediatric Neurology: Principles & Practice (6th ed.; pp. 424-430). Elsevier Inc. doi:10.1016/B978-0-323-37101-8.00052-7

Siddiqi, Z. A., Sanders, D. B., & Massey, J. M. (2006). Peripheral neuropathy in Krabbe disease: Electrodiagnostic findings. *Neurology*, 67(2), 263–267. doi:10.1212/01.wnl.0000230153.34613.84 PMID:16864819

Siddiqui, F. M., Dandapat, S., Banerjee, C., Zuurbier, S. M., Johnson, M., Stam, J., & Coutinho, J. M. (2015). Mechanical thrombectomy in cerebral venous thrombosis: Systematic review of 185 cases. *Stroke*, *46*(5), 1263–1268. doi:10.1161/STROKEAHA.114.007465 PMID:25899238

Sieradzan, K., Channon, S., Ramponi, C., Stern, G. M., Lees, A. J., & Youdim, M. B. (1995). The therapeutic potential of moclobemide, a reversible selective monoamine oxidase A inhibitor in Parkinson's disease. *Journal of Clinical Psychopharmacology*, *15*(4), 51S–59S. doi:10.1097/00004714-199508001-00010 PMID:7593732

Sies, H. (1997). Oxidative stress: Oxidants and antioxidants. *Experimental Physiology*, 82(2), 291–295. doi:10.1113/ expphysiol.1997.sp004024 PMID:9129943

Sigman, M., & Capps, L. (1997). Children with autism: A developmental perspective. Boston: Harvard University Press.

Silhavy, T. J., Kahne, D., & Walker, S. (2010). The bacterial cell envelope. *Cold Spring Harbor Perspectives in Biology*, 2(5), a000414. doi:10.1101/cshperspect.a000414 PMID:20452953

Simon, D. K., Lin, M. T., Zheng, L., Liu, G.-J., Ahn, C. H., Kim, L. M., ... Johns, D. R. (2004). Somatic mitochondrial DNA mutations in cortex and substantia nigra in aging and Parkinson's disease. *Neurobiology of Aging*, 25(1), 71–81. doi:10.1016/S0197-4580(03)00037-X PMID:14675733

Simon, D., Pulst, S., Sutton, J., Browne, S., Beal, M., & Johns, D. (1999). Familial multisystem degeneration with parkinsonism associated with the 11778 mitochondrial DNA mutation. *Neurology*, *53*(8), 1787–1787. doi:10.1212/WNL.53.8.1787 PMID:10563629

Simuni, T., Luo, S. T., Chou, K. L., Fernandez, H., He, B., & Parashos, S. (2013). Rankin scale as a potencial measure of global disability in early Parkinson's disease. *Journal of Clinical Neuroscience*, 20(9), 1200–1203. doi:10.1016/j. jocn.2012.10.030 PMID:23810387

Sinclair, A. J., Bayer, A. J., Johnston, J., Warner, C., & Maxwell, S. R. (1998). Altered plasma antioxidant status in subjects with Alzheimer's disease and vascular dementia. *International Journal of Geriatric Psychiatry*, *13*(12), 840–845. doi: PMID:9884908

Singer, H. S., Mink, J. W., Gilbert, D. L., & Jankovic, J. (2010). Inherited Metabolic Disorders Associated with Extrapyramidal Symptoms. *Movement Disorders in Childhood*, 164-204.

Singhal, S., Bevan, S., Barrick, T., Rich, P., & Markus, H. S. (2004). The influence of genetic and cardiovascular risk factors on the CADASIL phenotype. *Brain*, *127*(9), 2031–2038. doi:10.1093/brain/awh223 PMID:15229130

Singh, S. K., Srivastav, S., Yadav, A. K., Srikrishna, S., & Perry, G. (2016). Overview of Alzheimer's Disease and Some Therapeutic Approaches Targeting Aβ by Using Several Synthetic and Herbal Compounds. *Oxidative Medicine and Cellular Longevity*, 2016, 1–22. doi:10.1155/2016/7361613 PMID:27034741

Singleton, A. B., Farrer, M., Johnson, J., Singleton, A., Hague, S., Kachergus, J., ... Lincoln, S. (2003). α-Synuclein locus triplication causes Parkinson's disease. *Science*, *302*(5646), 841–841. doi:10.1126cience.1090278 PMID:14593171

Sirven, J. I., & Shafer, P. O. (2014). *What is Epilepsy?* Epilepsy Foundation and Epilepsy Therapy Project. Available at: http://www.epilepsy.com/learn/about-epilepsy-basics/what-epilepsy

Sitburana, O., & Ondo, W. G. (2009). Brain magnetic resonance imaging (MRI) in parkinsonian disorders. *Parkinsonism & Related Disorders*, *15*(3), 165–174. doi:10.1016/j.parkreldis.2008.04.033 PMID:19059803

Skovronsky, D. M., Moore, D. B., Milla, M. E., Doms, R. W., & Lee, V. M. (2000). Protein kinase C-dependent alphasecretase competes with beta-secretase for cleavage of amyloid-beta precursor protein in the trans-golgi network. *The Journal of Biological Chemistry*, 275(4), 2568–2575. doi:10.1074/jbc.275.4.2568 PMID:10644715

Sleegers, K., Cruts, M., & Van Broeckhoven, C. (2010). Molecular pathways of frontotemporal lobar degeneration. *Annual Review of Neuroscience*, *33*(1), 71–88. doi:10.1146/annurev-neuro-060909-153144 PMID:20415586

Slomianka, L., Danscher, G., & Frederickson, C. J. (1990). Labeling of the neurons of origin of zinc-containing pathways by intraperitoneal injections of sodium selenite. *Neuroscience*, *38*(3), 843–854. doi:10.1016/0306-4522(90)90076-G PMID:2176723

Small, D. H., & Cappai, R. (2006). Alois Alzheimer and Alzheimer's disease: A centennial perspective. *Journal of Neurochemistry*, 99(3), 708–710. doi:10.1111/j.1471-4159.2006.04212.x PMID:17076655

Small, G. W., Bookheimer, S. Y., Thompson, P. M., Cole, G. M., Huang, S., Kepe, V., & Barrio, J. R. (2008). Current and future uses of neuroimaging for cognitively impaired patients. *Lancet Neurology*, 7(2), 161–172. doi:10.1016/S1474-4422(08)70019-X PMID:18207114

Smith, D. G., Cappai, R., & Barnham, K. J. (2007). The redox chemistry of the Alzheimer's disease amyloid β peptide. *Biochimica et Biophysica Acta (BBA)-. Biomembranes, 1768*(8), 1976–1990. doi:10.1016/j.bbamem.2007.02.002

Smith, E. E., Cieslak, A., Barber, P., Chen, J., Chen, Y. W., Donnini, I., ... Hachinski, V. (2017). Therapeutic Strategies and Drug Development for Vascular Cognitive Impairment. *Journal of the American Heart Association*, 6(5), e005568. doi:10.1161/JAHA.117.005568 PMID:28476873

Smith, E. E., Schneider, J. A., Wardlaw, J. M., & Greenberg, S. M. (2012). Cerebral microinfarcts: The invisible lesions. *Lancet Neurology*, *11*(13), 272–282. doi:10.1016/S1474-4422(11)70307-6 PMID:22341035

Smithers, G., Finch, S., Doyle, W., Lowe, C., Bates, C. J., Prentice, A., & Clarke, P. C. (1998). The national diet and nutrition survey: People aged 65 years and over. *Nutrition & Food Science*, *98*(3), 133–134. doi:10.1108/00346659810209791

Smith, M. A., Richey, G. P. P., Sayre, L. M., Anderson, V. E., Beal, M. F., & Kowal, N. (1996). Test for oxidative damage in Alzheimer's. *Nature*, *382*(6587), 120–121. doi:10.1038/382120b0 PMID:8700201

Smith, R. A., Curtain, R., Ovcaric, M., Tajouri, L., Macmillan, J., & Griffiths, L. (2008). Investigation of the NOTCH3 and TNFSF7 genes on C19p13 as candidates for migraine. *The Open Neurology Journal*, 2(1), 1–7. doi:10.2174/1874205X00802010001 PMID:19018300

Smith, W. W., Pei, Z., Jiang, H., Dawson, V. L., Dawson, T. M., & Ross, C. A. (2006). Kinase activity of mutant LRRK2 mediates neuronal toxicity. *Nature Neuroscience*, *9*(10), 1231–1233. doi:10.1038/nn1776 PMID:16980962

Snow, B. J., Rolfe, F. L., Lockhart, M. M., Frampton, C. M., O'Sullivan, J. D., Fung, V., ... Taylor, K. M. (2010). A double-blind, placebo-controlled study to assess the mitochondria-targeted antioxidant MitoQ as a disease-modifying therapy in Parkinson's disease. *Movement Disorders*, 25(11), 1670–1674. doi:10.1002/mds.23148 PMID:20568096

Sode, K., Ochiai, S., Kobayashi, N., & Usuzaka, E. (2007). Effect of reparation of repeat sequences in the human alpha-synuclein on fibrillation ability. *International Journal of Biological Sciences*, *3*(1), 1–7. doi:10.7150/ijbs.3.1 PMID:17200685

Sohal, R. S., Ku, H. H., Agarwal, S., Forster, M. J., & Lal, H. (1994). Oxidative damage, mitochondrial oxidant generation and antioxidant defenses during aging and in response to food restriction in the mouse. *Mechanisms of Ageing and Development*, 74(1), 121–133. doi:10.1016/0047-6374(94)90104-X PMID:7934203

Sohal, R. S., & Sohal, B. H. (1991). Hydrogen peroxide release by mitochondria increases during aging. *Mechanisms of Ageing and Development*, *57*(2), 187–202. doi:10.1016/0047-6374(91)90034-W PMID:1904965

Solís-Herrera, A., & Ashraf, G. M., Esparza, M., Arias, R. I., Bachurin, S. O., Barreto, G. E., & Aliev, G. (2015). Biological activities of QIAPI 1 as a melanin precursor and its therapeutic effects in wistar rats exposed to arsenic poisoning. *Central Nervous System Agents in Medicinal Chemistry*, *15*(2), 99–108. doi:10.2174/18715249156661504241138 31 PMID:25909193

Somlyo, A. P., Bond, M., & Somlyo, A. V. (1985). Calcium content of mitochondria and endoplasmic reticulum in liver frozen rapidly *in vivo. Nature*, *314*(6012), 622–625. doi:10.1038/314622a0 PMID:3990795

Song, D. D., Shults, C. W., Sisk, A., Rockenstein, E., & Masliah, E. (2004). Enhanced substantia nigra mitochondrial pathology in human α -synuclein transgenic mice after treatment with MPTP. *Experimental Neurology*, *186*(2), 158–172. doi:10.1016/S0014-4886(03)00342-X PMID:15026254

Sonnen, J. A., Larson, E. B., Crane, P. K., Haneuse, S., Li, G., Schellenberg, G. D., ... Montine, T. J. (2007). Pathological correlates of dementia in a longitudinal, population-based sample of aging. *Annals of Neurology*, *62*(4), 406–413. doi:10.1002/ana.21208 PMID:17879383

Soong, N. W., Hinton, D. R., Cortopassi, G., & Arnheim, N. (1992). Mosaicism for a specific somatic mitochondrial DNA mutation in adult human brain. *Nature Genetics*, 2(4), 318–323. doi:10.1038/ng1292-318 PMID:1303287

Soontornniyomkij, V., Lynch, M. D., Mermash, S., Pomakian, J., Badkoobehi, H., Clare, R., & Vinters, H. V. (2010). Cerebral microinfarcts associated with severe cerebral beta-amyloid angiopathy. *Brain Pathology (Zurich, Switzerland)*, 20(2), 459–467. doi:10.1111/j.1750-3639.2009.00322.x PMID:19725828

Sorwell, K. G., & Urbanski, H. F. (2013). Causes and consequences of age-related steroid hormone changes: Insights gained from nonhuman primates. *Journal of Neuroendocrinology*, 25(11), 1062–1069. doi:10.1111/jne.12064 PMID:23796387

558

Sosa-Ortiz, A. L., Acosta-Castillo, I., & Prince, M. J. (2012). Epidemiology of dementias and Alzheimer's disease. *Archives of Medical Research*, 43(8), 600–608. doi:10.1016/j.arcmed.2012.11.003 PMID:23159715

Soto, C. (2011). Prion hypothesis: The end of the controversy? *Trends in Biochemical Sciences*, *36*(3), 151–158. doi:10.1016/j.tibs.2010.11.001 PMID:21130657

Soto, C., & Castilla, J. (2004). The controversial protein-only hypothesis of prion propagation. *Nature Medicine*, *10*(July), 63–67. doi:10.1038/nm1069 PMID:15272271

Soto, C., & Estrada, L. D. (2008). Protein misfolding and neurodegeneration. *Archives of Neurology*, 65(2), 184–189. doi:10.1001/archneurol.2007.56 PMID:18268186

Sotres-Bayon, F., & Quirk, G. J. (2010). Prefrontal control of fear: More than just extinction. *Current Opinion in Neurobiology*, 20(2), 231–235. doi:10.1016/j.conb.2010.02.005 PMID:20303254

Sparks, D. L., Scheff, S. W., Liu, H., Landers, T. M., Coyne, C. M., & Hunsaker, J. C. III. (1995). Increased incidence of neurofibrillary tangles (NFT) in non-demented individuals with hypertension. *Journal of the Neurological Sciences*, *131*(2), 162–169. doi:10.1016/0022-510X(95)00105-B PMID:7595642

Spencer, T., Biederman, J., Wilens, T., Harding, M., O'Donneel, D., & Griffin, S. (1996). Pharmacotherapy of attentiondeficit hyperactivity disorder across the life cycle. *Journal of the American Academy of Child and Adolescent Psychiatry*, *35*(4), 409–432. doi:10.1097/00004583-199604000-00008 PMID:8919704

Sperlágh, B., & Sylvester Vizi, E. (2011). The role of extracellular adenosine in chemical neurotransmission in the hippocampus and Basal Ganglia: Pharmacological and clinical aspects. *Current Topics in Medicinal Chemistry*, *11*(8), 1034–1046. doi:10.2174/156802611795347564 PMID:21401497

Spezio, M. L., Huang, P. Y., Castelli, F., & Adolphs, R. (2007). Amygdala damage impairs eye contact during conversations with real people. *The Journal of Neuroscience*, *27*(15), 3994–3997. doi:10.1523/JNEUROSCI.3789-06.2007 PMID:17428974

Spillantini, M. G., Bird, T. D., & Ghetti, B. (1998). Frontotemporal dementia and Parkinsonism linked to chromosome 17: A new group of tauopathies. *Brain Pathology (Zurich, Switzerland)*, 8(2), 387–402. doi:10.1111/j.1750-3639.1998. tb00162.x PMID:9546295

Spillantini, M. G., Schmidt, M. L., Lee, V. M. Y., Trojanowski, J. Q., Jakes, R., & Goedert, M. (1997). [alpha]-Synuclein in Lewy bodies. *Nature*, *388*(6645), 839–840. doi:10.1038/42166 PMID:9278044

Spratley, S. J., & Deane, J. E. (2016). New therapeutic approaches for Krabbe disease: The potential of pharmacological chaperones. *Journal of Neuroscience Research*, *94*(11), 1203–1219. doi:10.1002/jnr.23762 PMID:27638604

Spratley, S. J., Hill, C. H., Viuff, A. H., Edgar, J. R., Skjødt, K., & Deane, J. E. (2016). Molecular Mechanisms of Disease Pathogenesis Differ in Krabbe Disease Variants. *Traffic (Copenhagen, Denmark)*, *17*(8), 908–922. doi:10.1111/tra.12404 PMID:27126738

Sprengers, M., Vonck, K., Carrette, E., & Boon, P. (2017). Deep brain and cortical stimulation for epilepsy. *Cochrane Database of Systematic Reviews*, 7. PMID:28718878

Springer, S., Erlewin, R., Nagaele, T., Becker, I., Auer, D., Grodd, W., & Krägeloh-Mann, I. (2000). Alexander Disease - Classification revisited and isolation of a neonatal form. *Neuropediatrics*, *31*(2), 86–92. doi:10.1055-2000-7479 PMID:10832583

Spuch, C., Ortolano, S., & Navarro, C. (2012, May). Lafora progressive myoclonus epilepsy: Recent insights into cell degeneration. *Recent Patents on Endocrine, Metabolic & Immune Drug Discovery*, 6(2), 99–107. doi:10.2174/187221412800604617 PMID:22369717

Sreedharan, J., Blair, I. P., Tripathi, V. B., Hu, X., Vance, C., Rogelj, B., ... Shaw, C. E. (2008). TDP-43 mutations in familial and sporadic amyotrophic lateral sclerosis. *Science*, *319*(5870), 1668–1672. doi:10.1126cience.1154584 PMID:18309045

Srivasthava, N. S. (2002, Nov. 15). Alexander disease. In Gene Reviews. Seattle, WA: University of Washington.

St George, R. J., Nutt, J. G., Burchiel, K. J., & Horak, F. B. (2010, October). A meta-regression of the long-term effects of deep brain stimulation on balance and gait in PD. *Neurology*, 75(14), 1292–1299. doi:10.1212/WNL.0b013e3181f61329 PMID:20921515

Stamenkovic, S., Ducic, T., Stamenkovic, V., Kranz, A., & Andjus, P. R. (2017). Imaging of glial cell morphology, SOD1 distribution and elemental composition in the brainstem and hippocampus of the ALS hSOD1 G93A rat. *Neuroscience*, *357*, 37–55. doi:10.1016/j.neuroscience.2017.05.041 PMID:28576725

Stamer, K., Vogel, R., Thies, E., Mandelkow, E., & Mandelkow, E. M. (2002). Tau blocks traffic of organelles, neurofilaments, and APP vesicles in neurons and enhances oxidative stress. *The Journal of Cell Biology*, *156*(6), 1051–1063. doi:10.1083/jcb.200108057 PMID:11901170

Stam, J. (2005). Thrombosis of the cerebral veins and sinuses. *The New England Journal of Medicine*, 352(17), 1791–1798. doi:10.1056/NEJMra042354 PMID:15858188

Starr, P. A. (2002). Placement of deep brain stimulators into the subthalamic nucleus or Globus pallidus internus: Technical approach. *Stereotactic and Functional Neurosurgery*, 79(3-4), 118–145. doi:10.1159/000070828 PMID:12890973

States, U., Gambetti, P., Hunter, S., Maddox, R. A., Crockett, L., Zaki, S. R., & Schonberger, L. B. (2005). Variant Creutzfeldt-Jakob Disease. *Centres for Disease Control and Prevention*, 11(9), 9–12.

Stayte, S., & Vissel, B. (2014). Advances in non-dopaminergic treatments for Parkinson's disease. *Frontiers in Neuroscience*, *8*, 113. PMID:24904259

Stefani, A., Peppe, A., Pierantozzi, M., Galati, S., Moschella, V., Stanzione, P., & Mazzone, P. (2009). Multi-target strategy for Parkinsonian patients: The role of deep brain stimulation in the centromedian-parafascicularis complex. *Brain Research Bulletin*, 78(2-3), 113–118. doi:10.1016/j.brainresbull.2008.08.007 PMID:18812214

Stefani, M. (2004). Protein misfolding and aggregation: New examples in medicine and biology of the dark side of the protein world. *Biochimica et Biophysica Acta (BBA)- Molecular Basis of Disease*, *1739*(1), 5–25. doi:10.1016/j. bbadis.2004.08.004 PMID:15607113

Stefanis, L. (2012). α-Synuclein in Parkinson's Disease. *Cold Spring Harbor Perspectives in Medicine*, 2(2), a009399. doi:10.1101/cshperspect.a009399 PMID:22355802

Steffenburg, S., & Gillberg, C. (1986). Autism and autistic like condition in Swedish rural and urban areas: A population study. *The British Journal of Psychiatry*, *149*(1), 81–87. doi:10.1192/bjp.149.1.81 PMID:3779317

Steffens, S., & Pacher, P. (2015). The activated endocannabinoid system in atherosclerosis: Driving force or protective mechanism? *Current Drug Targets*, *16*(4), 334–341. doi:10.2174/1389450115666141202113225 PMID:25469884

Stein, J. F. (2009). Akinesia, motor oscillations and the pedunculopontine nucleus in rats and men. *Experimental Neurology*, *215*(1), 1–4. doi:10.1016/j.expneurol.2008.09.022 PMID:18977223

Stephens, S., Kenny, R. A., Rowan, E., Allan, L., Kalaria, R. N., Bradbury, M., & Ballard, C. G. (2004). Neuropsychological characteristics of mild vascular cognitive impairment and dementia after stroke. *International Journal of Geriatric Psychiatry*, 19(11), 1053–1057. doi:10.1002/gps.1209 PMID:15481073

Sterky, F. H., Lee, S., Wibom, R., Olson, L., & Larsson, N. G. (2011). Impaired mitochondrial transport and Parkinindependent degeneration of respiratory chain-deficient dopamine neurons in vivo. *Proceedings of the National Academy* of Sciences of the United States of America, 108(31), 12937–12942. doi:10.1073/pnas.1103295108 PMID:21768369

Stewart, H. G., Andersen, P. M., Eisen, A., & Weber, M. (2006). Corticomotoneuronal dysfunction in ALS patients with different SOD1 mutations. *Clinical Neurophysiology*, *117*(8), 1850–1861. doi:10.1016/j.clinph.2006.04.004 PMID:16793335

Stigler, K. A., McDonald, B. C., Anand, A., Saykin, A. J., & McDougle, C. J. (2011). Structural and functional magnetic resonance imaging of autism spectrum disorders. *Brain Research*, *1380*, 146–161. doi:10.1016/j.brainres.2010.11.076 PMID:21130750

Stoessl, A. J. (2014, January). Developments in neuroimaging: Positron emission tomography. *Parkinsonism & Related Disorders*, 20(Suppl 1), S180–S183. doi:10.1016/S1353-8020(13)70042-7 PMID:24262176

Storey & Cappai. (1999). The amyloid precursor protein of Alzheimer's disease and the Abeta peptide. *Neuropathology and Applied Neurobiology*, 25(2), 81–97. 10.1046/j.1365-2990.1999.00164.x

Strauss, I., Kalia, S. K., & Lozano, A. M. (2014, January). Where are we with surgical therapies for Parkinson's disease? *Parkinsonism & Related Disorders*, 20(Suppl 1), S187–S191. doi:10.1016/S1353-8020(13)70044-0 PMID:24262178

Strauss, K. M., Martins, L. M., Plun-Favreau, H., Marx, F. P., Kautzmann, S., Berg, D., ... Kruger, R. (2005). Loss of function mutations in the gene encoding Omi/HtrA2 in Parkinson's disease. *Human Molecular Genetics*, *14*(15), 2099–2111. doi:10.1093/hmg/ddi215 PMID:15961413

Strey, C. W., Spellman, D., Stieber, A., Gonatas, J. O., Wang, X., Lambris, J. D., & Gonatas, N. K. (2004). Dysregulation of stathmin, a microtubule-destabilizing protein, and up-regulation of Hsp25, Hsp27, and the antioxidant peroxiredoxin 6 in a mouse model of familial amyotrophic lateral sclerosis. *American Journal of Pathology*, *165*(5), 1701–1718. doi:10.1016/S0002-9440(10)63426-8 PMID:15509539

Stuart, J. A., & Brown, M. F. (2006). Mitochondrial DNA maintenance and bioenergetics. *Biochimica et Biophysica Acta (BBA)-. Bioenergetics*, *1757*(2), 79–89. doi:10.1016/j.bbabio.2006.01.003

Stuss, D. T., & Knight, R. T. (2013). *Principles of frontal lobe function*. Oxford, UK: Oxford University Press. doi:10.1093/med/9780199837755.001.0001

Su, B., Wang, X., Lee, H. G., Tabaton, M., Perry, G., Smith, M. A., & Zhu, X. (2010). Chronic oxidative stress causes increased tau phosphorylation in M17 neuroblastoma cells. *Neuroscience Letters*, *468*(3), 267–271. doi:10.1016/j.neulet.2009.11.010 PMID:19914335

Su, B., Wang, X., Zheng, L., Perry, G., Smith, M. A., & Zhu, X. (2010). Abnormal mitochondrial dynamics and neurodegenerative diseases. *Biochimica et Biophysica Acta (BBA)-. Molecular Basis of Disease*, *1802*(1), 135–142. doi:10.1016/j. bbadis.2009.09.013 PMID:19799998

Suh, S. W., Jensen, K. B., Jensen, M. S., Silva, D. S., Kesslak, P. J., Danscher, G., & Frederickson, C. J. (2000). Histochemically-reactive zinc in amyloid plaques, angiopathy and degenerating neurons of Alzheimer's disease brains. *Brain Research*, 852(2), 274–278. doi:10.1016/S0006-8993(99)02096-X PMID:10678753 Su, J. H., Deng, G., & Cotman, C. W. (1997). Transneuronal degeneration in the spread of Alzheimer's disease pathology: Immunohistochemical evidence for the transmission of tau hyperphosphorylation. *Neurobiology of Disease*, *4*(5), 365–375. doi:10.1006/nbdi.1997.0164 PMID:9440125

Sultana, R., Boyd-Kimball, D., Poon, H. F., Cai, J., Pierce, W. M., Klein, J. B., ... Butterfield, D. A. (2006). Oxidative modification and down-regulation of Pin1 in Alzheimer's disease hippocampus: A redox proteomics analysis. *Neurobiology of Aging*, *27*(7), 918–925. doi:10.1016/j.neurobiolaging.2005.05.005 PMID:15950321

Sultana, R., Perluigi, M., & Butterfield, D. A. (2013). Lipid peroxidation triggers neurodegeneration: A redox proteomics view into the Alzheimer disease brain. *Free Radical Biology & Medicine*, 62, 157–169. doi:10.1016/j.freeradbiomed.2012.09.027 PMID:23044265

Sulzer, D. (2007). Multiple hit hypotheses for dopamine neuron loss in Parkinson's disease. *Trends in Neurosciences*, *30*(5), 244–250. doi:10.1016/j.tins.2007.03.009 PMID:17418429

Sulzer, D., & Zecca, L. (1999). Intraneuronal dopamine-quinone synthesis: A review. *Neurotoxicity Research*, 1(3), 181–195. doi:10.1007/BF03033289 PMID:12835101

Sumanaweera, T. S., Glover, G. H., Hemler, P. F., van den Elsen, P. A., Martin, D., Adler, J. R., & Napel, S. (1995, July). MR geometric distortion correction for improved frame-based stereotaxic target localization accuracy. *Magnetic Resonance in Medicine*, *34*(1), 106–113. doi:10.1002/mrm.1910340116 PMID:7674887

Summers, P. M., Hartmann, D. A., Hui, E. S., Nie, X., Deardorff, R. L., McKinnon, E. T., ... Jensen, J. H. Sh. (2017). Functional deficits induced by cortical microinfarcts. *Journal of Cerebral Blood Flow and Metabolism*, *37*(11), 3599–3614. doi:10.1177/0271678X16685573 PMID:28090802

Sundaramoorthy, V., Walker, A. K., Tan, V., Fifita, J. A., Mccann, E. P., Williams, K. L., ... Atkin, J. D. (2015). Defects in optineurin-and myosin VI-mediated cellular trafficking in amyotrophic lateral sclerosis. *Human Molecular Genetics*, 24(13), 3830–3846. doi:10.1093/hmg/ddv126 PMID:25859013

Su, Z., Zhang, Y., Gendron, T. F., Bauer, P. O., Chew, J., Yang, W. Y., ... Desaro, P. (2014). Discovery of a biomarker and lead small molecules to target r(GGGGCC)-associated defects in c9FTD/ALS. *Neuron*, *83*(5), 1043–1050. doi:10.1016/j. neuron.2014.07.041 PMID:25132468

Suzuki, K. (2003). Globoid Cell Leukodystrophy (Krabbes Disease): Update. *Journal of Child Neurology*, *18*(9), 595–603. doi:10.1177/08830738030180090201 PMID:14572137

Suzuki, K., Iseki, E., Katsuse, O., Yamaguchi, A., Katsuyama, K., Aoki, I., ... Kosaka, K. (2003). Neuronal accumulation of alpha- and beta-synucleins in the brain of a GM2 gangliosidosis mouse model. *Neuroreport*, *14*(4), 551–554. doi:10.1097/00001756-200303240-00004 PMID:12657883

Suzuki, K., Iseki, E., Togo, T., Yamaguchi, A., Katsuse, O., Katsuyama, K., ... Hirayasy, Y. (2007). Neuronal and glial accumulation of alpha- and beta-synucleins in human lipidoses. *Acta Neuropathologica*, *114*(5), 481–489. doi:10.100700401-007-0264-z PMID:17653558

Suzuki, K., Sugihara, G., Ouchi, Y., Nakamura, K., Futatsubashi, M., Takebayashi, K., ... Mori, N. (2013). Microglial activation in young adults with autism spectrum disorder. *JAMA Psychiatry*, *70*(1), 49–58. doi:10.1001/jamapsychia-try.2013.272 PMID:23404112

Suzuki, N., Maroof, A. M., Merkle, F. T., Koszka, K., Intoh, A., & Armstrong, I., ... Eggan, K. (2013). The mouse C9ORF72 ortholog is enriched in neurons known to degenerate in ALS and FTD. *Nature Neuroscience*, 16.

562

Suzuki, Y., Isogai, K., Teramoto, T., Tashita, H., Shimozawa, N., Nishimura, M., & Kondo, N. (2000). JULY). Bone marrow transplantation for the treatment of X-linked adrenoleukodystrophy. *Journal of Inherited Metabolic Disease*, 23(5), 453–458. doi:10.1023/A:1005656029200 PMID:10947199

Sweeney, P., Park, H., Baumann, M., Dunlop, J., Frydman, J., Kopito, R., ... Hodgson, R. (2017). Protein misfolding in neurodegenerative diseases: Implications and strategies. *Translational Neurodegeneration*, *6*(1), 6. doi:10.118640035-017-0077-5 PMID:28293421

Sweeten, T. L., Posey, D. J., Shekhar, A., & McDougle, C. J. (2002). The amygdala and related structures in the pathophysiology of autism. *Pharmacology, Biochemistry, and Behavior*, *71*(3), 449–455. doi:10.1016/S0091-3057(01)00697-9 PMID:11830179

Swerdlow, R. H., Burns, J. M., & Khan, S. M. (2014). The Alzheimer's disease mitochondrial cascade hypothesis: Progress and perspectives. *Biochimica et Biophysica Acta (BBA)-. Molecular Basis of Disease*, *1842*(8), 1219–1231. doi:10.1016/j.bbadis.2013.09.010 PMID:24071439

Swerdlow, R. H., & Khan, S. M. (2004). A "mitochondrial cascade hypothesis" for sporadic Alzheimer's disease. *Medical Hypotheses*, 63(1), 8–20. doi:10.1016/j.mehy.2003.12.045 PMID:15193340

Swerdlow, R. H., Parks, J. K., Miller, S. W., Davis, R. E., Tuttle, J. B., Trimmer, P. A., ... Parker, W. D. (1996). Origin and functional consequences of the complex I defect in Parkinson's disease. *Annals of Neurology*, *40*(4), 663–671. doi:10.1002/ana.410400417 PMID:8871587

Swerdlow, R., Parks, J., Cassarino, D., Maguire, D., Maguire, R., Bennett, J., ... Parker, W. (1997). Cybrids in Alzheimer's disease: A cellular model of the disease? *Neurology*, *49*(4), 918–925. doi:10.1212/WNL.49.4.918 PMID:9339668

Swillen, A., Devriendt, K., Legius, E., Prinzie, P., Vogels, A., & Ghesquiere, P. (1999). The behavioral phenotype in velo-cardio-facial syndrome (VCFS): From infancy to adolescence. *Genetic Counseling (Geneva, Switzerland)*, *10*, 79–88. PMID:10191433

Szatmari, P., Jones, M. B., Fisman, S. F., Tuff, L., Bartolucci, G., Mahoney, W. J., & Bryson, S. E. (1995). Parents and collateral relatives of children with pervasive developmental disorders: A family history study. *American Journal of Medical Genetics*, 60(4), 282–289. doi:10.1002/ajmg.1320600405 PMID:7485262

Szatmari, P., MacLean, J. E., Jones, M. B., Bryson, S. E., Zwaigenbaum, L., Bartolucci, G., ... Tuff, L. (2000). The familial aggretion of the lesser varient in biological and non-biological relatives of PDD probands: A family history study. *Journal of Child Psychology and Psychiatry, and Allied Disciplines*, *41*(5), 579–586. doi:10.1111/1469-7610.00644 PMID:10946750

Szewczyk, B. (2013). Zinc homeostasis and neurodegenerative disorders. Frontiers in Aging Neuroscience, 19.

Tabrizi, S. J., Cleeter, M. W. J., Xuereb, J., Taanman, J. W., Cooper, J. M., & Schapira, A. H. V. (1999). Biochemical abnormalities and excitotoxicity in Huntington's disease brain. *Annals of Neurology*, 45(1), 25–32. doi: PMID:9894873

Tagliavini, F., Pilleri, G., Bouras, C., & Constantinidis, J. (1984). The basal nucleus of Meynert in idiopathic Parkinson's disease. *Acta Neurologica Scandinavica*, 70(1), 20–28. doi:10.1111/j.1600-0404.1984.tb00798.x PMID:6475484

Tai, C. H., Wu, R. M., Lin, C. H., Pan, M. K., Chen, Y. F., Liu, H. M., ... Tseng, S.-H. (2010, November). Deep brain stimulation therapy for Parkinson's disease using frameless stereotaxy: Comparison with frame-based surgery. *European Journal of Neurology*, *17*(11), 1377–1385. doi:10.1111/j.1468-1331.2010.03035.x PMID:20443976

Tai, H. C., & Schuman, E. M. (2008). Ubiquitin, the proteasome and protein degradation in neuronal function and dysfunction. *Nature Reviews. Neuroscience*, 9(11), 826–838. doi:10.1038/nrn2499 PMID:18931696 Tait, S. W., & Green, D. R. (2010). Mitochondria and cell death: Outer membrane permeabilization and beyond. *Nature Reviews. Molecular Cell Biology*, *11*(9), 621–632. doi:10.1038/nrm2952 PMID:20683470

Takalo, M., Salminen, A., Soininen, H., Hiltunen, M., & Haapasalo, A. (2013). Protein aggregation and degradation mechanisms in neurodegenerative diseases. *American Journal of Neurodegenerative Disease*, 2(1), 1. PMID:23516262

Takamine, K., Okamoto, K., Fujita, Y., Sakurai, A., Takatama, M., & Gonatas, N. K. (2000). The involvement of the neuronal Golgi apparatus and trans-Golgi network in the human olivary hypertrophy. *Journal of the Neurological Sciences*, *182*(1), 45–50. doi:10.1016/S0022-510X(00)00447-0 PMID:11102638

Takeda, A. (2000). Movement of zinc and its functional significance in the brain. *Brain Research. Brain Research Reviews*, *34*(3), 137–148. doi:10.1016/S0165-0173(00)00044-8 PMID:11113504

Takeuchi, H., Kobayashi, Y., Ishigaki, S., Doyu, M., & Sobue, G. (2002). Mitochondrial localization of mutant superoxide dismutase 1 triggers caspase-dependent cell death in a cellular model of familial amyotrophic lateral sclerosis. *The Journal of Biological Chemistry*, 277(52), 50966–50972. doi:10.1074/jbc.M209356200 PMID:12393885

Tamagno, E., Guglielmotto, M., Aragno, M., Borghi, R., Autelli, R., Giliberto, L., ... Tabaton, M. (2008). Oxidative stress activates a positive feedback between the γ - and β -secretase cleavages of the β -amyloid precursor protein. *Journal of Neurochemistry*, *104*(3), 683–695. PMID:18005001

Tamano, H., & Takeda, A. (2011). Dynamic action of neurometals at the synapse. *Metallomics*, 3(7), 656–661. doi:10.1039/ c1mt00008j PMID:21409223

Tam, C., Burton, E., McKeith, I., Burn, D., & O'brien, J. (2005). Temporal lobe atrophy on MRI in Parkinson disease with dementia A comparison with Alzheimer disease and dementia with Lewy bodies. *Neurology*, *64*(5), 861–865. doi:10.1212/01.WNL.0000153070.82309.D4 PMID:15753423

Tanaka, M. (2002). Mitochondrial genotypes and cytochrome b variants associated with longevity or Parkinson's disease. *Journal of Neurology*, 249. PMID:12375058

Tan, E. K. (2003). Dopamine agonists and their role in Parkinson's disease treatment. *Expert Review of Neurotherapeutics*, *3*(6), 805–810. doi:10.1586/14737175.3.6.805 PMID:19810883

Tang, G., Perng, M. D., Wilk, S., Quinlan, R., & Goldman, J. E. (2010). Oligomers of Mutant Glial Fibrillary Acidic Protein (GFAP) Inhibit the Proteasome System in Alexander Disease Astrocytes, and the Small Heat Shock Protein αB-Crystallin Reverses the Inhibition. *The Journal of Biological Chemistry*, 285(14), 10527–10537. doi:10.1074/jbc. M109.067975 PMID:20110364

Tang, Y.-P., & Gershon, E. S. (2003). Genetic studies in Alzheimer's disease. *Dialogues in Clinical Neuroscience*, 5(1), 17–26. PMID:22033785

Tang, Y.-P., & Gershon, E. S. (2003). Genetic studies in Alzheimer's disease. *Journal of Clinical Neuroscience*, 5, 17–26. PMID:22033785

Tanner, C. M. (1989). The role of environmental toxins in the etiology of Parkinson's disease. *Trends in Neurosciences*, *12*(2), 49–54. doi:10.1016/0166-2236(89)90135-5 PMID:2469210

Tanner, C. M. (2003). Is the cause of Parkinson's disease environmental or hereditary? Evidence from twin studies. *Advances in Neurology*, *91*, 133. PMID:12442672

Tanner, C. M., Ottman, R., & Goldman, S. M. (1999). Parkinson disease in twins: An etiologic study. *Journal of the American Medical Association*, 281(4), 341–346. doi:10.1001/jama.281.4.341 PMID:9929087

Tan, W., Pasinelli, P., & Trotti, D. (2014). Role of mitochondria in mutant SOD1 linked amyotrophic lateral sclerosis. *Biochimica et Biophysica Acta (BBA)-. Molecular Basis of Disease*, 1842(8), 1295–1301. doi:10.1016/j.bbadis.2014.02.009 PMID:24568860

Tasakis, R. N. S., & Tsolaki, M. (2015). Mitochondria; pathogenesis and dysfunction in Alzheimer's disease. *Hellenic Journal of Nuclear Medicine*, *18*, 10.

Tasker, R. R., & Kiss, Z. H. (1995, January). The role of the thalamus in functional neurosurgery. *Neurosurgery Clinics of North America*, 6(1), 73–104. PMID:7696876

Tatemichi, T. K., Desmond, D. W., & Prohovnik, I. (1995). Strategic infarcts in vascular dementia. A clinical and brain imaging experience. *Arzneimittel-Forschung*, *45*(3A), 371–385. PMID:7763329

Tatton, N. A., & Kish, S. J. (1997). In situ detection of apoptotic nuclei in the substantia nigra compacta of 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine-treated mice using terminal deoxynucleotidyl transferase labelling and acridine orange staining. *Neuroscience*, 77(4), 1037–1048. doi:10.1016/S0306-4522(96)00545-3 PMID:9130785

Taylor, R. W., & Turnbull, D. M. (2005). Mitochondrial DNA mutations in human disease. *Nature Reviews. Genetics*, *6*(5), 389–402. doi:10.1038/nrg1606 PMID:15861210

Taymans, J. M., Vancraenenbroeck, R., Ollikainen, P., Beilina, A., Lobbestael, E., De Maeyer, M., ... Cookson, M. R. (2011). LRRK2 kinase activity is dependent on LRRK2 GTP binding capacity but independent of LRRK2 GTP binding. *PLoS One*, *6*(8), e23207. doi:10.1371/journal.pone.0023207 PMID:21858031

Teive, H. A., Germiniani, F. M., Della Coletta, M. V., & Werneck, L. C. (2001, September). (2001-a). Tics and Tourette syndrome: Clinical evaluation of 44 cases. *Arquivos de Neuro-Psiquiatria*, *59*(3B), 725–728. doi:10.1590/S0004-282X2001000500014 PMID:11593273

Teive, H. A., Zavala, J. A., Iwamoto, F. M., Sá, D., Carraro, H. Jr, & Werneck, L. C. (2001, September). (2001 –b). As contribuições de Charcot e de Marsden para o desenvolvimento dos distúrbios do movimento nos séculos XIX e XX. *Arquivos de Neuro-Psiquiatria*, *59*(3-A), 633–636. doi:10.1590/S0004-282X2001000400031 PMID:11588652

Teixeira, M. J., & Fonoff, E. T. (2004). Tratamento cirúrgico da Doença de Parkinson. Rev Med (São Paulo), 83(1-2), 1–16.

Temel, Y., Ackermans, L., Celik, H., Spincemaille, G. H., van der Linden, C., Walenkamp, G. H., & ... (2004). Management of hardware infections following deep brain stimulation. *Acta Neurochirurgica*, *146*(4), 355–361. doi:10.100700701-004-0219-2 PMID:15057529

Temme, C., Weissbach, R., Lilie, H., Wilson, C., Meinhart, A., Meyer, S., ... Wahle, E. (2009). The Drosophila melanogaster gene cg4930 encodes a high affinity inhibitor for endonuclease G. *The Journal of Biological Chemistry*, 284(13), 8337–8348. doi:10.1074/jbc.M808319200 PMID:19129189

Termsarasab, P., Thammongkolchai, T., & Frucht, S. J. (2016). Medical treatment of dystonia. *Journal of Clinical Movement Disorders*, *3*(1), 19. doi:10.118640734-016-0047-6 PMID:28031858

Terribilli, D., Schaufelberger, M. S., Duran, F. L. S., Zanetti, M. V., Curiati, P. K., Menezes, P. R., ... Busatto, G. F. (2011). Age-related gray matter volume changes in the brain during non-elderly adulthood. *Neurobiology of Aging*, *32*(2-6), 354–368. doi:10.1016/j.neurobiolaging.2009.02.008 PMID:19282066

Tezel, G. (2006). Oxidative stress in glaucomatous neurodegeneration: Mechanisms and consequences. *Progress in Retinal and Eye Research*, 25(5), 490–513. doi:10.1016/j.preteyeres.2006.07.003 PMID:16962364

Tezenas, D., Montcel, S., Mendizibal, H., Ayme, S., Levy, A., & Philip, N. (1996). Prevalence of 22q11 microdeletion. *Journal of Medical Genetics*, *33*, 719.

Thal, D. R., Walter, J., Saido, T. C., & Fändrich, M. (2015). Neuropathology and biochemistry of Abeta and its aggregates in Alzheimer's disease. *Acta Neuropathologica*, *129*(2), 167–182. doi:10.100700401-014-1375-y PMID:25534025

The Huntington's Disease Collaborative Research Group. (1993). A novel gene containing a trinucleotide repeat that is expanded and unstable on Huntington's disease chromosomes. *Cell*, 72(6), 971-983.

Thenganatt, M. A., & Jankovic, J. (2016). Recent Advances in Understanding and Managing Tourette Syndrome. *F1000 Research*, *5*, F1000. PMID:26918185

Thenganatt, M. A., & Louis, E. D. (2012). Distinguishing essential tremor from Parkinson's disease: Bedside tests and laboratory evaluations. *Expert Review of Neurotherapeutics*, *12*(6), 687–696. doi:10.1586/ern.12.49 PMID:22650171

Theuns, J., & Van Broeckhoven, C. (2000). Transcriptional regulation of Alzheimer's disease genes: Implications for susceptibility. *Human Molecular Genetics*, *9*(16), 2383–2394. doi:10.1093/hmg/9.16.2383 PMID:11005793

Thevathasan, W., Mazzone, P., Jha, A., Djamshidian, A., Dileone, M., Di Lazzaro, V., & Brown, P. (2010). Spinal cord stimulation failed to relieve akinesia or restore locomotion in Parkinson disease. *Neurology*, *74*(16), 1325–1327. doi:10.1212/WNL.0b013e3181d9ed58 PMID:20404313

Thrower, J. S., Hoffman, L., Rechsteiner, M., & Pickart, C. M. (2000). Recognition of the polyubiquitin proteolytic signal. *The EMBO Journal*, *19*(1), 94–102. doi:10.1093/emboj/19.1.94 PMID:10619848

Tian, R., Wu, X., Hagemann, T. L., Sosunov, A. A., Messing, A., McKhann, G. M., & Goldman, J. E. (2010). Alexander disease mutant GFAP compromises glutamate transport in astrocytes. *Journal of Neuropathology and Experimental Neurology*, *69*(4), 335–345. doi:10.1097/NEN.0b013e3181d3cb52 PMID:20448479

Tieu, K., Perier, C., Caspersen, C., Teismann, P., Wu, D. C., Yan, S. D., ... Przedborski, S. (2003). D-β-Hydroxybutyrate rescues mitochondrial respiration and mitigates features of Parkinson disease. *The Journal of Clinical Investigation*, *112*(6), 892–901. doi:10.1172/JCI200318797 PMID:12975474

Tintner, R., & Jankovic, J. (2003). Dopamine agonists in Parkinson's disease. *Expert Opinion on Investigational Drugs*, *12*(11), 1803–1820. doi:10.1517/13543784.12.11.1803 PMID:14585056

Toescu, E. C., & Verkhratsky, A. (2007). The importance of being subtle: Small changes in calcium homeostasis control cognitive decline in normal aging. *Aging Cell*, 6(3), 267–273. doi:10.1111/j.1474-9726.2007.00296.x PMID:17517038

Toledo, J. B., Arnold, S. E., Raible, K., Brettschneider, J., Xie, S. X., Grossman, M., ... Trojanowski, J. Q. (2013). Contribution of cerebrovascular disease in autopsy confirmed neurodegenerative disease cases in the National Alzheimer's Coordinating centre. *Brain*, *136*(Pt 9), 2697–2706. doi:10.1093/brain/awt188 PMID:23842566

Tong, Y., Giaime, E., Yamaguchi, H., Ichimura, T., Liu, Y., Si, H., ... Shen, J. (2012). Loss of leucine-rich repeat kinase 2 causes age-dependent bi-phasic alterations of the autophagy pathway. *Molecular Neurodegeneration*, 7(1), 2. doi:10.1186/1750-1326-7-2 PMID:22230652

Toren, P., Eldar, S., Sela, B. A., Wolmer, L., Weitz, R., Inbar, D., ... Laor, N. (1996). Zinc deficiency in attention-deficit hyperactivity disorder. *Biological Psychiatry*, 40(12), 1308–1310. doi:10.1016/S0006-3223(96)00310-1 PMID:8959299

Tóth, K. (2011). Zinc in neurotransmission. *Annual Review of Nutrition*, 21(1), 139–153. doi:10.1146/annurev-nu-tr-072610-145218 PMID:21548772

Tran, H. T., Chung, C. H., Iba, M., Zhang, B., Trojanowski, J. Q., Luk, K. C., & Lee, V. M. Y. (2014). A-synuclein immunotherapy blocks uptake and templated propagation of misfolded α-synuclein and neurodegeneration. *Cell Reports*, 7(6), 2054–2065. doi:10.1016/j.celrep.2014.05.033 PMID:24931606

566

Traynor, B. J., Alexander, M., Corr, B., Frost, E., & Hardiman, O. (2003). An outcome study of riluzole in amyotrophic lateral sclerosis-a population-based study in Ireland, 1996–2000. *Journal of Neurology*, *250*(4), 473–479. doi:10.100700415-003-1026-z PMID:12700914

Traynor, B. J., Codd, M. B., Corr, B., Forde, C., Frost, E., & Hardiman, O. (1999). Incidence and prevalence of ALS in Ireland, 1995-1997: A population-based study. *Neurology*, *52*(3), 504–509. doi:10.1212/WNL.52.3.504 PMID:10025778

Traynor, B. J., Codd, M. B., Corr, B., Forde, C., Frost, E., & Hardiman, O. (2000). Amyotrophic lateral sclerosis mimic syndromes: A population-based study. *Archives of Neurology*, *57*(1), 109–113. doi:10.1001/archneur.57.1.109 PMID:10634456

Tretiakoff, C. (1919). Contribution à l'étude de l'anatomie du locus niger de Soemmering avec quelques déductions relatives à la pathogenie des troubles du tonus musculaire et de la maladie de Parkinson. Paris: Thèse de.

Treuheit, M. J., Kosky, A. A., & Brems, D. N. (2002). Inverse relationship of protein concentration and aggregation. *Pharmaceutical Research*, *19*(4), 511–516. doi:10.1023/A:1015108115452 PMID:12033388

Trevelyan, A. J., Kirby, D. M., Smulders-Srinivasan, T. K., Nooteboom, M., Acin-Perez, R., Enriquez, J. A., ... Turnbull, D. M. (2010). Mitochondrial DNA mutations affect calcium handling in differentiated neurons. *Brain*, *133*(3), 787–796. doi:10.1093/brain/awq023 PMID:20207702

Trevitt, C. R., & Singh, P. N. (2003). Variant Creutzfeldt-Jakob disease : Pathology epidemiology, and public health implications 1–4. *The American Journal of Clinical Nutrition*, 1986(3), 651–656. doi:10.1093/ajcn/78.3.651S PMID:12600856

Troiano, A. R., Teive, H. A., Fabiani, G. B., Zavala, J. A., Sã, D. S., Ferminiani, F. M., ... Werneck, L. C. (2004). Clinical response to long action propranolol in 40 patients diagnosed with essential tremor with no previous treatment: An open, non-controlled study. *Arquivos de Neuro-Psiquiatria*, *62*(1), 86–90. doi:10.1590/S0004-282X2004000100015 PMID:15122439

Troncoso, J. C., Zonderman, A. B., Resnick, S. M., Crain, B., Pletnikova, O., & O'Brien, R. J. (2008). Effect of infarcts on dementia in the Baltimore longitudinal study of aging. *Annals of Neurology*, *64*(2), 168–176. doi:10.1002/ana.21413 PMID:18496870

Trottier, Y., Biancalana, V., & Mandel, J. L. (1994). Instability of CAG repeats in Huntington's disease: Relation to parental transmission and age of onset. *Journal of Medical Genetics*, *31*(5), 377–382. doi:10.1136/jmg.31.5.377 PMID:8064815

Truban, D., Hou, X., Caulfield, T. R., Fiesel, F. C., & Springer, W. (2016). PINK1, Parkin, and Mitochondrial Quality Control: What can we Learn about Parkinson's Disease Pathobiology? *Journal of Parkinson's Disease*, 7(1), 13–29. doi:10.3233/JPD-160989 PMID:27911343

Truong-Tran, A. Q., Ho, L. H., Chai, F., & Zalewski, P. D. (2000). Zinc and health: Current Status and future directions. *The Journal of Nutrition*, *130*, 1459S–1466S. doi:10.1093/jn/130.5.1459S PMID:10801960

Tsai, R. M., & Boxer, A. L. (2014). Clinical Trials: Past, current and future for atypical parkinsonian syndromes. *Seminars in Neurology*, *34*(2), 225–234. doi:10.1055-0034-1381739 PMID:24963682

Tsuboi, Y., & Dickson, D. W. (2005). Dementia with Lewy bodies and Parkinson's disease with dementia: Are they different? *Parkinsonism & Related Disorders*, *11*, S47–S51. doi:10.1016/j.parkreldis.2004.10.014 PMID:15885629

Tsuboi, Y., Uchikado, H., & Dickson, D. W. (2007). Neuropathology of Parkinson's disease dementia and dementia with Lewy bodies with reference to striatal pathology. *Parkinsonism & Related Disorders*, *13*, S221–S224. doi:10.1016/S1353-8020(08)70005-1 PMID:18267239

Tsujimoto, Y. (2000). Mitochondria and cell death. Cell Death and Differentiation, 7(1), 134-135. doi:10.1038j.cdd.4400645

Tsuji, S. (2007). 4 - Leukodystrophies. In S. Gilman (Ed.), *Neurobiology of Disease* (pp. 43–49). Burlington: Academic Press. doi:10.1016/B978-012088592-3/50006-2

Turnbull, J., DePaoli-Roach, A. A., Zhao, X., Cortez, M. A., Pencea, N., Tiberia, E., ... Minassian, B. A. (2011). PTG depletion removes lafora bodies and rescues the fatal epilepsy of lafora disease. *PLOS Genetics*, 7(4), e1002037. doi:10.1371/journal.pgen.1002037 PMID:21552327

Turner, M. R., Bowser, R., Bruijn, L., Dupuis, L., Ludolph, A., McGrath, M., . . . Pullman, S. L. (2013). Mechanisms, models and biomarkers in amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration, 14*(sup1), 19-32.

Turner, B. J., & Atkin, J. D. (2006). ER stress and UPR in familial amyotrophic lateral sclerosis. *Current Molecular Medicine*, *6*(1), 79–86. doi:10.2174/156652406775574550 PMID:16472115

Tutar, L., & Tutar, Y. (2010). Heat shock proteins; an overview. *Current Pharmaceutical Biotechnology*, *11*(2), 216–222. doi:10.2174/138920110790909632 PMID:20170474

Tyas, S. L., Manfreda, J., Strain, L. A., & Montgomery, P. R. (2001). Risk factors for Alzheimer's disease: A populationbased, longitudinal study in Manitoba, Canada. *International Journal of Epidemiology*, *30*(3), 590–597. doi:10.1093/ ije/30.3.590 PMID:11416089

Tyler, M., Danilov, Y., Bach-y-Rita, P., & Bach-y-Rita, J. (2007). *Systems and methods for altering brain and body functions and for treating conditions and diseases of the same*. Google Patents.

Tyler, S. J., Dawbarn, D., Wilcock, G. K., & Allen, S. J. (2002). alpha- and beta-secretase: Profound changes in Alzheimer's disease. *Biochemical and Biophysical Research Communications*, 299(3), 373–376. doi:10.1016/S0006-291X(02)02635-9 PMID:12445809

Ubhi, K., Low, P., & Masliah, E. (2011). Multiple System Atrophy: A Clinical and Neuropathological Perspective. *Trends in Neurosciences*, *34*(11), 581–590. doi:10.1016/j.tins.2011.08.003 PMID:21962754

Ungerstedt, U. (1968). 6-Hydroxy-dopamine induced degeneration of central monoamine neurons. *European Journal of Pharmacology*, 5(1), 107–110. doi:10.1016/0014-2999(68)90164-7 PMID:5718510

Ungvari, Z., Orosz, Z., Labinskyy, N., Rivera, A., Xiangmin, Z., Smith, K., & Csiszar, A. (2007). Increased mitochondrial H 2 O 2 production promotes endothelial NF-κB activation in aged rat arteries. *American Journal of Physiology. Heart and Circulatory Physiology*, 293(1), H37–H47. doi:10.1152/ajpheart.01346.2006 PMID:17416599

Uttara, B., Singh, A. V., Zamboni, P., & Mahajan, R. T. (2009). Oxidative stress and neurodegenerative diseases: A review of upstream and downstream antioxidant therapeutic options. *Current Neuropharmacology*, 7(1), 65–74. doi:10.2174/157015909787602823 PMID:19721819

Vajda, F. J. (2002). Neuroprotection and neurodegenerative disease. *Journal of Clinical Neuroscience*, 9(1), 4–8. doi:10.1054/jocn.2001.1027 PMID:11749009

Valastyan, J. S., & Lindquist, S. (2014). Mechanisms of protein-folding diseases at a glance. *Disease Models & Mechanisms*, 7(1), 9–14. doi:10.1242/dmm.013474 PMID:24396149

Valencia, A., Sapp, E., Kimm, J. S., McClory, H., Reeves, P. B., Alexander, J., ... Kegel, K. B. (2012). Elevated NADPH oxidase activity contributes to oxidative stress and cell death in Huntington's disease. *Human Molecular Genetics*, 22(6), 1112–1131. doi:10.1093/hmg/dds516 PMID:23223017

Valera, E., Compagnoni, G. M., & Masliah, E. (2016, February). (2016-a). Novel treatment strategies targeting alphasynuclein in multiple system atrophy as a model of synucleinopathy. *Neuropathology and Applied Neurobiology*, 42(1), 95–106. doi:10.1111/nan.12312 PMID:26924723

Valera, E., Spencer, B., & Masliah, E. (2016, January). (2016-b). Immunotherapeutic Approaches Targeting Amyloid- β , α -Synuclein, and Tau for the Treatment of Neurodegenerative Disorders. *Neurotherapeutics; the Journal of the American Society for Experimental NeuroTherapeutics, 13*(1), 179–189. doi:10.100713311-015-0397-z PMID:26494242

Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T., Mazur, M., & Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The International Journal of Biochemistry & Cell Biology*, *39*(1), 44–84. doi:10.1016/j.biocel.2006.07.001 PMID:16978905

Valla, J., Schneider, L., Niedzielko, T., Coon, K. D., Caselli, R., Sabbagh, M. N., ... Reiman, E. M. (2006). Impaired platelet mitochondrial activity in Alzheimer's disease and mild cognitive impairment. *Mitochondrion*, *6*(6), 323–330. doi:10.1016/j.mito.2006.10.004 PMID:17123871

Van Asch, C. J., Luitse, M. J., Rinkel, G. J., van der Tweel, I., Algra, A., & Klijn, C. J. (2010). Incidence, case fatality, and functional outcome of intracerebral haemorrhage over time, according to age, sex, and ethnic origin: A systematic review and meta-analysis. *Lancet Neurology*, *9*(2), 167–176. doi:10.1016/S1474-4422(09)70340-0 PMID:20056489

Van Deerlin, V. M., Leverenz, J. B., Bekris, L. M., Bird, T. D., Yuan, W., Elman, L. B., ... Yu, C. E. (2008). TARDBP mutations in amyotrophic lateral sclerosis with TDP-43 neuropathology: A genetic and histopathological analysis. *Lancet Neurology*, 7(5), 409–416. doi:10.1016/S1474-4422(08)70071-1 PMID:18396105

Van Dellen, A., Grote, H. E., & Hannan, A. J. (2005). Gene–environment interactions, neuronal dysfunction and pathological plasticity in Huntington's disease. *Clinical and Experimental Pharmacology & Physiology*, *32*(12), 1007–1019. doi:10.1111/j.1440-1681.2005.04313.x PMID:16445565

Van Den Eeden, S. K., Tanner, C. M., Bernstein, A. L., Fross, R. D., Leimpeter, A., Bloch, D. A., & Nelson, L. M. (2003). Incidence of Parkinson's disease: Variation by age, gender, and race/ethnicity. *American Journal of Epidemiology*, *157*(11), 1015–1022. doi:10.1093/aje/kwg068 PMID:12777365

van der Knaap, M. S., & Valk, J. (2005). X-linked adrenoleukodystrophy. In U. Heilmann (Ed.), *Magnetic Resonance of Myelination and Myelin Disorders* (3rd ed.; pp. 176–190). Berlin: Springer. doi:10.1007/3-540-27660-2_21

van der Knaap, M.S., Naidu, S., & Breiter, S. N., Blaser, S., Stroink, H., Springer, S., ... Powers, J.M. (2001). Alexander disease: Diagnosis with MR imaging. *Ameican Journal of Neuroradiology*., *22*, 541–552. PMID:11237983

Van Dooren, T., Princen, K., De Witte, K., & Griffioen, G. (2014). Derailed Intraneuronal Signalling Drives Pathogenesis in Sporadic and Familial Alzheimer's Disease. *BioMed Research International*. Retrieved September 5, 2017, from https://www.hindawi.com/journals/bmri/2014/167024/

Van Egmond, W. N., & van Haastert, P. J. (2010). Characterization of the Roco protein family in Dictyostelium discoideum. *Eukaryotic Cell*, 9(5), 751–761. doi:10.1128/EC.00366-09 PMID:20348387

Van Everbroeck, B., Dobbeleir, I., De Waele, M., De Deyn, P., Martin, J.-J., & Cras, P. (2004). Differential diagnosis of 201 possible Creutzfeldt-Jakob disease patients. *Journal of Neurology*, 251(3), 298–304. doi:10.100700415-004-0311-9 PMID:15015009

Van Houten, B., Woshner, V., & Santos, J. H. (2006). Role of mitochondrial DNA in toxic responses to oxidative stress. *DNA Repair*, 5(2), 145–152. doi:10.1016/j.dnarep.2005.03.002 PMID:15878696 Van Paesschen, W., Revesz, T., Duncan, J. S., King, M. D., & Connelly, A. (1997). Quantitative neuropathology and quantitative magnetic resonance imaging for the hippocampus in temporal lobe epilepsy. *Annals of Neurology*, *42*(5), 756–766. doi:10.1002/ana.410420512 PMID:9392575

Van Raamsdonk, J. M., Vega, I. E., & Brundin, P. (2017). Oxidative stress in neurodegenerative disease: Causation or association? *Oncotarget*, 8(7), 10777–10778. doi:10.18632/oncotarget.14650 PMID:28099897

Van Veluw, S. J., Shih, A. Y., Smith, E. E., Chen, C., Schneider, J. A., Wardlaw, J. M., ... Biessels, G. J. (2017). Detection, risk factors, and functional consequences of cerebral microinfarcts. *Lancet Neurology*, *16*(9), 730–740. doi:10.1016/S1474-4422(17)30196-5 PMID:28716371

Vanacore, N., Bonifati, V., Fabbrini, G., Colosimo, C., Marconi, R., Nicholl, D., ... Meco, G. (2000). Smoking habits in multiple system atrophy and progressive supranuclear palsy. European Study Group on Atypical Parkinsonisms. *Neurology*, *54*(1), 114–119. doi:10.1212/WNL.54.1.114 PMID:10636135

Vance, C., Rogelj, B., Hortobágyi, T., De Vos, K. J., Nishimura, A. L., Sreedharan, J., & Ganesalingam, J. (2009). Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science*, *323*(5918), 1208–1211. doi:10.1126cience.1165942 PMID:19251628

Van-Kooten, I. A., Palmen, S. J., von Cappeln, P., Steinbusch, H. W., Korr, H., Heinsen, H., ... Schmitz, C. (2008). Neurons in the fusiform gyrus are fewer and smaller in autism. *Brain*, *131*(Pt 4), 987–999. doi:10.1093/brain/awn033 PMID:18332073

Vargas, D. L., Nascimbene, C., Krishnan, C., Zimmerman, A. W., & Pardo, C. A. (2005). Neuroglial activation and neuroinflammation in the brain of patients with autism. *Annals of Neurology*, *57*(1), 304. doi:10.1002/ana.20315 PMID:15546155

Vasaikar, S. V., Padhi, A. K., Jayaram, B., & Gomes, J. (2013). NeuroDNet - an open source platform for constructing and analyzing neurodegenerative disease networks. *BMC Neuroscience*, *14*(1), 3. doi:10.1186/1471-2202-14-3 PMID:23286825

Vassal, F., Coste, J., Derost, P., Mendes, V., Gabrillargues, J., Nuti, C., ... Lemaire, J.-J. (2012). Direct stereotactic targeting of the ventrointermediate nucleus of the thalamus based on anatomic 1.5-T MRI mapping with a white matter attenuated inversion recovery (WAIR) sequence. *Brain Stimulation*, 5(4), 625–633. doi:10.1016/j.brs.2011.10.007 PMID:22405744

Vassar, R., Bennett, B. D., Babu-Khan, S., Kahn, S., Mendiaz, E. A., Denis, P., ... Citron, M. (1999). Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. *Science (New York, N.Y.)*, 286(5440), 735–41. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/10531052

Vassar, R. (2004). BACE1: The β-Secretase Enzyme in Alzheimer's Disease. *Journal of Molecular Neuroscience*, 23(1–2), 105–114. doi:10.1385/JMN:23:1-2:105 PMID:15126696

Vassar, R. (2014). BACE1 inhibitor drugs in clinical trials for Alzheimer's disease. *Alzheimer's Research & Therapy*, 6(9), 89. doi:10.118613195-014-0089-7 PMID:25621019

Ved, R., Saha, S., Westlund, B., Perier, C., Burnam, L., Sluder, A., ... Liu, L. (2005). Similar patterns of mitochondrial vulnerability and rescue induced by genetic modification of α -synuclein, parkin, and DJ-1 in Caenorhabditis elegans. *The Journal of Biological Chemistry*, 280(52), 42655–42668. doi:10.1074/jbc.M505910200 PMID:16239214

Velasco, F., Jimenez, F., Perez, M. L., Carrillo-Ruiz, J. D., Velasco, A. L., Ceballos, J., & (2001, August). Electrical stimulation of the prelemniscal radiation in the treatment of Parkinson's disease: An old target revised with new techniques. *Neurosurgery*, 49(2), 293–306. PMID:11504105

Velusamy, T., Panneerselvam, A. S., Purushottam, M., Anusuyadevi, M., Pal, P. K., Jain, S., ... Kandasamy, M. (2017). Protective Effect of Antioxidants on Neuronal Dysfunction and Plasticity in Huntington's Disease. *Oxidative Medicine and Cellular Longevity*. PMID:28168008

Vemuri, P., Lesnick, T. G., Przybelski, S. A., Knopman, D. S., Preboske, G. M., Kantarci, K., ... Jack, C. R. Jr. (2015). Vascular and amyloid pathologies are independent predictors of cognitive decline in normal elderly. *Brain*, *138*(3), 761–771. doi:10.1093/brain/awu393 PMID:25595145

Venugopal, C., Demos, C. M., Rao, K. S. J., Pappolla, M. A., & Sambamurti, K. (2008). Beta-secretase: Structure, function, and evolution. *CNS & Neurological Disorders - Drug Targets*, 7(3), 278–294. doi:10.2174/187152708784936626 PMID:18673212

Vermeer, S. E., Koudstaal, P. J., Oudkerk, M., Hofman, A., & Breteler, M. M. (2002). Prevalence and risk factors of silent brain infarcts in the population-based Rotterdam Scan Study. *Stroke*, *33*(1), 21–25. doi:10.1161/hs0102.101629 PMID:11779883

Vermeer, S. E., Longstreth, W. T. Jr, & Koudstaal, P. J. (2007). Silent brain infarcts: A systematic review. *Lancet Neurology*, *6*(7), 611–619. doi:10.1016/S1474-4422(07)70170-9 PMID:17582361

Vermeer, S. E., Prins, N. D., den Heijer, T., Hofman, A., Koudstaal, P. J., & Breteler, M. M. (2003). Silent brain infarcts and the risk of dementia and cognitive decline. *The New England Journal of Medicine*, *348*(13), 1215–1222. doi:10.1056/NEJMoa022066 PMID:12660385

Vetrugno, V., Puopolo, M., Cardone, F., Capozzoli, F., Ladogana, A., & Pocchiari, M. (2015). The future for treating Creutzfeldt–Jakob disease. *Expert Opinion on Orphan Drugs*, *3*(1), 57–74. doi:10.1517/21678707.2015.994605

Vetter, I. R., & Wittinghofer, A. (2001). The guanine nucleotide-binding switch in three dimensions. *Science*, 294(5545), 1299–1304. doi:10.1126cience.1062023 PMID:11701921

Vidakovic, A., Dragasevic, N., & Kostic, V. S. (1994). Hemiballism: Report of 25 cases. *Journal of Neurology, Neuro*surgery, and Psychiatry, 57(8), 945–949. doi:10.1136/jnnp.57.8.945 PMID:7914529

Vijay, K., Ashok, K., Sandeep, K.S., Sanjeev, K.T., Dinesh K, Ragni, S., Seema, D. (2016b). Zinc deficiency and its effect on the brain: An update. *International Journal of Genetics and Gene Therapy*. doi: 10.16966/2471-4968.105

Vijay, K., Neha, S., Tara, K., Asimul, I., Faizan, A., & Imtaiyaz, H. (2016a). Protein aggregation and neurodegenerative diseases: From theory to therapy. *European Journal of Medicinal Chemistry*, *124*, 1105–1120. doi:10.1016/j. ejmech.2016.07.054 PMID:27486076

Vijayvergiya, C., Beal, M. F., Buck, J., & Manfredi, G. (2005). Mutant superoxide dismutase 1 forms aggregates in the brain mitochondrial matrix of amyotrophic lateral sclerosis mice. *The Journal of Neuroscience*, *25*(10), 2463–2470. doi:10.1523/JNEUROSCI.4385-04.2005 PMID:15758154

Viña, J., Sastre, J., Pallardó, F., & Borrás, C. (2003). Mitochondrial theory of aging: Importance to explain why females live longer than males. *Antioxidants & Redox Signalling*, 5(5), 549–556. doi:10.1089/152308603770310194 PMID:14580309

Vitek, J. L., Bakay, R. A., & DeLong, M. R. (1997). Microelectrode-guided pallidotomy for medically intractable Parkinson's disease. *Advances in Neurology*, 74, 183–198. PMID:9348414

Vitner, E. B., Platt, F. M., & Futerman, A. H. (2010). Common and uncommon pathogenic cascades in lysosomal storage diseases. *The Journal of Biological Chemistry*, 285(27), 20423–20427. doi:10.1074/jbc.R110.134452 PMID:20430897

Vives-Bauza, C., Andreu, A. L., Manfredi, G., Beal, M. F., Janetzky, B., Gruenewald, T. H., & Lin, M. T. (2002). Sequence analysis of the entire mitochondrial genome in Parkinson's disease. *Biochemical and Biophysical Research Communications*, 290(5), 1593–1601. doi:10.1006/bbrc.2002.6388 PMID:11820805

Voisset, C., Saupe, S. J., Galons, H., & Blondel, M. (2009). Procedure for identification and characterization of drugs efficient against mammalian prion: From a yeast-based antiprion drug screening assay to in vivo mouse models. *Infectious Disorders Drug Targets*, 9(1), 31–39. doi:10.2174/1871526510909010031 PMID:19200013

Volkmar, F., Cook, E., Pomeroy, J., Realmuto, G., & Tanguay, P. (1999). the Work Group on Quality Issues, AACAP Practice parameters for the assessment and treatment of children, adolescents, and adults with autism and other pervasive developmental disorders. *Journal of the American Academy of Child and Adolescent Psychiatry*, *38*(Suppl 12), 32S–54S. doi:10.1016/S0890-8567(99)80003-3 PMID:10624084

von Jonquieres, G., Spencer, Z. H. T., & Rowlands, B. D. (2017). Uncoupling *N*-acetylaspartate from brain pathology: Implications for Canavan disease gene therapy. *Acta Neuropathologica*, *134*(627), 1–19. doi:10.100700401-017-1784-9 PMID:29116375

Vonsattel, J. P. G., & DiFiglia, M. (1998). Huntington disease. *Journal of Neuropathology and Experimental Neurology*, 57(5), 369. doi:10.1097/00005072-199805000-00001 PMID:9596408

Vonsattel, J. P., Keller, C., & Cortes Ramirez, E. P. (2011). Huntington's disease – neuropathology. *Handbook of Clinical Neurology*, *100*, 83–100. doi:10.1016/B978-0-444-52014-2.00004-5 PMID:21496571

Voon, V., Krack, P., Lang, A. E., Lozano, A. M., Dujardin, K., Schupbach, M., ... Moro, E. (2008). A multicentre study on suicide outcomes following subthalamic stimulation for Parkinson's disease. *Brain*, *131*(Pt 10), 2720–2728. doi:10.1093/brain/awn214 PMID:18941146

Voon, V., Kubu, C., Krack, P., Houeto, J. L., & Tröster, A. I. (2006). Deep brain stimulation: Neuropsychological and neuropsychiatric issues. *Movement Disorders*, 21(S14Suppl 14), S305–S327. doi:10.1002/mds.20963 PMID:16810676

Vossel, K. A., Zhang, K., Brodbeck, J., Daub, A. C., Sharma, P., Finkbeiner, S., ... Mucke, L. (2010). Tau Reduction Prevents Aβ-Induced Defects in Axonal Transport: Fig. 1. *Science*, *330*(6001), 198–198. doi:10.1126cience.1194653 PMID:20829454

Vrancic, D., Nanclares, V., Soares, D., Kulesz, A., Mordzinski, C., Plebst, C., & Starkstein, S. (2002). Sensitivity and specificity of the Autism Diagnostic Inventory-telephone screening in Spanish. *Journal of Autism and Developmental Disorders*, *32*(4), 313–320. doi:10.1023/A:1016335003256 PMID:12199136

Vucic, S., Burke, D., & Kiernan, M. C. (2007). Diagnosis of motor neuron disease. In M. C. Kiernan (Ed.), *The Motor Neuron Disease Handbook* (pp. 89–115). Sydney: Australasian Medical Publishing Company Limited.

Vucic, S., & Kiernan, M. C. (2009). Pathophysiology of degeneration in familial amyotrophic lateral sclerosis. *Current Molecular Medicine*, *9*(3), 255–272. doi:10.2174/156652409787847173 PMID:19355908

Vyas, S., Zaganjor, E., & Haigis, M. C. (2016). Mitochondria and cancer. *Cell*, *166*(3), 555–566. doi:10.1016/j. cell.2016.07.002 PMID:27471965

Waak, J., Weber, S. S., Görner, K., Schall, C., Ichijo, H., Stehle, T., & Kahle, P. J. (2009). Oxidizable residues mediating protein stability and cytoprotective interaction of DJ-1 with apoptosis signal-regulating kinase 1. *The Journal of Biological Chemistry*, 284(21), 14245–14257. doi:10.1074/jbc.M806902200 PMID:19293155

Wakabayashi, K., Tanji, K., Mori, F., & Takahashi, H. (2007). The Lewy body in Parkinson's disease: Molecules implicated in the formation and degradation of α -synuclein aggregates. *Neuropathology*, 27(5), 494–506. doi:10.1111/j.1440-1789.2007.00803.x PMID:18018486

Walia, V. (2017). Possible Role of Serotonin and Selective Serotonin Reuptake Inhibitors in Suicidal Ideations and Attempts. *Journal of Pharmaceutical Sciences and Pharmacology*, *3*(1), 54–70. doi:10.1166/jpsp.2017.1076

Walia, V., Sharma, A., Gahlawat, M., & Dube, O. P. (2014). Dual Role of Nitric Oxide in the Pathogenesis of Parkinson's Disease. *Journal of Pharmaceutical Sciences & Pharmacology*, *1*(4), 243–253. doi:10.1166/jpsp.2014.1038

Walker, L. C., Diamond, M. I., Duff, K. E., & Hyman, B. T. (2013). Mechanisms of protein seeding in neurodegenerative diseases. *JAMA Neurology*, 70(3), 304–310. doi:10.1001/jamaneurol.2013.1453 PMID:23599928

Walker, Z., Jaros, E., Walker, R. W., Lee, L., Costa, D. C., Livingston, G., ... Katona, C. L. E. (2007). Dementia with Lewy bodies: A comparison of clinical diagnosis, FP-CIT single photon emission computed tomography imaging and autopsy. *Journal of Neurology, Neurosurgery, and Psychiatry*, 78(11), 1176–1181. doi:10.1136/jnnp.2006.110122 PMID:17353255

Wallace, D. C. (2005). A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: A dawn for evolutionary medicine. *Annual Review of Genetics*, *39*(1), 359–407. doi:10.1146/annurev.genet.39.110304.095751 PMID:16285865

Wallace, D. C. (2010). Mitochondrial DNA mutations in disease and aging. *Environmental and Molecular Mutagenesis*, *51*(5), 440–450. PMID:20544884

Walls, K. C., Coskun, P., Gallegos-Perez, J. L., Zadourian, N., Freude, K., Rasool, S., ... LaFerla, F. M. (2012). Swedish Alzheimer mutation induces mitochondrial dysfunction mediated by HSP60 mislocalization of amyloid precursor protein (APP) and beta-amyloid. *The Journal of Biological Chemistry*, 287(36), 30317–30327. doi:10.1074/jbc.M112.365890 PMID:22753410

Walsh, D. M., Lomakin, A., Benedek, G. B., Condron, M. M., & Teplow, D. B. (1997). Amyloid β-protein fibrillogenesis detection of a protofibrillar intermediate. *The Journal of Biological Chemistry*, 272(35), 22364–22372. doi:10.1074/jbc.272.35.22364 PMID:9268388

Walsh, D. M., & Selkoe, D. J. (2004). Deciphering the molecular basis of memory failure in Alzheimer's disease. *Neuron*, 44(1), 181–193. doi:10.1016/j.neuron.2004.09.010 PMID:15450169

Wang, H., Lim, P. J., Karbowski, M., & Monteiro, M. J. (2008). Effects of overexpression of huntingtin proteins on mitochondrial integrity. *Human Molecular Genetics*, *18*(4), 737–752. doi:10.1093/hmg/ddn404 PMID:19039036

Wang, H., O'Reilly, E. J., Weisskopf, M. G., Logroscino, G., McCullough, M. L., Thun, M. J., ... Ascherio, A. (2011). Smoking and risk of amyotrophic lateral sclerosis: A pooled analysis of 5 prospective cohorts. *Archives of Neurology*, *68*(2), 207–213. doi:10.1001/archneurol.2010.367 PMID:21320987

Wang, L., Li, J., Zhao, H., Hu, J., Ping, Y., Li, F., ... Li, X. (2016). Identifying the crosstalk of dysfunctional pathways mediated by lncRNAs in breast cancer subtypes. *Molecular BioSystems*, *12*(3), 711–720. doi:10.1039/C5MB00700C PMID:26725846

Wang, R. Y., Bodamer, O. A., Watson, M. S., & Wilcox, W. R. (2011). Lysosomal storage diseases: Diagnostic confirmation and management of presymptomatic individuals. *Genetics in Medicine*, *13*(5), 457–484. doi:10.1097/GIM.0b013e318211a7e1 PMID:21502868

Wang, T., & Hay, J. C. (2015). Alpha-synuclein toxicity in the early secretory pathway: How it drives neurodegeneration in Parkinsons disease. *Frontiers in Neuroscience*, 9. PMID:26617485

Wang, W. (2005). Protein aggregation and its inhibition in biopharmaceutics. *International Journal of Pharmaceutics*, 289(1-2), 1–30. doi:10.1016/j.ijpharm.2004.11.014 PMID:15652195

Wang, W. X., Rajeev, B. W., Stromberg, A. J., Ren, N., Tang, G., Huang, Q., ... Nelson, P. T. (2008). The expression of microRNA miR-107 decreases early in Alzheimer's disease and may accelerate disease progression through regulation of β -site amyloid precursor protein-cleaving enzyme 1. *The Journal of Neuroscience*, 28(5), 1213–1223. doi:10.1523/JNEUROSCI.5065-07.2008 PMID:18234899

Wang, W., & Roberts, C. J. (2010). Aggregation of therapeutic proteins. John Wiley & Sons. doi:10.1002/9780470769829

Wang, X. F., Li, S., Chou, A. P., & Bronstein, J. M. (2006). Inhibitory effects of pesticides on proteasome activity: Implication in Parkinson's disease. *Neurobiology of Disease*, 23(1), 198–205. doi:10.1016/j.nbd.2006.02.012 PMID:16626962

Wang, X., & Michaelis, E. K. (2010). Selective neuronal vulnerability to oxidative stress in the brain. *Frontiers in Aging Neuroscience*, *2*, 12. PMID:20552050

Wan, L., Ottinger, E., Cho, S., & Dreyfuss, G. (2008). Inactivation of the SMN complex by oxidative stress. *Molecular Cell*, *31*(2), 244–254. doi:10.1016/j.molcel.2008.06.004 PMID:18657506

Ward, R. J., Zucca, F. A., Duyn, J. H., Crichton, R. R., & Zecca, L. (2014). The role of iron in brain ageing and neurodegenerative disorders. *Lancet Neurology*, *13*(10), 1045–1060. doi:10.1016/S1474-4422(14)70117-6 PMID:25231526

Waris, G., & Ahsan, H. (2006). Reactive oxygen species: Role in the development of cancer and various chronic conditions. *Journal of Carcinogenesis*. 5. doi:10.1186/1477-3163-5-14

Wąsik, A., & Antkiewicz-Michaluk, L. (2017). The mechanism of neuroprotective action of natural compounds. *Pharmacological Reports*, 69(5), 851–860. doi:10.1016/j.pharep.2017.03.018 PMID:28623709

Waters, C. (2000). Catechol-O-Methyltransferase (COMT) Inhibitors in Parkinson's Disease. *Journal of the American Geriatrics Society*, 48(6), 692–698. doi:10.1111/j.1532-5415.2000.tb04732.x PMID:10855610

Watson, R. E., McKim, J. M., Cockerell, G. L., & Goodman, J. I. (2004). The value of DNA methylation analysis in basic, initial toxicity assessments. *Toxicological Sciences*, 79(1), 178–188. doi:10.1093/toxsci/kfh099 PMID:15103049

Watt, N. T., Taylor, D. R., Gillott, A., Thomas, D. A., Perera, W. S. S., & Hooper, N. M. (2005). Reactive oxygen speciesmediated β-cleavage of the prion protein in the cellular response to oxidative stress. *The Journal of Biological Chemistry*, 280(43), 35914–35921. doi:10.1074/jbc.M507327200 PMID:16120605

Watts, R. L., Jankovic, J., Waters, C., Rajput, A., Boroojerdi, B., & Rao, J. (2007). Randomized, blind, controlled trial of transdermal rotigotine in early Parkinson disease. *Neurology*, *68*(4), 272–276. doi:10.1212/01.wnl.0000252355.79284.22 PMID:17202432

Webber, P. J., Smith, A. D., Sen, S., Renfrow, M. B., Mobley, J. A., & West, A. B. (2011). Autophosphorylation in the leucine-rich repeat kinase 2 (LRRK2) GTPase domain modifies kinase and GTP-binding activities. *Journal of Molecular Biology*, *412*(1), 94–110. doi:10.1016/j.jmb.2011.07.033 PMID:21806997

Weber, W., & Newmark, S. (2007). Complementary and alternative medical therapies for attention-deficit/hyperactivity disorder and autism. *Pediatric Clinics of North America*, 54(6), 983–1006. doi:10.1016/j.pcl.2007.09.006 PMID:18061787

Wei, M. C., Zong, W. X., Cheng, E. H. Y., Lindsten, T., Panoutsakopoulou, V., Ross, A. J., ... Korsmeyer, S. J. (2001). Proapoptotic BAX and BAK: A requisite gateway to mitochondrial dysfunction and death. *Science*, 292(5517), 727–730. doi:10.1126cience.1059108 PMID:11326099

574

Weimer, J. M., Kriscenski-Perry, E., Elshatory, Y., & Pearce, D. A. (2002). The neuronal ceroid lipofuscinoses: Mutations in different proteins result in similar disease. *Neuromolecular Medicine*, *1*(2), 111–124. doi:10.1385/NMM:1:2:111 PMID:12025857

Weisman, H., Qureshi, I. A., Leckman, J. F., Scahill, L., & Bloch, M. H. (2013). Systematic Review: Pharmacological Treatment of Tic Disorders – Efficacy of Antipsychotic and Alpha-2 Adrenergic Agonist Agents. *Neuroscience and Biobehavioral Reviews*, *37*(6), 1162–1171. doi:10.1016/j.neubiorev.2012.09.008 PMID:23099282

Weiss, J. H., Hartley, D. M., Koh, J. Y., & Choi, D. W. (1993). AMPA receptor activation potentiates zinc neurotoxicity. *Neuron*, *10*(1), 43–49. doi:10.1016/0896-6273(93)90240-R PMID:7678965

Weiss, J. H., Sensi, S. L., & Koh, J. Y. (2000). Zn²⁺: A novel ionic mediator of neural injury in brain disease. *Trends in Pharmacological Sciences*, *21*(10), 395–401. doi:10.1016/S0165-6147(00)01541-8 PMID:11050320

Weleber, R. G. (1998). The dystrophic retina in multisystem disorders: The electroretinogram in neuronal ceroid lipofuscinoses. *Eye (London, England)*, *12*(3), 580–590. doi:10.1038/eye.1998.148 PMID:9775220

Wenger, D. A. (2000 Jun 19). Krabbe Dsease. In M. P. Adam, H. H. Ardinger, & R. A. Pagon (Eds.), *GeneReviews* (pp. 1993–2017). Seattle, WA: University of Washington.

Wenning, G. K., Ben Shlomo, Y., Magalhaes, M., Daniel, S. E., & Quinn, N. P. (1994). Clinical features and natural history of multiple system atrophy. An analysis of 100 cases. *Brain*, *117*(Pt4), 835–845. doi:10.1093/brain/117.4.835 PMID:7922469

Wenning, G. K., Seppi, K., Scherfler, C., Stefanova, N., & Puschban, Z. (2001). Multiple system atrophy. *Seminars in Neurology*, *21*(01), 33–40. doi:10.1055-2001-13117 PMID:11346023

Wen, W., & Sachdev, P. (2004). The topography of white matter hyperintensities on brain MRI in healthy 60-to 64-yearold individuals. *NeuroImage*, 22(1), 144–154. doi:10.1016/j.neuroimage.2003.12.027 PMID:15110004

Wen, W., Sachdev, P. S., Li, J. J., Chen, X., & Anstey, K. J. (2009). White matter hyperintensities in the forties: Their prevalence and topography in an epidemiological sample aged 44-48. *Human Brain Mapping*, *30*(9), 1155–1167. doi:10.1002/hbm.20586 PMID:18465744

Wesnes, K. A., McKeith, I., Edgar, C., Emre, M., & Lane, R. (2005). Benefits of rivastigmine on attention in dementia associated with Parkinson disease. *Neurology*, 65(10), 1654–1656. doi:10.1212/01.wnl.0000184517.69816.e9PMID:16301500

West, A. B., Dawson, V. L., & Dawson, T. M. (2007). The role of Parkin in Parkinson's disease. *Neurological Disease* & *Therapy*, 83, 199.

West, A. B., Moore, D. J., Biskup, S., Bugayenko, A., Smith, W. W., Ross, C. A., ... Dawson, T. M. (2005). Parkinson's disease-associated mutations in leucine-rich repeat kinase 2 augment kinase activity. *Proceedings of the National Academy of Sciences of the United States of America*, 102(46), 16842–16847. doi:10.1073/pnas.0507360102 PMID:16269541

Westover, M. B., Bianchi, M. T., Yang, C., Schneider, J. A., & Greenberg, S. M. (2013). Estimating cerebral microinfarct burden from autopsy samples. *Neurology*, *80*(15), 1365–1369. doi:10.1212/WNL.0b013e31828c2f52 PMID:23486880

Whalley, K. (2017). Neurodegenerative disease: Probing prions. Nature Reviews. Neuroscience, 18(1), 5-5. PMID: 27974838

White, A. B., Givogri, M. I., Lopez-Rosas, A., Cao, H., Breemen, R. V., Thinakaran, G., & Bongarzone, E. R. (2009). Psychosine Accumulates in Membrane Microdomains in the Brain of Krabbe Patients, Disrupting the Raft Architecture. *The Journal of Neuroscience*, *29*(19), 6068–6077. doi:10.1523/JNEUROSCI.5597-08.2009 PMID:19439584

Whitehouse, P. J., Hedreen, J. C., White, C. L., & Price, D. L. (1983). Basal forebrain neurons in the dementia of Parkinson disease. *Annals of Neurology*, *13*(3), 243–248. doi:10.1002/ana.410130304 PMID:6847136

Whitney, E. R., Kemper, T. L., Bauman, M. L., Rosene, D. L., & Blatt, G. J. (2008). Cerebellar Purkinje cells are reduced in a subpopulation of autistic brains: A stereological experiment using calbindin-D28k. *Cerebellum (London, England)*, 7(3), 406–416. doi:10.100712311-008-0043-y PMID:18587625

Whitwell, J. L., Jack, C. R. Jr, Boeve, B. F., Parisi, J. E., Ahlskog, J. E., Drubach, D. A., ... Josephs, K. A. (2010). Imaging correlates of pathology in corticobasal syndrome. *Neurology*, 75(21), 1879–1887. doi:10.1212/WNL.0b013e3181feb2e8 PMID:21098403

Wicks, P., Abrahams, S., Masi, D., Hejda-Forde, S., Leigh, P. N., & Goldstein, L. H. (2007). Prevalence of depression in a 12-month consecutive sample of patients with ALS. *European Journal of Neurology*, *14*(9), 993–1001. doi:10.1111/j.1468-1331.2007.01843.x PMID:17718691

Wicks, P., Abrahams, S., Papps, B., Al-Chalabi, A., Shaw, C. E., Leigh, P. N., & Goldstein, L. H. (2009). SOD1 and cognitive dysfunction in familial amyotrophic lateral sclerosis. *Journal of Neurology*, 256(2), 234–241. doi:10.100700415-009-0078-0 PMID:19252762

Wiedemann, F. R., Manfredi, G., Mawrin, C., Beal, M. F., & Schon, E. A. (2002). Mitochondrial DNA and respiratory chain function in spinal cords of ALS patients. *Journal of Neurochemistry*, *80*(4), 616–625. doi:10.1046/j.0022-3042.2001.00731.x PMID:11841569

Wigram, T., & Gold, C. (2006). Music therapy in the assessment and treatment of autistic spectrum disorder: Clinical application and research evidence. *Child: Care, Health and Development, 32*(5), 535–542. doi:10.1111/j.1365-2214.2006.00615.x PMID:16919132

Wijesekera, L. C., & Leigh, P. N. (2009). Amyotrophic lateral sclerosis. *Orphanet Journal of Rare Diseases*, 4(1), 3. doi:10.1186/1750-1172-4-3 PMID:19192301

Wikgren, M., Maripuu, M., Karlsson, T., Nordfjäll, K., Bergdahl, J., Hultdin, J., ... Adolfsson, R. (2012). Short telomeres in depression and the general population are associated with a hypocortisolemic state. *Biological Psychiatry*, *71*(4), 294–300. doi:10.1016/j.biopsych.2011.09.015 PMID:22055018

Wilbourn, A. J. (1998). Clinical neurophysiology in the diagnosis of amyotrophic lateral sclerosis : The Lambert and the El Escorial criteria. *Journal of the Neurological Sciences*, *160*, 25–29. doi:10.1016/S0022-510X(98)00194-4 PMID:9851644

Wild, R., Pettit, T. A., & Burns, A. (2003). Cholinesterase inhibitors for dementia with Lewy bodies. *The Cochrane Library*. PMID:12917981

Wilesmith, J. W., Wells, G. A., Cranwell, M. P., & Ryan, J. B. (1988). Bovine spongiform encephalopathy: Epidemiological studies. *The Veterinary Record*, *123*(25), 638–644. PMID:3218047

Wilkie, S., van Schalkwyk, M. C., Hobbs, S., Davies, D. M., van der Stegen, S. J., Pereira, A. C., ... Maher, J. (2012). Dual targeting of ErbB2 and MUC1 in breast cancer using chimeric antigen receptors engineered to provide complementary signaling. *Journal of Clinical Immunology*, *32*(5), 1059–1070. doi:10.100710875-012-9689-9 PMID:22526592

Wilkinson, C. W., Peskind, E. R., & Raskind, M. A. (1997). Decreased hypothalamic-pituitary adrenal axis sensitivity to cortisol feedback inhibition in human aging. *Neuroendocrinology*, 65(1), 79–90. doi:10.1159/000127167 PMID:9032777

Williams, D. R., Holton, J. L., Strand, K., Revesz, T., & Lees, A. J. (2007). Pure akinesia with gait freezing: A third clinical phenotype of progressive supranuclear palsy. *Movement Disorders*, 22(15), 2235–2241. doi:10.1002/mds.21698 PMID:17712855

576

Williams-Gray, C. H., Evans, J. R., Goris, A., Foltynie, T., Ban, M., Robbins, T. W., ... Barker, R. A. (2009). The distinct cognitive syndromes of Parkinson's disease: 5 year follow-up of the CamPaIGN cohort. *Brain*, *132*(11), 2958–2969. doi:10.1093/brain/awp245 PMID:19812213

Williamson, T. L., & Cleveland, D. W. (1999). Slowing of axonal transport is a very early event in the toxicity of ALSlinked SOD1 mutants to motor neurons. *Nature Neuroscience*, 2(1), 50–56. doi:10.1038/4553 PMID:10195180

Williams, R. E., & Mole, S. E. (2012). New nomenclature and classification scheme for the neuronal ceroid lipofuscinoses. *Neurology*, *79*(2), 183–191. doi:10.1212/WNL.0b013e31825f0547 PMID:22778232

Wilson, A. C., Dugger, B. N., & Dickson, D. W. (2011). TDP-43 in aging and Alzheimer's disease - a review. *International Journal of Clinical and Experimental Pathology*, 4(2), 147–155. PMID:21326809

Wiltgen, B. J., Royle, G. A., Gray, E. E., Abdipranoto, A., Thangthaeng, N., Jacobs, N., ... Vissel, B. (2010). A role for calcium-permeable AMPA receptors in synaptic plasticity and learning. *PLoS One*, *5*(9), 12818. doi:10.1371/journal. pone.0012818 PMID:20927382

Wingo, T. S., Cutler, D. J., Yarab, N., Kelly, C. M., & Glass, J. D. (2011). The Heritability of Amyotrophic Lateral Sclerosis in a Clinically Ascertained United States Research Registry. *Public Library of Science One*, *6*(11), e27985. PMID:22132186

Winklhofer, K. F. (2007). The *Parkin* protein as a therapeutic target in Parkinson's disease. *Expert Opinion on Therapeutic Targets*, *11*(12), 1543–1552. doi:10.1517/14728222.11.12.1543 PMID:18020977

Winklhofer, K. F., & Haass, C. (2010). Mitochondrial dysfunction in Parkinson's disease. *Biochimica et Biophysica Acta*, *1802*(1), 29–44. doi:10.1016/j.bbadis.2009.08.013 PMID:19733240

Winner, B., Jappelli, R., Maji, S. K., Desplats, P. A., Boyer, L., Aigner, S., ... Tzitzilonis, C. (2011). In vivo demonstration that α-synuclein oligomers are toxic. *Proceedings of the National Academy of Sciences of the United States of America*, *108*(10), 4194–4199. doi:10.1073/pnas.1100976108 PMID:21325059

Wippold, F. J., Perry, A., & Lennerz, J. (2006, May). Neuropathology for the Neuroradiologist: Rosenthal Fibers. *AJNR*. *American Journal of Neuroradiology*, 27(5), 958–961. PMID:16687524

Wisniewski, K. E., Kaczmarski, A., Kida, E., Connell, F., Kaczmarski, W., Michalewski, M., ... Zhong, N. (2017). Reevaluation of Neuronal Ceroid Lipofuscinoses: Atypical Juvenile Onset May Be the Result of CLN2 Mutations. *Molecular Genetics and Metabolism*, 66(4), 248–252. doi:10.1006/mgme.1999.2814 PMID:10191110

Withers, G. S., George, J. M., Banker, G. A., & Clayton, D. F. (1997). Delayed localization of synelfin (synuclein, NACP) to presynaptic terminals in cultured rat hippocampal neurons. *Brain Research. Developmental Brain Research*, *99*(1), 87–94. doi:10.1016/S0165-3806(96)00210-6 PMID:9088569

Witwer, A., & Lecavalier, L. (2005). Treatment incidence and patterns in children and adolescents with autism spectrum disorders. *Journal of Child and Adolescent Psychopharmacology*, *15*(4), 671–681. doi:10.1089/cap.2005.15.671 PMID:16190798

Wolfe, M. S. (2008). Gamma-secretase: Structure, function, and modulation for Alzheimer's disease. *Current Topics in Medicinal Chemistry*, 8(1), 2–8. doi:10.2174/156802608783334024 PMID:18220927

Wolf, O. T., Convit, A., de Leon, M. J., Caraos, C., & Qadri, S. F. (2002). Basal hypothalamo-pituitary-adrenal axis activity and corticotropin feedback in young and older men: Relationships to magnetic resonance imaging-derived hip-pocampus and cingulate gyrus volumes. *Neuroendocrinology*, 75(4), 241–249. doi:10.1159/000054715 PMID:11979054

Wollenweber, F. A., Buerger, K., Mueller, C., Ertl-Wagner, B., Malik, R., Dichgans, M., ... Opherk, C. (2014). Prevalence of cortical superficial siderosis in patients with cognitive impairment. *Journal of Neurology*, 261(2), 277–282. doi:10.100700415-013-7181-y PMID:24221645

Wolozin, B., & Behl, C. (2000). Mechanisms of neurodegenerative disorders: Part 1: protein aggregates. Archives of Neurology, 57(6), 793–796. doi:10.1001/archneur.57.6.793 PMID:10867775

Wong, B. S., Brown, D. R., Pan, T., Whiteman, M., Liu, T., Bu, X., ... Rubenstein, R. (2001). Oxidative impairment in scrapie-infected mice is associated with brain metals perturbations and altered antioxidant activities. *Journal of Neurochemistry*, *79*(3), 689–698. doi:10.1046/j.1471-4159.2001.00625.x PMID:11701772

Wong, B. S., Chen, S. G., Colucci, M., Xie, Z., Pan, T., Liu, T., ... Brown, D. R. (2001). Aberrant metal binding by prion protein in human prion disease. *Journal of Neurochemistry*, 78(6), 1400–1408. doi:10.1046/j.1471-4159.2001.00522.x PMID:11579148

Wong, B.-S., Wang, H., Brown, D. R., & Jones, I. M. (1999). Selective oxidation of methionine residues in prion proteins. *Biochemical and Biophysical Research Communications*, 259(2), 352–355. doi:10.1006/bbrc.1999.0802 PMID:10362513

Wong, K., Sidransky, E., Verma, A., Mixon, T., Sandberg, G. D., Wakefield, L. K., ... Schiffmann, R. (2004). Neuropathology provides clues to the pathophysiology of Gaucher disease. *Molecular Genetics and Metabolism*, 82(3), 192–207. doi:10.1016/j.ymgme.2004.04.011 PMID:15234332

Wong, Y. C., & Krainc, D. (2017). α-synuclein toxicity in neurodegeneration: Mechanism and therapeutic strategies. *Nature Medicine*, 23(2), 1–13. doi:10.1038/nm.4269 PMID:28170377

Woodin, M., Wang, P. P., Aleman, D., McDonald-McGinn, D., Zackai, E., & Moss, E. (2001). Neuropsychological profile of children and adolescents with the 22q11.2 microdeletion. *Genetics in Medicine*, *3*(1), 34–39. doi:10.1097/00125817-200101000-00008 PMID:11339375

Wood-Kaczmar, A., Gandhi, S., & Wood, N. W. (2006). Understanding the molecular causes of Parkinson's disease. *Trends in Molecular Medicine*, *12*(11), 521–528. doi:10.1016/j.molmed.2006.09.007 PMID:17027339

Wood-Kaczmar, A., Gandhi, S., Yao, Z., Abramov, A. S., Miljan, E. A., Keen, G., ... Downward, J. (2008). PINK1 is necessary for long term survival and mitochondrial function in human dopaminergic neurons. *PLoS One*, *3*(6), 2455. doi:10.1371/journal.pone.0002455 PMID:18560593

Wood, L. B., Winslow, A. R., & Strasser, S. D. (2015). Systems biology of neurodegenerative diseases. *Integrative Biology*, 7(7), 758–775. doi:10.1039/C5IB00031A PMID:26065845

Woollacott, M., & Shumway-Cook, A. (2002). Attention and the control of posture and gait: A review of an emerging area of research. *Gait & Posture*, *16*(1), 1–14. doi:10.1016/S0966-6362(01)00156-4 PMID:12127181

Worby, C. A., Gentry, M. S., & Dixon, J. E. (2008). Malin decreases glycogen accumulation by promoting the degradation of protein targeting to glycogen (PTG). *The Journal of Biological Chemistry*, 283(7), 4069–4076. doi:10.1074/jbc. M708712200 PMID:18070875

World Health Organization. (2003). WHO manual for surveillance of human transmissible spongiform encephalopathies, including variant Creutzfeldt-Jakob disease. WHO.

World Health Organization. (2008). International Health Regulations (2005). World Health Organization.

World Health Organization. (2010). WHO Tables on tissue infectivity distribution in transmissible spongiform encephalopathies. Updated 2010. World Health Organization.

World Health Organization. (2016). Dementia Fact Sheet. WHO. Retrieved from http://www.who.int/mediacentre/factsheets/fs362/en/

Worms, P. M. (2001). The epidemiology of motor neuron diseases: A review of recent studies. *Journal of the Neurological Sciences*, *191*(1-2), 3–9. doi:10.1016/S0022-510X(01)00630-X PMID:11676986

Wright, A., & Vissel, B. (2012). The essential role of AMPA receptor GluR2sub-unit RNA editing in the normal and diseased brain. *Frontiers in Molecular Neuroscience*, *5*, 34. doi:10.3389/fnmol.2012.00034 PMID:22514516

Wszolek, Z. K., Tsuboi, Y., Ghetti, B., Pickering-Brown, S., Baba, Y., & Cheshire, W. P. (2006). Frontotemporal dementia and parkinsonism linked to chromosome 17 (FTDP-17). *Orphanet Journal of Rare Diseases*, *1*(1), 30. doi:10.1186/1750-1172-1-30 PMID:16899117

Wunderlich, V., Schütt, M., Böttger, M., & Graffi, A. (1970). Preferential alkylation of mitochondrial deoxyribonucleic acid by N-methyl-N-nitrosourea. *The Biochemical Journal*, *118*(1), 99–109. doi:10.1042/bj1180099 PMID:5472159

Wyss-Coray, T., & Mucke, L. (2002). Inflammation in neurodegenerative disease—a double-edged sword. *Neuron*, *35*(3), 419–432. doi:10.1016/S0896-6273(02)00794-8 PMID:12165466

Xiang, W., Schlachetzki, J. C., Helling, S., Bussmann, J. C., Berlinghof, M., Schäffer, T. E., ... Becker, C. M. (2013). Oxidative stress-induced posttranslational modifications of alpha-synuclein: Specific modification of alphasynuclein by 4-hydroxy-2-nonenal increases dopaminergic toxicity. *Molecular and Cellular Neurosciences*, 54, 71–83. doi:10.1016/j. mcn.2013.01.004 PMID:23369945

Xia, P., Chen, H. S., Zhang, D., & Lipton, S. A. (2010). Memantine preferentially blocks extrasynaptic over synaptic NMDA receptor currents in hippocampal autapses. *The Journal of Neuroscience*, *30*(33), 11246–11250. doi:10.1523/JNEUROSCI.2488-10.2010 PMID:20720132

Xiong, Y., Coombes, C. E., Kilaru, A., Li, X., Gitler, A. D., Bowers, W. J., ... Moore, D. J. (2010). GTPase activity plays a key role in the pathobiology of LRRK2. *PLOS Genetics*, 6(4), e1000902. doi:10.1371/journal.pgen.1000902 PMID:20386743

Xi, Z., van Blitterswijk, M., Zhang, M., McGoldrick, P., McLean, J. R., Yunusova, Y., ... Rogaeva, E. (2015). Jump from Pre-mutation to Pathologic Expansion in C9orf72. *American Journal of Human Genetics*, *96*(6), 962–970. doi:10.1016/j. ajhg.2015.04.016 PMID:26004200

Xi, Z., Zinman, L., Moreno, D., Schymick, J., Liang, Y., Sato, C., ... Rogaeva, E. (2013). Hypermethylation of the CpG island near the G4C2 repeat in ALS with a C9orf72 expansion. *American Journal of Human Genetics*, 92(6), 981–989. doi:10.1016/j.ajhg.2013.04.017 PMID:23731538

Xu, H., Gao, H.-L., Zheng, W., Xin, N., Chi, Z.-H., Bai, S.-L., & Wang, Z.-Y. (2011). Lactational zinc deficiency-induced hippocampal neuronal apoptosis by a BDNF-independent TrkB signaling pathway. *Hippocampus*, 21(5), 495–501. doi:10.1002/hipo.20767 PMID:20101602

Xu, J., Kurup, P., Zhang, Y., Goebel-Goody, S. M., Wu, P. H., Hawasli, A. H., ... Lombroso, P. J. (2009). Extrasynaptic NMDA receptors couple preferentially to excitotoxicity via calpain-mediated cleavage of STEP. *The Journal of Neuroscience*, *29*(29), 9330–9343. doi:10.1523/JNEUROSCI.2212-09.2009 PMID:19625523

Yabe, I., Soma, H., Takei, A., Fujiki, N., Yanagihara, H., & Sasaki, H. (2006). MSA-C is the predominant clinical phenotype of MSA in Japan, analysis of 142 patients with probable MSA. *Journal of the Neurological Sciences*, 249(2), 115–121. doi:10.1016/j.jns.2006.05.064 PMID:16828805

Yakes, F. M., & Van Houten, B. (1997). Mitochondrial DNA damage is more extensive and persists longer than nuclear DNA damage in human cells following oxidative stress. *Proceedings of the National Academy of Sciences of the United States of America*, 94(2), 514–519. doi:10.1073/pnas.94.2.514 PMID:9012815

Yang, H., & Hu, H. Y. (2016). Sequestration of cellular interacting partners by protein aggregates: Implication in a loss-of-function pathology. *The FEBS Journal*, 283(20), 3705–3717. doi:10.1111/febs.13722 PMID:27016044

Yang, J. L., Sykora, P., Wilson, D. M. III, Mattson, M. P., & Bohr, V. A. (2011). The excitatory neurotransmitter glutamate stimulates DNA repair to increase neuronal resiliency. *Mechanisms of Ageing and Development*, *132*(8-9), 405–411. doi:10.1016/j.mad.2011.06.005 PMID:21729715

Yang, J., Wong, A., Wang, Z., Liu, W., Au, L., Xiong, Y., ... Mok, V. C. (2015). Risk factors for incident dementia after stroke and transient ischemic attack. *Alzheimer's & Dementia*, 11(1), 16–23. doi:10.1016/j.jalz.2014.01.003 PMID:24603162

Yang, Y., Gehrke, S., Imai, Y., Huang, Z., Ouyang, Y., Wang, J. W., ... Lu, B. (2006). Mitochondrial pathology and muscle and dopaminergic neuron degeneration caused by inactivation of Drosophila Pink1 is rescued by Parkin. *Proceedings of the National Academy of Sciences of the United States of America*, *103*(28), 10793–10798. doi:10.1073/pnas.0602493103 PMID:16818890

Yan, R., & Vassar, R. (2014). Targeting the β secretase BACE1 for Alzheimer's disease therapy. *Lancet Neurology*, 13(3), 319–329. doi:10.1016/S1474-4422(13)70276-X PMID:24556009

Yasuda, H., Yoshida, K., Yasuda, Y., & Tsutsui, T. (2011). Infantile zinc deficiency: Association with autism spectrum disorders. *Scientific Reports*, *1*(1), 129. doi:10.1038rep00129 PMID:22355646

Yates, P. A., Villemagne, V. L., Ellis, K. A., Desmond, P. M., Masters, C. L., & Rowe, C. C. (2014). Cerebral microbleeds: A review of clinical, genetic, and neuroimaging associations. *Frontiers in Neurology*, 6(4), 205. PMID:24432010

Yavich, L., Jakala, P., & Tanila, H. (2006). Abnormal compart-mentalization of norepinephrine in mousedentate gyrus in alpha-synuclein knockout and A30P transgenic mice. *Journal of Neurochemistry*, *99*(3), 724–732. doi:10.1111/j.1471-4159.2006.04098.x PMID:16824047

Yen, T. C., Su, J. H., King, K. L., & Wei, Y. H. (1991). Ageing-associated 5 kb deletion in human liver mitochondrial DNA. *Biochemical and Biophysical Research Communications*, *178*(1), 124–131. doi:10.1016/0006-291X(91)91788-E PMID:2069552

Yergin-Allsop, M., Rice, C., Karapurkar, T., Doemberg, N., Boyle, C., & Murphy, C. (2003). Prevalence of autism in a US metropolitan community. *Journal of the American Medical Association*, 289, 49–55. PMID:12503976

Yin, H., & Zhu, M. (2012). Free radical oxidation of cardiolipin: Chemical mechanisms, detection and implication in apoptosis, mitochondrial dysfunction and human diseases. *Free Radical Research*, *46*(8), 959–974. doi:10.3109/10715 762.2012.676642 PMID:22468920

Yirmiya, N., Sigman, M., Kasari, C., & Mundy, P. (1992). Empathy and cognition in high-functioning children with autism. *Child Development*, *63*(1), 150–160. doi:10.2307/1130909 PMID:1551323

Yokoseki, A., Shiga, A., Tan, C. F., Tagawa, A., Kaneko, H., Koyama, A., ... Onodera, O. (2008). TDP-43 mutation in familial amyotrophic lateral sclerosis. *Annals of Neurology*, *63*(4), 538–542. doi:10.1002/ana.21392 PMID:18438952

Yokota, T., Sugawara, K., Ito, K., Takahashi, R., Ariga, H., & Mizusawa, H. (2003). Down regulation of DJ-1 enhances cell death by oxidative stress, ER stress, and proteasome inhibition. *Biochemical and Biophysical Research Communications*, *312*(4), 1342–1348. doi:10.1016/j.bbrc.2003.11.056 PMID:14652021

Yong, J., Wan, L., & Dreyfuss, G. (2004). Why do cells need an assembly machine for RNA–protein complexes? *Trends in Cell Biology*, *14*(5), 226–232. doi:10.1016/j.tcb.2004.03.010 PMID:15130578

Yoo, H. K., Joung, Y. S., Lee, J. S., Song, D. H., Lee, Y. S., Kim, J.-W., ... Cho, S. C. (2013). A multicenter, randomized, double-blind, placebo-controlled study of aripiprazole in children and adolescents with Tourette's disorder. *The Journal of Clinical Psychiatry*, 74(8), e772–e780. doi:10.4088/JCP.12m08189 PMID:24021518

Yoshida, T., Sasaki, M., Yoshida, M., Namekawa, M., Okamoto, Y., Tsujino, S., ... Nakagawa, M. (2011). Nationwide survey of Alexander disease in Japan and proposed new guidelines for diagnosis. *Journal of Neurology*, 258(11), 1998–2008. doi:10.100700415-011-6056-3 PMID:21533827

Yoshii, S. R., Kishi, C., Ishihara, N., & Mizushima, N. (2011). Parkin mediates proteasome-dependent protein degradation and rupture of the outer mitochondrial membrane. *The Journal of Biological Chemistry*, 286(22), 19630–19640. doi:10.1074/jbc.M110.209338 PMID:21454557

Yoshita, M., Taki, J., & Yamada, M. (2001). A clinical role for [(123)I]MIBG myocardial scintigraphy in the distinction between dementia of the Alzheimer's-type and dementia with Lewy bodies. *Journal of Neurology, Neurosurgery, and Psychiatry*, 71(5), 583–588. doi:10.1136/jnnp.71.5.583 PMID:11606666

Youdim, M. B. H., & Bakhle, Y. S. (2006). Monoamine oxidase: Isoforms and inhibitors in Parkinson's disease and depressive illness. *British Journal of Pharmacology*, *147*(1), S287–S296. PMID:16402116

Youle, R. J., & Narendra, D. P. (2011). Mechanisms of mitophagy. *Nature Reviews. Molecular Cell Biology*, *12*(1), 9–14. doi:10.1038/nrm3028 PMID:21179058

Younan, N. D., Nadal, R. C., Davies, P., Brown, D. R., & Viles, J. H. (2012). Methionine oxidation perturbs the structural core of the prion protein and suggests a generic misfolding pathway. *The Journal of Biological Chemistry*, 287(34), 28263–28275. doi:10.1074/jbc.M112.354779 PMID:22654104

Young, B., Runge, J. W., Waxman, K. S., Harrington, T., Wilberger, J., Muizelaar, J. P., ... Kupiec, J. W. (1996). Effects of pegorgotein on neurologic outcome of patients with severe head injury: A multicenter, randomized controlled trial. *Journal of the American Medical Association*, 276(7), 538–543. doi:10.1001/jama.1996.03540070034027 PMID:8709402

Yu, L., Xie, B., Yin, X., Liang, M., Evans, A. C., Wang, J., & Dai, C. (2013). Reduced cortical thickness in primary open-angle glaucoma and its relationship to the retinal nerve fiber layer thickness. *PLoS One*, 8(9), e73208. doi:10.1371/ journal.pone.0073208 PMID:24019910

Yun, S.-W., Gerlach, M., Riederer, P., & Klein, M. A. (2006). Oxidative stress in the brain at early preclinical stages of mouse scrapie. *Experimental Neurology*, 201(1), 90–98. doi:10.1016/j.expneurol.2006.03.025 PMID:16806186

Zaccai, J., McCracken, C., & Brayne, C. (2005). A systematic review of prevalence and incidence studies of dementia with Lewy bodies. *Age and Ageing*, *34*(6), 561–566. doi:10.1093/ageing/afi190 PMID:16267179

Zaleska, M. M., & Floyd, R. A. (1985). Regional lipid peroxidation in rat brain in vitro: Possible role of endogenous iron. *Neurochemical Research*, *10*(3), 397–410. doi:10.1007/BF00964608 PMID:4000395

Zaleska, M. M., Nagy, K., & Floyd, R. A. (1989). Iron-induced lipid peroxidation and inhibition of dopamine synthesis in striatum synaptosomes. *Neurochemistry*, *14*(7), 597–605. doi:10.1007/BF00964867 PMID:2550829

Zambrano, C. A., Egaña, J. T., Núñez, M. T., Maccioni, R. B., & González-billault, C. (2004). Oxidative stress promotes tau dephosphorylation in neuronal cells: The roles of cdk5 and PP1. *Free Radical Biology & Medicine*, *36*(11), 1393–1402. doi:10.1016/j.freeradbiomed.2004.03.007 PMID:15135175

Zangaglia, R., Martignoni, E., Glorioso, M., Ossola, M., Riboldazzi, G., Calandrella, D., ... Pacchetti, C. (2007). Macrogol for the treatment of constipation in Parkinson's disease. A randomized placebo-controlled study. *Movement Disorders*, 22(9), 1239–1244. doi:10.1002/mds.21243 PMID:17566120

Zanusso, G., Monaco, S., Pocchiari, M., & Caughey, B. (2016). Advanced tests for early and accurate diagnosis of Creutzfeldt-Jakob disease. *Nature Reviews. Neurology*, *12*(6), 325–333. doi:10.1038/nrneurol.2016.65 PMID:27174240

Zarei, S., Carr, K., Reiley, L., Diaz, K., Guerra, O., Altamirano, P. F., ... Chinea, A. (2015). A comprehensive review of amyotrophic lateral sclerosis. *Surgical Neurology International*, 6. PMID:26629397

Zarranz, J. J., Alegre, J., Gómez-Esteban, J. C., Lezcano, E., Ros, R., Ampuero, I., ... Llorens, V. (2004). The new mutation, E46K, of α -synuclein causes parkinson and Lewy body dementia. *Annals of Neurology*, 55(2), 164–173. doi:10.1002/ana.10795 PMID:14755719

Zempel, H., Luedtke, J., Kumar, Y., Biernat, J., Dawson, H., Mandelkow, E., & Mandelkow, E.-M. (2013). Amyloid-β oligomers induce synaptic damage via Tau-dependent microtubule severing by TTLL6 and spastin. *The EMBO Journal*, *32*(22), 2920–2937. doi:10.1038/emboj.2013.207 PMID:24065130

Zempel, H., Thies, E., Mandelkow, E., & Mandelkow, E.-M. (2010). A Oligomers Cause Localized Ca2+ Elevation, Missorting of Endogenous Tau into Dendrites, Tau Phosphorylation, and Destruction of Microtubules and Spines. *The Journal of Neuroscience*, *30*(36), 11938–11950. doi:10.1523/JNEUROSCI.2357-10.2010 PMID:20826658

Zeng, B. J., Pastores, M. G., Leone, P., Raghavan, S., Wang, H. Z., Ribeiro, A. L., Torres, P., ... Kolodny, H. E. (2006). Mutation Analysis of the Aspartoacylase Gene in Non-Jewish Patients with Canavan Disease. Academic Press.

Zerr, I., Bodemer, M., Gefeller, O., Otto, M., Poser, S., Wiltfang, J., & Weber, T. (1998). Detection of 14-3-3 protein in the cerebrospinal fluid supports the diagnosis of Creutzfeldt-Jakob disease. *Annals of Neurology*, 43(1), 32–40. doi:10.1002/ana.410430109 PMID:9450766

Zesiewicz, T. A., Baker, M. J., Dunne, P. B., & Hauser, R. A. (2001). Diffuse Lewy body disease. *Current Treatment Options in Neurology*, *3*(6), 507–518. doi:10.100711940-001-0013-x PMID:11581527

Zhang, B., Tu, P. H., Abtahian, F., Trojanowski, J. Q., & Lee, V. M. Y. (1997). Neurofilaments and orthograde transport are reduced in ventral root axons of transgenic mice that express human SOD1 with a G93A mutation. *The Journal of Cell Biology*, *139*(5), 1307–1315. doi:10.1083/jcb.139.5.1307 PMID:9382875

Zhang, C., & Saunders, A. J. (2007). Therapeutic targeting of the alpha-secretase pathway to treat Alzheimer's disease. *Discovery Medicine*, 7(39), 113–117. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/18093473 PMID:18093473

Zhang, J., Fitsanakis, V. A., Gu, G., Jing, D., Ao, M., Amarnath, V., & Montine, T. J. (2003). Manganese ethylene-bisdithiocarbamate and selective dopaminergic neurodegeneration in rat: A link through mitochondrial dysfunction. *Journal of Neurochemistry*, *84*(2), 336–346. doi:10.1046/j.1471-4159.2003.01525.x PMID:12558996

Zhang, Q.-G., Laird, M. D., Han, D., Nguyen, K., Scott, E., Dong, Y., ... Brann, D. W. (2012). Critical role of NADPH oxidase in neuronal oxidative damage and microglia activation following traumatic brain injury. *PLoS One*, *7*(4), e34504. doi:10.1371/journal.pone.0034504 PMID:22485176

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Zhang, Q., Raoof, M., Chen, Y., Sumi, Y., Sursal, T., Junger, W., ... Hauser, C. J. (2010). Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature*, *464*(7285), 104–107. doi:10.1038/nature08780 PMID:20203610

Zhang, Y., Gao, J., Chung, K. K., Huang, H., Dawson, V. L., & Dawson, T. M. (2000). Parkin functions as an E2dependent ubiquitin–protein ligase and promotes the degradation of the synaptic vesicle-associated protein, CDCrel-1. *Proceedings of the National Academy of Sciences of the United States of America*, 97(24), 13354–13359. doi:10.1073/ pnas.240347797 PMID:11078524

Zhang, Z., Lotti, F., Dittmar, K., Younis, I., Wan, L., Kasim, M., & Dreyfuss, G. (2008). SMN deficiency causes tissuespecific perturbations in the repertoire of snRNAs and widespread defects in splicing. *Cell*, *133*(4), 585–600. doi:10.1016/j. cell.2008.03.031 PMID:18485868

Zheng, C. Y., Seabold, G. K., Horak, M., & Petralia, R. S. (2011). MAGUKs, synaptic development, and synaptic plasticity. *The Neuroscientist*, *17*(5), 493–512. doi:10.1177/1073858410386384 PMID:21498811

Zheng, Z., & Diamond, M. I. (2012). Huntington disease and the huntingtin protein. *Progress in Molecular Biology and Translational Science*, *107*, 189–214. doi:10.1016/B978-0-12-385883-2.00010-2 PMID:22482451

Zhou, M., Xu, S., Mi, J., Uèda, K., & Chan, P. (2013). Nuclear translocation of alpha-synuclein increases susceptibility of MES23.5 cells to oxidative stress. *Brain Research*, *1500*, 19–27. doi:10.1016/j.brainres.2013.01.024 PMID:23337620

Zhou, Q., Homma, K. J., & Poo, M. (2004). Shrinkage of dendritic spines associated with long-term depression of hippocampal synapses. *Neuron*, 44(5), 749–757. doi:10.1016/j.neuron.2004.11.011 PMID:15572107

Zhu, X., Perry, G., Moreira, P. I., Aliev, G., Cash, A. D., Hirai, K., & Smith, M. A. (2006). Mitochondrial abnormalities and oxidative imbalance in Alzheimer disease. *Journal of Alzheimer's Disease*, *9*(2), 147–153. doi:10.3233/JAD-2006-9207 PMID:16873962

Zieren, N., Duering, M., Peters, N., Reyes, S., Jouvent, E., Hervé, D., ... Dichgans, M. (2013). Education modifies the relation of vascular pathology to cognitive function: Cognitive reserve in cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy. *Neurobiology of Aging*, *34*(4), 400–407. doi:10.1016/j.neurobiolag-ing.2012.04.019 PMID:22626524

Zoccolella, S., Beghi, E., Palagano, G., Fraddosio, A., Guerra, V., Samarelli, V., ... Logroscino, G. (2008). Analysis of survival and prognostic factors in amyotrophic lateral sclerosis: A population based study. *Journal of Neurology, Neurosurgery, and Psychiatry*, *79*(1), 33–37. doi:10.1136/jnnp.2007.118018 PMID:17550991

Zonneveld, H. I., Goos, J. D., Wattjes, M. P., Prins, N. D., Scheltens, P., van der Flier, W. M., ... Barkhof, F. (2014). Prevalence of cortical superficial siderosis in a memory clinic population. *Neurology*, *82*(8), 698–704. doi:10.1212/WNL.000000000000150 PMID:24477113

Zorov, D. B., Juhaszova, M., & Sollott, S. J. (2014). Mitochondrial reactive oxygen species (ROS) and ROS-induced ROS release. *Physiological Reviews*, *94*(3), 909–950. doi:10.1152/physrev.00026.2013 PMID:24987008

Zu, T., Liu, Y., Banez-Coronel, M., Reid, T., Pletnikova, O., Lewis, J., ... Subramony, S. H. (2013). RAN proteins and RNA foci from antisense transcripts in C9ORF72 ALS and frontotemporal dementia. *Proceedings of the National Academy of Sciences*, *110*, E4968–E4977.

Zuber, C., Ludewigs, H., & Weiss, S. (2007). Therapeutic approaches targeting the prion receptor LRP/LR. *Veterinary Microbiology*, *123*(4), 387–393. doi:10.1016/j.vetmic.2007.04.005 PMID:17498894

Zwaigenbaum, L., Bryson, S., Rogers, T., Roberts, W., Brian, J., & Szatmari, P. (2005). Behavioral manifestations of autism in the first year of life. *International Journal of Developmental Neuroscience*, 23(2-3), 143–152. doi:10.1016/j. ijdevneu.2004.05.001 PMID:15749241

Zweig, R. M., Jankel, W. R., Hedreen, J. C., Mayeux, R., & Price, D. L. (1989). The pedunculopontine nucleus in Parkinson's disease. *Annals of Neurology*, 26(1), 41–46. doi:10.1002/ana.410260106 PMID:2549845

Zweig, Y. R., & Galvin, J. E. (2014). Lewy body dementia: The impact on patients and caregivers. *Alzheimer's Research* & *Therapy*, 6(2), 21. doi:10.1186/alzrt251 PMID:25031635

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